Supplemental Table S1: Patient details.

ID	Age	Sex	Diagnosis	Treatment History	Mutations
Hu-56	58	F	Right base of tongue	Transoral robotic resection/neck dissection	unknown
			squamous cell carcinoma	(pT1N2bcM0) and adjuvant chemoradiation	
			(P16+)		
Hu-66	60	М	Pancreatic head	Neoadjuvant Gemcitabine/Abraxane followed by	KRAS G12V, CDKN2A/B
			adenocarcinoma	pancreaticoduodenectomy (pT3N1cM0) and	loss of exons 2-3, TP53
				adjuvant Capecitabine/Gemcitabine	G244C

TCR	Coreceptor	v	J	CDR3α	CDR3β	
		TRAV14/DV4*02,				
F29	CD8	TRBV7-6*01	TRAJ58*01, TRBJ1-5*01	CAMREPRATSGSRLTF	CASSLGEGFSNQPQHF	
		TRAV9-2*01,				
F12	CD4	TRBV6-1*01	TRAJ40*01, TRBJ1-2*01	CALSHYSGTYKYIF	CASSEGTDYGYTF	
		TRAV13-2,				
KRAS	CD4	TRBV5-4	TRAJ53, TRBJ1-1	CAENSGGSNYKLTF	CASSWERAGKAFF	

Supplemental Table S2: TCR gene sequences.

Peptide ID	AA Sequence
E6-1	MHQKRTAMFQDPQER
E6-2	RTAMFQDPQERPRKL
E6-3	FQDPQERPRKLPQLC
E6-4	QERPRKLPQLCTELQ
E6-5	RKLPQLCTELQTTIH
E6-6	QLCTELQTTIHDIIL
E6-7	ELQTTIHDIILECVY
E6-8	TIHDIILECVYCKQQ
E6-9	IILECVYCKQQLLRR
E6-10	CVYCKQQLLRREVYD
E6-11	KQQLLRREVYDFAFR
E6-12	LRREVYDFAFRDLCI
E6-13	VYDFAFRDLCIVYRD
E6-14	AFRDLCIVYRDGNPY
E6-15	LCIVYRDGNPYAVCD
E6-16	YRDGNPYAVCDKCLK
E6-17	NPYAVCDKCLKFYSK
E6-18	VCDKCLKFYSKISEY
E6-19	CLKFYSKISEYRHYC
E6-20	YSKISEYRHYCYSLY
E6-21	SEYRHYCYSLYGTTL
E6-22	HYCYSLYGTTLEQQY
E6-23	SLYGTTLEQQYNKPL
E6-24	TTLEQQYNKPLCDLL
E6-25	QQYNKPLCDLLIRCI
E6-26	KPLCDLLIRCINCQK
E6-27	DLLIRCINCQKPLCP
E6-28	RCINCQKPLCPEEKQ
E6-29	CQKPLCPEEKQRHLD
E6-30	LCPEEKQRHLDKKQR
E6-31	EKQRHLDKKQRFHNI
E6-32	HLDKKQRFHNIRGRW
E6-33	KQRFHNIRGRWTGRC
E6-34	HNIRGRWTGRCMSCC
E6-35	GRWTGRCMSCCRSSR
E6-36	GRCMSCCRSSRTRRE
E6-37	SCCRSSRTRRETQL

Supplemental Table S3: Individual peptides contained within E6 pepmix.

B Cell Line	DQA1	DQA2	DQB1	DQB2	DRB1	DRB2	DRB3	DRB4	DRB5
Hu-56	01:02	05:05	03:01	05:02	11:01	16:01			
Hu-195	01:04	05:05	03:01	06:01	13:03	14:04			
KAS011	01:02	01:02	05:02	05:02	16:01	16:01			
Hu-66	01:02	05:01	02:01	06:02	03:01	15:01	01:01		01:01
Hu-37	01:02	03:03	02:02	06:02	04:03	15:01		01:03	01:01
Hu-159	05:05	02:02	03:19	07:01	11:02	15:01	03:01	01:01	
Hu-198	01:03	05:01	02:01	06:03	03:01	13:01			
D66	N/A	N/A	05:01	06:02	01:01	15:01			01:01
D8	01:02	01:03	06:02	06:03	13:01	15:01			01:01
GM3107	N/A	N/A	06:02	06:02	15:01	15:01			01:01

Supplemental Table S4: HLA class II genotyping of B cell lines.

