

## **Figure Legends**

Supplemental Figure 1. SDS-PAGE analysis following capture by IMAC of full-length, His-tagged GRB2 A) WT, B) N188D/N214D, C) V123D, and D) V122P/V123P. Loading order was as follows: bulk lysate (lane 1); post-capture flow-through (lane 2); eluate (lanes 3-5 and 7-10 or 11); molecular weight ladder (lane 6). Red stars denote target protein in lysate, absent thereafter in flow-through. Analyses representative of at least 3 independent experiments.

Supplemental Figure 2. SDS PAGE analysis of fractions spanning chromatographic peaks are shown for A) GRB2 WT, B) N188D/N214D, C) V123D putative monomer, and D) V122P/V123P putative dimer.

Supplemental Figure 3. SEC-MALS elutions of GRB2 dimers in full, with light scattering and refractive index plotted against elution time for A) WT dimer, B) N188D/N214D dimer, and C) V122P/V123P dimer. Dashed lines denote the molecular weights of GRB2 monomer (~28 kDa) and dimer (~56 kDa).

Supplemental Figure 4. Guinier analyses of the low-q regions, along with  $R^2$  are shown for GRB2 A) WT monomer, B) WT dimer, C) N188D/N214D monomer, D) N188D/N214D dimer, E) V123D putative monomer, and F) V122P/V123P putative dimer.

Supplemental Figure 5. Western immunoblot of whole-cell lysates (WCL) of transfected HuT78 T cells probing for GRB2 and GAPDH (control). Loading order was as follows: molecular weight ladder (lane 1); WCL of GRB2 knockdown (lane 2); WCL of GRB2 WT addback (lane 3); WCL of GRB2 V123D addback (lane 4); WCL of GRB2 V123N addback (lane 5); and WCL of GRB2 V122P/V123P addback (lane 6). Note: GRB2 V123N mutant was not used in these studies.

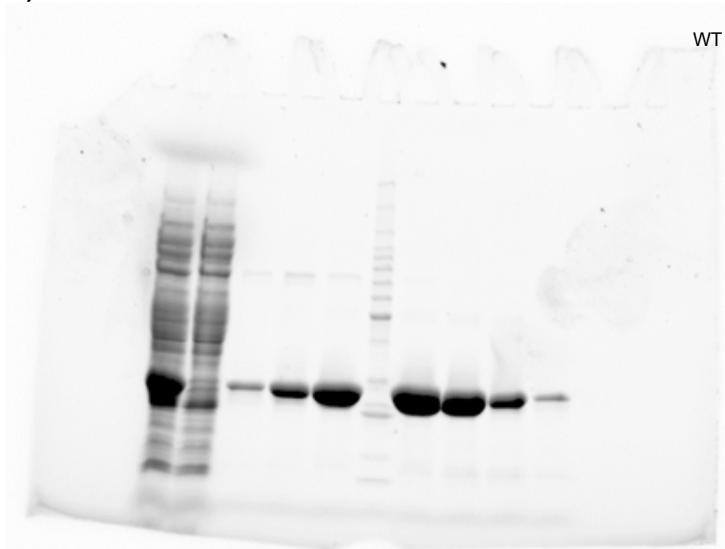
Supplemental Figure 6. Western immunoblot of whole-cell lysates (WCL) of transfected HuT78 T cells probing for phospho-LAT (pY226) and GAPDH (control).

Supplementary Table 1. Mammalian GRB2 structures available in the PDB.

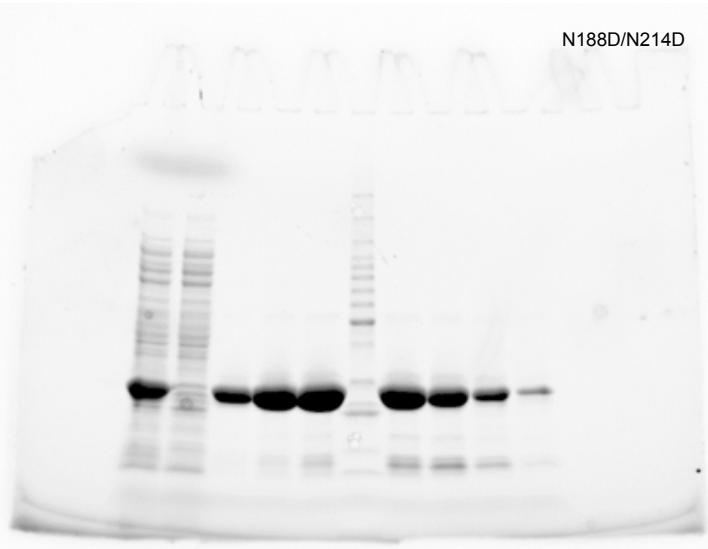
Supplemental Table 2. SEC-MALS-SAXS experimental parameters.

