

---

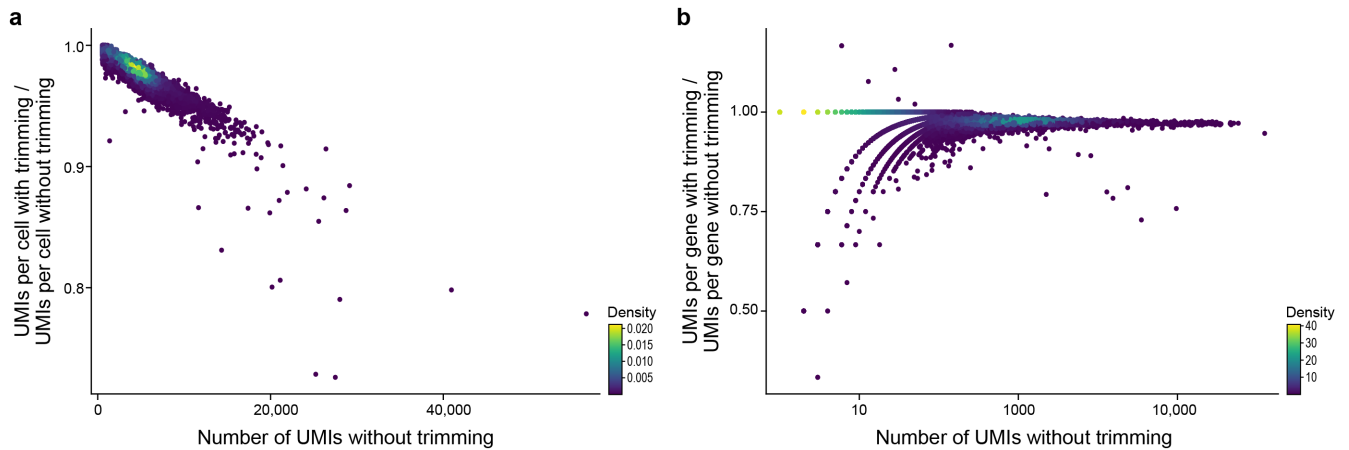
**Supplementary information**

---

**Mostly natural sequencing-by-synthesis for  
scRNA-seq using Ultima sequencing**

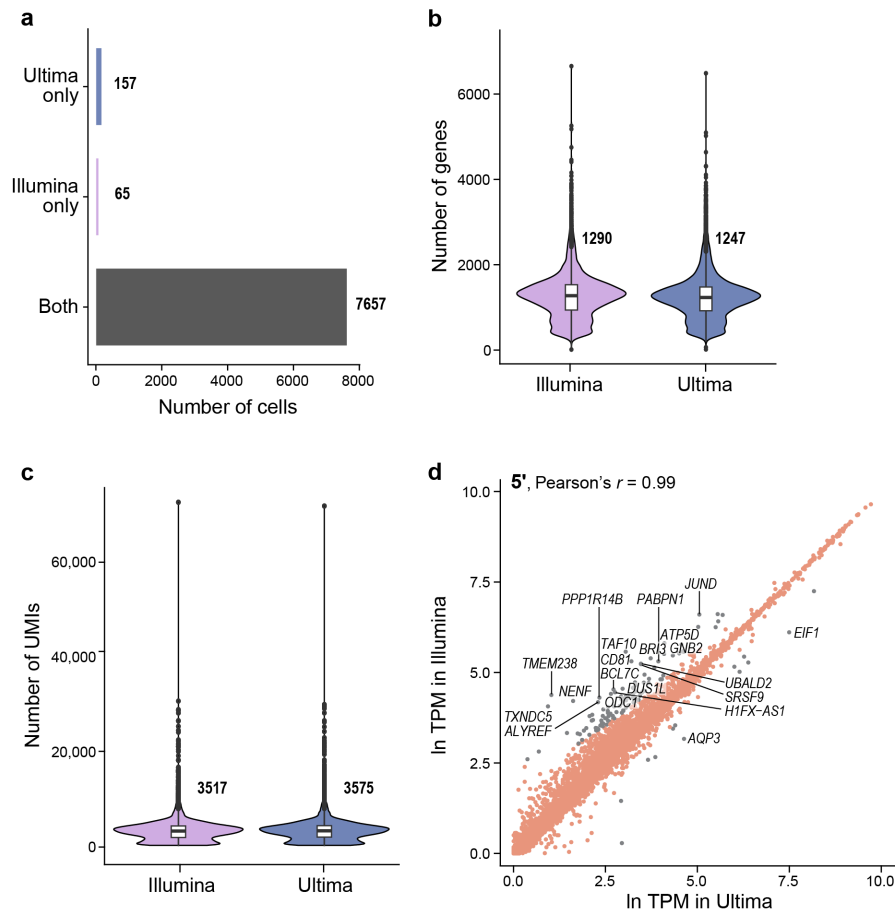
---

In the format provided by the  
authors and unedited



**Supplementary Figure 1. Relationship between UMI coverage and UMI trimming.**

The number of UMIs per (a) cell and (b) gene for 3' PBMC Illumina data with both full-length UMIs and UMIs trimmed to 9 bases. Ratio between the two (the number of UMIs per cell or gene with trimmed UMIs divided by the number of UMIs per cell or gene with untrimmed UMIs) is plotted versus the number of UMIs without trimming. Each point is colored by its local density in the graph.

