Supplementary Information for

mTORC1 upregulates B7-H3/CD276 to inhibit antitumor T cells and drive tumor immune evasion

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Supplementary Fig. 1



Supplementary Fig. 1. B7-H3 expression is regulated by mTORC1, but not mTORC2, STAT3 or YY1.

a-c, Suppression of B7-H3 protein expression in A549 cells (**a**), T47D cells (**b**), and PC3 cells (**c**) after treatment with Rapamycin (Rapa) or Torin 1 for 24 hrs (n = 3).

d-f, Suppression of B7-H3 mRNA expression in A549 cells (d), T47D cells (e), and PC3 cells (f) (n = 3) treated as in **a-c**. Means \pm SD, one-way ANOVA with Dunnett's test, * p < 0.05.

g, Suppression of B7-H3 with downregulation of Raptor, mTOR, and S6K. *Tsc2-/-* 105K cells with stable reconstitution of TSC2 or empty vector (EV) were transfected with non-targeting control siRNA (Ctrl) or SMARTpool siRNA targeting Raptor, Rictor, mTOR, S6K, or 4E-BP1 for 48 hr (n = 3).

h, B7-H3 mRNA expression in the cells from (g) (n = 3). Means \pm SD, two-way ANOVA with Holm-Sidak's multiple comparisons test, **** *p* < 0.0001.

i, Knockout of Raptor in iRapKO MEFs decreases B7-H3 expression. iRapKO or control (Ctrl) MEFs were treated with 1 μ M 4-hydroxytamoxifen (4-OHT) for 72 hr (n = 3).

j, Knockout of Rictor in iRicKO MEFs does not affect B7-H3 expression. iRicKO or control (Ctrl) MEFs were treated with 1 μ M 4-hydroxytamoxifen (4-OHT) for 72 hr (n = 3).

k, Immunoblot analysis of HEK293T cells transfected with wild-type mTOR (FLAG-mTOR-WT), constitutive active mutant mTOR in expression vectors followed by whole-cell-lysis 48 hr after transfection (n = 3).

I, Immunoblot analysis of WT MEFs that were starved of amino acid for 16 hr and re-stimulated with MEM amino acid for 24 hr (n = 3).

m,**n**, B7-H3 protein expression in *Tsc2-/-* 105K cells (**m**) and *Tsc2* KO MEFs (**n**) following transfection with control siRNA or SMARTpool siRNA targeting STAT3 for 48 hr (n = 3).

o, B7-H3 mRNA expression in *Tsc2-/-* 105K cells with stable reconstitution of TSC2 or empty vector (EV) following transfection with control siRNA (Ctrl) or SMARTpool siRNAs targeting YY1 for 48 hr (n = 3).

Source data and exact p values are provided in the Source data file.

Supplementary Fig. 2





Supplementary Fig. 2. S6K forms a complex with YY2 in the nucleus.

a,b, YY2 mRNA expression in *Tsc2*-WT and *Tsc2* KO MEFs (**a**) and *Tsc2-/-* 105K cells with stable reconstitution of TSC2 or empty vector (EV) (**b**) following 20 nM Rapamycin (Rapa) or vehicle control for 24 hr (n = 3). Means ± SD, two-tailed unpaired student's t-test.

c,d, YY1 mRNA expression in *Tsc2*-WT and *Tsc2* KO MEFs (**c**) and *Tsc2-/-* 105K cells with stable reconstitution of TSC2 or empty vector (EV) (**d**) following 20 nM Rapamycin (Rapa) or vehicle control for 24 hr (n = 3). Means \pm SD, two-tailed unpaired student's t-test.

e, Cytoplasmic and nuclear fractions of *Tsc2* KO MEFs following 10 μ M PF-4708671 treatment for the indicated time points (n = 3).

f, Immunofluorescent staining of HeLa cells stably expressing EGFP-YY2 and treated with 20 nM rapamycin (Rapa), 500 nM Torin 1, 30 μ M PF-4708671 or vehicle for 2 hr. Cells were fixed and stained with pS6 (S235/S236) and DAPI (n = 3). Scale bar = 100 μ m.

