

Supplemental Figure S1.

Determine the optimized concentration of AKTi-1/2 to generate Tcm-like CAR-T cells. (A) Schematic diagram of plasmids expressing a second-generation CAR against human Carbonic Anhydrase 9 (hCA9). (B) Representative flow cytometry histogram of GFP+CAR-T cells expanded in 0-2 μ M AKTi-1/2. AKTi-1/2 was maintained in the entire period (**Mode1**) or after retroviral transduction (**Mode2**). (C) Western blotting of CAR-T cells during the course of the culture in 1 μ M AKTi-1/2 or vehicle. p-AKT, Phospho-Akt (Ser473). (D) Normalized count of T cells expanded in 0-2 μ M AKTi-1/2 for six days. T cells were divided equally among the groups on day 2 after retroviral transduction and T cell count were measured by flow cytometry on day 6. Normalize the T cell count of vehicle group to 100. (E) KEGG pathway analysis of the down-regulated genes of CD8+CAR-T cells expanded in 1 μ M AKTi-1/2 compared to vehicle. A one-way ANOVA was used for multiple comparisons in panel D, and P-values were adjusted by Tukey method. P<0.05 was considered statistically significant.



Supplemental Figure S2.

Ex vivo AKT inhibition promoted T cells expansion in vivo but had limited impact on tumor-infiltration. (**A**) Representative flow cytometry histogram of GFP+CAR-T cells. (**B**) Schematics of experiment; 5×10⁶ T cells with 50% CAR-T cells were adoptively transferred into tumor-bearing nude mice. (**C**) Picture and weight of inguinal lymph nodes on day 12. (**D**) Representative flow cytometry plot identifying T cells and GFP+CAR-T cells. (**E**) Percentages of T cells in CD45+ immune cells and percentages of CAR-T cells in T cells. Cells were isolated from inguinal lymph nodes 12 days after CAR-T therapy. (**F**) Representative flow cytometry plot of tumor-infiltrating CD8+T cells and GFP+CAR-T cells 12 days after CAR-T therapy. (**G**) Tumor infiltrating T cells per mm³ of tumor on day12. (**H**) Tumor infiltrating CAR-T cells per mm³ of tumor on day12. (**I**) Tumor progression. Tumor volume was calculated in cubic centimeter (1/2×length×width×width). **CTRL**, no adoptive therapy; **Ctrl-T**, T cells without retroviral transduction; **CAR-T**, normal CAR-T cells; **AKTi**, T cells were adjusted by Tukey method. An unpaired 2-tailed Student's t-test was used between two groups in panels C, E, G and H. P<0.05 was considered statistically significant.



Supplemental Figure S3.

Runx3-overexpression cooperated with AKT inhibition to generate CAR-T cells with both tissue-resident and central memory characteristics. (**A**) Representative flow cytometry plot of CD44+CD62L+ Tcm-like CAR-T cells. **CAR-T**, normal CAR-T cells; **CAR-RUNX3-T**, Runx3-overexpressed CAR-T cells; **AKTi**, T cells were expended in 1µM AKTi-1/2. (**B**) PCA analysis of the transcriptome data of CAR-T cells. **RUNX3-T**, Runx3overexpressed CAR-T cells. (**C-E**) Heatmap of the expression of core genes up-regulated or down-regulated in Trm cells, as quantified by mRNA-seq. (**C**) Validation of core Trm markers. Kid_D35, CD8+T cells isolated from kidneys 35 days after adoptive therapy; IEL, intraepithelial lymphocytes; Tcm, central memory T cells isolated from spleens; Tem, effector memory T cells isolated from spleens. (**D-E**) Analysis of the expression of core Trm markers in our CD4+CAR-T cells.



Supplemental Figure S4.

