

Cell Genomics, Volume 3

Supplemental information

**ClonoCluster: A method for using clonal
origin to inform transcriptome clustering**

Lee P. Richman, Yogesh Goyal, Connie L. Jiang, and Arjun Raj

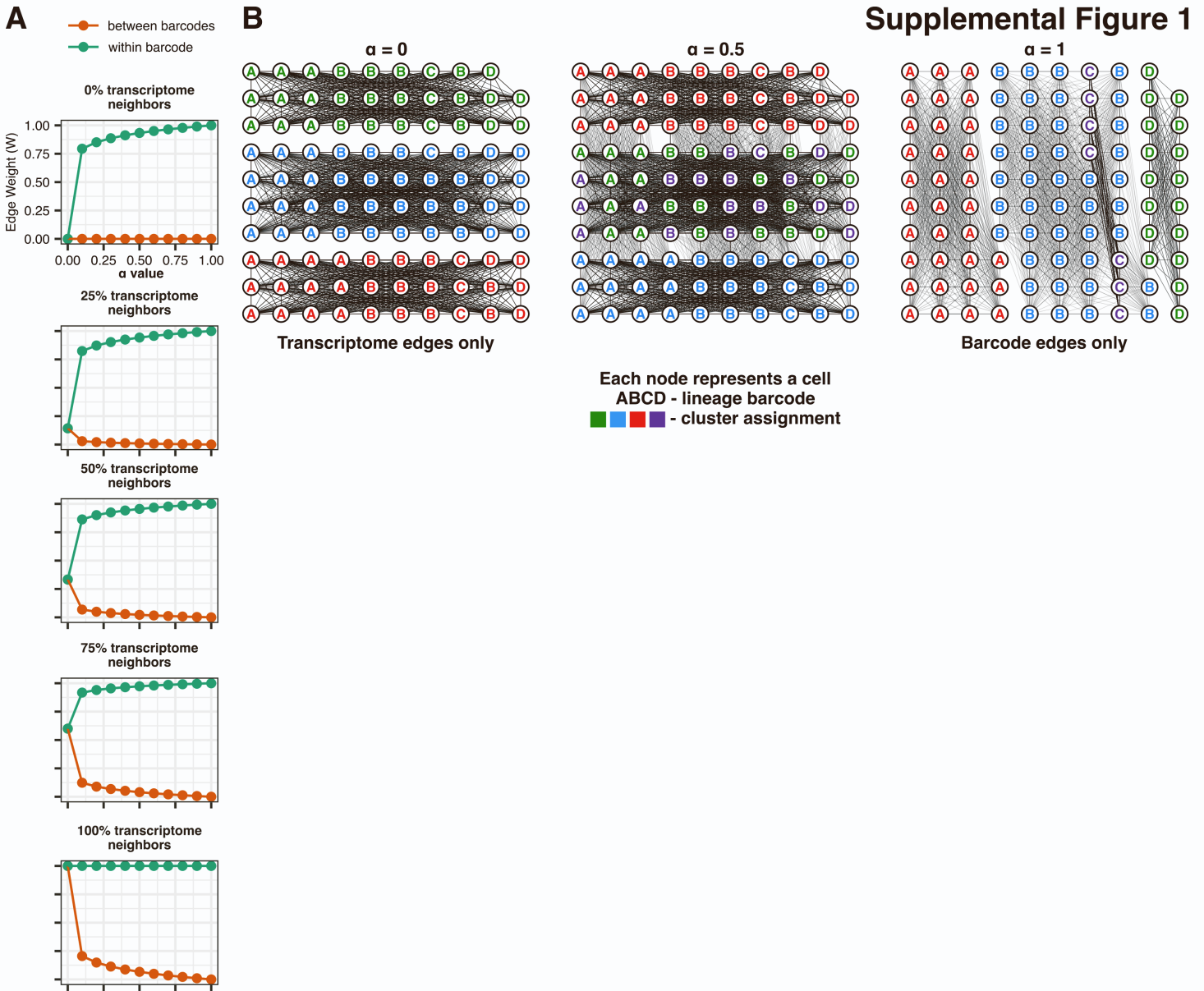


Figure S1, Related to Figure 1: α modifies network graph edge weights to create Louvain clusters that increasingly reflect clone barcode assignments.

