

1 Supplementary information for:  
2

3 **Longitudinal single-cell profiling of chemotherapy response in acute**  
4 **myeloid leukemia**  
5

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21 **Supplementary Figure 1**

22 **A** Flow cytometry gating and sorting strategies for patient sample bone marrow (PB for PT06 at diagnosis) prior to single-  
23 cell RNA sequencing. Samples processed with each sorting strategy are reported on the left. Strategy1:  
24 Singlets/Physical/CD45<sup>+/low</sup>/Recovery of CD117<sup>+</sup>34<sup>-</sup> (Gate CD117<sup>+</sup>) & CD34<sup>+</sup> cells. Strategy2:  
25 Singlets/Physical/Live/FITC<sup>-</sup> (CD3<sup>-</sup>CD19<sup>-</sup>CD235a<sup>-</sup>)/CD45<sup>+/low</sup>/ Recovery of CD34<sup>+</sup> cells; cells not within CD34<sup>+</sup> gate  
26 are further divided into recovery gates: CD117<sup>+</sup> & inverse CD117<sup>+</sup> gate (NOT-CD117<sup>+</sup>). Strategy3:  
27 Singlets/Physical/CD45<sup>+/low</sup>FITC<sup>-</sup> (AnnexinV<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD235a<sup>-</sup>)/ Recovery of CD34<sup>+</sup> cells; CD117<sup>+</sup>34<sup>-</sup> cells (Gate  
28 CD117<sup>+</sup>); and remaining CD34<sup>-</sup>117<sup>-</sup> cells (Gate P4).

29 **B** Table summarizing patients' clinical features at diagnosis. UPN: unique patient number; WHO diagnosis: clinical entity  
30 according to WHO 2016; ELN 2017: risk stratification according to European Leukemia Net recommendations 2017;  
31 Clinical outcome was dichotomized as Favorable, if the patient is alive, and Adverse, if dead at the time of writing; ASCT,  
32 autologous stem cell transplantation; HSCT, allogeneic stem cell transplantation; FAB, French-American-British  
33 classification; BM blast %, blast count on bone marrow by morphological analysis; for *NPM1*, the mutation type is  
34 reported; or *FLT3/ITD*, the allelic ratio is reported when available; n.a. not available.

35 **C** Table reporting differentially expressed genes (DEGs) between cells harboring the *NPM1* mutA transcript (MUT) and  
36 those classified as AML by the *NPM1*-MF algorithm within each cluster identified by unsupervised clustering at a  
37 resolution of 0.6. We identified a non-negligible number of DEGs only for cluster 7, where we observed higher  
38 expression of erythroid lineage transcripts within MUT blasts with respect to other AML classified cells (Supplementary  
39 Data 2).

40 **D** Left: Human engraftment over time in PB of NSG mice transplanted with diagnosis BMDCs of PT01 (black) or PBMCs  
41 of PT06 (grey). Right: Human graft composition for PT01 and PT06 at 10 weeks post-transplant. Data are presented as  
42 mean +/- SEM. n=6 PDX from 2 patients.

43 **E** Left: UMAP plot of the total merged *NPM1*<sup>mut</sup> AML dataset prior to harmonization process. Right: *NPM1*<sup>mut</sup> AML  
44 dataset post harmonization<sup>1</sup> for correction of patient- and chromium chemistry-derived batch effects. Cells are colored by  
45 10x genomics chromium chemistry used for library production (v2 or v3) or Patient ID.

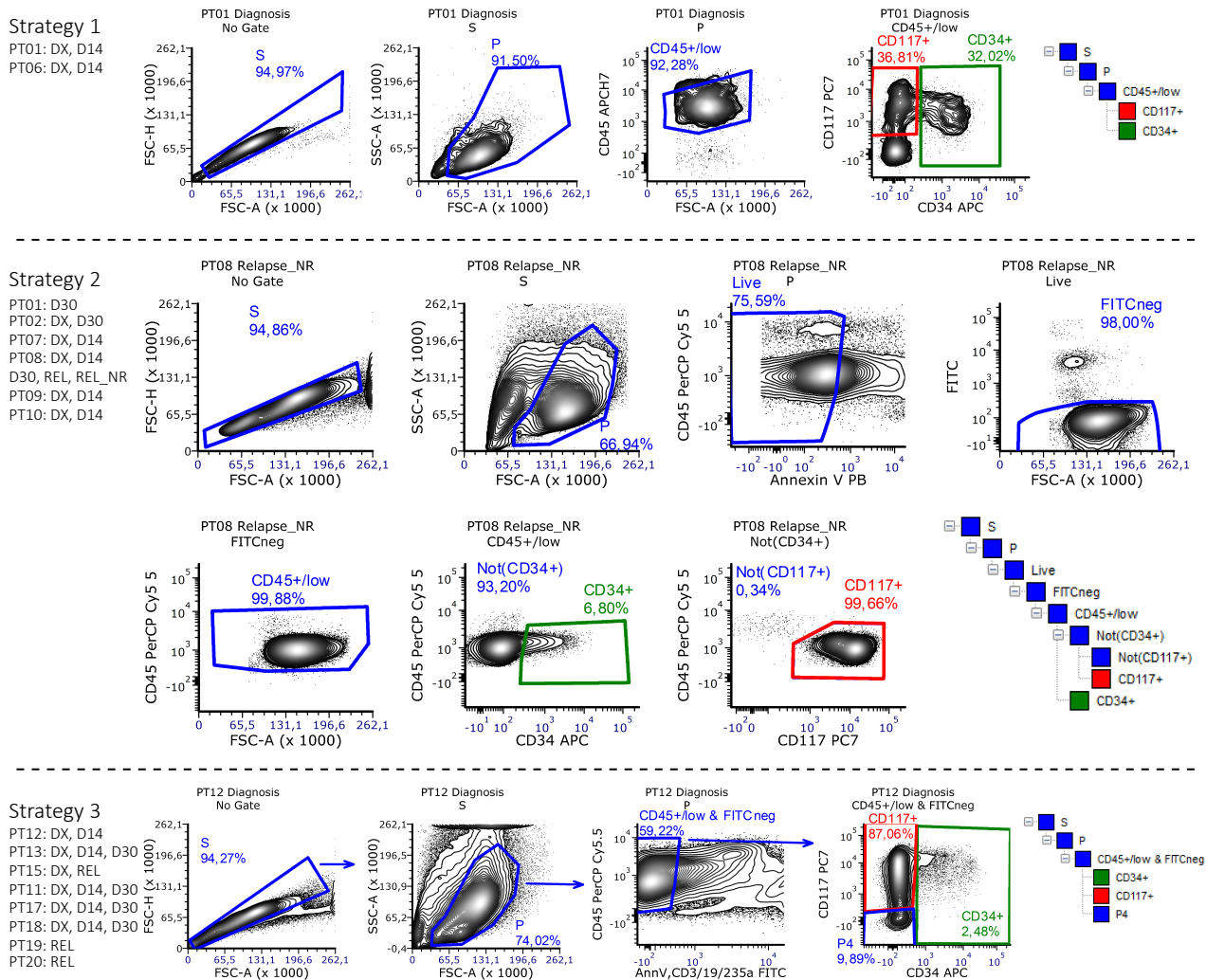
46 **F** Single-cell expression heatmap of the top 10 marker genes of each cluster identified at a resolution of 0.6 for the  
47 *NPM1*<sup>mut</sup> AML dataset. Each column represents a single cell transcriptome and is annotated on top for cluster, patient,  
48 and sampling timepoint variables.

49 **G** Single-cell expression heatmap of the top 10 marker genes of each cluster identified at a resolution of 0.6 for the del(7)  
50 AML dataset. Each column represents a single cell transcriptome and is annotated on top for cluster, patient, and sampling  
51 timepoint variables.

52 Source data are provided as a Source Data file

# Supplementary Figure 1:

**A**



**B**

UPN	WHO diagnosis	Gender	Age at diagnosis (years)	ELN 2017	Clinical Outcome	Karyotype	FAB	BM blast %	DNMT3A	IDH2	NPM1	FLT3ITD
PT01	AML NPM1	M	52	INT	Adverse	46, XY	n.a.	92	n.a.	n.a.	A	0,67
PT02	AML NPM1	F	24	INT	Adverse	46, XX	M5a	70	n.a.	n.a.	A	0,77
PT03	AML NPM1	F	36	INT	Adverse	46, XX	M2	76	n.a.	n.a.	A	pos
PT04	AML NPM1	F	48	n.a.	Favorable	46, XX	M1	86	n.a.	n.a.	A	pos
PT06	AML NPM1	F	76	FAV	Favorable	46, XX	M2	75	n.a.	n.a.	A	0,31
PT07	AML NPM1	M	42	FAV	Favorable	46, XY	M5	58	n.a.	n.a.	A	neg
PT08	AML NPM1	M	52	FAV	Adverse	46, XY	M4	19+14	R882H	R140Q	A	neg
PT09	AML NPM1	F	66	INT	Adverse	46, XX	M4	35	n.a.	n.a.	A	5,07
PT10	AML NPM1	F	65	FAV	Adverse	na	M5b	21	n.a.	neg	A	neg
PT12	AML NPM1	M	44	FAV	Favorable	46, XY	M5a	32	neg	n.a.	A	neg
PT13	AML NPM1	M	72	n.a.	Adverse	46, XY, t(10;13)	M2	82	n.a.	n.a.	A	pos
PT15	AML NPM1	F	48	FAV	Adverse	46, XX	M5	38	n.a.	n.a.	A	neg
PT16	AML NPM1	M	55	INT	Adverse	46, XY, del(10)q?	n.a.	76	n.a.	n.a.	A	0,53
PT11	AML DEL(7)	M	19	ADV	Adverse	45, XY-7	M5a	73	n.a.	n.a.	WT	neg
PT17	AML DEL(7)	M	33	ADV	Adverse	45, XY-7	M5b	38	n.a.	neg	WT	neg
PT18	AML DEL(7)	M	73	ADV	Adverse	45, XY-7	n.a.	42	n.a.	neg	WT	neg
PT19	AML NPM1	F	65	INT	Adverse	46, XX	n.a.	n.a.	n.a.	n.a.	A	pos
PT20	AML NPM1	F	59	FAV	Favorable	46, XX	n.a.	40	n.a.	n.a.	A	0,052

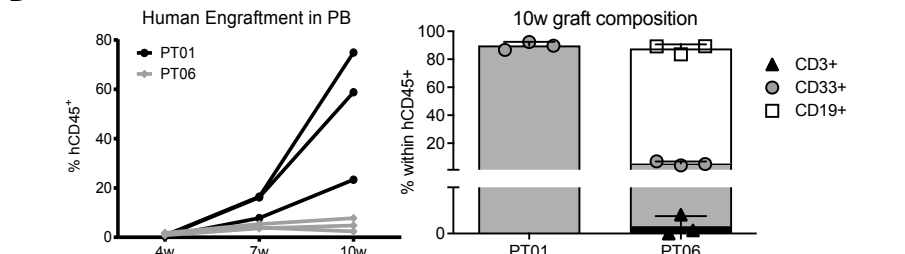
**C**

**INTRA-Cluster comparison of AML cells harboring the NPM1mut transcript (MUT cells) vs other cells classified as AML by NPM1-MF**