# Supplemental Figure 1

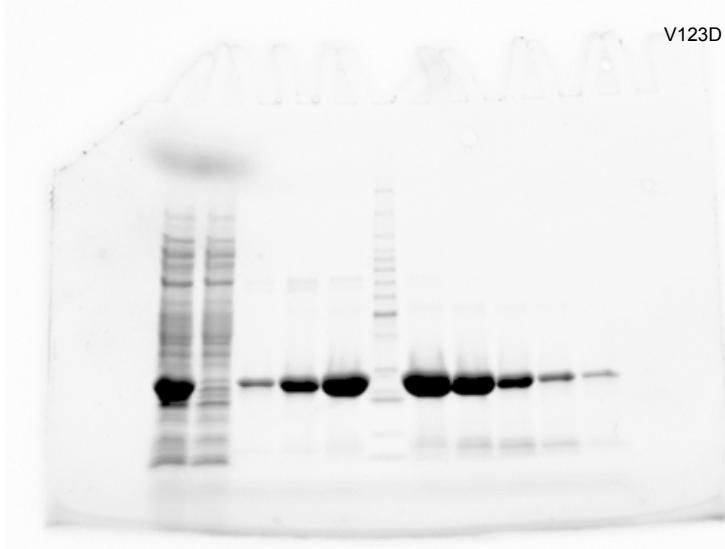
A)



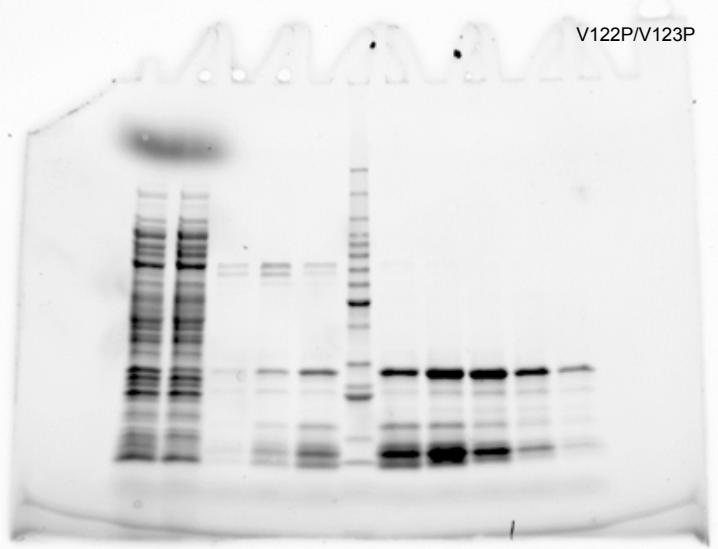
B)



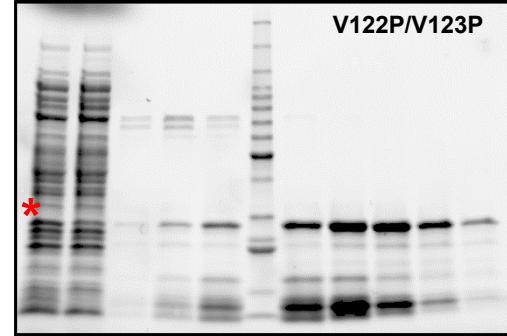
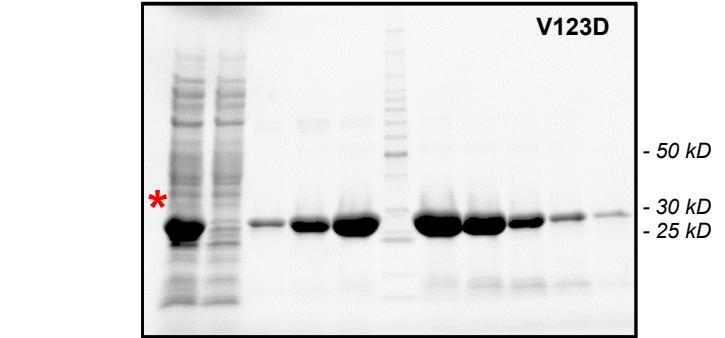
C)



D)

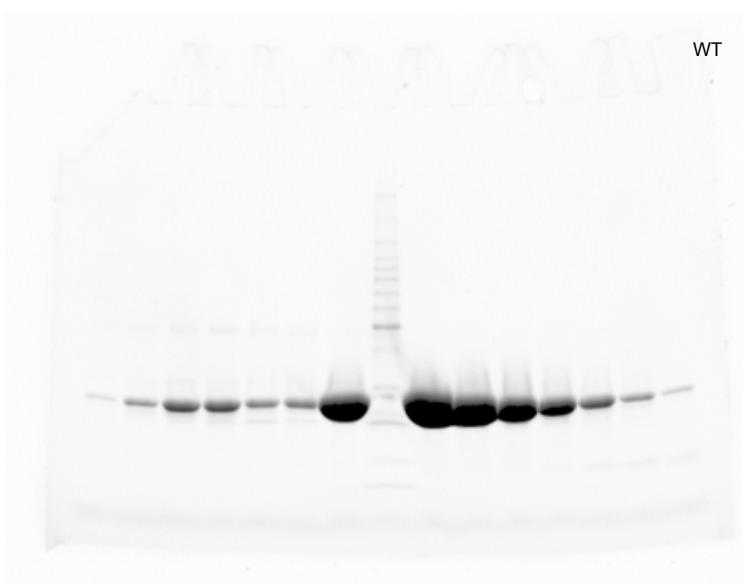


E)