g, Immunofluorescent staining of parental HeLa cells with anti-YY2 and S6K antibodies (n = 3). White arrows indicate co-localization of YY2 and S6K. Top panel: scale bar = 100 μ m, bottom panel: scale bar = 20 μ m.

h, Left panel: Representative images of in situ proximity ligation assay between S6K and YY2 (red) in HeLa cells. Data are representative of three independent experiments. Scale bar = 30 μ m. Right panel: Quantification of the number of PLA puncta per nucleus. Column 1, IgG; column 2, mouse anti-YY2 and IgG; column 3, rabbit anti-S6K antibody and IgG; column 4, mouse anti-YY2 and rabbit anti-S6K antibody and IgG; column 4, mouse anti-YY2 and rabbit anti-S6K antibodies. n = 30 cells. Means ± SD, one-way ANOVA with Dunnett's multiple comparisons test, **** p < 0.0001.

Source data are provided in the Source data file.



Supplementary Fig. 3. The effects of B7-H3 inhibition *in vitro* and *in vivo*.

a,b, Immunoblot of sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* 105K cells (a) or *Tsc2-/-* TTJ cells
(b) (n = 3).

c,d, Cell proliferation assessed by crystal violet staining of the cells in **(c)** and **(d)** (n = 8). Means ± SD, one-way ANOVA with Dunnett's multiple comparisons test.

e,f, Representative images of the cells in a, b grown in soft agar for 3 weeks (n = 3).

g, Immunoblot analysis of whole-cell lysates derived from sgCtrl, sgCd276 (1), and sgCd276 (2) *Tsc2-* /- 105K cells (n = 3).

h, Subcutaneous growth of sgCtrl, sg*Cd*276 (1), and sg*Cd*276 (2) *Tsc*2-/- 105K cells in wild-type C57BL/6J mice. n = 7 mice for sgCtrl and sg*Cd*276 (1), n = 8 mice for sg*Cd*276 (2). Means \pm SD, nonparametric Kruskal-Wallis's test with Dunn's multiple comparisons test, * *p* < 0.05, ** *p* < 0.01.

i, Experimental design for treatment of *Tsc2-/-* 105K tumors with isotype control antibody (Isotype IgG) or anti-B7-H3 antibody (α -B7-H3) (Created with Biorender.com).

j, Growth of *Tsc2-/-* 105K tumors treated as depicted in i. n = 9 mice for Isotype IgG, n = 10 mice for anti-B7-H3 antibody. Means \pm SD, Mann-Whitney U Test, ** p < 0.01, **** p < 0.0001.

k,**l**, Flow cytometry analysis of tumor-infiltrating CD4⁺ (**k**) and CD8⁺ (**l**) T cells from *Tsc2-/-* 105K tumors treated as depicted in (I). 10 mice per treatment group. Means \pm SD, Mann-Whitney U Test, * *p* < 0.05. **m**, Experimental design for *Tsc2^{+/-}* mice treated with isotype control antibody (Isotype IgG) or anti-B7-H3 antibody (α -B7-H3) (Created with Biorender.com).

n, Representative kidneys of $Tsc2^{+/-}$ mice treated as depicted in **m**.

o, Representative H&E images of $Tsc2^{+/-}$ mice treated as depicted in **m**.

p, Gross tumor score of kidneys of $Tsc2^{+/-}$ mice treated as depicted in **m**. n = 14 kidneys per treatment group, each dot represents one kidney. Means ± SD, Mann-Whitney U Test, *** *p* < 0.001.

q, Microscopic tumor score of $Tsc2^{+/-}$ mice treated as depicted in **m**. n = 14 kidneys per treatment group, each dot represents one kidney. Means ± SD, Mann-Whitney U Test, **** p < 0.0001.

Source data and exact p values are provided in the Source data file.



