NQO1 regulates cell cycle progression at the G2/M phase

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Supplementary Table 1 and Figures

Supplementary Table 1. Status of NRF2 and KEAP1 in the cell lines used in this study or causes of activation of Nrf2.

Cell line	Cancer type	Status of NRF2 and KEAP1 or causes of activation of Nrf2	Reference
A549	Lung cancer	KEAP1 G333C (homo) KEAP1 promoter methylation	A
RKO	Colon cancer	Wild type Nrf2 (low basal level) Wild type KEAP1	В
MDA-MB-231	Breast cancer	Wild type Nrf2 (low basal level) Wild type KEAP1	С
U87-MG	Glioblastoma	WASP-interacting protein (WIP) PTEN in the wild type of EGFR	D E
PC3	Prostate cancer	Wild type KEAP1 GRP/Bip translocation	F G
MIA PaCa-2	Pancreatic cancer	Wild type Nrf2 (low basal level) Wild type KEAP1 (high level)	н

References for Supplementary Table 1

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RKO/pshCont cells



RKO/pshNQO1 cells



В

MDA-MB-231/pNQO1



Figure S1. NQO1 regulates cancer cell proliferation. (A-B) RKO/pshCont and RKO/pshNQO1 cells **(A)** and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells **(B)** were seeded in 8-well chamber slides and cell cycle progression determined via confocal microscopy as indicated. White arrows signify the cells monitored for analysis of cell cycle progression.

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Figure S2. Quantification of cell cycle analysis. (A) Quantification of cell cycle analysis of RKO/pshCont cells and RKO/pshNQO1 cells in the flow cytometry analysis results of Figure 2A. **(B)** Quantification of cell cycle analysis of MDA-MB-231/pNQO1 cells and MDA-MB-231/pCont cells in the flow cytometry analysis results of Figure 2B.



Figure S3. Effect of NQO1 on CDK1 expression in cancer cells. (A-B) RKO/pshCont and RKO/pshNQO1 cells (A) and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells (B) were synchronized at the G1/S boundary via double-thymidine blocking, released from synchronization, and harvested at the indicated times. Whole-cell lysates were analyzed by immunoblotting for CDK1, NQO1, and β -actin. AS indicates asynchronization.



Figure S4. Quantification of cell cycle analysis. (A) Quantification of cell cycle analysis of RKO/pshCont cells and RKO/pshNQO1 cells in the flow cytometry analysis results of Figure 2G. **(B)** Quantification of cell cycle analysis of MDA-MB-231/pNQO1 cells and MDA-MB-231/pCont cells in the flow cytometry analysis results of Figure 2H.



Figure S5. Effect of NQO1 on CKS1 expression in cancer cells. (A-B) The relative mRNA and protein expression levels of NQO1 and the relative protein expression levels of CKS1 in RKO/pshCont and RKO/pshNQO1 cells (A) and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells (B). *NQO1* expression was examined via qPCR using *18S rRNA* as the internal control. All data are presented as mean ± SEM. *** *P* < 0.001 with unpaired *t*-test, **** *P* < 0.0001 with unpaired *t*-test. Protein expression was examined via immunoblotting. Whole-cell lysates were analyzed by immunoblotting for NQO1, CKS1, and β-actin. (C) The relative mRNA and protein expression was examined via qPCR using *18S rRNA* as the internal control expression levels of CKS1 in A549, MIA-PaCa-2, PC3, and U87-MG cells transfected with siCont or siNQO1. *NQO1* expression was examined via qPCR using *18S rRNA* as the internal control. All data are presented as mean ± SEM. * *P* < 0.001 with unpaired *t*-test *A*, **** *P* < 0.001 with unpaired *t*-test. Protein expression was examined via qPCR using *18S rRNA* as the internal control. All data are presented as mean ± SEM. * *P* < 0.05 with unpaired *t*-test, *** *P* < 0.01 with unpaired *t*-test. Protein expression was examined via qPCR using *18S rRNA* as the internal control. All data are presented as mean ± SEM. * *P* < 0.05 with unpaired *t*-test, *** *P* < 0.01 with unpaired *t*-test. Protein expression was examined via immunoblotting. Whole-cell lysates were analyzed by immunoblotting for NQO1, CKS1, and β-actin.



Figure S6. Effect of NQO1 on *CKS1B* mRNA stability in cancer cells. (A-B) RKO/pshCont and RKO/pshNQO1 cells (A) and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells (B) were incubated with 5 µg/ml actinomycin D for 2 h and harvested. *CKS1B* expression was examined via qPCR using *18S rRNA* as the internal control. All data are presented as mean \pm SEM. ** *P* < 0.01 with ANOVA, **** *P* < 0.001 with ANOVA.



