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Supplemental information

Progressive development

of melanoma-induced cachexia

differentially impacts organ systems in mice

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Supplemental Figure S1, Related to Figure 1. The secretome of the mouse stroma is influenced by but differs from that of adjacent human cancer cells. Analysis of the transcriptional changes induced in the mouse stroma (microenvironment) by human cachectic versus non-cachectic melanoma xenografts. Compared to secreted factors expressed by human cachectic melanomas (Fig. 1), the mouse stroma expresses a largely different panel of secreted factors that are upregulated (A) and downregulated (B) by vicinity to cachectic melanoma cells compared to non-cachectic melanoma cells. (C) Analysis of the overall similarity between secreted factors significantly expressed in the cachectic versus non-cachectic cancer cells (x axis) and stroma (y axis) indicates minimal overlap (R²=0.1384). In (A-B), secreted factors with p<0.05 and Log2R>1 (A) and Log2R<-1 (B) are shown. For comparison in (C), secreted factors with p<0.05 in both conditions are shown, irrespective of the Log2R values.



Supplemental Figure S2, Related to Figure 2. The cachexia-non-inducing ("non-cachectic") melanoma tumors do not induce body and tissue wasting. (A) Analysis of mice injected with cachexia-non-inducing ("non-cachectic") melanoma cells. After 13 weeks from cancer cell injection, there is no decline in the tumor-free body mass, even if the tumor mass has reached a substantial size (~4g) at that stage (B). (C) There is a minor but significant increase in the length of the tibia bone from 2 to 13 weeks post cancer cell injection. (D-G) The weight of several skeletal muscles (tibialis anterior, gastrocnemius, soleus, diaphragm) remains unchanged even after 13 weeks from cancer cell injection. (H-J) Likewise, the weight of the heart (H) and liver (I) does not change, whereas (J) there is a trend towards a decline in the weight of the white adipose tissue (WAT). (K) The weight of the brown adipose tissue (BAT) declines from 2 to 4 weeks post cancer cell injection but then remains substantially stable. The graphs report the mean \pm SD, the n is indicated; *p<0.05 and **p<0.01.



Supplemental Figure S3, Related to Figure 4. Principal Component Analysis (PCA) of RNA-seq from distinct tissues at different time points from implantation of cachexia-inducing ("cachectic") and cachexia-non-inducing ("non-cachectic") melanoma xenografts. Principal Component Analysis (PCA) of RNA-seq data from: brain regions, i.e. cerebellum, hippocampus, and neocortex; skeletal muscles, i.e. gastrocnemius, soleus, and tibialis anterior; brown and white adipose tissues; heart; and liver. PCA indicates that RNA-seq is reproducible because samples of the same group cluster together. Moreover, although cachectic samples co-cluster with control mock-injected and non-cachectic samples at an early time point (2 weeks from tumor implantation), they progressively diverge from control samples at later time points.



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Supplemental Figure S4, Related to Figure 4. The cachexia-inducing ("cachectic") melanoma tumors induce transcriptional changes in the liver, brown adipose, and white adipose tissue.

(A-D) Transcriptional changes induced in the liver. (A) Heatmap of 890 genes that are most highly modulated in the liver of cachectic mice, compared to controls. The average z-scores are color-coded (downregulated genes are shown in blue; upregulated genes are shown in red). (B) Upregulated genes include secreted proteins, protease inhibitors, lipocalins, and cell adhesion proteins. (C) Cachexia in the liver is characterized primarily by downregulation of gene expression, and categories such as "lipid metabolism", "endoplasmic reticulum", and "peroxisome" are over-represented. Genes modulated with p<0.05 and Log2R>1 (B) and Log2R<-1 (C) in the liver of cachectic melanomas versus control at 8 weeks post tumor implantation are shown. (D) Examples of genes downregulated by cachexia in the liver include regulators of lipid metabolism and peroxisomal proteins: *Fads2*, *Acsl1*, *Ecl1*, *Ddc*, *Pex6*, *Cat*, *Hadh*, and *Lpin2*.