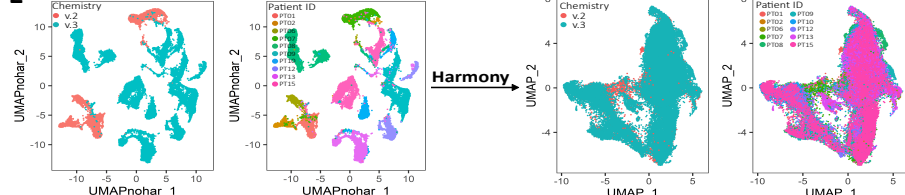
Cluster	#DEGs logFC >0 & p.adj <0.05	DEGs:
0	2	NPM1, RPL31
1	2	NPM1, RPS4Y1
2	4	RPS4Y1, ELANE, SPINK2, RPS10
3	2	NPM1, RPS4Y1
4	8	NPM1, RPL10, RPS3, RPS18, RPS5, RPL10A, RPS10, RPS26
5	1	NPM1
6	1	NPM1
7	224	See Supplementary Table 2, "Intra-cluster 7 MUT vs Rest"
8	1	NPM1
9	2	NPM1, REXO2
10	0	
11	2	NPM1, RPL13A
12	0	

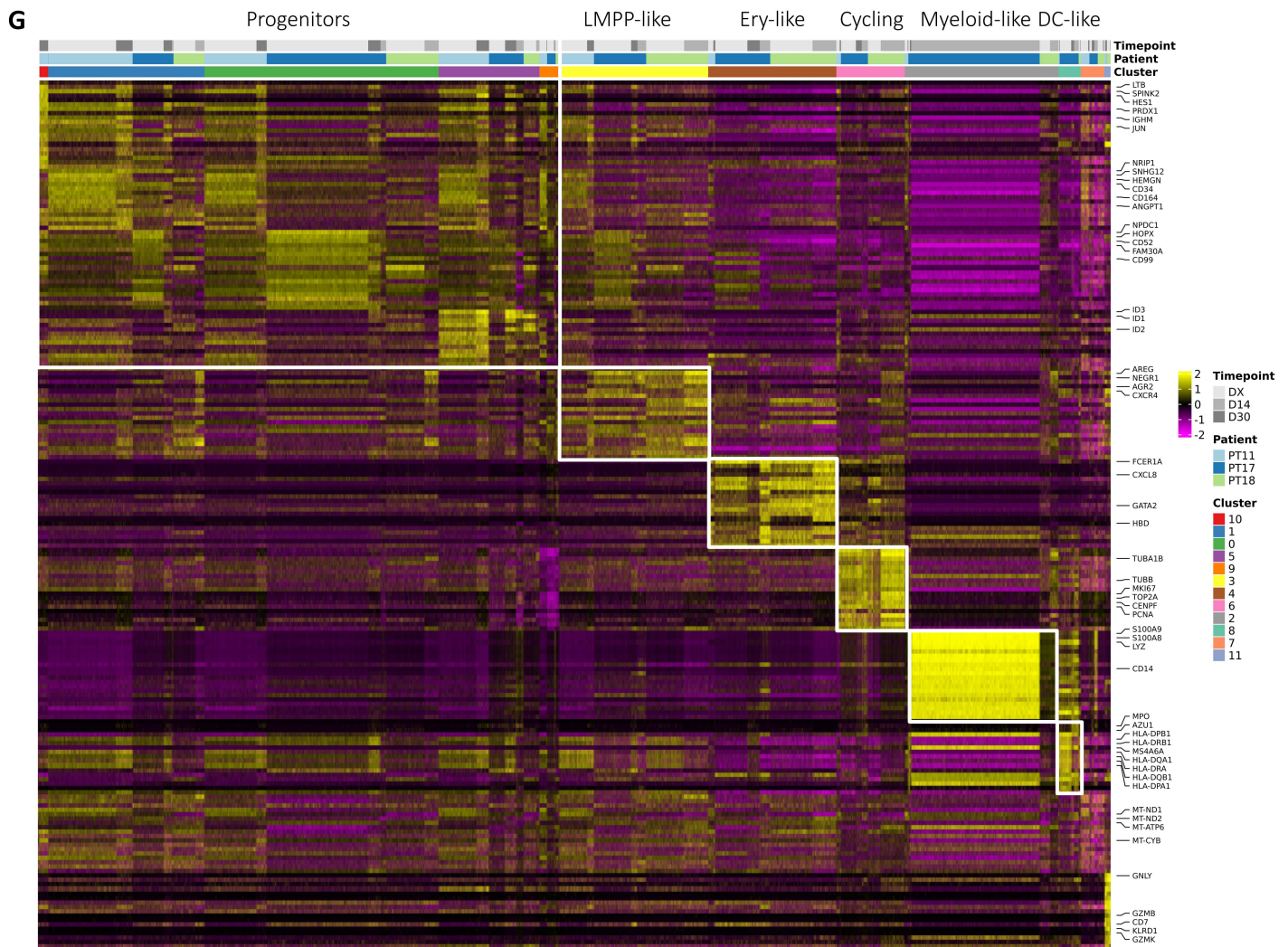
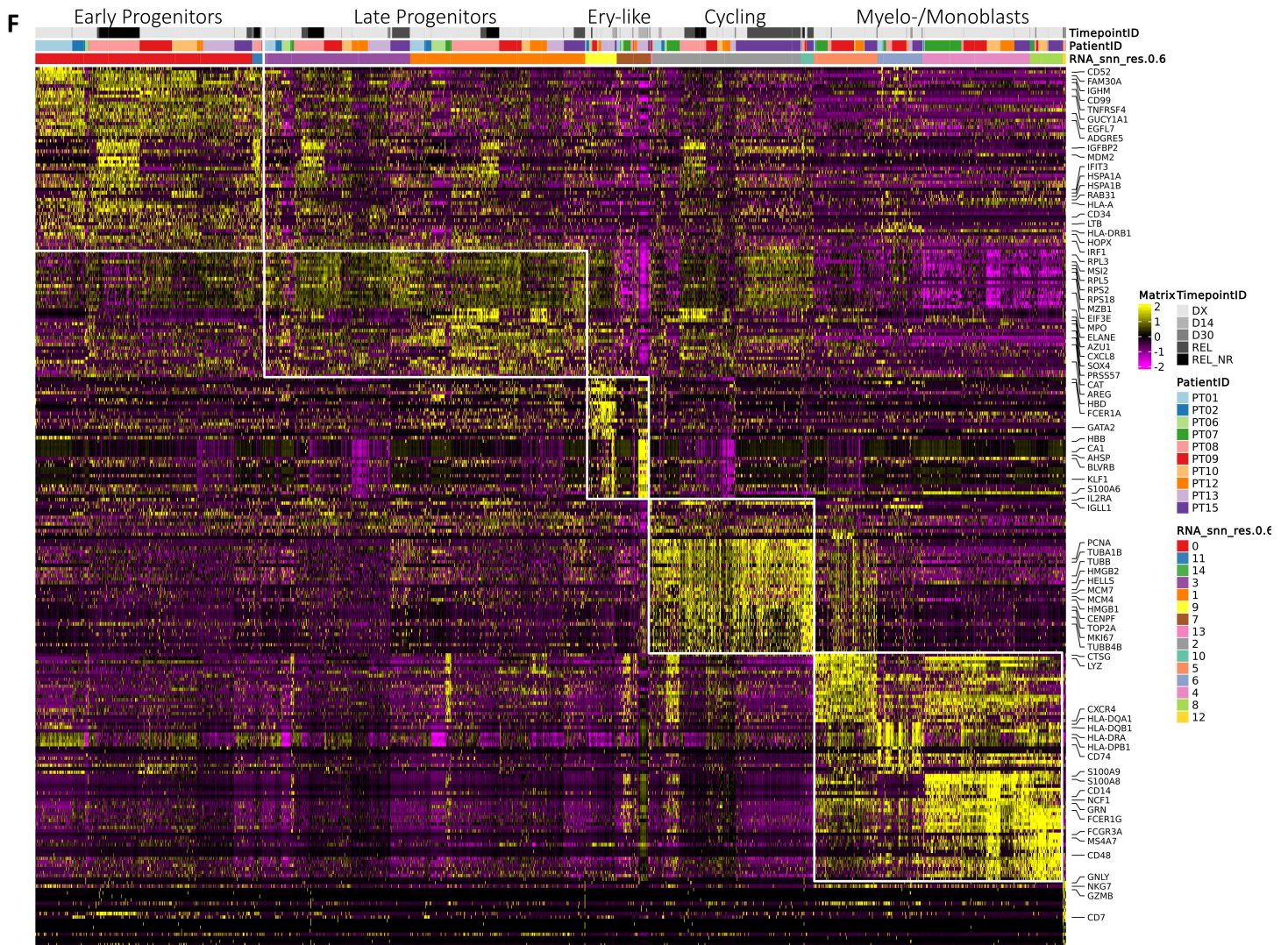
DEGs were computed with a logistic regression framework in Seurat to determine differentially expressed genes while controlling for PatientID and Timepoint latent variables.

**D**



**E**





53 **Supplementary Figure 2**

54 We transduced primary AML cells from 4 patients with the miR-126 reporter and analyzed xenografts (Fig. 3A). The  
55 miR-126 reporter identified intra-tumor heterogeneity (ITH), with the GFP<sup>low</sup>/miR-126<sup>high</sup> cell subpopulation enriched in  
56 CD34<sup>+</sup>38<sup>-</sup> cells (Supplementary Fig. 3C) and confirmed to express higher miR-126 RNA than its GFP<sup>high</sup> counterpart  
57 (Supplementary Fig. 3D). miR-126 intra-tumor heterogeneity (ITH) was maintained over serial xenotransplantation  
58 passages and in paired diagnosis/relapse samples from PT03 (Supplementary Fig. 2A). Taking advantage of semi-  
59 randomly distributed lentiviral integration sites, we confirmed that sorted GFP<sup>high</sup>/miR-126<sup>low</sup> and GFP<sup>low</sup>/miR-126<sup>high</sup>  
60 subpopulations were clonally related, with 88% of genomic integration sites retrieved from the rare GFP<sup>low</sup>/miR-126<sup>high</sup>  
61 population also found in GFP<sup>high</sup>/miR-126<sup>low</sup> blasts. Moreover, a higher proportion of integration sites found in the  
62 GFP<sup>low</sup>/miR-126<sup>high</sup> population was propagated in serial transplantation assays compared to integration sites found in  
63 GFP<sup>high</sup>/miR-126<sup>low</sup> blasts only (Supplementary Fig. 2B,C).