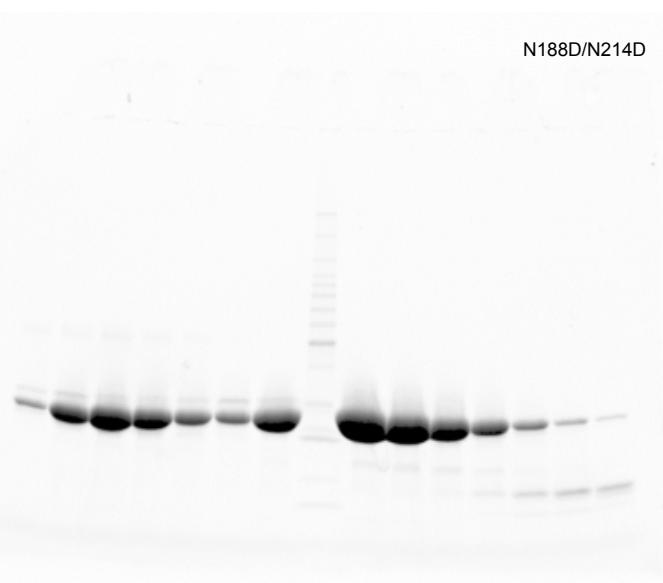


## Supplemental Figure 2

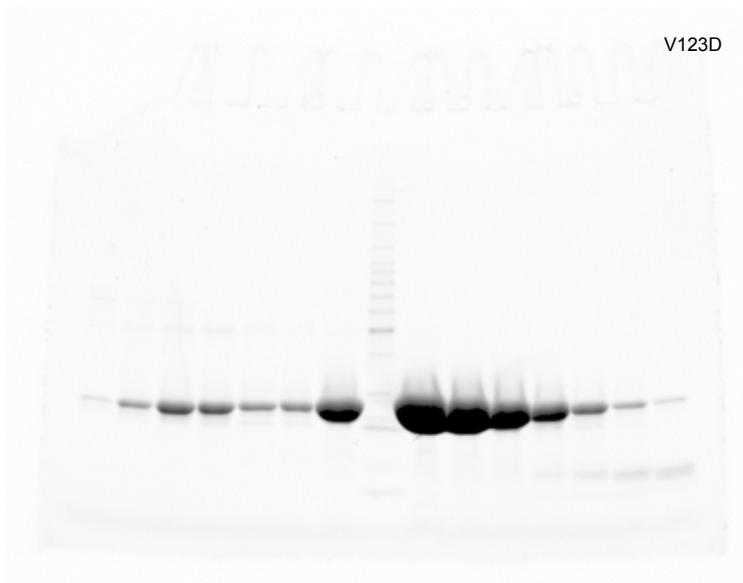
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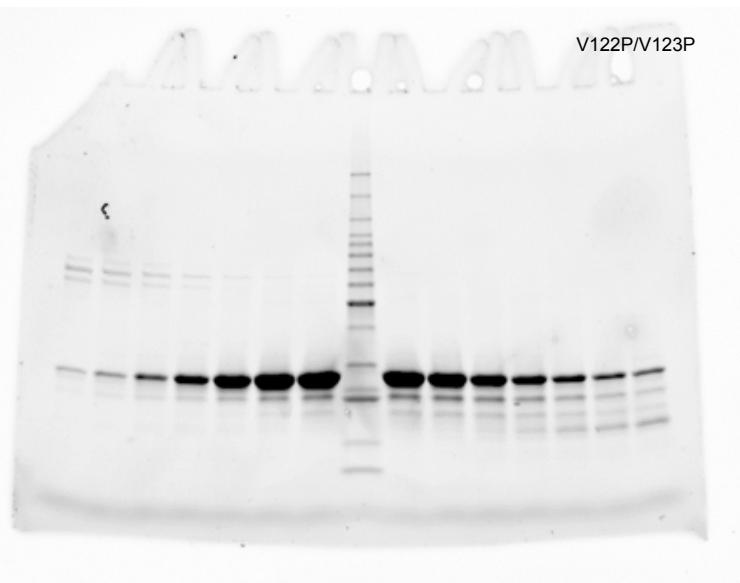
B)



C)