A. Output edge weight of the ClonoCluster model for graph edge weights between two cells for different proportions of shared nearest neighbors in transcriptome space with fixed $\beta = 0.1$.

B. Network graphs with modified edge weights and cluster assignments at three α values for simulated data.

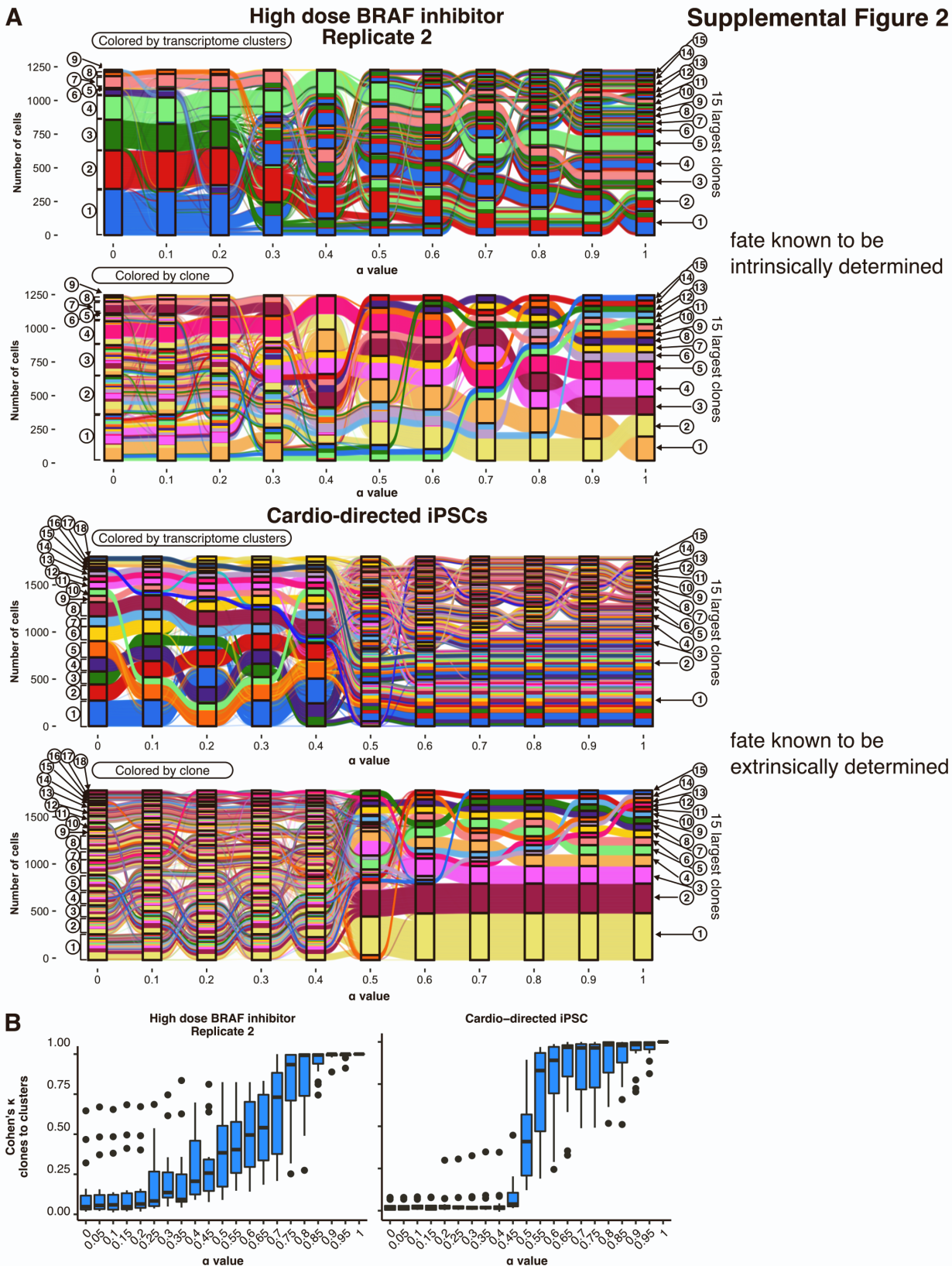


Figure S2, Related to Figure 1: Intrinsic vs extrinsic determinants of cell fate stratify the effect of α on clone-cluster congruence.

A. Sankey diagrams for the 15 largest clone clusters in two samples across α values, a clonal melanoma cell line treated with high dose ($1\mu\text{M}$) vemurafenib¹¹ with known intrinsic determinants of cell fate and directed differentiation of induced pluripotent stem cells (iPSCs) towards a cardiomyocyte fate¹⁸ with previously described dominance of extrinsic determinants of cell fate.

B. Cohen's κ for interrater reliability for classification of a cell by clone barcode and cluster assignment for the top 15 largest clone clusters across α values for the samples in (A).

