1	Interpreting random forest analysis of ecological models to move from
2	nrediction to explanation
2	prediction to explanation
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10	Supplementary Information
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12	S1 - Model development S2 - Analyzing Simulation Desults w/ Dandom Forests
13	S2 - Anaryzing Simulation Results w/ Random Porests
14	55 - Supplementary Analysis Results
10	S1 Model development
10	SI - Model development In this section we use the online database COMPADRE to investigate model structure, plausible
18	narameter values and simulation design R scripts to run our analysis are presented in SI github. The
19	analysis presented below was completed in the later part of 2021 and COMPADRE is a growing database.
20	Therefore, attempts to recreate our analysis at a later date are likely to reflect changes made to the
21	database.
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23	S1.1 - Model structure
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25	Three distinct stages were used as the ontogenetic structure of the plant population in our model:
26	fecund adults (F), non-fecund seedlings (S_2), and the seed bank (S_1) (Fig 1). While no single ontogenetic
21	structure can perfectly represent all plant species, three stages is a suitable baseline for the purposes of this study. First, a three store structure enotes a treatable dimensionality in our model which allows for
20 20	an in depth analysis of interdependent dynamic effects horn out of interacting demographic/ecological
30	rates Importantly this allows us to more clearly graphically represent the consequences and details of
31	interacting rates which would be less apparent in higher dimensional ontogenetic formulations. Second.
32	using the growing global database of stage-structured plant demographic data, COMPADRE ⁴² , we can see
33	that the three stage ontogeny is well-represented in plant taxa empirically. Of all the documented plant
34	taxa within the COMPADRE database, three-stage plant structures represent roughly 34% of families,
35	18% of genera, and 14% of species. This includes abundant, species-rich, economically important, and
36	geographically wide-spread plant families such as Asteraceae, Brassicaceae, Orchidaceae, Rosaceae, etc.
37	The plant population in the model experiences density dependent restrictions on the production of
38	seeds via the maximum function which restricts seed production to a minimum of zero in relation to the
39	density of F. Stage transitions are also modified by density dependent pressure with limited density
40 11	dependent effects from younger via the parameter ϵ (see Eq1 & Table 1). Finally, consumer pressure
41 12	from the heroivore population (<i>H</i>) can differentially focus its heroivory on either the security stage $(a_2 > 0, a_1 = 0)$ the focus of a dult stage $(a_2 = 0, a_1 > 0)$ or both $(a_1 > 0, a_2 > 0)$. Herbivory occurs under a
43	Type II functional response on each stage, where consumption on each stage is affected by the handling
44	time required to consume both stages. All stages experience a background mortality rate. Seeds (S.)
45	experience a low level amplification of background mortality linked to adult (F) density under the
46	assumption that sufficiently high populations of conspecific mature plants reduce resources for seeds
47	(through lack of nutrients, shading, etc.) ⁴³ or increase frequencies of exploitative interactions not
	- ^

explicitly unaccounted for in the model (e.g. soil pathogens)^{44,45}. Parameters mediating all density 48

49 dependent effects and trophic interactions are listed and described in Table 1.

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$$\frac{dF}{dt} = \underbrace{\frac{g_{2F}S_2}{1 + \alpha_{g2}(F + \epsilon S_2)}}_{Y_{12}} - \underbrace{\frac{\theta_F}{consumption}}_{Y_{2F}} - \underbrace{\frac{\theta_F}{a_FFH}}_{Y_{2F}} - \underbrace{\frac{\theta_F}{a_FFH}}_{Q_2F} - \underbrace{\frac{\theta_F}{a_FF}}_{\theta_2}$$

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$$\frac{dS_2}{dt} = \underbrace{\frac{g_{12}S_1}{1 + \alpha_{g1}(F + \epsilon(S_1 + S_2))}}_{\delta} - \underbrace{\frac{g_{2F}S_2}{1 + \alpha_{g2}(F + \epsilon S_2)}}_{\gamma_{12}} - \underbrace{\frac{g_{2F}S_2}{1 + \alpha_{g2}(F + \epsilon S_2)}}_{\gamma_{12}} - \underbrace{\frac{g_{2F}S_2}{1 + \alpha_{F}h_FF + \alpha_2h_2S_2}}_{\rho_{12}} - \underbrace{\frac{g_{2F}S_2}{1 + \alpha_{F}h_FF + \alpha_{F}h_FF + \alpha_{F}h_FF + \alpha_{F}h_FF + \alpha_{F}h_FF + \alpha_{F}h_FF + \alpha_$$