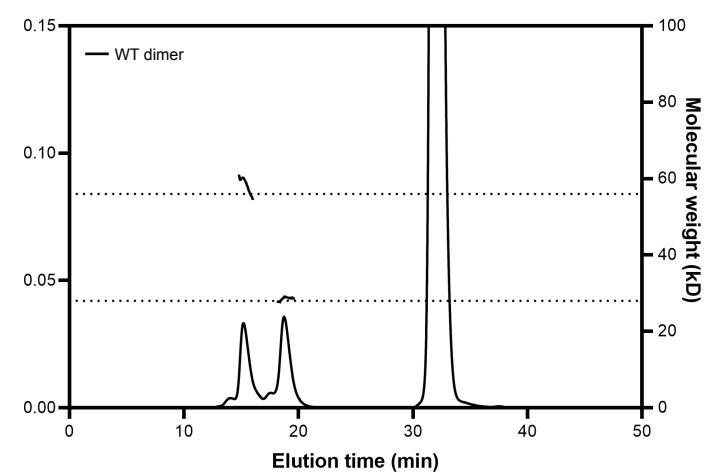
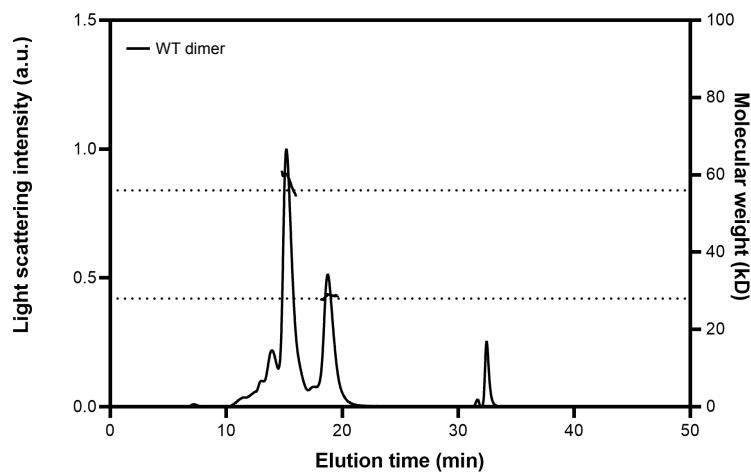


D)

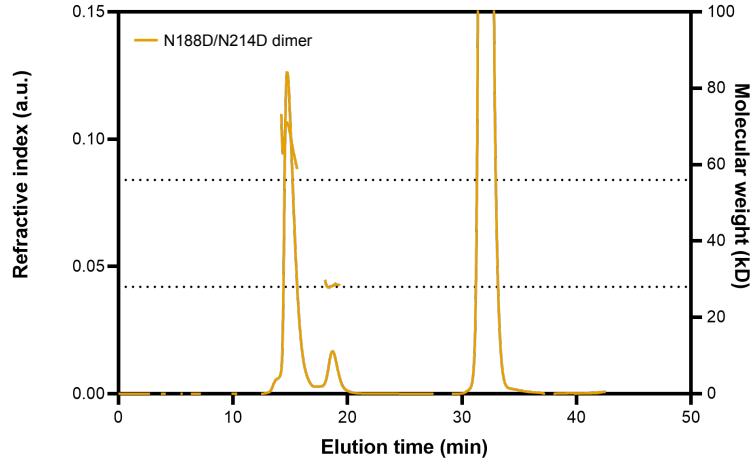
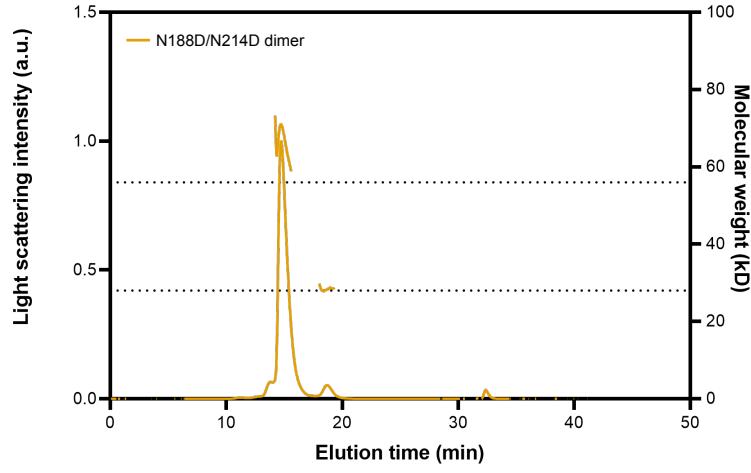


# Supplemental Figure 3

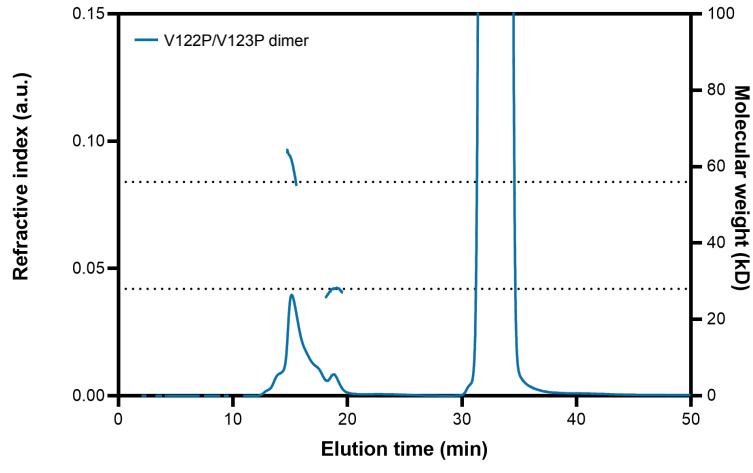
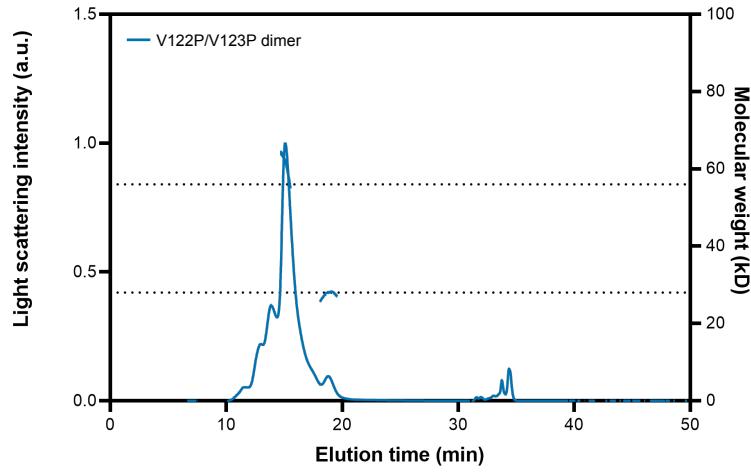
A)