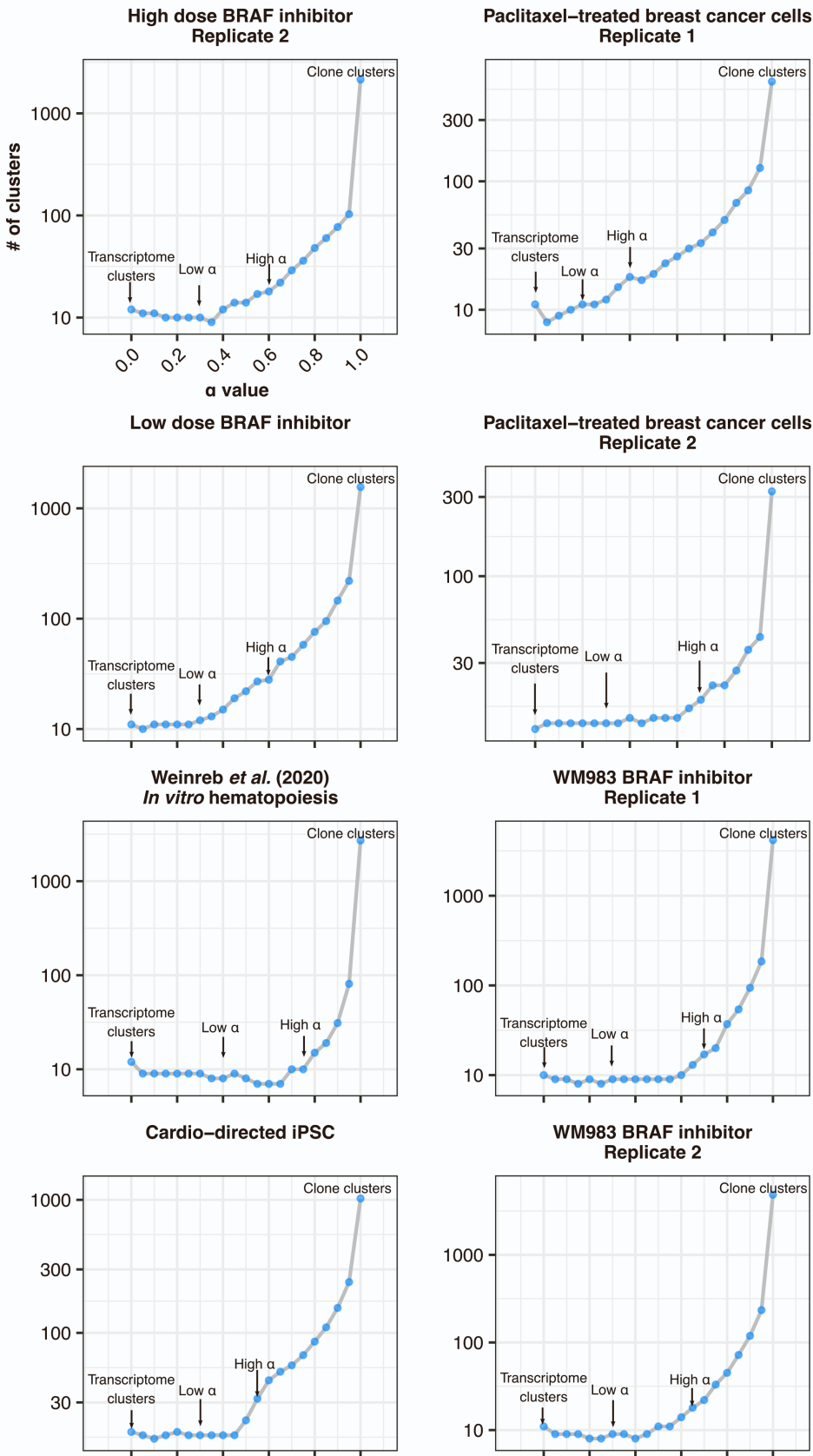


Figure S3, Related to Figure 1: The number of clusters returned by community detection at fixed resolution approaches the number of unique clone barcodes in the data as α approaches 1. Plots showing the number of clusters returned by community detection at constant resolution with increasing α in each dataset. At $\alpha = 1$, the network graph is composed of like-barcoded cells (a clone cluster) connected only to each other and community detection returns the number of unique clone clusters in the data as the number of clusters.

