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Supplemental information

Platelet-instructed SPP1⁺ macrophages drive

myofibroblast activation in fibrosis in a

CXCL4-dependent manner

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Supplemental Figure 1: Single cell RNA analysis of a time-series of myocardial infarction. Related to Fig. 1.

A) UMAP embedding of all (52497) interstitial cells from murine heart tissue at different timepoints after MI from Forte et al., 2020. Cd45⁺ immune cells (circled in red) were subsetted for further analysis. Labels refer to clusters. Res-like Mac: resident-like macrophages, *Spp1* Mac: *Spp1*⁺ macrophages, Ly6c2hi Mono: Ly6c2hi monocytes, Prolif. Mac: Proliferating macrophages, T- & NK-cells: T-cells and natural killer cells, EC: Endothelial cells, SMC: Smooth muscle cells, Progen. Fibro: Progenitor-like fibroblasts, Ifn Fibro: Interferon-Induced fibroblasts, Prolif. Fibro: Proliferating fibroblasts, EndD. Fibro: Endocardial-derived fibroblasts.
B) Dotplot of the top 3 specific genes for interstitial cell clusters shown in Fig. S1A. C) Dotplot of the top 3 specific genes for immune cell subclusters shown in Fig. 1A E) Featureplot of the core matrisome score on the UMAP embedding shown in Fig. 1A. F) Core matrisome score stratified by immune cell type.



Symphony transferred Cluster Labels obtained from monocyte-derived Macrophages

Supplemental Figure 2: Characterisation of cardiac *Spp1*⁺ macrophages. Related to Fig. 1.

A) PROGENy pathway analysis of cardiac immune cell clusters shown in Fig. 1A. **B)** DoRothEA transcription factor analysis of cardiac immune cell clusters shown in Fig. 1A. **C)** *Trem2* expression stratified by immune cell clusters of 1A). **D)** Pseudotime Trajectory for infiltrating MPC on the PHATE embedding shown in Fig. 1D. **E)** Heatmap of the top genes correlating with ECM regulator scores in immune cells from Fig. 1A, *Cxcl4* is ranked #7. **F)** Symphony Reference Map of monocyte-derived macrophage activation states from Sanin et al. with Query Mapping of cardiac macrophages after myocardial infarction (MI) from Forte et al. into the same low dimensional space. **G)** Symphony Transfer of Cluster Labels of monocyte-derived activation states to cardiac macrophages. For C p-values from MAST (Seurat, FindAllMarkers) are displayed. ****P<0.0001.



Supplemental Figure 3: Quality Control of single nuclear RNA sequencing of WT and *Cxcl4^{-/-}* mice after IRI or sham surgery. Related to Fig. 3. A) Representative flow cytometric plots for isolation of DAPI⁺ nuclei. B) UMI Count, gene count, mitochondrial gene (mt.gene) fraction as well as scrublet doublet score for individual snRNA-seq libraries. C) UMAP embedding of 66235 nuclei split by genotype and surgery. Labels refer to clusters. Podo: podocytes, PT: proximal tubule, DTL: descending thin limb, TAL: thick ascending limb, DCT: distal convoluted tubule, IC: intercalated cells, PC: principal cells, DME: deep medullary epithelium, Injured Tub: injured tubular cells, Endo: endothelial cells, Fibro: fibroblasts, VSMC: vascular smooth muscle cells, Peri: pericytes. D) Dotplot of the top 3 specific genes for cell clusters shown in Fig. 3A. E) Symphony Reference Map of murine kidneys after IRI from Kirita et al. with Query Mapping of kidneys from WT or *Cxcl4^{-/-}* mice after sham or IRI surgery into the same low dimensional space. F) Symphony Transfer of Cluster Labels from Kirita et al.



Supplemental Figure 4: Compositional validation and marker gene analysis of single nuclear RNA sequencing data of WT and $Cxcl4^{-/-}$ mice after IRI or sham surgery. Related to Fig. 3. A) Representative images of immunofluorescence-stained sections for Lotus tetragonolobus lectin (LTL) and Kidney Injury Molecule -1 (KIM1), PDGFRA and Uromodulin (UMOD) or CD68 from WT or *Cxcl4^{-/-}* kidneys after Sham or IRI surgery. Scale bar = 50µm. B) Quantification of KIM1⁺ injured tubular cells based on the staining in A. C) Quantification of LTL⁺ proximal tubular cells based on the staining in A. D) Quantification of PDGFRA⁺ fibroblasts based on the staining in A. E) Quantification of Uromodulin⁺ thick ascending loop cells based on the staining in A. F) Quantification of CD68⁺ macrophages based on the staining in A. G) Fn1, Spp1, and C1qa expression stratified by immune cell cluster for sub-clustered renal snRNA leukocytes as well as sub-clustered cardiac immune cells from Forte et al. H) snRNA-seq Arg1 expression stratified by clusters. I) Irf2, Ccr2, NIrp1b, and Trem2 expression in renal snRNA leukocytes stratified by immune cell clusters. For B-F a twoway ANOVA was computed using Tukey corrections. For G p-values from MAST (Seurat, FindAllMarkers) are displayed. *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001. For B-F n = 8 WT Sham ; 5 *Cxcl4^{-/-}* Sham; 8 WT IRI; 5 *Cxcl4^{-/-}* IRI.