$$\frac{dS_1}{dt} = \underbrace{\operatorname{Max}(F(r_F - \alpha_F F), 0)}_{0} - \underbrace{\frac{g_{12}S_1}{1 + \alpha_{g1}(F + \epsilon(S_1 + S_2))}}_{0} - \underbrace{\frac{g_{12}S_1}{S_1(d_S + \alpha_{FS} F)}}_{0}$$

$$\frac{dH}{dt} = c_{FH} \frac{a_FFH}{1 + a_Fh_F + a_Fh_S} + c_{2H} \frac{a_2S_2H}{1 + a_Fh_F + a_Fh_S} - \frac{background}{d_HH}$$

$$\frac{1}{dt} = c_{FH} \frac{1}{1 + a_F h_F F + a_2 h_2 S_2} + c_{2H} \frac{1}{1 + a_F h_F F + a_2 h_2 S_2} - c_{2H} \frac{1}{1 + a_F h_F F + a_F h_F F$$

55 S1.2 - Model Parameters

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57 The COMPADRE database also allows for further insight into our model. We can use the 58 available data to inform certain model parameters. The COMPADRE reproductive and stage transition 59 rates mirror our demographic parameters. Production of stage 1 individuals from stage 3 corresponds to 60 seed production (r_F) . Transition from stage 1 to 2 corresponds to seed germination (g_{12}) and transition from stage 2 to 3 corresponds to seedling maturation (g_{2F}) . However, these rates represent data from a 61 62 range of different studies using a range of experimental, natural, and agricultural plant demographic data. 63 Additionally, these studies involve transition rates which emerge from both competition and trophic 64 interactions. Therefore, measurable field rates are frequently emergent from environmental conditions and 65 are not necessarily the inherent demographic rates. As such, we do not intend to use these empirical rates 66 to determine exact values of our model's demographic parameters. Instead, we use the ranges of each 67 empirical rate to inform plausible ranges for their corresponding model parameter to be analyzed in 68 parameter sweeps of model simulations.

69 In doing so, we limited our survey of values from each parameter to those greater than 0 as the 70 effect of setting any demographic parameter to 0 in the model is obviously a steady fall to extinction (Fig 71 S1). Starting with rates of reproduction (Fig S1a), we see a large range of reproduction into the first stage 72 from the third, from 0 to roughly 100. However, we limited our preliminary analysis to $r_F < 10$ as 87% 73 of sampled values fall within this range (Fig S1a, Insert). The rates of transition between stages are much 74 more evenly distributed from values >0 to 1. While not a completely even distribution across the range, we take this as sufficient reason to study the full range of values, $0.1 < g_{12} < 0.9$ and $0.1 < g_{2F} < 0.9$. 75 76 With these results in mind (Fig S1), ranges for these three parameters are provided in Table 1 along with

- 77 both parameter definitions and the ranges/values used for every model parameter.
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Fig S1: Distribution of empirically measured reproduction and stage transition rates from three-stage plant matrix models in the COMPADRE database (2021). a) Reproduction into 1st stage by 3rd stage. b) Transition rates from stage 1 to 2. Transition rates from stage 2 to 3.

S1.3 - Simulation Design & Output

86 Potentially the most important insight garnered from available data in COMPADRE, we see 87 absolutely no correlation between any of the measured rates. There is no discernable relationship between 88 empirical rates of reproduction and maturation from either stage 1 to 2 or 2 to 3 (visualized in Fig S2a & 89 b). Also, we see no correlation between the stage transition rates (visualized in Fig S2c). The lack of any 90 relationships in the values of any rate prompted a full factorial investigation of the model's demographic 91 rates (r_F, g_{12}, g_{2F}) ; see Table 1 for value range), without any necessary covariation in parameter values. 92 In other words, all demographic parameter values were factorially tested against each other in a large 93 parameter sweep without the need to change any one parameter value in concert with another. Past work 94 also indicates interactivity between demographic rates and trophic interactions in driving model 95 dynamics¹⁹, so we considered trophic interactions in our simulation design by including rates of herbivory $(a_F \& a_2)$ into what becomes a five dimensional fully factorial parameter sweep. Herbivore attack rate 96 97 ranges were chosen heuristically (Table 1). Herbivore attack rates on either consumed plant stage (see Fig 98 1d-1i) vary factorially in the parameter sweep as herbivores can range from focusing their attack on either 99 stage to splitting their consumption between stages to varying degrees.