Supplemental Figure 4

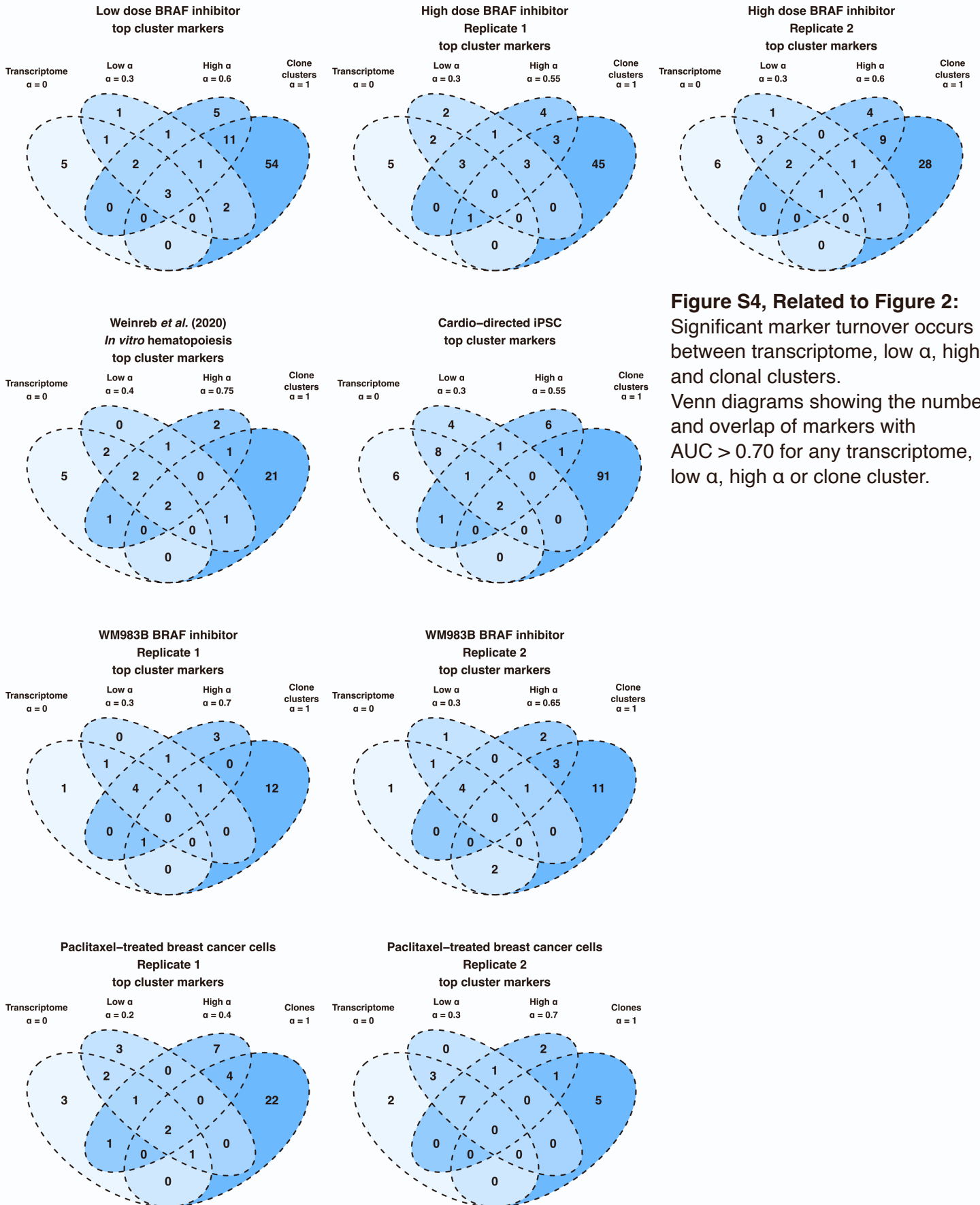