Supplemental Figure 5: Cxcl4 is critical for platelet-monocyte interaction and mediates profibrotic fibroblast activation. Related to Figs. 4 and 5. A) Experimental design of murine PBMC co-culture with WT or Cxcl4^{-/-} platelets with subsequent FACS isolation of CD11b⁺ monocytes for Bulk RNA sequencing. B) Representative flow cytometric plots for quantification of APC⁺CD11b⁺ monocytes in WT PBMC co-cultured with CMFDA⁺ WT or Cxcl4^{-/-} platelets (Plt) after stimulation with Vehicle or LPS and Thrombin. C) Confocal imaging of CMPTX⁺ WT PBMC after 48 hours co-culture with CMFDA⁺ WT or Cxcl4^{-/-} platelets. Scale Bar=10 µm. **D**) Automated quantification of CMFDA⁺CMPTX⁺ plate-adherent platelet-monocytes aggregates compared to total plate-adherent monocytes from (B) [n = 4]. E) Counts per Million (CPM) for individual samples of FACS-isolated CD11b⁺ monocytes co-cultured with either WT or Cxcl4⁻⁻ /- platelets. F) Generalised (glmpca) Principal Component Analysis (PCA) of FACS-isolated CD11b⁺ monocytes co-cultured with either WT or $Cxcl4^{-/-}$ platelets. **G)** PROGENy analysis of differentially expressed genes in CD11b⁺ monocytes co-cultured with either WT or $Cxcl4^{-/-}$ platelets. H) Schematic for isolation and immortalization of cardiac Gli1⁺ fibroblasts. I) Representative flow cytometric plots for FACS of CMPTX⁺Gli1⁺ fibroblasts (CMPTX⁺Gli1⁺ Fibro). J) Gating strategy and representative flow cytometric plots for analysis of CXCL4 expression in peripheral blood as shown in Fig 5B. K) RT-qPCR analysis for Col1a1 and Fn1 in IRI and sham kidneys of ^{HSC}WT and ^{HSC}Cxcl4^{-/-} mice. All quantitative data are shown as mean±SD. For D a two-tailed unpaired t-test was performed. For K a two-way ANOVA was computed using Tukey correction for multiple testing. *P<0.05, **P<0.01, ***P<0.001.



Supplemental Figure 6: Ligand Receptor Interaction analysis of snRNA WT and Cxcl4^{-/}

- kidneys after IRI. Related to Fig. 6. A) Network plots of inferred number of ligand receptor interactions split by genotype. Labels refer to clusters. DCT: distal convoluted tubule, Endo: endothelial cells, Fibro: fibroblasts, IC: intercalated cells, Tub: tubular cells, Mac: macrophages, PC: principal cells, Podo: podocytes, PT: proximal tubular cells, TAL: thick ascending limb, VSMC: vascular smooth muscle cells, Peri: pericytes. B) Inferred macrophage (Mac) Ligand-Receptor (LR) interaction weight stratified by cell clusters and genotype. Labels refer to clusters. C) Dot-Plot for all inferred ligand receptor interactions of macrophages and fibroblasts stratified by genotype and cell-cell interaction. D) Network plots for inferred *Spp1* ligand receptor interaction activity split by genotype. F) Dotplot of the top 3 specific genes for fibroblast sub-clusters shown in Fig. 5F. G) *Dapk2* Expression stratified by fibroblast sub-clusters. H) Fibroblast sub-cluster composition in percent stratified by genotype and surgery. For G p-values from MAST (Seurat, FindAllMarkers) are displayed. ****P<0.0001.</p>



Supplemental Figure 7: Single cell RNA analysis of MPC in human CKD and heart failure. Related to Fig. 7. A) UMAP of all (51849) CD10 negative cells from Kuppe et al.. Mononuclear phagocytes (MPC, circled in red) were sub-clustered for further analysis. Labels refer to clusters. Art Endo: arterial endothelium. CD - PC: Collecting Duct - Principal cells, CNT: connecting tubule, DC: dendritic cells, DTL: descending thin limb, DCT: distal convoluted tubule, Fibro: fibroblast, Glom Capill: glomerular capillaries, Inj Endo: injured endothelium, Inj Tub: injured tubule, IC: intercalated cells, Lymp Endo: lymphatic endothelium, Mac: macrophages, Mono: monocytes, Myofibro: myofibroblasts, Podo: podocytes, PT: proximal tubule, SMC: smooth muscle cells, TAL: thick ascending limb, Uro Cells: urothelial cells, Ven Endo: venular endothelium. B) Dotplot of the top 3 specific genes for MPC cell clusters shown in Fig. 6A. Mono: monocytes, Mac: macrophages, cDC: conventional dendritic cells, Res-like Mac: Resident-like macrophages. C) APOE and SPP1 expression in kidney MPC stratified by cell type. D) RNA-ISH for SPP1 and COL1A1 with CD68 immunofluorescent staining in human kidney nephrectomies (n=41). Representative images of kidneys with low and high fibrosis as determined by COL1A1 expression are shown. SPP1+CD68+ macrophages are circled in white, while SPP1⁻CD68⁺ macrophages are circled in orange. Scale bar = 30 µm. E) UMAP of all (58786) immune cells from Rao et al.. MPC (circled in red) were sub-clustered for further analysis. Labels refer to clusters. Mac: macrophages, DC: dendritic cells, Mono: monocytes, Prolif Cells: proliferating cells. F) Dotplot of the top 3 specific genes for immune cell clusters shown in E. G) Dotplot of the top 3 specific genes for MPC clusters shown in Fig. 6E. H) FN1 and SPP1 expression in cardiac MPC stratified by cell type. (I) TREM2 and CD9 expression in human kidney MPC stratified by cell type. (J) TREM2 and CD9 expression in human cardiac MPC stratified by cell type. K) Symphony Reference Map of renal immune cells in human chronic kidney disease (CKD) with Query Mapping of cardiac immune cells in human heart failure (HF) from Rao et al. into the same low dimensional space. L) Symphony Transfer of Cluster Labels. For C, H, I and J p-values from MAST (Seurat, FindAllMarkers) are displayed. ****P<0.0001.