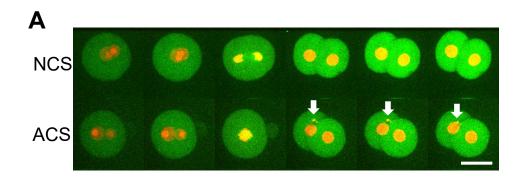
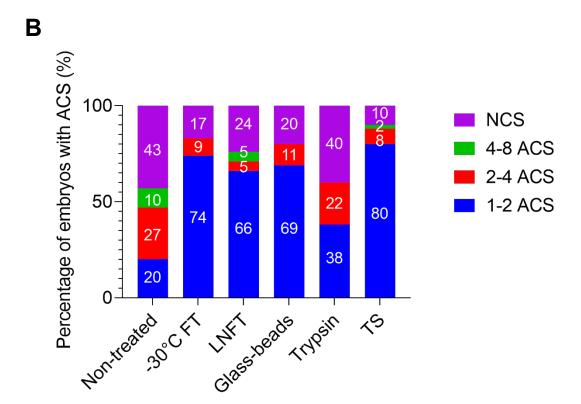
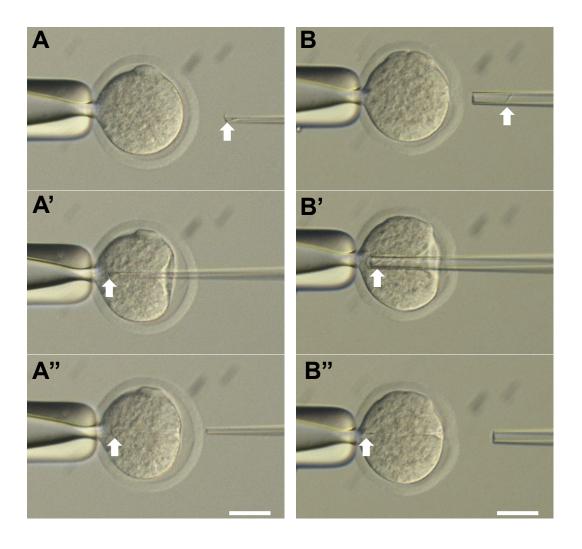


Supplementary Fig. 1. Morphology of mouse and rat sperm after tail removal treatment. (A-F) Bright field images of mouse sperm after tail removal treatment are shown; (A) non-treated control. (B) -30° C freeze-thawed, (C) liquid nitrogen freeze-thawed, (D) glass bead-homogenized, (E) trypsin-treated, and (F) TrypLE<sup>TM</sup> Select-treated. (G-H) Bright field images of rat sperm after treatment. (G) Non-treated, (H) Sonicated, (I) Trypsin-treated. Arrows indicate sperm that have lost their tail. Scale bars in A-F: 20 μm, in G-I: 15 μm.





**Supplementary Fig. 2.** Abnormal chromosome segregation found in ICSI embryos generated with treated sperm. (**A**) Timelapse images of ICSI embryos with NCS and ACS. Arrows indicate abnormal located chromosomes. Red: H2B-mRFP1, Green: EGFP-tubulin. Scale bar: 40 μm. (**B**) ACS frequency of ICSI embryos using treated sperms. ACS was analyzed at the 1–2, 2–4, and 4–8 cell divisions for each embryo. The percentages of ACS at each division in ICSI embryos from each treated and non-treated control sperm are shown. ACS: abnormal chromosome segregation, NCS: normal chromosome segregation.



**Supplementary Fig. 3.** Rat ICSI procedure. (**A**–**A**") The conventional rat ICSI process using a thin injection needle. (**B**–**B**") Rat ICSI using pre-activated oocytes and a thick needle. In **A**–**A**" and **B**–**B**", arrows indicate sperm head. Scale bars in **A**–**A**" and **B**–**B**": 40 μm.

## Supplementary Tables Supplementary Table 1. *In vitro* development of ICSI embryos using sperm with tails artificially removed

Treatment	No.	of 1	No. of embryos developed to						
	oocytes	Ī	pn (%)	2-cell (%)**	4-cell (%)**	Morula (%)**	Blastocyst (%)**		
Piezo-cut	40	3	35 (88)	33 (94)	31 (88)	29 (83)	24 (69) <sup>a</sup>		
−30°CFT	95	8	85 (89)	82 (96)	75 (88)	70 (82)	50 (59)		
LNFT	80	-	72 (90)	69 (96)	62 (86)	55 (76)	43 (60)		
Glass-beads	100*	Ģ	94 (94)	87 (93)	76 (81)	62 (66)	39 (41) b		
Trypsin	87*	-	70 (80)	68 (97)	67 (96)	65 (93)	47 (67)		
TS	92*	8	85 (92)	84 (99)	69 (81)	63 (74)	45 (53)		

<sup>\*</sup> These oocytes were activated with  $SrCl_2$ . \*\*The percentages relative to no. of embryos that developed into pronucleus (pn). Statistical analysis of blastocyst rate was performed between piezo-cut control and each treatment. Significant  $\chi^2$  comparisons a  $\nu s$ . b, P < 0.05.

**Supplementary Table 2**. Comparison of time burden between conventional ICSI and modified ICSI (pre-activated oocytes injected with trypsin-treated sperm) performed by operators with varying experience

Operator	Type of sperm	No. of oocytes used	No. of oocytes survived (%)*	No. of e develor pn (%)**	•	Required time (min) for injection	Required time (min)/ No. of oocyte used	No. of embryos transferred	No. of offspring (%)***
A	Piezo-cut	30	26 (87)	26 (100)	25 (96)	18.9	0.63	N.D.	N.D.
	Trypsin	30	30 (100)	30 (100)	29 (97)	16.1	0.53	N.D.	N.D.
В	Piezo-cut	30	23 (77)	23 (100)	23 (100)	19.9	0.66	N.D.	N.D.
	Trypsin	29	27 (93)	27 (100)	24 (89)	18.6	0.64	N.D.	N.D.
C	Piezo-cut	30	22 (73)	22 (100)	22 (100)	35.2	1.17	N.D.	N.D.
	Trypsin	30	27 (90)	27 (100)	25 (93)	30.8	1.02	N.D.	N.D.
D	Piezo-cut	30	20 (67)	19 (95)	17 (90)	28.4	0.94	N.D.	N.D.
	Trypsin	30	29 (97)	29 (100)	29 (100)	26.8	0.89	N.D.	N.D.
E	Piezo-cut	50	45 (90)	45 (100)	43 (96)	32.5	0.72	43	18 (42)
	Trypsin	50	50 (100)	50 (100)	49 (98)	23.5	0.47	24	9 (39)
F	Piezo-cut	40	36 (90)	35 (97)	31(86)	142.9	3.97	31	8 (26)
	Trypsin	40	36 (90)	33 (92)	30 (83)	117.3	3.26	14	5 (36)
G	Piezo-cut	36	32 (89)	24 (75)	23 (72)	151.5	4.73	23	2 (9)
	Trypsin	36	35 (97)	29 (83)	28 (80)	114.1	3.26	28	2 (7)
Н	Piezo-cut	40	24 (60)	23 (96)	22 (92)	103.8	4.33	22	6 (27)
	Trypsin	40	36 (90)	36 (100)	35 (97)	74.5	2.07	28	7 (25)

<sup>\*</sup> Percentages relative to the number of oocytes used. \*\*Percentages relative to the number of ooocytes survived. \*\*\* Percentages relative to the number of embryos transferred.

N.D.: not determined.