Supplementary Fig. 4. B7-H3 deficiency in *Tsc2-/-* TTJ tumor cells suppresses subcutaneous tumor growth and lung tumor burden in models of metastasis.

a, Subcutaneous tumor growth of sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* TTJ tumors in WT C57BL/6J mice. n = 6 mice per group, means \pm SD, one-way ANOVA with Dunnett's multiple comparisons test, ** *p* < 0.01, *** *p* < 0.001.

b, Tumor-free survival curve of mice in **a**. Log-rank analysis.

c, Representative H&E images of lungs with sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* 105K tumors in WT C57BL/6J mice.

d, Percentage of lung area occupied by tumor per lung in (c). n = 9 mice for sh-NC and sh-B7-H3 (2), n = 10 mice for sh-B7-H3 (1), means ± SD, one-way ANOVA with Dunnett's multiple comparisons test, **** p < 0.0001.

e, Representative H&E images of lungs with sh-NC, sh-B7-H3 (1), or sh-B7-H3 (2) *Tsc2-/-* TTJ tumors in WT C57BL/6J mice.

f, Percentage of lung area occupied by tumor per lung in (E). n = 6-9 mice per group, means \pm SD, one-way ANOVA with Dunnett's multiple comparisons test, **** *p* < 0.0001.

g-i, Percentage of indicated cell types within CD45⁺ TILs from sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* TTJ tumors. n = 7 tumors for T cells, n = 10 tumors for CD4⁺ T and NK cells, n = 6 tumors for CD8⁺ T cells in sh-NC, n = 10 tumors for all cell types in sh-B7-H3 (1), and sh-B7-H3 (2), means ± SD, two-way ANOVA with Holm-Sidak's multiple comparisons test, ** p < 0.01, **** p < 0.0001.

j-l, Representative images of CD4 and CD8 immunohistochemical staining on sections from *Tsc2-/-* sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) TTJ subcutaneous tumors (j). Scale bar = 100 μ m. Quantification of CD8⁺ (k) or CD4⁺ (l) T cells in each group (n = 6/group). Means ± SD, one-way ANOVA with Dunnett's multiple comparisons test, **** *p* < 0.0001.

m, Representative images of F4/80, CD31, and pS6 (S235/236) immunohistochemical staining on tumor sections from *Tsc2-/-* 105K subcutaneous tumors transduced with sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) (n = 3). Scale bar = 100 μ m.

n, Representative images of E-cadherin and STAT1 immunofluorescent staining on subcutaneous TTJ tumor sections of the indicated groups (n = 3). Scale bar = 100 μ m. White arrows indicate cells with nuclear STAT1.

Source data and exact p values are provided in the Source data file.



Supplementary Fig. 5. The effects of B7-H3 suppression *in vivo* and *in vitro* relating to IFN-γ.

a, Top 10 activated gene signatures identified by gene set enrichment analysis in B7-H3 knockdown *Tsc2-/-* 105K subcutaneous tumors compared to controls (n = 3). ES = Enrichment Score, NES = Normalized Enrichment Score, NOM = Nominal, FDR = False Discovery Rate.

b, Immunoblot analysis of sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* 105K cells treated with IFN- γ for the indicated time points and concentrations (n = 3).

c, Immunoblot analysis of sgCtrl, sgCd276 (1), and sgCd276 (2) *Tsc2-/-* 105K cells treated with IFN- γ for the indicated time points and concentrations (n = 3).

d, Immunoblot analysis of cells in (**b**) co-cultured with CD3/CD28 activated splenic CD3⁺ T cells at the indicated tumor: T-cell ratio. CD3 T only was used as a control (n = 3).

e-g, Immunoblot analysis of *Tsc2-/-* 105K cells (**e**), *Tsc2-/-* TTJ cells (**f**), and *TSC2-/-* 621-101 cells (**g**) treated with IFN-γ for the indicated concentrations and time durations (n = 3).

h,i, Flow cytometry analysis of MHC-II expression on sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* 105K cells. Cells were treated with 10ng/ml of IFN- γ for 48 hr. (h). Mean fluorescent intensity (MFI) of MCH-II measured from each group of cells (n = 3) (i). Means ± SD, one-way ANOVA with Dunnett's multiple comparisons test.

j-I, Immunoblot analysis of *Tsc2-/-* 105K cells (j), *Tsc2-/-* TTJ cells (k), and *TSC2-/-* 621-101 cells (l) treated with IFN- γ for the indicated concentrations and time durations (n = 3).

m,**n** Left panels: Representative flow cytometry plots showing the percentage of IFN- γ /TNF- α -positive splenic CD8⁺ T cells (**m**) or CD4⁺ T cells (**n**) incubated with conditioned media isolated from cells in (**b**). Middle and right panels: Quantification of the left panel. Means ± SD, one-way ANOVA with Dunnett's multiple comparisons test (n = 3).