64 **A** Representative FACS plots exemplifying miR-126 sensor-specific GFP downregulation and the relative enrichment in  
65 immature CD34<sup>+</sup> blasts within GFP<sup>low</sup>/miR-126<sup>high</sup> (green histogram) vs GFP<sup>high</sup>/miR-126<sup>low</sup> (grey histogram) subsets  
66 across multiple patients (PT01, PT02, PT03, PT04), disease status (PT03-Diagnosis, PT03-Relapse) and serial passaging  
67 in xenografts (Passage #1,#2,#3).

68 **B** Experimental design for lentiviral vector integration site analysis in xenotransplanted AML samples. BM-derived  
69 human AML cells from primary recipients were FACS-sorted to isolate transduced cells (top) or GFP<sup>high/low</sup> populations  
70 (bottom). Transduced cells were then transplanted in secondary recipients. Lentiviral integrations sites (IS) were retrieved  
71 from gDNA of engrafting human cells and compared amongst groups.

72 **C** Top: Venn diagram illustrating IS sharing among primary GFP<sup>low</sup>/miR-126<sup>high</sup> and GFP<sup>high</sup>/miR-126<sup>low</sup> subsets and  
73 secondary recipient engrafting cells. Bottom: Plot reporting the percentage of shared IS.

# Supplementary Figure 2:

**A**

## PT03 AML =

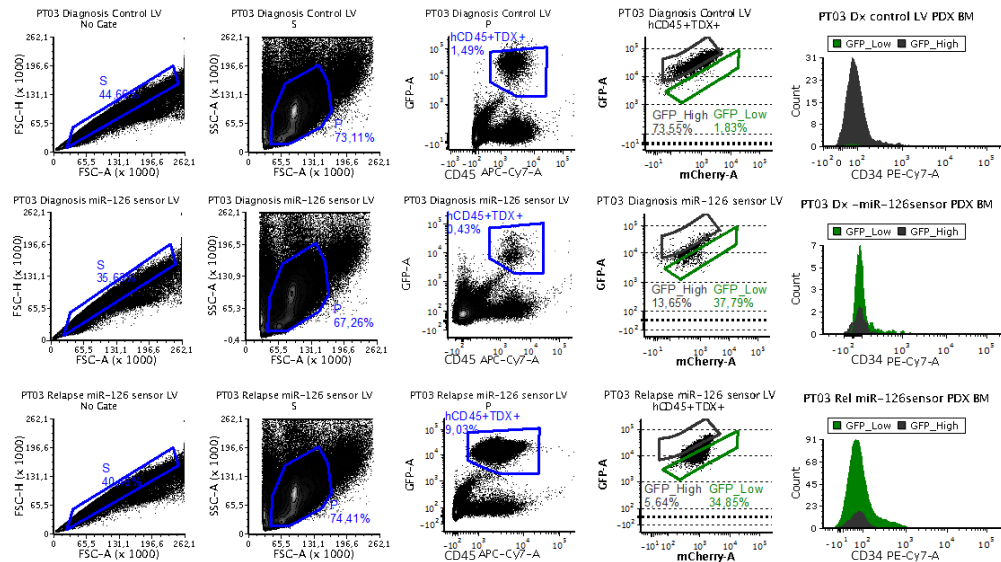
**Diagnosis**

Control LV=

miR-126 sensor

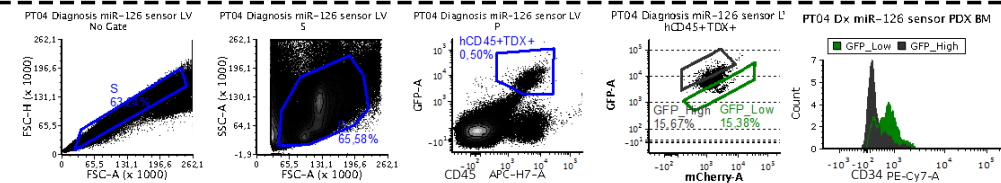
**Relapse**

mir-126 sensor



## PT04 AML =

mir-126 sensor

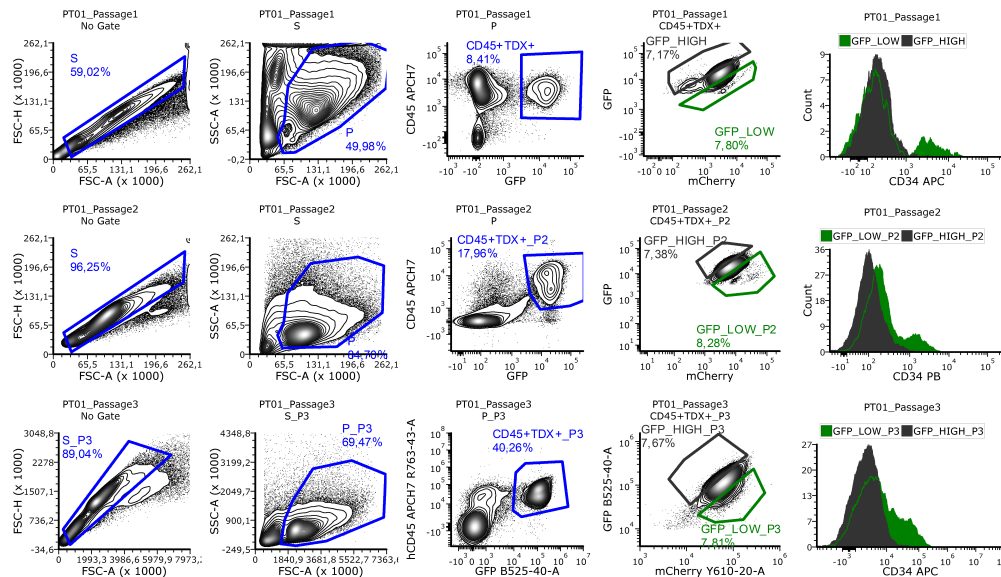


## PT01 AML =

Passage #1

Passage #2

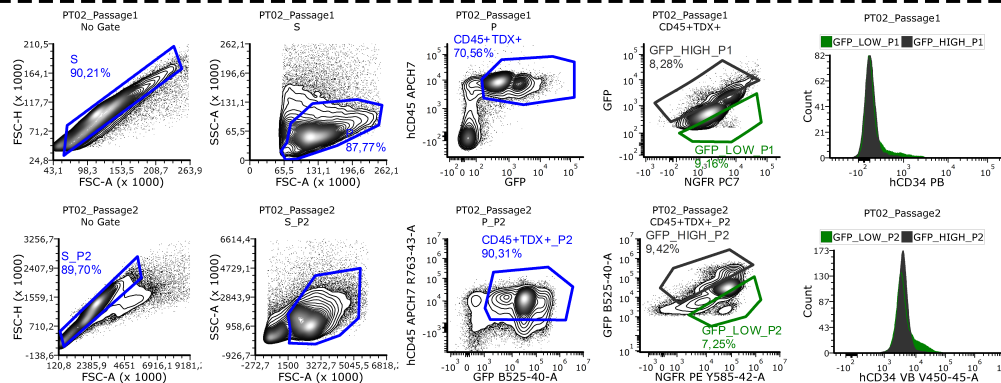
Passage #3



## PT02 AML =

Passage #1

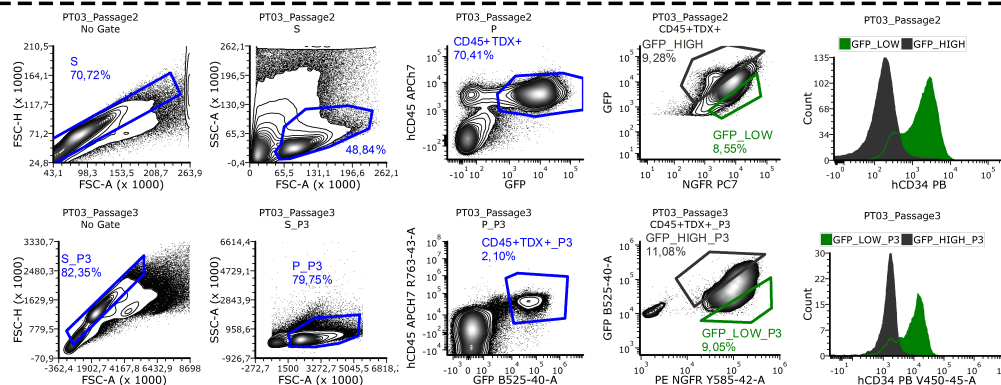
Passage #2

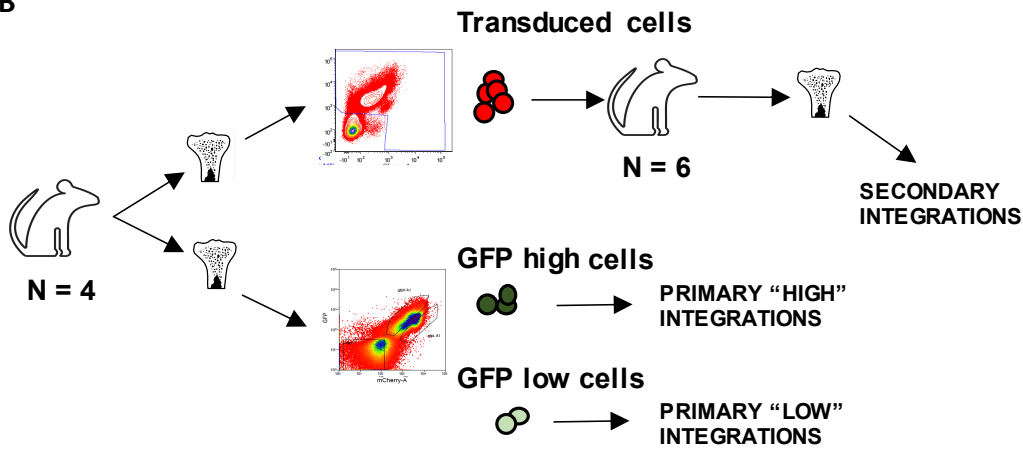
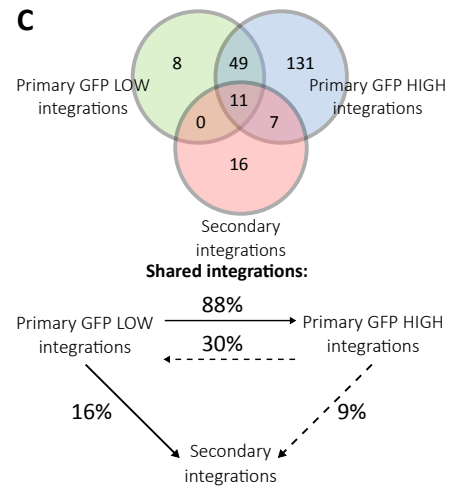


## PT03 AML =

Passage #2

Passage #3



**B****C**

74 **Supplementary Figure 3**

75 **A Left:** Table summarizing the fraction of engrafting animals per cell dose for GFP<sup>low</sup>/miR-126<sup>high</sup> or GFP<sup>high</sup>/miR-126<sup>low</sup>  
76 fractions from PT01, PT02, PT03 and PT16 xenografted AML. L.I.C., estimated leukemia initiating cell frequency.  
77 Within each disease, the fold L.I.C. enrichment in GFP<sup>low</sup> over GFP<sup>high</sup> subpopulations and p-value is reported. p-values  
78 were adjusted with Holm's correction to account for multiple testing. The lower limit of engrafting cell dose was not  
79 reached for PT02 at 500 cells/mouse (L.I.C. of 1/1). Engraftment was defined as >0.1% hCD45<sup>+</sup> blasts in mouse BM at  
80 16 +/- 2 weeks from transplant. n.a., not analyzable. Right: LDA transplantation dose response plots with estimated mean  
81 L.I.C. frequency (full line) and 95% Wald confidence intervals (dashed lines) for tested samples reported in A. Triangles  
82 indicate assays (transplant doses) with 100% success. For PT02, top 2 fully engrafting doses are not shown for plotting  
83 purposes.