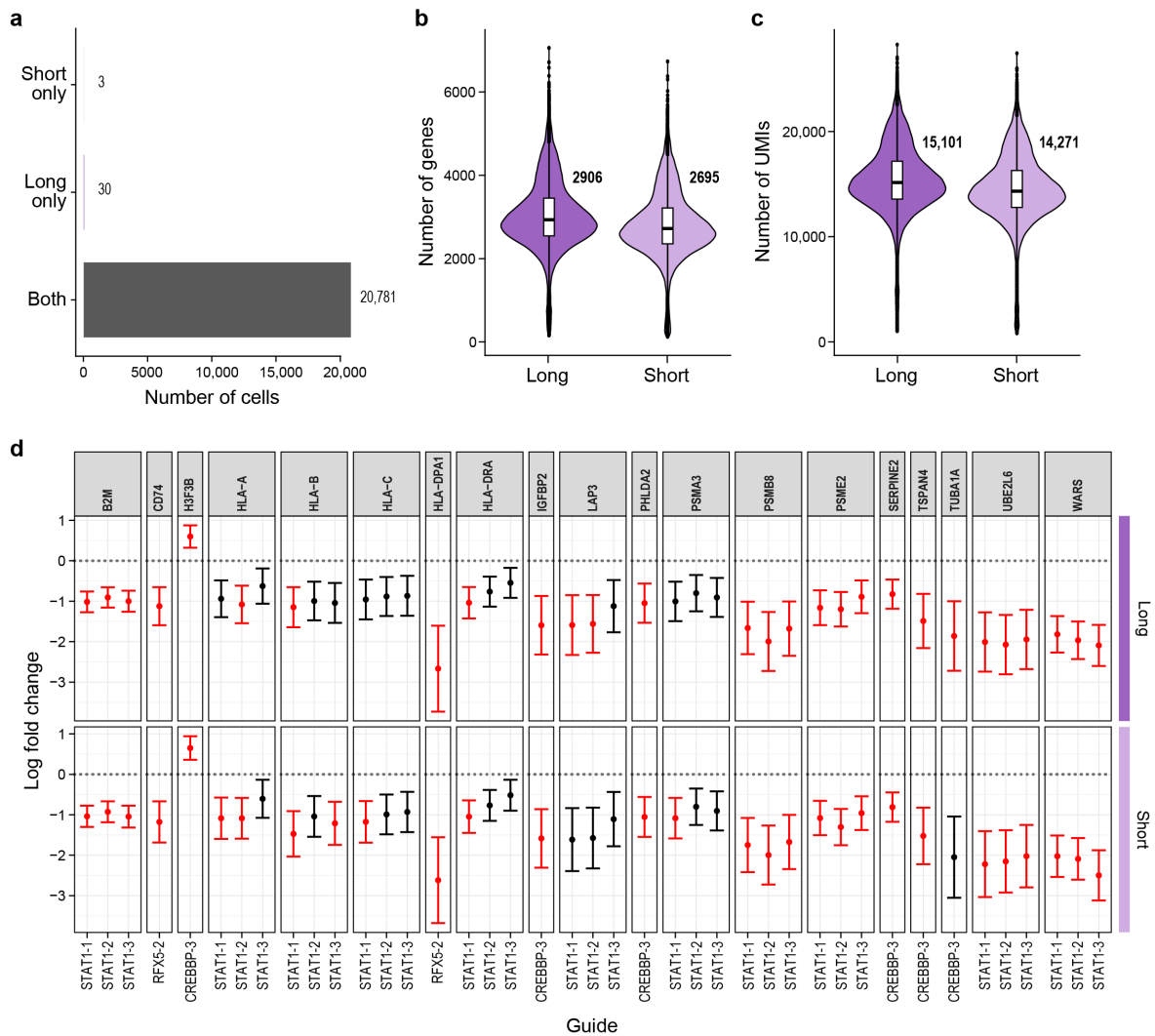


**Supplementary Figure 2. Quality metrics for 5' Ultima vs single-end Illumina data.**

(a) Number of cells identified by Cell Ranger only in Ultima, only in Illumina, or both.

Distribution of the number of genes (b) or UMIs (c) per cell. (d) Scatter plots with one point for each gene as in Fig. 2e. For all these analyses, reads were sampled so that Illumina and Ultima have the same number of reads.

# Supp. Fig. 3



## Supplementary Figure 3. Perturb-seq data with shortened Read 2 length.

(a) Number of cells identified only in the shorter Illumina data, only in longer Illumina data, or in both. Distribution of the number of genes (b) or UMIs (c) per cell in short and long Illumina data. (d) We extracted all gene/guide pairs from our DE analysis with FDR < 0.05 in either shorter or longer Illumina data. Data processed and shown here as in Fig. 4f.