Runx3-overexpression enhanced the cytotoxic potential of CD4+CAR-T cells. (A) Volcano plot of differentially expressed genes between Runx3-OE CD4+CAR-T cells and normal CD4+CAR-T cells, as quantified by mRNA-seq (n=2 biological replicates each). **AKTi**, CAR-T cells were expanded in 1 μ M AKTi-1/2. **CAR-T**, normal CAR-T cells; **CAR-RUNX3-T**, Runx3-overexpressed CAR-T cells. (**B-C**) KEGG pathway analysis of the up-regulated genes of Runx3-OE CD4+CAR-T cells compared to normal CD4+CAR-T cells. (**D**) Medium fluorescence intensity of Granzyme B measured by flow cytometry. (**E**) Representative flow cytometry plot of IFN γ and TNF α expression in CAR-T cells. CAR-T cells of all groups were co-incubation with target cells for 6 hours in the presence of IL-2. A one-way ANOVA was used for multiple comparisons in panel D, and P-values were adjusted by Tukey method. P<0.05 was considered statistically significant.



Supplemental Figure S5.

Runx3-overexpression inhibited terminal differentiation of CD8+CAR-T cells induced by tonic signaling. (A) Schematic diagram of plasmids. The 2nd generation CAR-T cells were based on CD28 co-stimulation domain and the 3rd generation were based on CD28 and 4-1BB co-stimulation domains. The CAR and murine Runx3 (or GFP) were separated by a P2A ribosomal skip sequence. (B) Representative flow cytometry plot and percentages of INF γ and TNF α expression in CD8+CAR-T cells. **CAR-T**, normal CAR-T cells; **CAR-RUNX3-T**, Runx3-overexpressed CAR-T cells; **AKTi**, T cells were expended in 1 μ M AKTi-1/2. (C) Representative flow cytometry histogram and percentages of INF γ expression in CD8+CAR-T cells. A one or two-way ANOVA was used for multiple comparisons in panels B and C, and P-values were adjusted by Tukey method. P<0.05 was considered statistically significant.



Supplemental Figure S6.

Runx3-overexpression enhanced the anti-tumor activity of AKTi-generated CAR-T cells in vivo. (A) Schematics of experiment; sorted CAR-T cells were adoptively transferred into tumor-bearing nude mice. (B) Tumor progression. Tumor volume was calculated in cubic centimeter (1/2×length×width×width). CTRL, no adoptive therapy; Ctrl-T, T cells without retroviral transduction; CAR-T+AKTi, normal CAR-T cells expended in 1µM AKTi-1/2; CAR-RUNX3-T+AKTi, Runx3-overexpressed CAR-T cells expended in 1µM AKTi-1/2. (C) Survival curve. (D) The picture of surviving mice 90 days after CAR-T therapy with 10x10^6 normal or Runx3-overexpressed CAR-T cells expended in 1µM AKTi-1/2. P values of survival curve in panel C were calculated by log-rank test analysis. A two-way ANOVA was used for multiple comparisons in panel B, and P-values were adjusted by Tukey method. P<0.05 was considered statistically significant.



Supplemental Figure S7.

Runx3-OE enriched T cell population with Trm characteristics in vivo. (**A**) Expression of representative markers associated with Trm and Tcm phenotypes on different T cell population. T cells were isolated from lymph nodes 14 days after CAR-T therapy. The expression of markers was quantified by CyTOF. (**B**) Percentages of Tem1 and Tem2 cells in CD8+CAR-T cells identified by CyTOF. **CAR-T**, normal CAR-T cells; **CAR-RUNX3-T**, Runx3-overexpressed CAR-T cells; **AKTi**, T cells were expended in 1µM AKTi-1/2. (**C**) Percentages of cluster 2 and 3 cells expressing some Trm markers (CLA, CXCR6 and CXCR3) in CD8+CAR-T cells. A two-way ANOVA was used for multiple comparisons in panel B, and P-values were adjusted by Tukey method. An unpaired 2-tailed Student's t-test was used between two groups in panel C. P<0.05 was considered statistically significant.



Supplemental Figure S8.