Peptide Sequence	Gene	aaPos	aaRef	aaAlt	mut.pos
FGLATEKSRWSGSHQFEQLS	BRAF	600	V	E	6
EDLTVKIGDFGLAT <mark>E</mark> KSRWS	BRAF	600	V	E	15
VVVGADGVGKSALTIQLIQN	KRAS	12	G	D	6
MTEYKLVVVGADGVGKSALT	KRAS	12	G	D	12
VVVGA <mark>V</mark> GVGKSALTIQLIQN	KRAS	12	G	V	6
MTEYKLVVVGAVGVGKSALT	KRAS	12	G	V	12
LSEITKQEKDFLWSHRHYCV	PIK3CA	545	E	К	6
LKAISTRDPLSEIT <mark>K</mark> QEKDF	PIK3CA	545	E	К	15
PSGGDHFCLGQLSNVHRTEA	SMAD4	361	R	Н	6
IVTVDGYVDPSGGD <mark>H</mark> FCLGQ	SMAD4	361	R	Н	15

Supplemental Table S6: Peptides representing recurrent cancer mutations.



Supplemental Figure S1: Cellular composition and epitope specificity of F29 TIL and HPV RNA expression by Hu-56 cell line and primary tumor. A Representative flow cytometry plots demonstrating percentage of CD4⁺ and CD8⁺ TIL among F12 and F29. **B** Cytokine secretion assay results demonstrating secretion of IFN-γ by CD8⁺ TIL after stimulation with either E6 pepmix or E6₂₉₋₃₈. Representative FACS plots from two independent experiments. **C** Quantification of RNA transcript reads expressed as transcripts per million (TPM) mapping to the indicated HPV-16 genes in HNSCC-56 primary tumor, HNSCC-56 cell line, and an HPV⁻ control cell line.



Supplemental Figure S2: Cytokine production, surface phenotype, and epitope specificity of E6-specific CD4⁺ TIL. A Representative FACS plots demonstrating intracellular cytokine staining of unstimulated (grey) or E6-stimulated (dashed line) F12 REP TIL for indicated cytokines. **B** TNF- α + and – cells were gated as a surrogate for E6-specific and non-specific, respectively, to determine the relative abundance of intracellular granzyme B in these two subsets. Data are representative of mean +/- SEM from 2 independent experiments. * p<0.05 paired T-test. C Representative FACS plots of unstimulated (left) or E6-stimulated (right) F12 REP TIL stained for intracellular IFN-y and surface PD1. D Individual peptides from the E6 pepmix (Table S1) were pulsed onto autologous B-LCLs prior to culture with E6-specific CD4⁺ TIL. IFN-γ secretion was measured by ELISA. Data is representative of 2 independent experiments.



Supplemental Figure S3: Addition of anti-CD28 agonist or anti-PD-L1 blocking antibodies do not impact tumor recognition by CD4⁺ T cells. A Quantification of CD137 upregulation after coculture of E6-specific F12 TCR-engineered CD4⁺ T cells with HNSCC-56 CIITA tumor cells with or without 1 µg/ml E6₁₋₁₅ peptide, 10 µg/ml anti-PD-L1, or 5 µg/ml anti-CD28. Data represent two independent experiments. **B** Quantification of CD137 upregulation after coculture of KRAS^{G12V}-specific TCRengineered CD4⁺ T cells with NCI-H2444 CIITA tumor cells with or without 1 µg/ml KRAS^{G12V} peptide, 10 µg/ml anti-PD-L1, or 5 µg/ml anti-CD28. Data represent two independent experiments.



Supplemental Figure S4: Membrane-anchored KRAS^{G12V} is not sufficient to

promote direct recognition by CD4⁺ T cells. A Schematic of KRAS^{G12V}-MITD construct (top) and transduction efficiency of NCI-H2444 and DAN-G measured by eGFP expression (bottom). **B** FACS plots demonstrating CD137 upregulation after coculture of TCR-engineered CD4⁺ T cells with designated CIITA/HLA-DRB5*01:01-expressing tumor lines pulsed with DMSO, KRAS^{G12V} peptide, or expressing the endogenous KRAS^{G12V}-MITD construct. Data representative of 2 independent experiments.