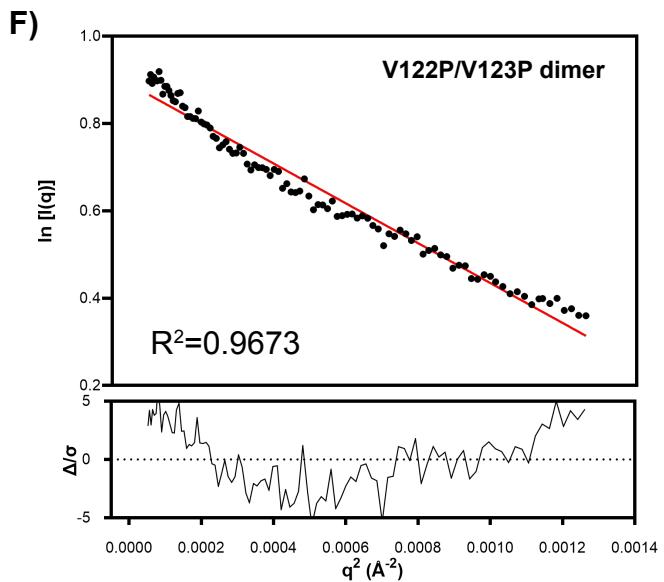
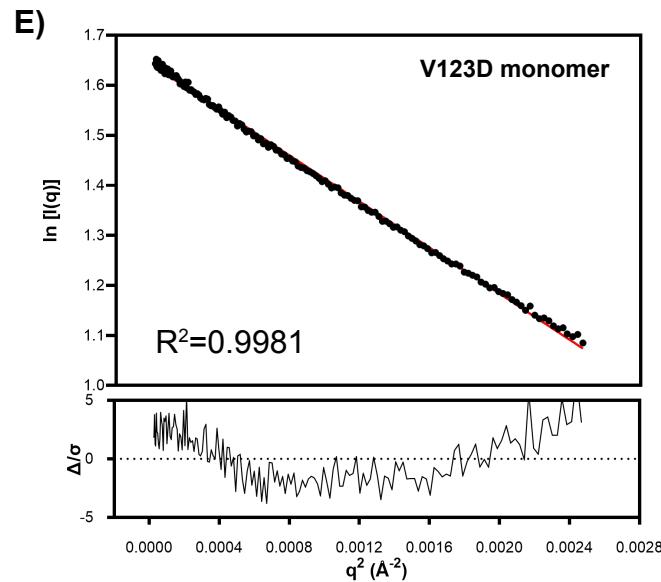
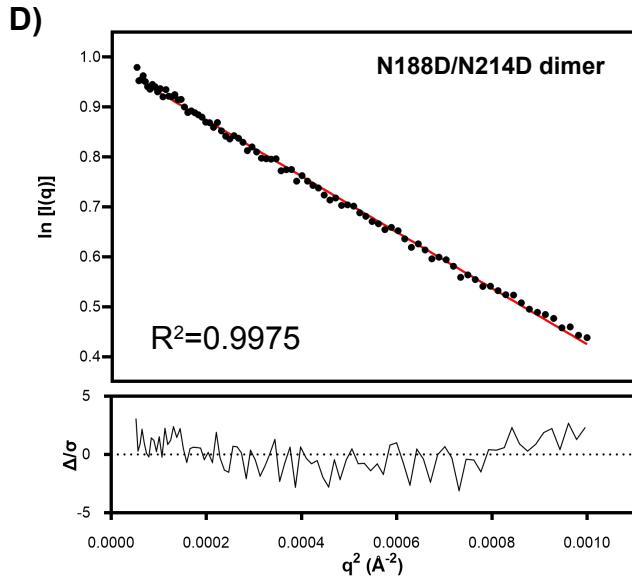
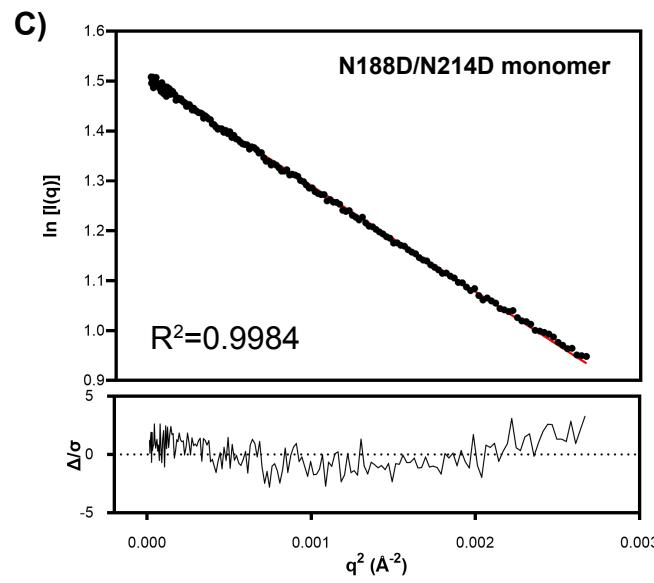
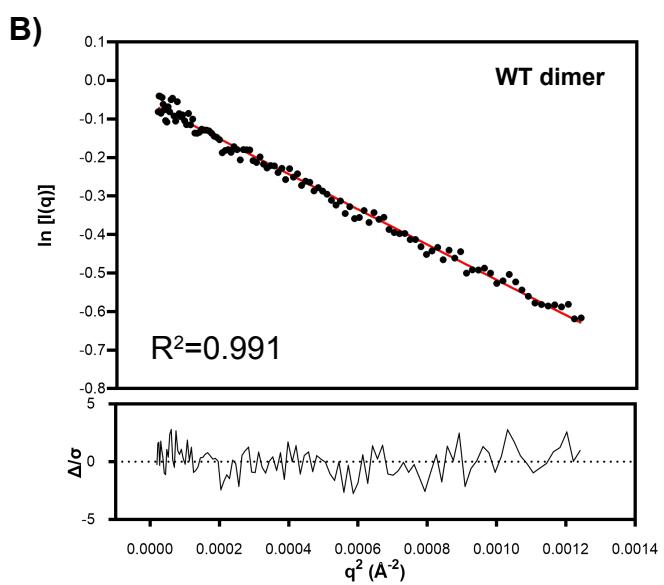
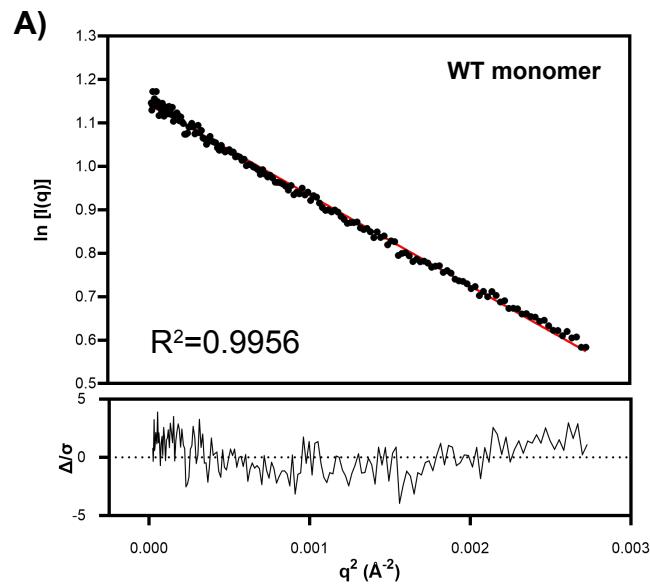
B)