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As a mode of sensitivity testing, we implemented the five parameter factorial sweep across
 multiple values for both density dependent parameters and handling times. Density dependent parameters

109 were varied because density dependence mediates stage transitions and therefore ontogenetic dynamics¹⁹.

110 Handling time was chosen for its role in mediating the trophic connections interacting with plant

ontogeny. This created eight unique instances of the full five parameter factorial sweeps (see values in
 Table 1), producing over 5.5 million unique simulations for analysis.

Each simulation outputs a number of initial conditions and post simulation factors detailing the results of the simulation. Initial conditions include all relevant parameter values and initial time

dependent variable densities (i.e., initial population densities such as F(t) at time 0). Post simulation

116 factors include equilibria values, equilibria linear stability, eigenvalues, periodicity, oscillating

117 populations' peak and trough densities, and finally the effective handling time of the herbivore

118 population. The effective handling time of the herbivore population is measured as the denominator of the

- 119 consumptive interaction in the model.
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121 S2 - Analyzing Simulation Results w/ Random Forests 122

Due to the large simulation dataset, we first produced an initial guided analysis using the Random 123 Forest based machine learning algorithm which can achieve high predictive power⁴⁶ and has shown 124 125 success in using permutation techniques to determine how much specific predictors contribute to that predictive ability (e.g., mean accuracy decrease⁴⁷). In our case, our five main parameters (see Table 1) 126 127 serve as our main random forest features/predictors. Random Forests can predict either categorical or 128 continuous variables and any post simulation factor can function as the predicted variable in the random 129 forest analysis. Our interest in trophic/demographic dynamics led us to use the simulation model's linear 130 stability as our predicted variables. We used a simple indicator, stable or unstable, for categorization 131 random forest tasks and model equilibria's eigenvalues for regression random forest tasks.

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133 S2.1 - Preparing data and training Random Forest134

135 Simulation data was split into "training" and "validation" (sometimes called test) data subsets for 136 hold out cross validation. We first created or "trained" the random forest on the training data. In Random 137 Forests, this process produces a series of unique categorization and regression trees that "vote" on the 138 outcome (our simulation model's stability) based on the values of any particular inputs (our model 139 parameters). As a default during training, random forest parameter "mtry" was set at floor(sqrt(p)) for 140 categorization tasks (stable vs unstable) and floor(p/3) for regression tasks (max eigenvalue) where p=#141 of features⁴⁸. Instances where a different p produced better results are noted in the text. The parameter 142 "intrees" (No. of trees) was varied from 300-600 with little to no effect on performance.

Once we have a trained random forest, we check its performance on the training data via the "out of the box" (OOTB) error rate. Given a sufficiently low error rate, we can begin to investigate feature importance. We measured the importance of individual features/parameters in our random forest with Mean Accuracy Decrease, which measures the loss in predictive accuracy by excluding each feature. The more the accuracy suffers, the more important the variable is for the successful classification/prediction. For a more detailed description of the mechanisms behind creating and training random forests, please see ref 14.

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151 S2.2 - Validating/Testing Trained Random Forest

Sufficiently high performing trained random forests were then used to predict simulation output in our validation data subsets which our random forests had not yet been exposed to. We analyzed the predictive power of categorization tasks by comparing their predicted output with data via the Area Under Curve the Receiver Operating Characteristic curve (AUC) metric (pROC package). In our case, the AUC metric measures how well the models are able to distinguish between stable and unstable results in the validation data subset. It varies between 0 and 1 with 1 indicating better predictions. Regression tasks were judged for accuracy using RMSE on maximum eigenvalue measurements between the predicted eigenvalue output and the simulation data. All AUC and RMSE values are from validation data unlessotherwise specified.

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163 S2.3 - Interpreting Feature Effects

165 Random Forest results can also be further interpreted with the H statistic which functions as a 166 measure of interactivity between features/predictors in driving prediction results⁴⁹. This can be done for each individual predictor as a measure of general interactivity with other predictors or be done with a 167 168 focus on particular predictors to study direct interactivity between specific predictor combinations. 169 Regardless, higher numbers indicate higher interaction strengths while lower numbers indicate less 170 interaction between predictors. While some have argued that the basic random forest measurements of 171 variable importance (e.g., mean accuracy decrease) "capture" the outcome of interactions between features in predictions, these metrics are not designed to detect interactions per se⁵⁰. Therefore, our use of 172 the H statistic helps us hone in on key interactions, a particularly useful outcome for any researcher 173 174 looking to implement our methods on a model with a larger parameter set.