(E-H) Transcriptional changes induced in the brown adipose tissue. (E) Heatmap of 1018 genes that are tightly associated with cachexia in the brown adipose tissue. The average z-scores are color-coded (downregulated genes are shown in blue; upregulated genes are shown in red). (F) Upregulated genes are the majority and include secreted proteins, complement pathway components, growth factors, and lysosomal proteins. (G) Few genes are downregulated by cachexia and these include secreted and transmembrane proteins. Genes modulated with p<0.05 and Log2R>1 (F) and Log2R<-1 (G) at 8 weeks post tumor implantation are shown. (H) Examples of genes upregulated by cachexia in the brown adipose tissue include lysosomal cathepsins *Ctsb* and *Ctsc*, and stearoyl-CoA desaturase-1 (*Scd1*).

(I-L) Transcriptional changes induced in the white adipose tissue. (I) Heatmap of 921 genes that are modulated by cachexia in the white adipose tissue. The average z-scores are color-coded (downregulated genes are shown in blue; upregulated genes are shown in red). (J) Upregulated genes include extracellular proteins, lipocalins, and regulators of lipid transport and metabolism. (K) Collagen, secreted proteins, and microtubule components are among downregulated genes. Genes modulated with p<0.05 and Log2R>1 (J) and Log2R<-1 (K) at 8 weeks post tumor implantation are shown. (L) Examples of genes upregulated by cachexia in the white adipose tissue include proteins involved in lipid metabolism: *Acot1*, *Acot3*, and *Apoe*.

In all graphs in (A-L), the bars represent SD, n is indicated; TPM indicates transcripts per million reads. All comparisons are significant (p<0.05) in cachectic versus control at 8 weeks post tumor cell injection.



Supplemental Figure S5, Related to Figure 4. The cachexia-inducing ("cachectic") melanoma tumors induce transcriptional changes in the heart and skeletal muscles.

Transcriptional changes induced in the heart. (A) Heatmap of 845 genes that are most highly modulated in the heart of cachectic mice, compared to controls. The average z-scores are color-coded (downregulated genes are shown in blue; upregulated genes are shown in red). Most of these genes are upregulated whereas fewer are downregulated. Upregulated genes include secreted proteins, complement pathway components, and lysosomal and immune proteins. Downregulated genes are enriched for voltage-gated channels and glycoproteins. Genes modulated with p<0.05 and Log2R>1 and Log2R<-1 in the heart of cachectic melanomas versus control at 8 weeks post tumor implantation are shown. Examples of genes upregulated by cachexia in the heart include lysosomal proteins, such as *Lamp2*, *Ctsc*, and *Ctss*, which may drive heart wasting via the autophagy/lysosome system. Upregulation of *Kcne4* and downregulation of *Scn4a*, *Cacna1s*, *Hcn2*, and *Clcn1* ion channels also occurs in cachexia, and these changes in ion channel expression have been previously linked with arrhythmias and heart dysfunction. Bars represent SD, n is indicated; TPM indicates transcripts per million reads. All comparisons are significant (p<0.05) in cachectic versus control at 8 weeks post tumor cell injection.

Transcriptional changes induced in the gastrocnemius skeletal muscle. (B) Heatmap of 930 genes that are most highly modulated in the gastrocnemius muscle by cachectic melanoma xenografts compared to controls. These consist primarily of cachexia-upregulated genes that include secreted, Jak/Stat, endoplasmic reticulum, and other upregulated proteins (p<0.05 and Log2R>1). Downregulated proteins (p<0.05 and Log2R<-1) include secreted proteins and phosphatases.

Transcriptional changes induced in the soleus skeletal muscle. (C) Heatmap of 609 genes that define the impact of melanoma-induced cachexia on the soleus, compared to controls. Upregulated genes include extracellular proteins, complement pathway components, lipocalins, and genes involved in triglyceride homeostasis. Downregulated genes are enriched for secreted and extracellular matrix proteins.