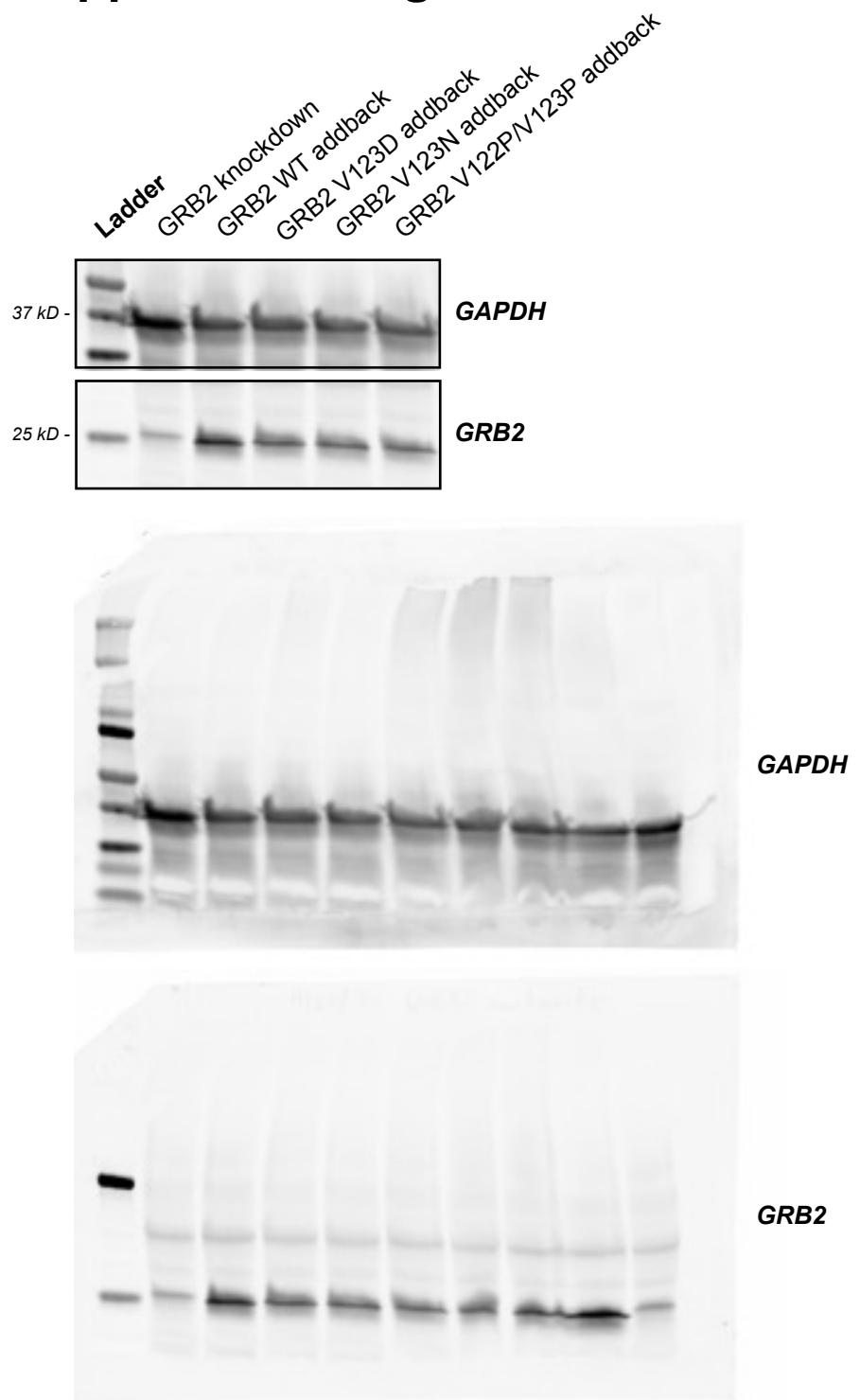
C)