Source data are provided in the Source data file.

Supplementary Fig. 6



b







Tumor





COBRO

Supplementary Fig. 6. Anti-tumor effect of B7-H3 inhibition depends more on CD4⁺ T cells compared to CD8⁺ T cells.

a, Growth of sh-NC or sh-B7-H3 (2) *Tsc2-/-* 105K tumors treated with isotype control, α -CD4 or α -CD8 antibodies (n = 10 mice/group).

b, Tumor volume of (A) at 25-day post-cell injection. n = 10 tumors for sh-NC treated with isotype control, sh-NC treated with α -CD4, and sh-B7-H3 (2) treated with α -CD8, and n = 8 tumors for the remaining groups, means ± SD, two-way ANOVA with Holm-Sidak's multiple comparisons test, * *p* < 0.05, ** *p* < 0.01, *** *p* < 0.001, **** *p* < 0.0001.

c, Representative flow cytometry plots showing that splenic CD8⁺ and CD4⁺ T cells were successfully depleted. (n = 5/group).

d, Representative immunohistochemical staining of CD8 and CD4 on tissue sections of sh-NC, sh-B7-H3 (1), or sh-B7-H3 (2) *Tsc2-/-* 105K tumors treated with the indicated antibody. Scale bar = 100 μ m. (n = 5/group).

e, Growth of sh-B7-H3 (1) *Tsc2-/-* 105K tumors in WT, *Cd4* KO, or *Cd8* KO mice. n = 9 mice for WT, *Cd8* KO, n = 9 mice for *Cd4* KO, means \pm SD, nonparametric Kruskal-Wallis test with Dunn's multiple comparisons test, ** p < 0.01.

f, Tumor volume of **e** at 42-day post-cell injection. n = 9 mice for WT, *Cd8* KO, n = 9 mice for *Cd4* KO, means \pm SD, nonparametric Kruskal-Wallis test with Dunn's multiple comparisons test, ** p < 0.01. **g**, Growth of sh-NC *Tsc2-/-* 105K tumors in WT, *Cd4* KO, or *Cd8* KO mice. n = 10 mice/group, means \pm SD, nonparametric Kruskal-Wallis test with Dunn's multiple comparisons test, * p < 0.05, *** p < 0.001. **h**, Tumor volume of **g** at 28-day post-cell injection. n = 10 mice/group, means \pm SD, nonparametric Kruskal-Wallis test with Dunn's test, * p < 0.05, *** p < 0.001. Source data and exact p values are provided in the Source data file.



Supplementary Fig. 7. Inhibition of B7-H3 increases the number of intratumoral CD38⁺CD39⁺CD4⁺ TILs and analysis of CD4⁺ T cells from advanced metastatic RCC patients.

a, UMAP plots showing co-expression of CD38 and CD39 proteins in CD4_Teff-Gzmk cells.

b, Spearman correlation analysis of CD38 protein expression versus other proteins detected by CITEseq in CD4_Teff-Gzmk cells.

c, Representative flow cytometry plots (left panel) and quantification of the percentage (right panel) of CD38 and CD39 on CD4⁺CD25⁻ TILs in sh-NC, sh-B7-H3 (1), and sh-B7-H3 (2) *Tsc2-/-* 105K tumors. n = 8 mice/group, mean \pm SD, two-way ANOVA with Holm-Sidak's multiple comparisons test, **** p < 0.0001.

d, Representative immunofluoresent images (left panel) and quantification (right panel) of CD38⁺CD39⁺CD4⁺ TILs in lungs of WT mice with sh-NC, sh-B7-H3 (1), or sh-B7-H3 (2) *Tsc2-/-* 105K tumors. n = 6 mice/group, mean ± SD, nonparametric Kruskal-Wallis test with Dunn's multiple comparisons test, * p < 0.05, ** p < 0.01. White arrows showing CD38⁺CD39⁺CD4⁺ TILs.