175 While the H statistic provides inference regarding which features have strong interactive effects, 176 other methods are required to determine how features (interacting or otherwise) actually affect the predicted outcomes. To do this we used two analytical techniques in the iml package in \mathbb{R}^{51} . First, we used 177 Partial Dependence plots (PD plots) to visualize the marginal effect of one or two features on the 178 predicted outcome of our random forests⁵². These PD plots can show whether the relationship between the 179 180 target and a feature is linear, monotonic or more complex. By focusing on two-feature partial dependence 181 plots, we can also see how these features interact in changing model predictions (e.g., Fig 2). Second, we 182 also used Individual Conditional Expectation (ICE) curves to uncover heterogeneous relationships by 183 showing individual instances of changing a feature's value at different permutations of the other 184 features⁵³. In using these ICE plots, we found quick evidence of the context dependent relationship of 185 features and their effect on model predictions (Fig S3), again helping us determine where interactions 186 matter.



- 191 b) Depiction of the high degree of heterogeneity in feature effects on random forest predictions 192 using the g2F parameter as an example. 193 The yellow line represents the average partial dependence effect (PD plot). The black lines show 194 an Individual Conditional Expectation (ICE) plot; instances of changing g2 in the context of differing 195 subsets of other parameters. Both the quantitative and qualitative range of differences seen in the ICE plot 196 indicate the fidelity lost in only examining the average effects and the need for a more fine scale look at 197 the heterogeneity in effect per parameter. Note, the right ICE plot only shows results for $a_2, a_F \le 1$ in 198 an effort to reduce the number of plotted lines for the sake of visibility. 199
- 200 S2.4 – Additional 201

202 Our random forests can produce a highly accurate level of predictive power. These levels of 203 predictive ability can induce questions of data leakage in producing and testing our models. We claim this 204 is not the case here and detail our reasons below. First, the two easiest sources of data leakage are 205 including the target variable as a feature in creating and training our models while the other is accidental 206 inclusion of test/validation data in our training data during model training. Neither of these occurred due 207 to simple due diligence in model creation. Our random forest code can be used to verify these claims. 208 Second, we have no "give away" features which are effectively tied to our target variables. Third, results 209 are not driven by particular outliers and are consistent across different subsets used as training and 210 test/validation datasets. Finally, models' variable importance changes in explanatory ways across 211 different subsets of data (e.g., different consumption allocations) and our results are supported by our 212 graphical analysis (Fig 2) which does not fall victim to data leakage issues. Therefore, we can have high 213 confidence that our random forest results do not reflect data leakage.

Despite the high levels of predictive power (e.g., results shown in Fig. 1), our random forests did 214 215 have limits on their immediate interpretability as noted by others (e.g. ref 54). Therefore, we use our 216 random forest models not as end points on their own, but as tools to direct our analysis across such a large 217 amount of simulation data. By determining feature importance, effect, and interactivity, we were able to 218 hone in specific subsections of the simulation model's parameter space and utilize graphical analysis to 219 expand our ecological understanding of our results.

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Figure S4: Maximum eigenvalue of the plant-herbivore system where the herbivore only eats the adult plant stage $(a_F > 0, a_2 = 0)$ across $\{g_{12}, g_{2F}, r_F\}$ parameter space. Maximum eigenvalue here dictates the 226

dynamic stability of the population trajectories of both species in the interaction. Positive values indicate
 instability and persistent oscillations while negative values indicate damped oscillations to stable
 population trajectories.



231 Figure S5: Box and whisker plots detailing range of H-statistic (y-axis) for each pair-wise 232 interaction of demographic rates (x-axis) in random forests run with set attack rates where a_2 and 233 a_F vary between 0.2 and 2.0 ($\alpha_{g1}=\alpha_{g2}=\alpha_F=0.1$; $h_2=h_F=0.5$). The H-statistic measures 234 interactivity between our demographic rates and our results here show g_{12} and g_{2F} to be the 235 most consistently interactive. Letters over boxes indicate significantly different groupings based 236 on the Tukey post hoc test. Boxes represent the interquartile range with the horizontal line 237 238 showing the median, the lower box showing the 25 percentile, and the upper box showing the 75 percentile. Upper and lower lines extending from the boxes show the most extreme values within 239 240 1.5 times the 75th and 25th percentile respectively. Outliers are shown as single dots. 241

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Seed Prodct. Rate,r_F
 Figure S6: Seed production effect on ecological factors with adult-only herbivores. The mean effect of
 raising seed production broken down to average constituent effects on ecological factors across all other
 simulation model parameters. Dots represent mean and error bars show standard deviation.