Figure S7. Effect of the TRE region of the *CKS1B* promoter on *CKS1B* expression in cancer cells. (A-B) RKO/pshCont cells (A) and MDA-MB-231/pNQO1 cells (B) were transfected with wild type pro1, TRE1 deletion mutant pro1, TRE2 deletion mutant pro1, or TRE1 and TRE2 deletion mutant pro1 and control pRL-*luc*. After 4 h, cells were washed with PBS and incubated with the appropriate medium for 48 h. Luciferase activity was normalized to that of *Renilla* (mean ± SEM). ** *P* < 0.01 with ANOVA. **** *P* < 0.001 with ANOVA. NS indicates no significance.



Figure S8. ChIP assay to verify the binding of c-Fos to TRE2 on CKS1B promoter. (A, B) RKO/pshCont cells **(A)** and MDA-MB-231 cells **(B)** were treated with formaldehyde for crosslinking, and DNA was extracted and immunoprecipitated with ant-c-Fos antibody and anti-normal IgG. After reversal of crosslinking, DNA was purified and used as a templated for PCR to amplify TRE2 site. The PCR products were electrophoresed on 1.5% agarose gel and visualized by staining with EtBr.



Figure S9. Effect of c-Fos on *CKS1B* expression in NQO1-expressing and NQO1deficient cancer cells. (A-B) RKO/pshCont and RKO/pshNQO1 cells (A) and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells (B) were transfected with siCont or sic-Fos, incubated for 1 day, transfected with pTRE-*luc* and the transfection control pRL*luc*. After 4 h, cells were washed with PBS and incubated with the appropriate medium for 48 h. Luciferase activity was normalized to that of *Renilla* (mean ± SEM). *** *P* < 0.001 with ANOVA. **** *P* < 0.001 with ANOVA. NS indicates no significance.



Figure S10. NQO1 and subcellular localization of c-Fos. Subcellular localization was visualized by confocal microscopy. RKO/pshCont cells and MDA-MB-231/pNQO1 cells were fixed for immunofluorescence analysis with the indicated antibodies. Nuclear staining is indicated by using DAPI.



Figure S11. Representative figures of Fig. 6C and D. (A-B) RKO/pshCont cells (A) and MDA-MB-231/pNQO1 cells (B) were transfected with siCont and pCont, sic-Fos and pCont or sic-Fos and pCKS1B, and RKO/pshNQO1 cells (A) or MDA-MB-231/pCont cells (B) transfected with pCont and siCont, pc-Fos and siCont or pc-Fos and siCKS1B. After 48 h of incubation, various quantities of cells were plated on an eight-well chamber slide, cultured for 16 h, and irradiated with 0 or 4 Gy, followed by culture for 24 h. Cells were fixed with 3.7% PFA and immunofluorescence performed using anti-phospho histone H2AX.

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Figure S12. Histopathological scoring criteria used for analysis of NQO1 and CKS1 staining in human clinical tumor tissue. IHC scores of NQO1 and CKS1 were defined by positive area score (5-25%,1; >25-50%,2; >50-75%,3; >75-100%,4).



Figure S13. Effect of NQO1 on mitotic spindle formation in cancer cells. (A-B) RKO/pshCont and RKO/pshNQO1 cells (A) and MDA-MB-231/pNQO1 and MDA-MB-231/pCont cells (B) were fixed with 3.7% PFA and immunofluorescence performed using anti- α -tubulin and anti-pericentrin.



Figure S14. Analysis of interaction between NQO1 and c-Jun. Whole-cell extracts of RKO/pshCont cells were immunoprecipitated with anti-NQO1 and anti-IgG (negative control) and analyzed by immunoblotting with anti-NQO1 anti-c-Jun antibodies.



Figure S15. Analysis of synthesized c-Fos peptide. (A) HPLC chromatogram of c-Fos peptide. **(B)** LC-Mass spectrum of c-Fos peptide.

Figure S16.



Related to Fig. 3f NQ01 CKS1

Related to Fig. 1B



Related to Fig. 2D



Related to Fig. S3B





Related to Fig. S5A

Related to Fig. S5B









Related to Fig. 4G



Related to Fig. 4H



NQ01 c-Fos CKS1 β-actin

Related to Fig. 4I

Related to Fig. 5A

c-Fos NQO1

Related to Fig. 5C



Related to Fig. 4J



Related to Fig. 5B



Related to Fig. 5D



NQO1-His₆

EGFP-c-Fos

Related to Fig. 5E







Related to Fig. 5H









Related to Fig. S13

c-Jun