84 **B** Flow cytometry evaluation of the percent of G0 (left), G1 (center), and G2M/S (right) blasts within sorted GFP<sup>low</sup>/miR-  
85 126<sup>high</sup> and GFP<sup>high</sup>/miR-126<sup>low</sup> subpopulations from BM aspirates of mice engrafted with PT01,PT03, and PT16 AML  
86 prior to in-vivo chemotherapy treatment. Full lines connect paired measurements from the same mice. Bottom row: Ki67  
87 /Hoechst flow cytometry plots for discrimination of G0, G1, G2M/S cell cycle phases in GFP<sup>high</sup>/miR-126<sup>low</sup> and  
88 GFP<sup>low</sup>/miR-126<sup>high</sup> AML subpopulations recovered from exemplary PDX. n=17 PDX from 3 patients over 3 independent  
89 experiments. Data are presented as mean +/- SD.

90 **C** Relative abundance of immature CD34<sup>+</sup>38<sup>-</sup> blasts within GFP<sup>low</sup>/miR-126<sup>high</sup> or GFP<sup>high</sup>/miR-126<sup>low</sup> subsets revealed  
91 by miR-126 reporter in *NPM1*<sup>mut</sup> AML xenografts. n=14 PDX from 4 patients over 4 independent experiments. Data are  
92 presented as standard Tukey boxplots: center: median, box: interquartile range (IQR), whiskers: IQR\*1.5.

93 **D** miR-126 expression measured by digital droplet PCR (ddPCR) in sorted GFP<sup>high/low</sup> populations from AML xenografts.  
94 Sample pairs sorted from the same mouse are connected by full lines. miR-126 levels are expressed as normalized  
95 copies/ng input cDNA. n=27 PDX from 4 patients over 4 independent experiments. Boxplot representation same as C.

96 **E** AML engraftment in pre-treatment BM aspirates of xenotransplanted NSGW41 mice. Percentage of transduced  
97 hCD45<sup>+</sup> cells on total physical gate cells by flow cytometry is reported. For PT01 and PT03 two independent experimental  
98 replicates are represented by different shapes (circles: replicate A, squares: replicate B). n=77 PDX from 5 patients over  
99 7 experiments. Boxplot representation same as C.

100 **F** Overall transgene ratio (TGR) (mean fluorescence intensity (MFI) ratio of GFP over mCherry or NGFR) of human,  
101 transduced BM AML cells at day 8 from start of treatment. n=78 PDX from 5 patients over 7 independent experiments.  
102 Boxplot representation same as C.

103 **G** TransGene ratio (GFP MFI/mCherry MFI or GFP MFI/NGFR MFI) density histogram of BM AML at single-cell  
104 resolution for each xenograft at day 8 from start of treatment. n=56 PDX from 5 patients over 5 independent experiments.

105 **H** Flow cytometry evaluation of the percent of G0 BM blasts from PT01 and PT03 PDX at baseline (B/L) or after in-vivo  
106 chemotherapy (Treat). Full lines connect paired measurements from the same mice. Right: Ki67/Hoechst flow cytometry  
107 plots for discrimination of G0, G1, G2M/S cell cycle phases prior to and after in vivo chemotherapy in two exemplary  
108 PDX. n=24 PDX from 2 patients over 2 independent experiments. Data are presented as mean +/- SD.

109 For panels **B**, **C**, **D** and **H** Comparisons between paired groups were performed with the paired two-sided Wilcoxon test.  
110 Since for n=5, the minimum achievable two-sided p-value of the test is 0.0625, another non-parametric test based on  
111 bootstrap sampling was employed for conditions with such sample size and such p-values are italicized. n.s.: p-value  
112 >0.05.

113 For panels **E** and **F**: comparisons between groups were performed by using linear mixed-effects (LME) models to account  
114 for mice with the same donor and for experimental replicates. p-values of post-hoc comparisons per patient were adjusted  
115 with Holm's correction in order to account for multiple testing. ns: not significant.

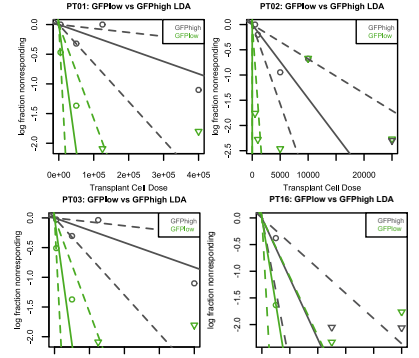
116 Source data are provided as a Source Data file.



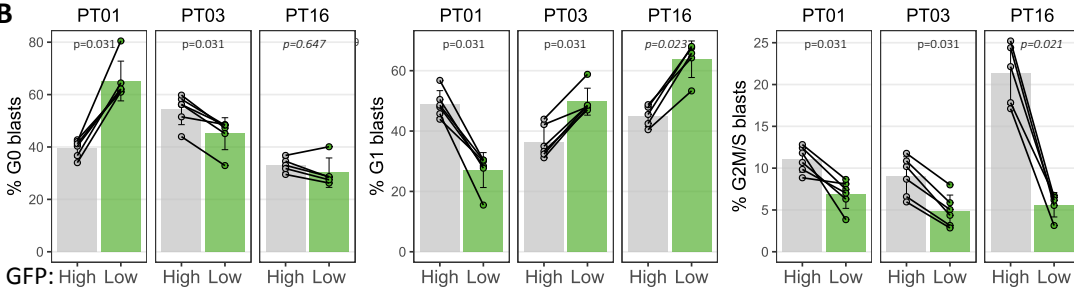
# Supplementary Figure 3:

**A**

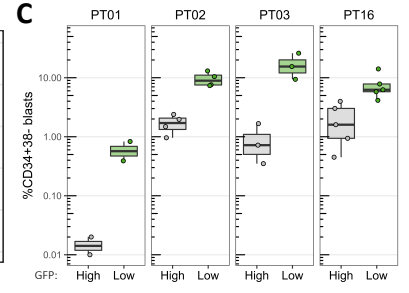
		PT01		PT02		PT03		PT16	
GFP population:		Low	High	Low	High	Low	High	Low	High
cell dose	4-5	* 10 <sup>5</sup>	3/3	2/3	3/3	3/3			
	1-2		4/4	0/4	3/3	4/4	3/3	3/3	5/5
	5		3/4	1/4			2/2	3/3	5/5
	2	* 10 <sup>4</sup>			5/5	5/5			
	1				1/1	1/2			4/5
	5	* 10 <sup>3</sup>	2/5	0/5	6/6	3/5	4/5	0/4	
	1			5/5	1/5				
	5	* 10 <sup>2</sup>		3/3	0/5	0/5	0/5		
L.I.C. Frequency		1/23518	1/481608	1/1	1/6823	1/3849	1/24469	1/6190	1/17621
Fold enrichment		20.5		n.a.		6.4		2.85	
p-value:		8.75 e-06		2.60 e-08		0.036		0.1526	
Adj. p-value:		2.63 e-05		1.04 e-07		0.072		0.1526	



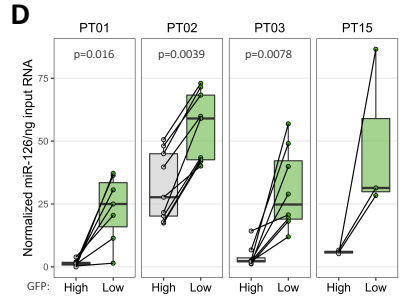
**B**



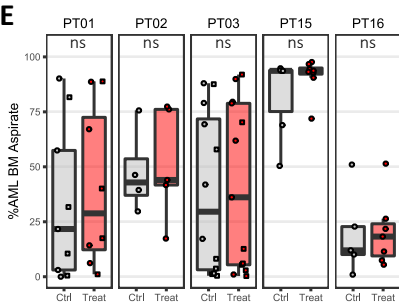
**C**



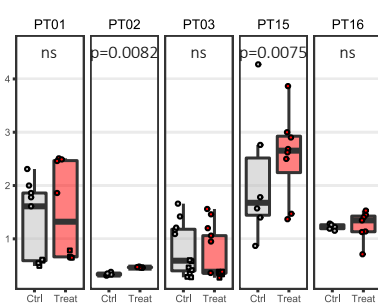
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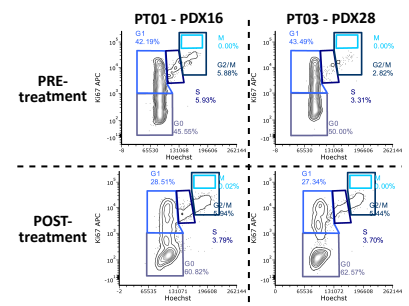
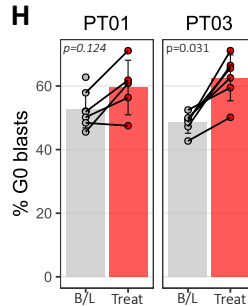
**E**