Transcriptional changes induced in the tibialis anterior skeletal muscle. (D) Heatmap of 870 genes that define the impact of melanoma-induced cachexia on the tibialis anterior, compared to controls. Upregulated genes, which are the majority, include ribosomal and secreted proteins, complement pathway components, and autophagy/lysosome components. Downregulated genes are enriched for proteins with immunoglobulin domains. Genes modulated with p<0.05 and Log2R>1 and Log2R<-1 in tibialis anterior muscles of cachectic versus control mice at 8 weeks post tumor implantation are shown.



Supplemental Figure S6, Related to Figures 4 and 5. (A-C) Examples of E3 ubiquitin ligases modulated by melanoma-induced cachexia in skeletal muscles. Cachexia prominently induces E3 ubiquitin ligases already implicated in atrophy (*Fbxo32, Trim63, Fbxo31, Socs3*) as well as other E3s not previously associated with wasting (*Lnx1* and *Mylip*) in the gastrocnemius (A), soleus (B), and tibialis anterior (C) skeletal muscles. Ubr4 is significantly modulated in the tibialis anterior and gastrocnemius but not in the soleus muscle. While these E3s were upregulated in skeletal muscle by cachexia, other E3s such as *Trim7* are significantly downregulated. Unless otherwise noted, all comparisons are significant (p<0.05) in cachectic versus control at the 8 weeks post tumor cell injection. The graphs report the mean \pm SD, the N is indicated. (D-E) The cachexia-non-inducing ("non-cachectic") melanoma tumors do not induce body and tissue wasting. (D) Heatmap of 586 genes (divided in 5 clusters) that are most highly modulated by LLC cancer cells in brain regions (cerebellum, hippocampus, neocortex) compared to controls. (E) Different gene categories are enriched in these 5 clusters, including secreted and extracellular proteins (clusters #1 and #2), proteins involved in immunity and chemotaxis (cluster #3), lipocalins and neuropeptides (cluster #4), and sterol biosynthesis (cluster #5).



Supplemental Figure S7, Related to Figure 7. Analysis of tibialis anterior muscles from mice implanted with cachectic melanomas and controls, and either treated with Lisinopril or mock-treated.

(A) Representative images of tibialis anterior (TA) muscle cross-sections from control mice and mice implanted with cachectic melanomas, and either mock-treated or treated with Lisinopril. (B) Frequency and gaussian distribution of Feret's minimal diameters for type 2a, 2x, and 2b myofibers from TA muscles sourced from control mice (gray), control mice treated with Lisinopril (green), mice implanted with cachectic melanoma xenografts (blue), and mice implanted with cachectic melanoma xenografts and treated with Lisinopril (orange). Melanoma xenografts reduce myofiber size but Lisinopril does not impact myofiber size. (C) Similar results are found with the analysis of the Feret's minimal diameter of different myofiber types from each TA muscle: there is significant decline in the size of type 2a, 2x, and 2b myofibers but this is not affected by Lisinopril. (D) Melanoma-induced cachexia tends to decrease the percentage of type 2a and type 2x myofibers and to correspondingly increase the percentage of type 2b myofibers. Whereas Lisinopril has no effect by itself, it significantly decreases the percentage of type 2a and 2x myofibers and increases the percentage of type 2b myofibers in conjunction with cachexia. (E) There are no significant changes in the total number of myofibers present in TA muscles in response to cachexia and/or Lisinopril treatment. In (C-E), the mean ±SD is shown with *p<0.05 and ****p<0.0001 with n(TA)=15 for control mice, n(TA)=14 for control mice treated with Lisinopril, n(TA)=10 for mice implanted with cachectic melanomas and mock-treated, and n(TA)=13 for mice implanted with cachectic melanomas and treated with Lisinopril.