Ex vivo AKT inhibition cooperated with Runx3-OE to promote CAR-T cell expansion and Runx3-OE further enhanced tumor-infiltrating CAR-T cell abundance in vivo. (A) Representative flow cytometry plot identifying T cells and CAR-T cells. Cells were isolated from inguinal lymph nodes 7 days after CAR-T therapy. (B) Picture of tumors 7 days after CAR-T therapy. CTRL, no adoptive therapy; Ctrl-T, T cells without retroviral transduction; CAR-T, normal CAR-T cells; CAR-RUNX3-T, Runx3-overexpressed CAR-T cells; AKTi, T cells were expended in 1µM AKTi-1/2. (C) Representative flow cytometry plot identifying tumor-infiltrating T cells. (D) Representative flow cytometry plot identifying tumor-infiltrating CAR-T and CAR-RUNX3-T cells. Fold Change was calculated through dividing the ratio of CAR-RUNX3-T to CAR-T on day8 or day30 by that on day0. Vehicle, T cells were expended without AKTi-1/2. (E) Representative flow cytometry plot identifying CAR-T and CAR-RUNX3-T cells in lymph nodes, spleens and blood. LN, lymph node. (F) Representative flow cytometry plot and (G) percentages of CCR2 expression on CAR-T cells 8 days after adoptive therapy. A two-way ANOVA was used for multiple comparisons in panels E and G, and P-values were adjusted by Tukey method. An unpaired 2-tailed Student's t test was used between two groups in panel D. P<0.05 was considered statistically significant.



Runx3-overexpression cooperated with AKT inhibition to enhance the persistence, cytotoxic potential and anti-tumor activity of human CAR-T cells as well. (A) Schematic diagram of plasmids expressing a thirdgeneration CAR based on CD28 and 41BB co-stimulation domains (CAR-T) against human Carbonic Anhydrase 9 or together with Runx3-overexpression (CAR-RUNX3-T). The CAR and human Runx3 were separated by a P2A ribosomal skip sequence. (B) Schema for the activation, lentiviral transduction (LV Td) and expansion of human peripheral blood lymphocytes in the presence of IL-2 (100IU/ml) and 1µM AKTi-1/2 or vehicle. (C) Representative flow cytometry plot and (D) percentages of CAR expression on CAR-T cells during the course of the culture. CAR-T, normal CAR-T cells; CAR-RUNX3-T, Runx3-overexpressed CAR-T cells; AKTi, T cells were expended in 1µM AKTi-1/2. (E) Representative flow cytometry plot and percentages of CD62L+GzmB- Tcm and CD62L-GzmB+ effector-type (Te) cells in CAR-T cells. (F) Representative flow cytometry plot and percentages of CD69+GzmB+ Trm-like cells and GzmB+ effector-type cells in CAR-T cells following 6 hours of co-incubation with hCA9-overexpressed Bxpc-3 cells. (G) Target-cell lysis activity of CAR-T cells following 20 hours of coincubation with hCA9-overexpressed Bxpc-3 cells. (H) Tumor progression in NCG mice. 8×10⁵ Sorted CAR-T cells were adoptively transferred into tumor-bearing NCG mice. Tumor volume was calculated in cubic centimeter (1/2×length×width×width). CTRL, no adoptive therapy; Ctrl-T, T cells without lentiviral transduction. A two-way ANOVA was used for multiple comparisons in panels D-H, and P-values were adjusted by Tukey method. P<0.05 was considered statistically significant.



Supplemental Figure S10.

Schematic diagram of the cooperation of Runx3-overexpression and ex vivo AKT inhibition to generate CAR-T cells with better persistence, tumor-residency and anti-tumor ability. (A-B) CAR-T cells expended in the presence of IL-2 exhibited mostly effector (Teff) phenotype with poor persistence. AKT inhibition promoted CAR-T cell CD62L+ central memory-like (Tcm) phenotype with better persistence but weaker cytotoxic potential compared to Teff. Runx3-overexpression promoted CAR-T cell tissue-resident memory (Trm) phenotype with better persistence, effector function and tumor residency ability. Runx3-overexpression cooperated with AKT inhibition to repress terminal differentiation induced by tonic signaling, which should promote CAR-T cell persistence. Runx3-overexpression cooperated with ex vivo AKT inhibition to generate CAR-T cells with both Tcm and Trm characteristics and enhanced antitumor activity, which responded to PD-1 blockade well.