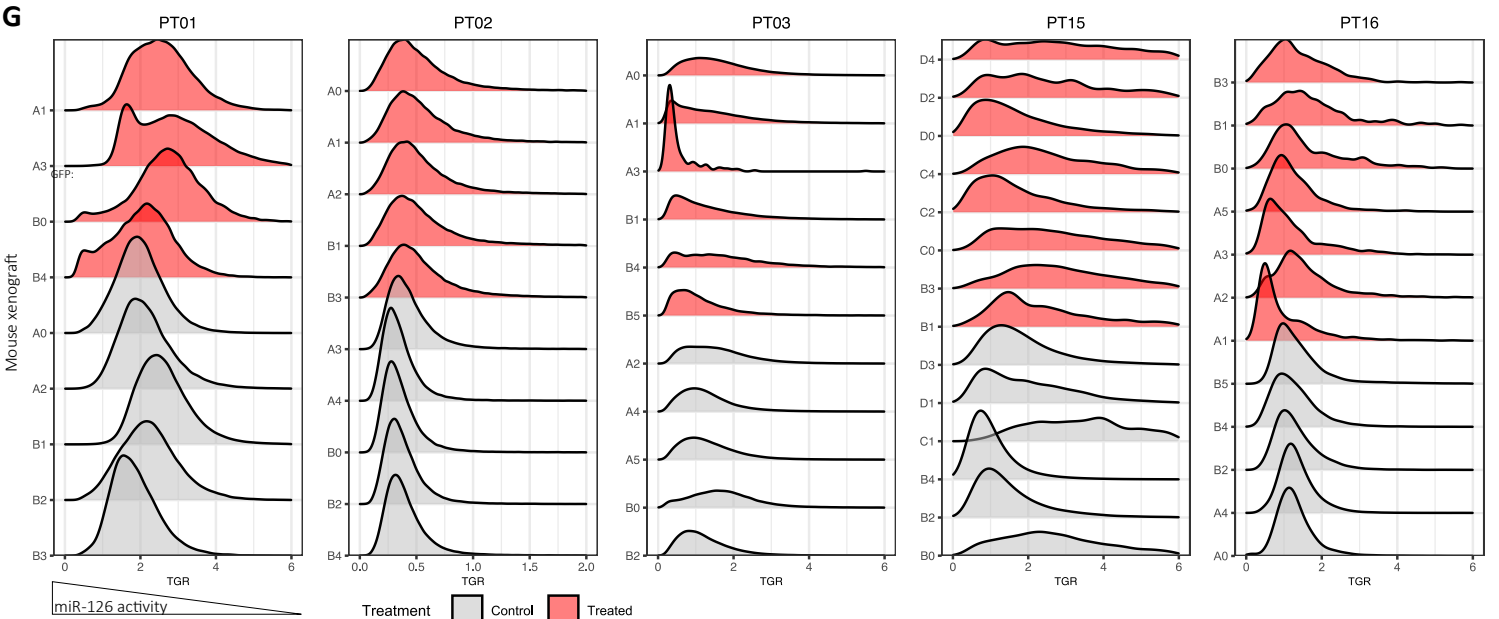
**F**



**H**



**G**



117 **Supplementary Figure 4**

118 **A** Exemplary flow cytometry gating and sorting strategy for enrichment of AML cells from BM of xenografts at day 8  
119 from start of treatment. Gating strategy: Singlets/Physical/human CD45<sup>+</sup> & murine CD45<sup>-</sup> cells were sorted for single-  
120 cell RNA sequencing and limiting dilution transplantation of chemotherapy-treated and control AML; miR-126 reporter  
121 transduced blasts (Transduced+) were brought forward and separated in top 15-20% GFP<sup>high</sup> & GFP<sup>low</sup> subpopulations  
122 for bulk RNA sequencing of both subpopulations as well as single-cell RNA sequencing of GFP<sup>low</sup> blasts.

123 **B** Expression of *ADGRG1* gene (normalized counts) for sequenced GFP<sup>low</sup>/miR-126<sup>high</sup> (green, L) and GFP<sup>high</sup>/miR-126<sup>low</sup>  
124 (grey, H) samples from control (C) or chemotherapy treated (T) AML patient xenograft. n=31 PDX from 4 patients over  
125 4 independent experiments. Data are presented as standard Tukey boxplots: center line: median, box: interquartile range,  
126 whiskers: IQR\*1.5.

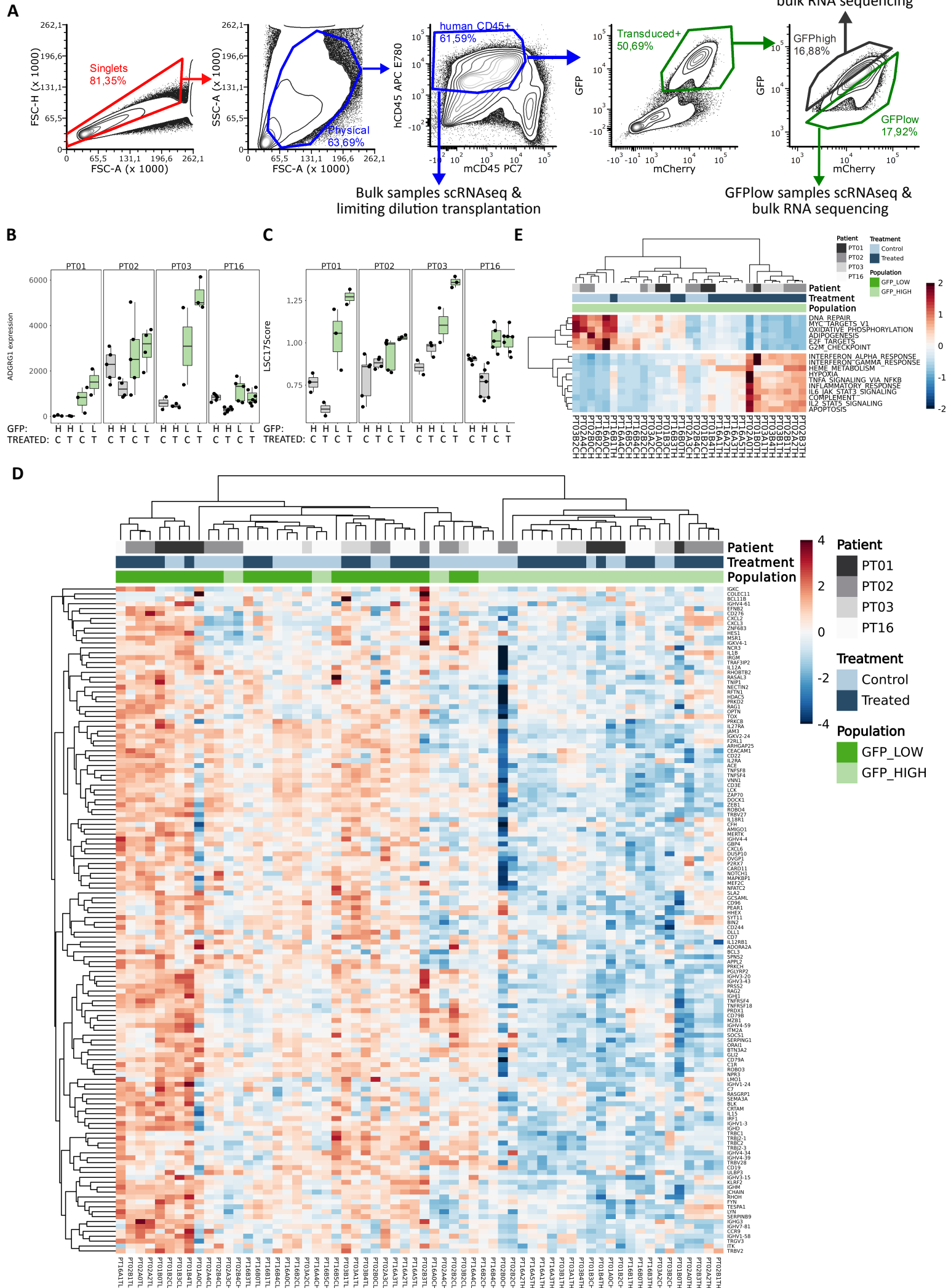
127 **C** Expression of LSC17 score for sequenced GFP<sup>low</sup>/miR-126<sup>high</sup> (green, L) and GFP<sup>high</sup>/miR-126<sup>low</sup> (grey, H) samples  
128 from control (C) or chemotherapy treated (T) AML patient xenograft. LSC17 score was calculated for each cell by  
129 multiplying each gene's rlog expression by the reported coefficient<sup>2</sup>. n=31 PDX from 4 patients over 4 independent  
130 experiments. Boxplot representation same as **B**.

131 **D** Heatmap of lymphoid-related genes within lymphoid Gene Ontologies - Biological Process (BP) categories enriched  
132 in miR-126<sup>high</sup> samples from the over-representation analysis (ORA) on DEGs from the GFP<sup>low</sup>/miR-126<sup>high</sup> vs  
133 GFP<sup>high</sup>/miR-126<sup>low</sup> bulk RNA sequencing comparison. Rows are genes, while columns represent single sorted samples.  
134 Treatment group, patient ID, and GFP sorting gate are annotated on the top of the heatmap.

135 **E** Heatmap of agglomerated z-scores for enriched Hallmark gene sets (MSigDB H group) (rows) from the chemotherapy-  
136 treated vs. control comparison in GFP<sup>high</sup>/miR-126<sup>low</sup> only blasts. Each column represents a single sorted sample.  
137 Treatment group, AML patient ID and GFP population (GFP<sup>high</sup>) are annotated on top.

138 Source data are provided as Source Data file.

**Supplementary Figure 4:**



139 **Supplementary Figure 5**

140 **A** Left: UMAP plot of the total merged single-cell RNA sequencing (scRNA-seq) xenograft AML dataset prior to the  
141 harmonization process<sup>1</sup>. Right: UMAP plot of the xenograft AML scRNA-seq dataset post harmonization process for  
142 correction of patient-derived variability. Cells are colored by Patient ID.

143 **B, C** UMAP plot of the xenograft AML scRNA-seq dataset colored by reference-based cell annotations.

144 **D** UMAP plot of xenograft AML scRNA-seq cells colored by unsupervised clustering (resolution = 1.2). The dashed line  
145 highlights localization of Cluster 1 HSC-like cells at resolution = 0.6.