# Supplemental Figure 4



# Supplemental Figure 5



# Supplemental Figure 6

Time (min) 0 2 5 15 0 2 5 15 0 2 5 15

LAT pY226

GAPDH

WT

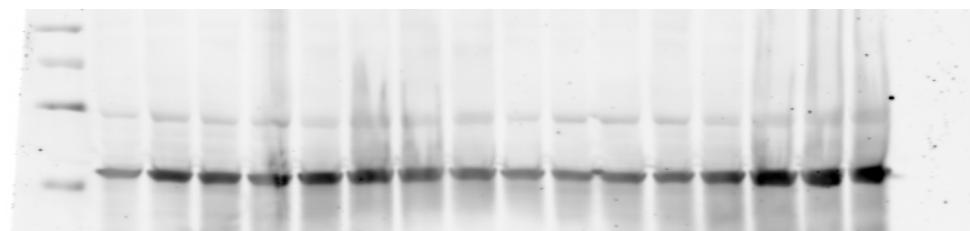
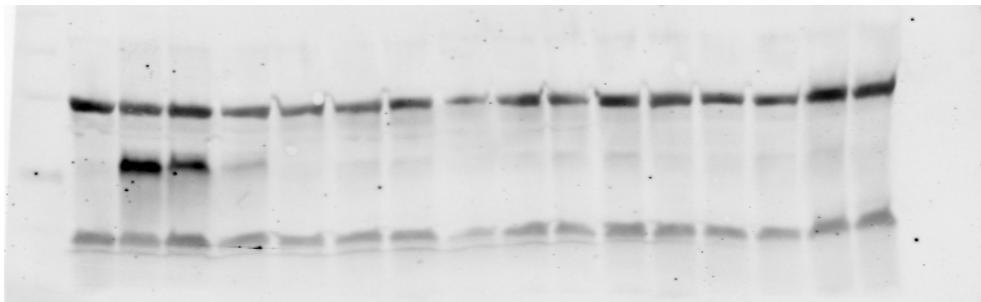
KD