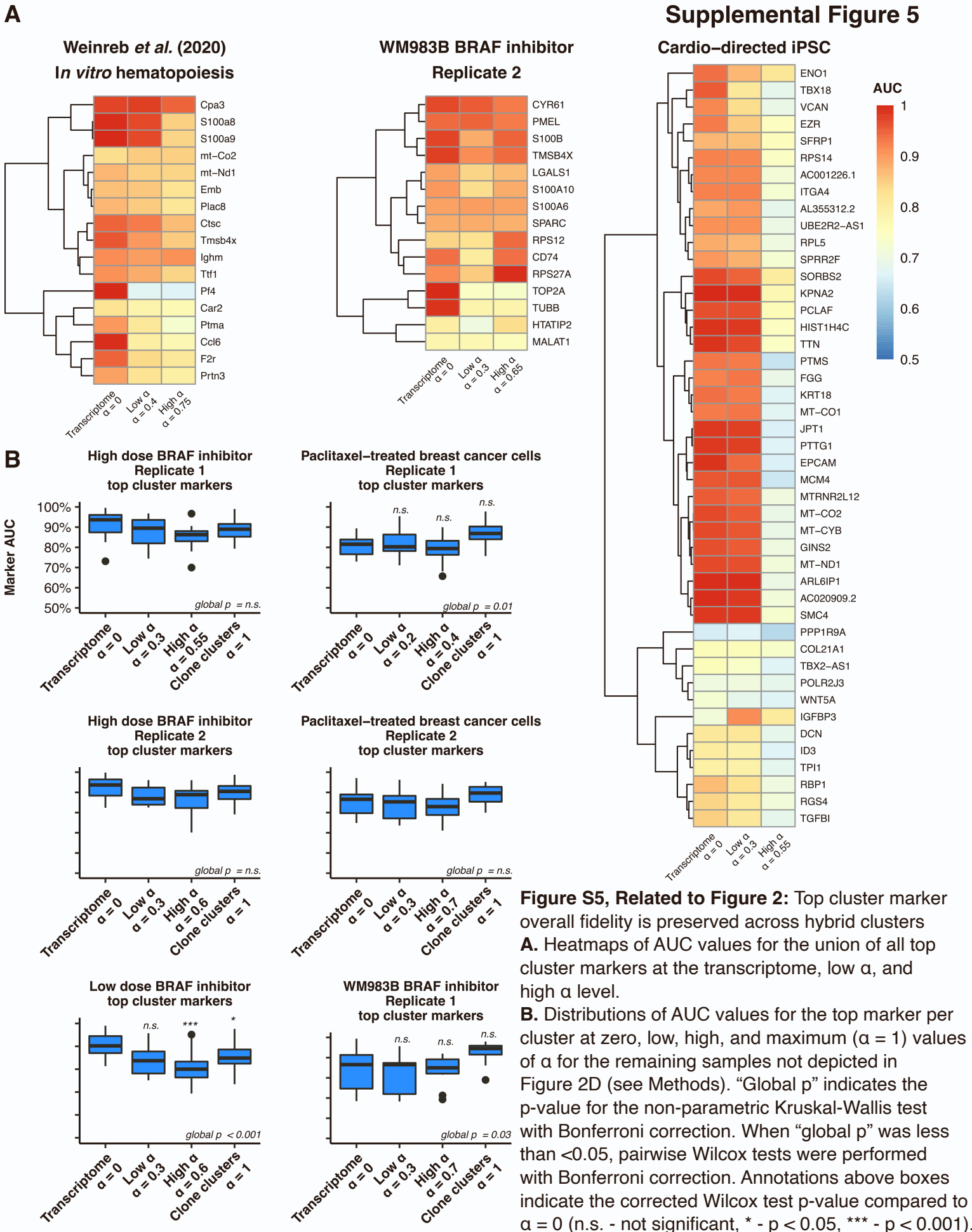