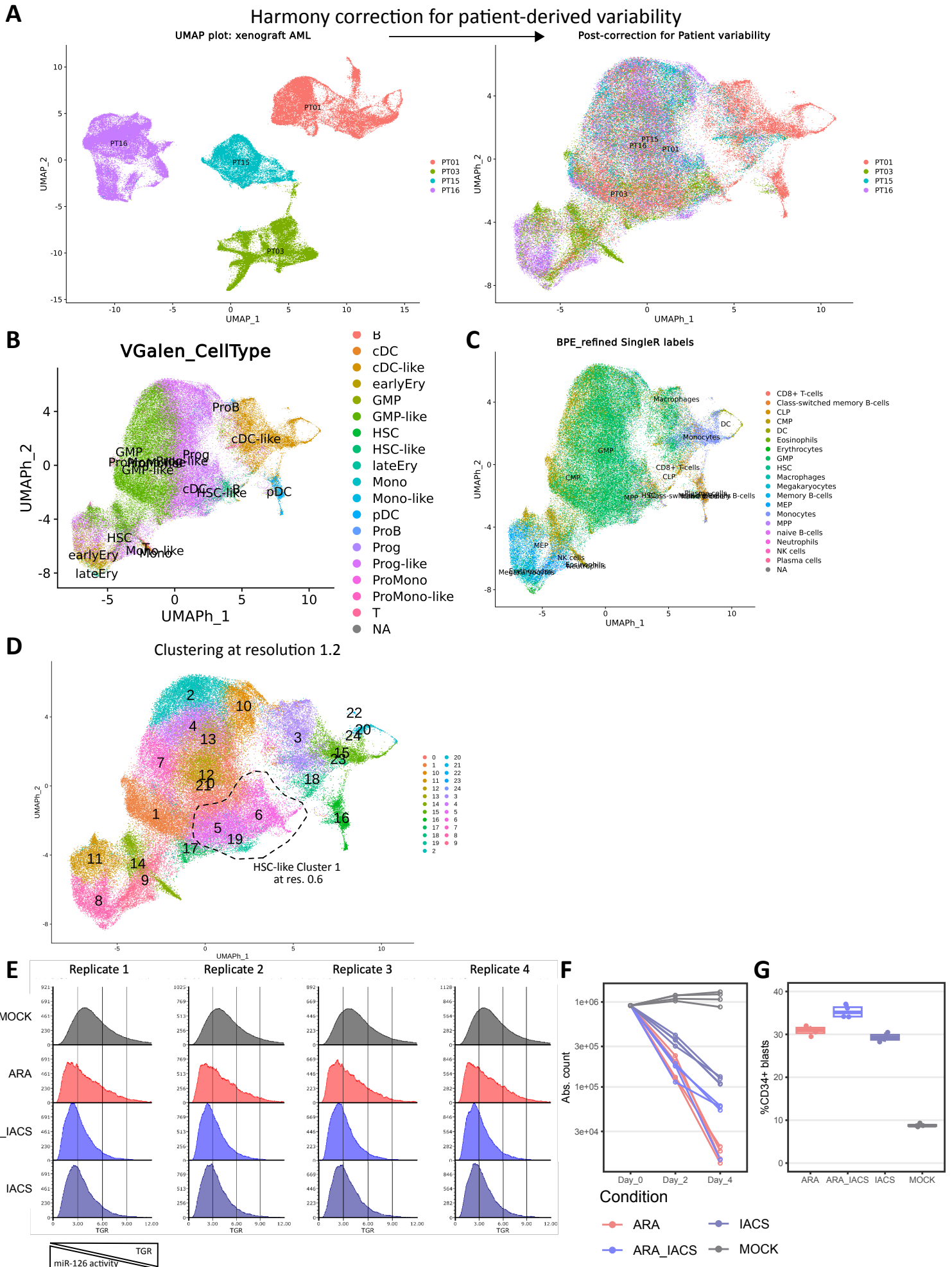
146 **E** TransGene Ratio of miR-126 sensor transduced AML blasts after in-vitro culture for 4 days exposed to Cytarabine  
147 (ARA, 50 ng/ml), IACS-010759 (IACS, 30  $\mu$ M), both (ARA\_IACS), or mock treatment (MOCK). n=4 technical  
148 replicates per condition.

149 **F** In vitro growth curves of miR-126 sensor transduced AML blasts exposed to Cytarabine (ARA, 50 ng/ml), IACS-  
150 010759 (IACS, 30  $\mu$ M), both (ARA\_IACS), or mock treatment (MOCK). n=4 technical replicates per condition.

151 **G** Percent of CD34+ blasts after in-vitro culture for 4 days exposed to Cytarabine (ARA, 50 ng/ml), IACS-010759 (IACS,  
152 30  $\mu$ M), both (ARA\_IACS), or mock treatment (MOCK). n=4 technical replicates per condition. Data are presented as  
153 standard Tukey boxplots: center line: median, box: interquartile range, whiskers: IQR\*1.5.

154 Source data are provided as Source Data file.

Supplementary Figure 5:



155 **Supplementary Figure 6**

156 **A** Correlation between module scores for LSC- and lymphoid-gene subsets. Cells are colored by patient outcomes (dark  
157 red for primary refractory, orange for early relapse after initial CR, green for persistent CR patients). Spearman rho  
158 correlation coefficient and p value are reported. Left: *NPM1*<sup>mut</sup> AML dataset. Right: del(7) AML dataset.

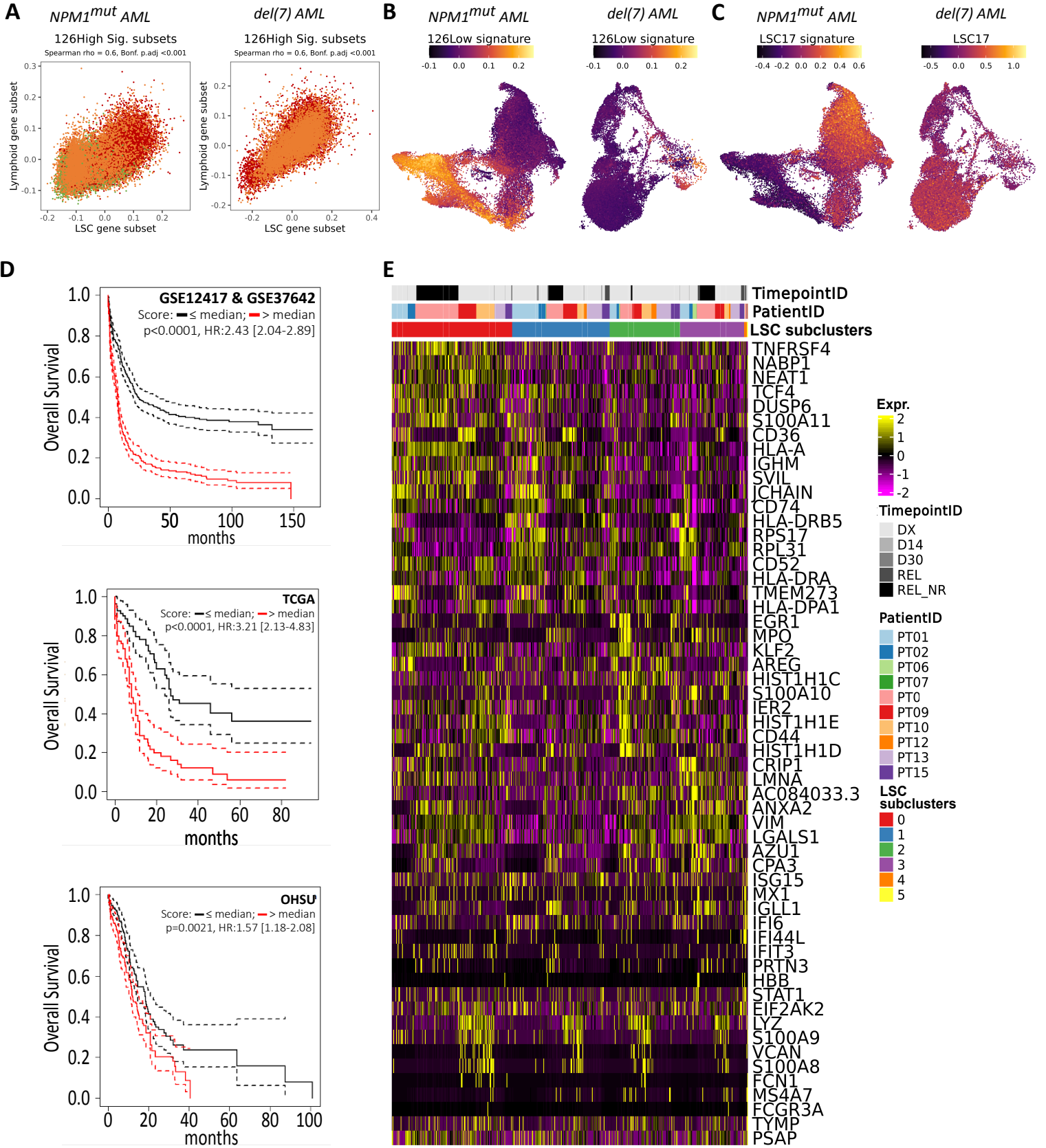
159 **B** Mapping of the expression of the 126Low module score on UMAP plots of *NPM1*<sup>mut</sup> (left) or del(7) (right) AML cells.

160 **C** Mapping of the LSC17 gene signature on UMAP plots of *NPM1*<sup>mut</sup> (left) or del(7) (right) AML cells. LSC17 score was  
161 calculated for each cell by multiplying each gene's scaled expression by the reported coefficient<sup>2</sup>.

162 **D** Kaplan-Meier plot with log rank test p-value for overall survival of AML patients from gene expression data of the  
163 microarray GSE1247 & GSE37642 (top), TCGA-AML (middle), and OSHU (bottom) cohorts. Log rank test p-value and  
164 Cox's proportional hazards model with lasso penalty hazard ratios (HR) with 95% confidence interval are reported. Risk  
165 groups are identified as follows: black line, when the value of the linear combination of selected genes within the 126High  
166 restricted gene list is less or equal to the median, red line when it is above the median.

167 **E** Single-cell expression heatmap of the top 10 marker genes of each LSC subcluster from the *NPM1*<sup>mut</sup> AML dataset.  
168 Each column represents a single cell transcriptome and is annotated on top for LSC subcluster, patient and sampling  
169 timepoint variables.

# Supplementary Figure 6:



170 **Supplementary Figure 7**

171 **A** Top to bottom: PT01, PT02, PT06, PT07. Left: UMAP plot of each single patient's BMBCs (PBMCs for diagnosis of  
172 PT06) from all sampled timepoints in scRNAseq passing quality control filtering and merged together. Cells are colored  
173 by cluster from unsupervised clustering at resolution 3.0. Middle: Heatmap reporting percent composition of each NPM1-  
174 MF category (*NPM1*<sup>mut</sup> (MUT); wild type (WT), Not Detected (ND); No Call) in each cluster identified at resolution 3.0.  
175 Left "AML" column denotes whether cells from that cluster (excluding those defined as WT) were considered as AML  
176 for further analysis based on NPM1-MF criteria (See methods: *NPM1* Mutation Finder). Right: Heatmap reporting  
177 percent composition of SingleR-based cell classification categories based on the BluePrint Encode main reference in each  
178 cluster identified at resolution 3.0.