Figure S7: Changes in F^* ratio in relation to ecological factors γ_{12}^* and γ_{2F}^* . The y axis, F^* Ratio is the percent makeup of the adult plant stage defined as F^* Ratio = $\frac{F^*}{S_1^* + S_2^* + F^*}$. Color contrast shows constituent changes in a) simulation model parameters $g_{12} + g_{2F}$, b) simulation model parameter r_F , c) stability of simulation model.





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272 Figure S10: Drivers of stabilization in $a_2 = 0.2 \& a_F = 1.0$ herbivory allocation when g_{2F} is high and 273 g_{12} is low. a) As g_{12} decreases and g_{2F} increases (measured by $g_{2F} - g_{12}$) this consequentially increases seedling maturation while limiting seed germination ($\gamma_{2F} - \gamma_{12}$), reducing the plant density held in the 274 seedling stages. b) As the plant density in the seedling stage decreases, so does the percent consumption of seedlings, measured here by a decrease in $\frac{\theta_2}{\theta_F + \theta_2}$. c) Reducing the percent of seedlings in the diet of the 275 276 277 herbivore makes the trophic interaction act more like single stage consumption. This increases the 278 probability of dampened oscillations to stability as shown via a generalized linear model (p<<2e-16, 279 beta=-8.003, Residual deviance: 3815.3 on 5598 degrees of freedom). 280



Figure S11: Heatmap colors represent ecological effects on stability via coefficients from partial least squares regression of ecological factors versus maximum eigenvalue across all specific combinations of herbivory on the adult and seedling stages when $h_2 = 0.5$, $h_F = 1$ and α_F , α_{g1} , $\alpha_{g2} = 0.6$. The ecological factors are a) L:D Ratio, b) γ_{12} , & c) γ_{2F} . Note, the gray square when both attack rates are set to 0 indicates no data given the lack of consumption. The figure depicts how a lower handling time for herbivory on seedlings versus adults can change the dynamic effect of γ_{12} such that increased germination can dampen oscillations more consistently across different herbivory allocations.



Figure S12: Generalized additive model derived representation of simulation results from $(a_2=0, a_F=1)$ using a 3-way tensor product smooth on g_1 , g_2 , and r_F . a) Results from categorization task (stable as 1 and unstable as 0). Producing this GAM took substantially more time to calculate than the random forest, but it does recreate our results. b) Results from regression task (maximum eigenvalue). "Hotter" colors indicate "lower" values. In the categorization task, this indicates higher probabilities of as unstable equilibrium (oscillations). In the regression task, conversely this indicates smaller eigenvalues. Overall, we can see how the GAMs can recreate the patterns found in our random forest analysis once we have developed the necessary parametric hypothesis from our random forest results.

S3.2 - Description of g_{12} & g_{2F} *vs* γ_{12} & γ_{2F}

303 Although the g_{12} & g_{2F} parameters and γ_{12} & γ_{2F} ecological sub-functions we labeled "factors" 304 (Table 1) are related, their relationship is not 1:1. Specifically, g_{12} and g_{2F} are parameters in the model 305 representing the per-capita germination/maturation rates of seeds/seedlings. The fixed values of these parameters are assigned for each simulation and are not dynamic. These parameters represent the rate ofmaturation without density dependent effects.

The γ_{12} and γ_{2F} sub-functions/components in the model represent the density of seeds/seedlings that germinate/mature to the seedling/adult stages over a timestep. The values of these sub-functions are an emergent property of the model resulting from the interaction between herbivore consumption and internal plant demography. For example, the parameter r_F affects the output of γ_{12} and γ_{2F} . Unlike g_{12} and g_{2F} , (which are fixed parameters), γ_{12} and γ_{2F} may increase with r_F as increased seed production boosts the flow of plant density through all stages. By increasing the number of plant individuals maturing, we observe a greater number of plant individuals replacing those lost to consumption. In our

analysis, γ_{12} and γ_{2F} (along with L:D ratio) are treated as explanatory variables which we label

316 "ecological factors," to differentiate them from other statistical inputs used in our other analyses.

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