Figure S4, Related to Figure 2: Significant marker turnover occurs between transcriptome, low α , high α , and clonal clusters. Venn diagrams showing the number and overlap of markers with AUC > 0.70 for any transcriptome, low α , high α or clone cluster.

Supplemental Figure 5



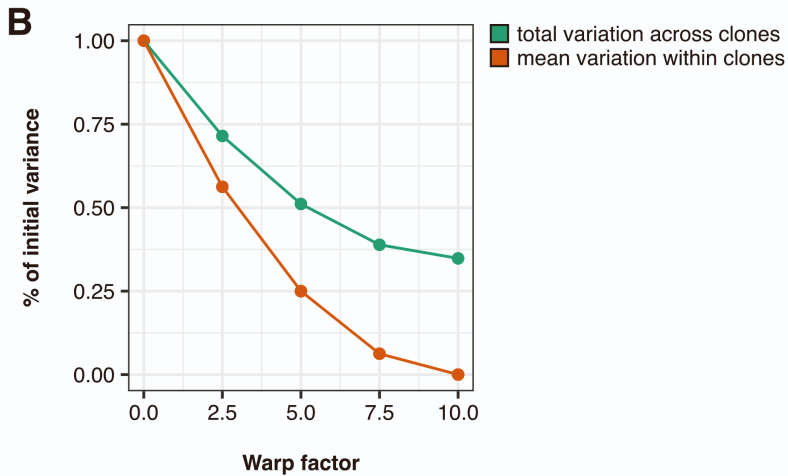
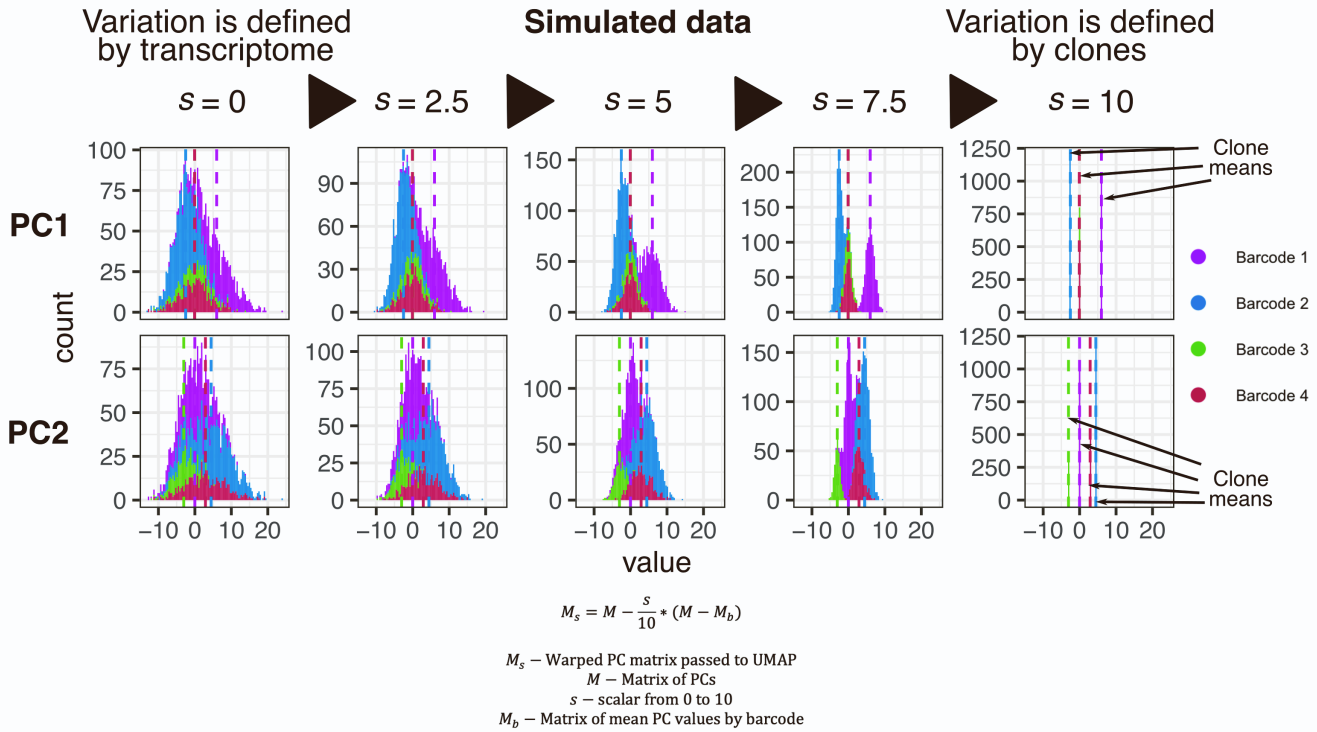


Figure S6, Related to Figure 4: Increasing Warp Factor, s , reduces the variation of principal component values around clone barcode means.

A. Histograms colored by clone barcodes demonstrating the distribution of PC values for two principal components for simulated data for 3,000 cells with four clone barcode assignments. Vertical dashed lines represent the mean values for each clone barcode at $s = 0$, equivalent to the unmodified PC means. At $s = 10$, modified PC values are equivalent to the assigned clone barcode means. Modified PC values are then passed to the UMAP implementation for visualization as in Figure 4.