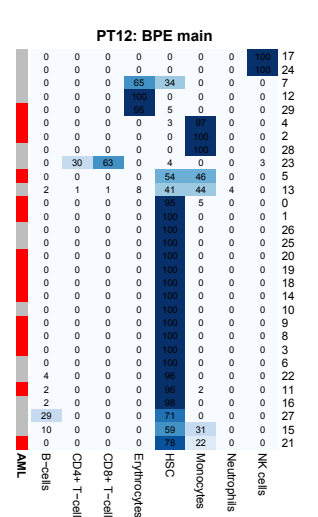
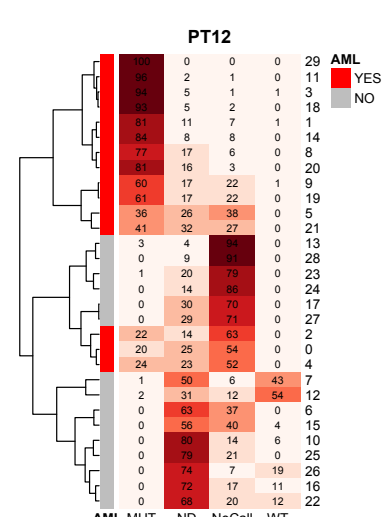
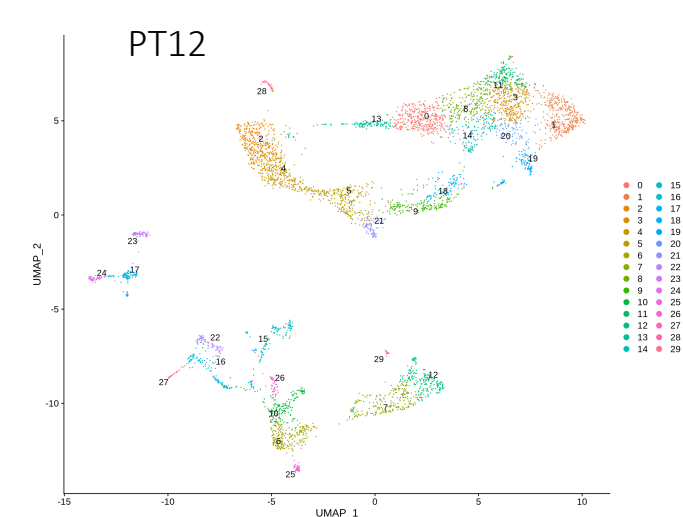
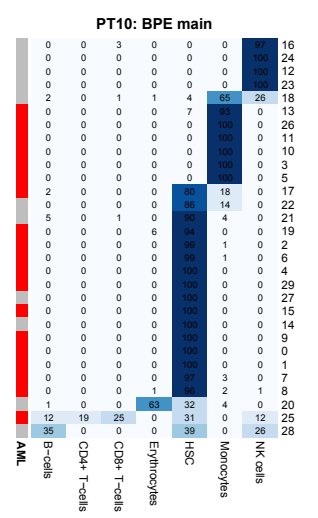
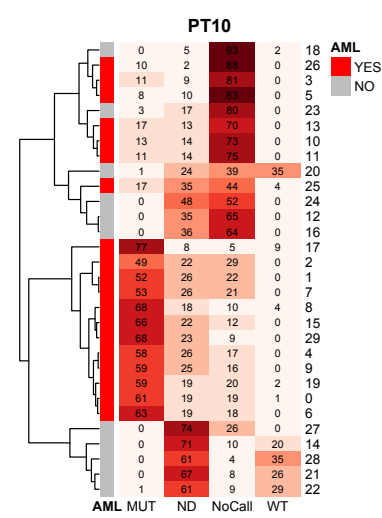
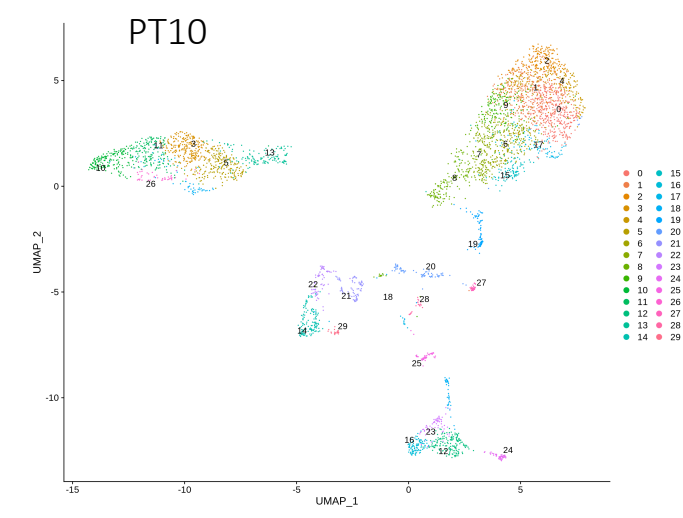
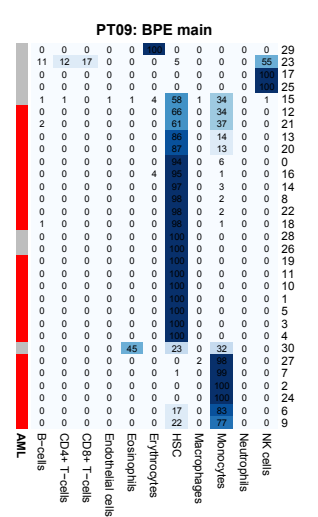
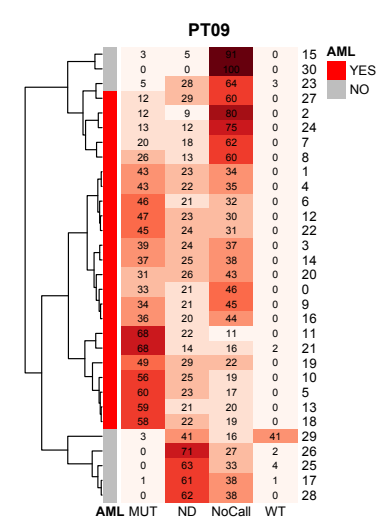
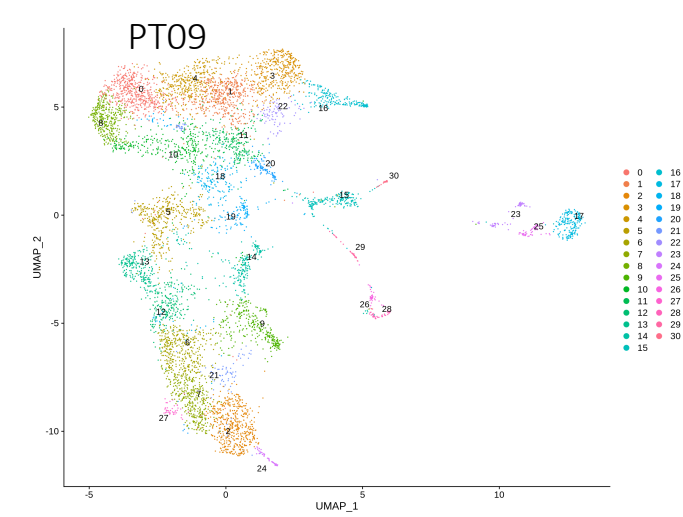
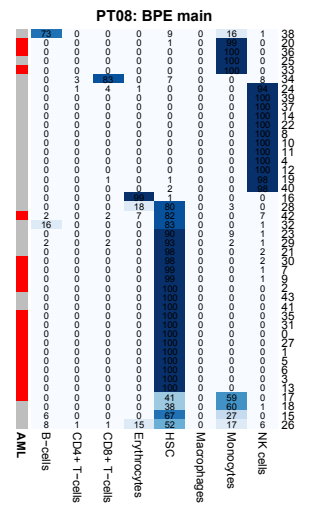
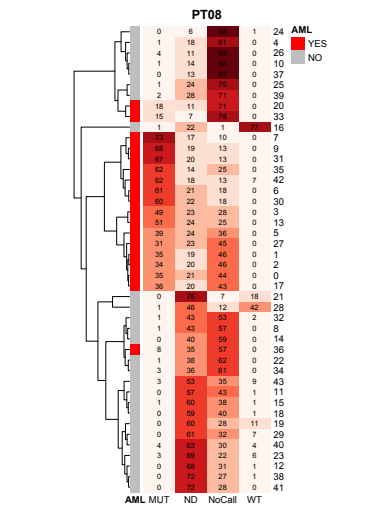
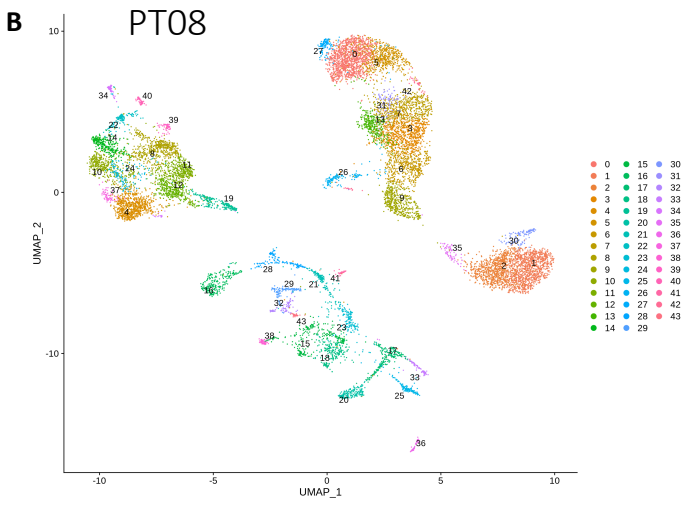
179 **B** Same as **A** for patients: PT08, PT09, PT10, PT12 (ordered top to bottom).