V123D  
monomer

V122P/V123P  
dimer

LAT pY226

GAPDH



# Supplemental Table 1

PDB ID	Resolution (Å)	Number of entities	Molecular weight (kD)	Domain(s) represented	Residues represented	Ligand	Reference (PMID)
1BM2	2.1	2	14.42	SH2	49-163	Yes	10090780
1BMB	1.8	2	15.57	SH2	49-168	Yes	10090780
1CJ1	3	1	142.16	SH2	57-152	Yes	10395476
1FYR*	2.4	2	55.52	SH2	50-161	Yes	11063574
1GRI*	3.1	1	50.48	SH2	1-217	No	7716522
1JYQ	2	2	23.67	SH2	60-151	Yes	11827484
1JYR	1.55	2	12.13	SH2	60-151	Yes	11827484
1JYU	2.75	1	11.07	SH2	60-151	No	11827484
1TZE	2.1	2	12.39	SH2	55-152	Yes	8673601
1ZFP	1.8	2	12.37	SH2	56-153	Yes	9642078
2AOA	1.99	1	25.74	SH2	55-153	Yes	16165154
2AOB	1.8	1	55.36	SH2	55-153	Yes	16165154
2H46*	1.9	1	13.78	SH2	53-162	No	17466257
2H5K	3.25	2	28.01	SH2	53-162	Yes	17466257
2HUW	1.9	1	28.52	SH2	53-162	Yes	17001728
3C7I	1.7	1	14.26	SH2	53-162	Yes	17001728
3IMD	2	1	28.8	SH2	53-163	Yes	19886660
3IMJ	2.02	1	28.6	SH2	53-163	Yes	19886660
3IN7	2	1	28.63	SH2	53-163	Yes	19886660
3IN8	1.7	1	14.35	SH2	53-163	Yes	19886660
3KFJ	2.02	1	14.41	SH2	53-163	Yes	19886660
3MXC	2	2	12.89	SH2	55-152	Yes	22001015
3MXY	2.3	2	12.86	SH2	55-152	Yes	22001015
3N7Y	2.02	2	36.44	SH2	55-152	Yes	21116482
3N84	2	2	83.89	SH2	53-163	Yes	21116482
3N8M	2	2	14.63	SH2	53-163	Yes	21116482
3OV1	1.6	2	14.51	SH2	53-163	Yes	22007755
3OVE	1.82	2	14.2	SH2	53-163	Yes	22007755
3S8L	1.71	2	14.38	SH2	53-163	Yes	22007755
3S8N	1.71	2	14.62	SH2	53-163	Yes	22007755
3S8O	1.85	2	14.34	SH2	53-163	Yes	22007755
3WA4	1.35	2	12.58	SH2	60-152	Yes	24098653
4P9V	1.64	2	14.45	SH2	53-163	Yes	24856058
4P9Z	1.8	2	14.69	SH2	53-163	Yes	24856058
5CDW	2.602	2	197.82	SH2	54-153	Yes	26645482
6ICG	1.15	1	22.57	SH2	60-152	No	30923665
6ICH*	2	1	11	SH2	60-152	No	30923665
6WM1	1.8	2	29.06	SH2	53-163	Yes	32916312
6WO2	2	2	28.92	SH2	53-163	Yes	32916312
1GCQ	1.68	2	22.15	C-SH3	159-217	Yes	11406576
2VVK	1.6	1	6.46	C-SH3	161-214	No	19523899
2VWF	1.58	2	8.39	C-SH3	159-214	No	19523899
2W0Z	1.7	2	7.57	C-SH3	159-214	No	19523899
6SDF	2.5	1	14.7	N-SH3	2-59	No	32510467

\*Domain-swapped dimer

## Supplemental Table 2

<b>A) Sample details</b>	
SEC Column	Superdex 75 increase 10/300
Loaded concentration (mg/ml)	8.28 (WT monomer), 4.46 (WT dimer), 8.37 (N188D/N214D monomer), 2.84 (N188D/N214D dimer), 8.68 (V123D monomer), 4.00 (V122P/V123P dimer)
Injection volume (ul)	250
Flow rate (ml/min)	0.5
Solvent (solvent blanks taken from SEC flow-through prior to elution of protein)	20 mM Tris pH 8.0. 150 mM NaCl, 1 mM DTT
<b>B) SAXS data collection parameters</b>	
Instrument	BioCAT facility at the Advanced Photon Source beamline 18ID with Pilatus3 1M (Dectris) detector
Wavelength (Å)	1.033
Beam size (um <sup>2</sup> )	150 (h) x 80 (v)
Camera length (m)	3.5
q measurement range (Å <sup>-1</sup> )	0.0045-0.35
Absolute scaling method	N/A
Basis for normalization to constant counts	To incident intensity, by ion chamber counter
Monitoring for radiation damage	Frame-by-frame comparison of data
Exposure time	0.5 sec exposure time with 1 sec total exposure period of entire SEC elution
Sample configuration	SEC-MALS-DLS-RI-SAXS. Size separation used a Superdex 75 increase 10/300 GL column and an Infinity II HPLC (Agilent Technologies). Flow was in line with the UV-MALS-DLS-RI instruments and SAXS after the column. UV data was measured in the Agilent, and MALS-DLS-RI data by DAWN HELEOS-II (17 MALS + 1 DLS channels) and Optilab T-rEX (RI) instruments (Wyatt). SAXS data was measured with sheath-flow cell in a 1.5 mm ID 1.52 mm OD quartz capillary, effective path length 0.49 mm.
Sample temperature (°C)	22
<b>C) Software employed for SAXS data reduction, analysis, and interpretation</b>	
SAXS data reduction	Radial averaging, background subtraction, frame comparison, averaging, and subtraction were made using BioXTAS RAW 2.0.2.
Basic analysis: Guinier, MW, Normalized Kratky, P(r)	Guinier plot and molecular weights were calculated using BioXTAS RAW 2.1.1, P(r) function using GNOM from ATSAS 3.0.
MALS-DLS-RI analysis	Astra 7.1.3 (Wyatt)