B. Mean variation of all PCs in simulated data within and between clone barcodes stratified by values of s .

Supplemental Figure 7

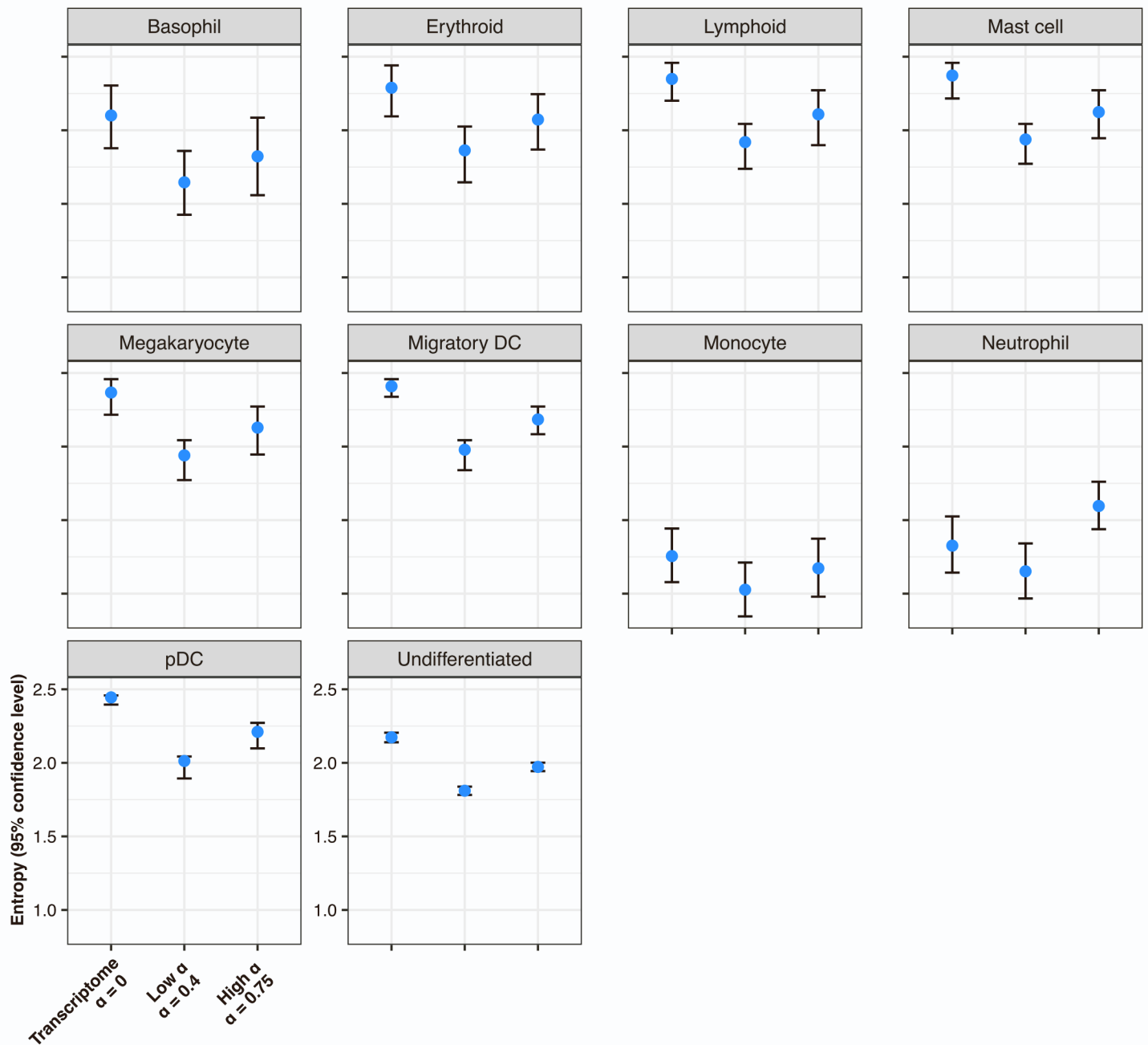


Figure S7, Related to Figure 5: Low α clusters reduce entropy of clustering for multiple hematopoietic cell types. Laplace entropy was calculated for the distribution of known cell types in hematopoiesis data¹² across transcriptome-only clusters, low α , and high α . Entropy is shown in bits, error bars represent 95% confidence intervals using the Monte Carlo method with 1,000 samplings of 33% of the cells.