180 **C** Same as **A** for patients: PT13, PT15, PT19, PT20 (ordered top to bottom)

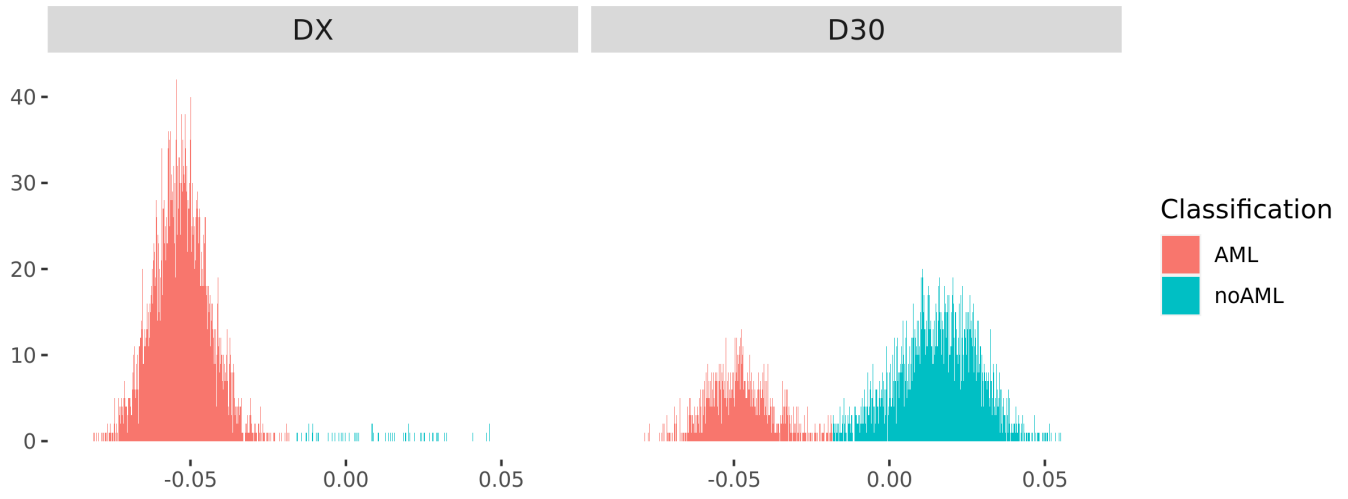
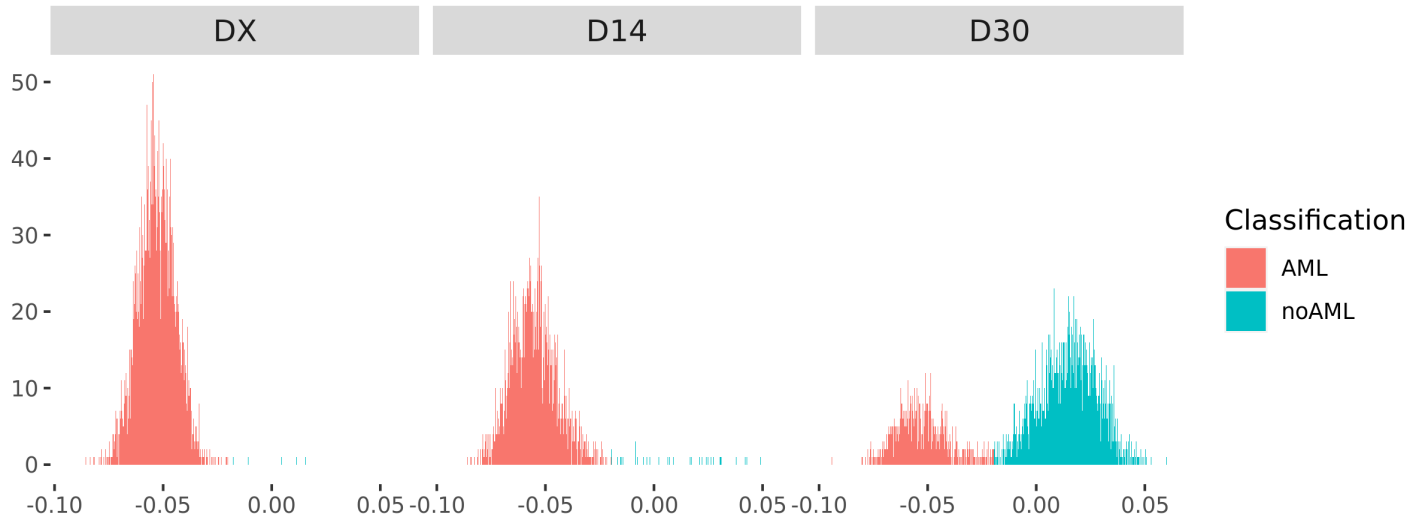
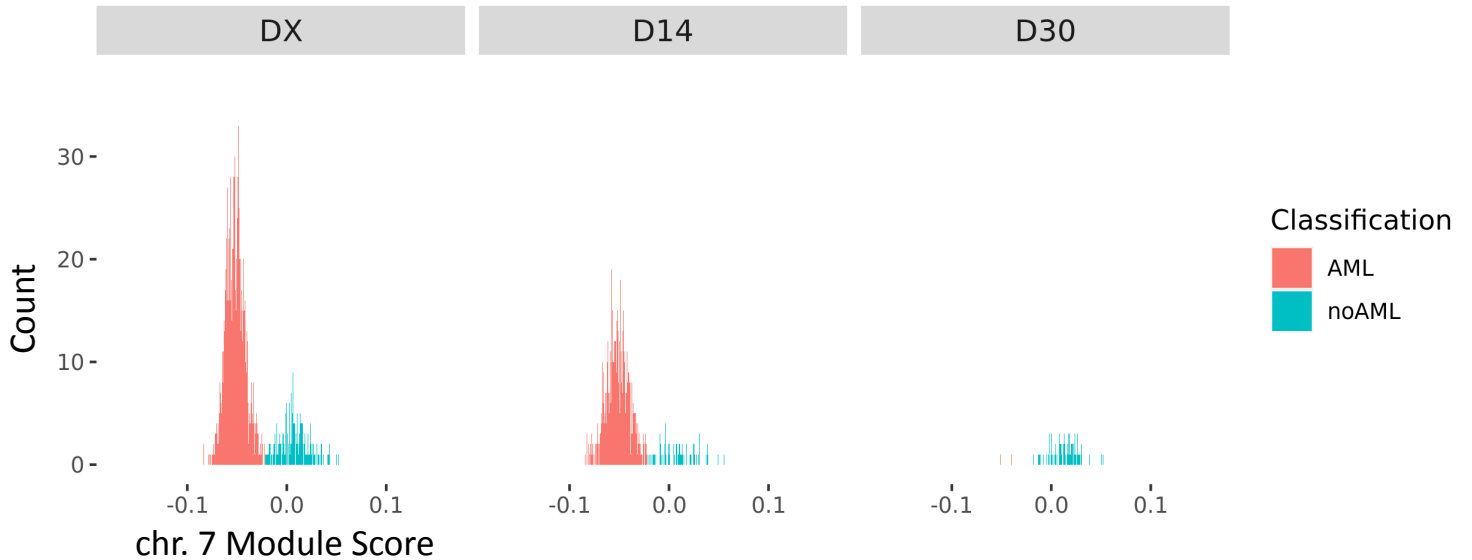
181 **D** Top to bottom: PT11, PT17, PT18. Distribution of Chromosome 7 module score values computed on the expression of  
182 genes located on Chromosome 7 in cells classified as AML (red), or non AML (blue) by k-means clustering ( $k = 2$ ).









**D****PT11****PT17****PT18**

183 **Supplementary Figure 8**

184 **A** Heatmap of k-scores for semantic similarities across Gene Ontology Biological Process enriched categories in miR-  
185 126<sup>high</sup> blasts in bulk RNA sequencing comparing miR-126<sup>high</sup> vs miR-126<sup>low</sup> subpopulations.

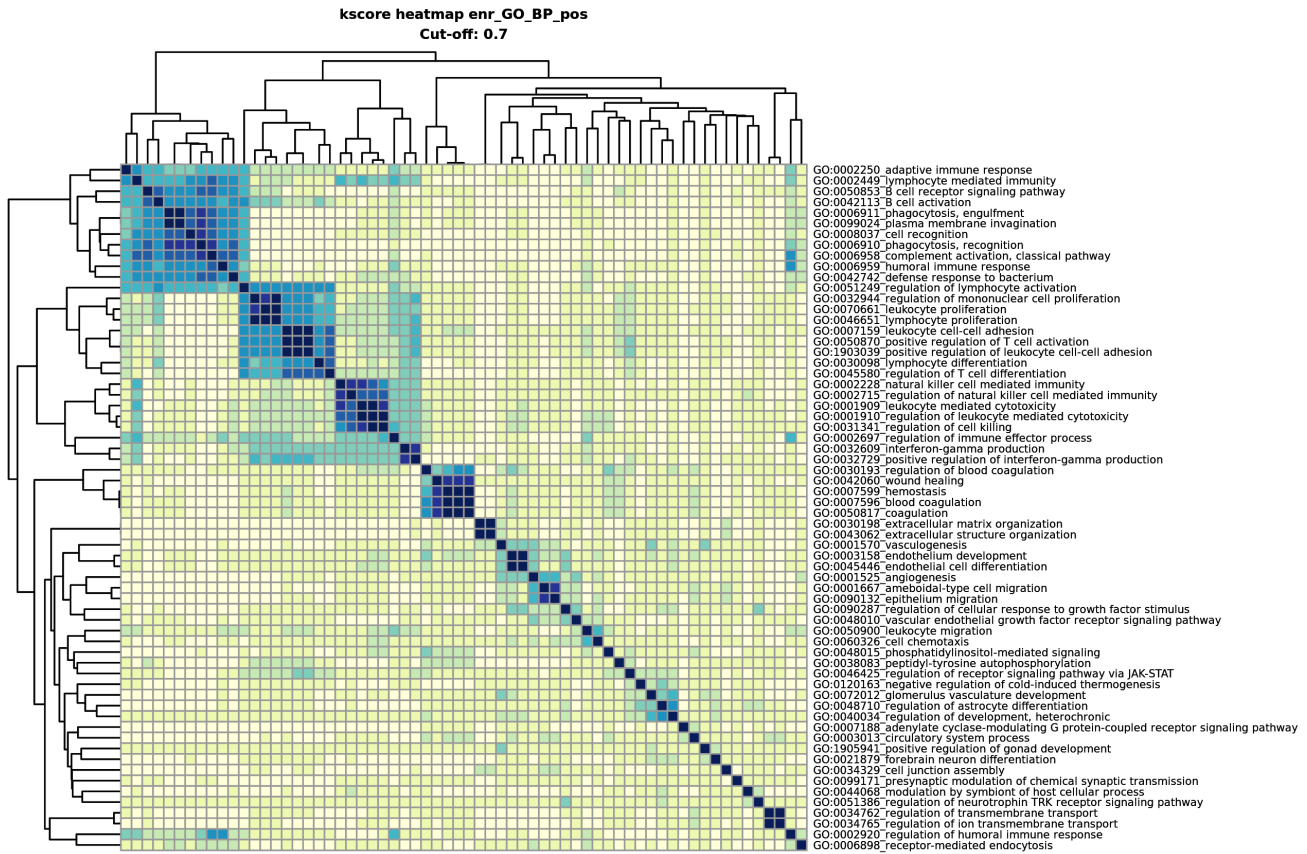
186 **B** Heatmap of k-scores for semantic similarities across Gene Ontology Biological Process enriched categories in miR-  
187 126<sup>low</sup> blasts after a pruning step to reduce redundant labels in bulk RNA sequencing comparing miR-126<sup>high</sup> vs miR-  
188 126<sup>low</sup> subpopulations.

189 **REFERENCES:**

- 190 1. Korsunsky, I. *et al.* Fast, sensitive and accurate integration of single-cell data with Harmony. *Nat. Methods* **16**,  
191 1289–1296 (2019).  
192 2. Ng, S. W. K. *et al.* A 17-gene stemness score for rapid determination of risk in acute leukaemia. *Nature* **540**,  
193 433–437 (2016).

Supplementary Figure 8:

A



B