## **Supplementary Tables**

**Supplementary Table 1.** Sequencing metrics.

**Supplementary Table 2.** Comparison of read mapping between Ultima and Illumina.

**Supplementary Table 3.** Differentially expressed genes between Ultima and Illumina with  
5 standard or extended references.

**Supplementary Table 4.** Antibody, Perturb-seq, and hashing DNA barcodes.

**Supplementary Table 5.** PCR primers to convert libraries for Ultima sequencing.

**Supplementary Table 6.** Commands for initial sequence processing.



Table S3

Method	Illumina read type	Illumina Reference	Ultima Reference**	Total outliers*	Outliers upregulated in Illumina	Outliers upregulated in Ultima
3'	Paired-end	Standard	Standard	182	131	51
3'	Paired-end	Standard	Extended by single cell	173	118	55
3'	Paired-end	Standard	Extended by bulk	177	117	60
3'	Paired-end	Extended by single cell	Extended by single cell	172	119	53
3'	Paired-end	Extended by bulk	Extended by bulk	183	135	48
5'	Paired-end	Standard	Standard	454	342	112
5'	Paired-end	Standard	Extended by single cell	352	233	119
5'	Paired-end	Standard	Extended by bulk	350	234	116
5'	Paired-end	Extended by single cell	Extended by single cell	346	247	109
5'	Paired-end	Extended by bulk	Extended by bulk	349	247	102
5'	Single-end	Standard	Standard	122	109	13
*Outliers are genes with absolute logFC>log(2) in Ultima relative to Illumina						
**Uses Ultima data downsampled to have the same number of reads as Illumina						

table S5

Name	Sequence	Notes
PS-SBC	5'-CCATCTCATCCCTGCGTGTCTCCGACTCAGCGGAAGAACCTCGATCTACACGACGCTCTTCCGATC*T-3'	IA region, Read 1 binding region
PB	5'-CCTATCCCCTGTGTGCCTTGGCAGTCTCAGCAGACGTGTGCTCTTCCGATC*T-3'	UBA region, Read 2 binding region
* = phosphorothioate bond modification protecting it from degradation		



Table S6

3' Ultima step 1	cutadapt -j 0 --discard-untrimmed --pair-filter any -a CTACACGACGCTCTCCGATCT;max_error_rate=0.2;min_overlap=10;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -U 50 -q 30 -A TTTTTTTTTTTT;max_error_rate=0.2;min_overlap=8;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -o <output_read1_long> -p <output_read2> --minimum-length 28:50 <input_fastq> <input_fastq>
3' Ultima step 2	cutadapt -j 0 --minimum-length 28 --maximum-length 28 --length 28 -o <output_read1> <output_read1_long>
3' Ultima step 3	zcat <output_read2>   awk '{if ((NR%4 == 2)    (NR%4 == 0)) {print substr(\$0,1+5,90)} else {print \$0}}'   seqkit -j 8 seq -p -r -t DNA   gzip > <output_read2_revcom>
5' Ultima step 1	cutadapt -j 0 --discard-untrimmed --pair-filter any -a CTACACGACGCTCTCCGATCT;max_error_rate=0.2;min_overlap=10;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -U 48 -q 30 -A ^TTTCTTATATGGG;max_error_rate=0.5;min_overlap=8;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -o <output_read1_long> -p <output_read2> --minimum-length 26:50 --maximum-length 390:315 <input_fastq> <input_fastq>
5' Ultima step 2	cutadapt -j 0 --minimum-length 26 --maximum-length 26 --length 26 -o <output_read1> <output_read1_long>
5' Ultima step 3	zcat <output_read2>   awk '{if ((NR%4 == 2)    (NR%4 == 0)) {print substr(\$0,1+3,90)} else {print \$0}}'   seqkit -j 8 seq -p -r -t DNA   gzip > <output_read2_revcom>
5' Illumina single-end step 1	cutadapt -j 0 --discard-untrimmed --pair-filter any -a CTACACGACGCTCTCCGATCT;max_error_rate=0.2;min_overlap=10;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -U 48 -q 30 -A ^TTTCTTATATGGG;max_error_rate=0.5;min_overlap=8;required...AGATCGGAAGAGCACACGTCTG;max_error_rate=0.2;min_overlap=6 -o <output_read1_long> -p <output_read2> --minimum-length 26:50 --maximum-length 390:315 <input_fastq> <input_fastq>
5' Illumina single-end step 2	cutadapt -j 0 --minimum-length 26 --maximum-length 26 --length 26 -o <output_read1> <output_read1_long>
5' Illumina single-end step 3	zcat <output_read2>   awk '{if ((NR%4 == 2)    (NR%4 == 0)) {print substr(\$0,1+3,90)} else {print \$0}}'   seqkit -j 8 seq -p -r -t DNA   gzip > <output_read2_revcom>