e, UMAP plot of CD4⁺ T cells combined from 8 metastatic RCC patients. 3 from kidneys, 3 from lymph nodes, 2 from visceral metastases.

f, Dotplot showing expression of selected genes from CD4⁺ T cells in e.

g,h, Proportions of each CD4⁺ T-cell clusters. ICB = Immune checkpoint blockade, NoICB = No Immune checkpoint blockade. ICB Response: PR = partial response, SD = stable disease, PD = progressive disease, and NE = not evaluable.

i, UMAP plots displaying expression of *CD38* and *ENTPD1* (CD39 gene name) in tumors according to their ICB exposure.

j, UMAP plots displaying expression of *CD38* and *ENTPD1* in tumors according to their ICB response. Source data and exact p values are provided in the Source data file.



Supplementary Fig 8. Gating strategy for *in vitro* CD3/CD28 activated T-cell IFN-γ and TNF-α analysis.



Supplementary Fig 9. Gating strategy for *ex vivo* tumor-infiltrating T-cell IFN- γ and TNF- α analysis.

Supplementary Table 1. List of antibodies used for CyTOF staining

Target	Clone	Supplier	Dilution
CD8a	53-6.7	Lederer Lab CyTOF Core	1:100
CD4	RM4-5	Lederer Lab CyTOF Core	1:100
CD11c	N418	Lederer Lab CyTOF Core	1:100
TCRβ	H57-597	Lederer Lab CyTOF Core	1:100
CD3	145-2C11	Lederer Lab CyTOF Core	1:100
CD19	6D5	Lederer Lab CyTOF Core	1:100
NK1.1	PK136	Lederer Lab CyTOF Core	1:100
CD45	30-F11	Lederer Lab CyTOF Core	1:100
CD11b	M1/70	Lederer Lab CyTOF Core	1:100
Singlec-F	E50-2440	Lederer Lab CyTOF Core	1:100
Ly6G	1A8	Lederer Lab CyTOF Core	1:100
Ly6C	HK1.4	Lederer Lab CyTOF Core	1:100

Supplementary Table 2. List of antibodies used for flow cytometry staining

Target	Clone	Supplier	Dilution
CD45	30-F11	BioLegend	1:100
IFNγ	XMG1.2	BioLegend	1:50
	MP6-		1:100
TNF-α	XT22	BioLegend	
CD4	RM4-5	BioLegend	1:100
CD8	53-6.7	BioLegend	1:100
CD3	17A2	BioLegend	1:100
CD38	90	BioLegend	1:100
CD39	Y23-1185	BD Biosciences	1:100

Supplementary Table 3. List of primers used for ChIP-qPCR

Primer	FW	RV
Cd276 promoter (mouse)	GAGTCCCTTCTTTCCTTGAGTC	CCCTTCACCTGTGTTTGTATCT

Supplementary Table 4. List of siRNAs used for transfection

siRNA	Cat #	Company
SMARTpools mTOR siRNA	L-065427-00-0005	Dharmacon
SMARTpools Raptor siRNA	L-058754-01-0005	Dharmacon
SMARTpools Rictor siRNA	L-064598-01-0005	Dharmacon
SMARTpools S6K siRNA	L-040893-00-0005	Dharmacon
SMARTpools 4E-BP1 siRNA	L-058681-01-0005	Dharmacon
SMARTpools YY2 siRNA	L-171481-00-0005	Dharmacon
SMARTpools control siRNA	D-001810-10-05	Dharmacon

Supplementary Table 5. List of probes used for qRT-PCR

Probe	Cat #	Company
Cd276 (mouse)	Mm00506020_m1	ThermoFisher Scientific
CD276 (human)	Hs00987207_m1	ThermoFisher Scientific
Yy2 (mouse)	Mm03059489_sH	ThermoFisher Scientific
YY2 (human)	Hs02597954_sH	ThermoFisher Scientific
Yy1 (mouse)	Mm00456392_m1	ThermoFisher Scientific
YY1 (human)	Hs00998747_m1	ThermoFisher Scientific