Dimeric and Trimeric Catenation of Giant Chiral [8+12] Imine Cubes Driven by Weak Supramolecular Interactions

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Table of Contents

1.	Materials and methods	2
2.	Synthesis and characterization	3
3.	1D and 2D NMR spectra	31
4.	DOSY spectra	195
5.	Mass spectra	206
6.	IR spectra	217
7.	UV spectra	228
8.	GPC data	239
9.	Variable concentration NMR studies	248
10.	Variable catalyst concentrations	252
11.	Different solvent NMR studies	253
12.	Thermodynamic data	267
13.	¹ H NMR assignment for dimeric catenane	287
14.	Variable temperature NMR studies with trimeric catenane	333
15.	¹ H NMR kinetic studies	338
16.	Catenation studies without acid catalyst	341
17.	Mechanistic studies of catenanes by mass spectrometry	342
18.	Models of Weak interactions	347
19.	Computational details	348
20.	References	361

1. Materials and Methods

All reagents for synthesis were purchased from commercial suppliers and used without further purification. Chloroform was passed through the basic alumina column (Type IV) before it was used it for all the cage and catenane reactions. All air-sensitive reactions were performed using oven-dried glassware under an inert atmosphere of argon. Syringe or cannula was used to transfer air-sensitive solvents, reagents and solutions. Analytical thin layer chromatography (TLC) was performed on MERCK precoated silica gel 60 F_{254} TLC plates. All the compounds were visualized under UV light.

For flash column chromatography silica gel 60 (40-63 µm/230-400 mesh ASTM) was purchased from Macherey-Nagel. NMR spectra were recorded on a Bruker Avance III 300 (300 MHz) or Bruker Avance DRX 300 (300 MHz) or Bruker Avance III 400 (400 MHz) or Bruker Avance III 500 (500 MHz) or Bruker Avance III 600 (600 MHz) or Bruker Avance III 700 (700 MHz) spectrometer. The NMR spectra were referenced using residual solvent peak as the standard. Chemical shift values are denoted in parts per million (δ), coupling constants (J) are reported in Hertz (Hz), and spin multiplicities are reported as singlet (s), broad singlet (bs), doublet (d), triplet (t), quartet (q), and multiplet (m). Recycling gel permeation chromatography (rGPC) was carried out using Shimadzu DGU-20A3R degassing unit, LC-20AD pump unit, CTO-20AC column oven, CBM-20A communication bus module, SPD-M20A diode array detector, FRC-10A fraction collector, FCV-20AH2 valve unit, a PSS SDV (20 x 50 mm) pre-column and SDV 100 Å (20 x 300 mm) and 500 Å (20 x 300 mm) columns. For HRMS measurement, Fourier Transform Ion Cyclotron Resonance (FTICR) mass spectrometer solariX (Bruker Daltonik GmbH, Bremen, Germany) equipped with a 7.0 T superconducting magnet and interfaced to an Apollo II Dual ESI/MALDI source was used. A tims-TOF MALDI spectra were collected on Bruker timsTOF fleX instrument from Bruker Daltonics with DCTB (trans-2-[3-(4-tert-butylphenyl)-2-methyl-2propenylidene]malononitrile) as a matrix. MALDI-TOF MS spectra were collected from a Bruker Daltonik Reflex III, on a Bruker ApexQe or on a Bruker AutoFlex Speed TOF with DCTB as a matrix. Elemental analysis was performed in Microanalytical Laboratory of the University of Heidelberg using an Elementar Vario EL machine. Melting points were measured using a Büchi Melting Point B-545 instrument. CompactStar CS4 centrifuge machine from VWR was used to collect the precipitate of the cages.

2. Synthesis

a. Synthesis of TBTQ and dialdehyde linkers

Compound 1

TBTQ 1 was synthesized according to literature known procedures.^[1a]

Compounds 2 and 3



1,4-dibromo-2,5-dimethoxybenzene,^[1b] 2,5-dimethoxyterephthalaldehyde,^[1c] 2,5dihydroxyterephthalaldehyde^[1d] were synthesized according to literature known procedures. All obtained analytical data were consistent with those in literature.

Compound 5



2,5-dimethylterephthalaldehyde ^[2,3] was obtained according to a literature known procedure. All obtained analytical data were consistent with those in literature.

Compound 6



1,4-diethyl-2,5-diiodobenzene^[2] 2,5-diethylterephthalaldehyde^[2] were synthesized according to literature known procedures. All obtained analytical data were consistent with those in literature.

Compounds 7 and 8



2,5-diethoxyterephthalaldehyde^[4] and 2,5-isopropoxyterephthalaldehyde^[4] were obtained according to literature known procedures. All obtained analytical data were consistent with the literature.

Compounds 9 and 10



2,5-bis(methylthio)terephthalaldehyde^[5] and 2,5-bis(tert-butylthio)terephthalaldehyde^[5] were synthesized according to literature known procedure. All obtained analytical data were consistent with those in literature.

Compound 11



2.5-Dibromo-terephthaldehyde^[6] was obtained according to literature known procedure. All obtained analytical data were consistent with those in literature.

Compound 12



2-methoxy terephthaldehyde **12** was synthesized according to a literature procedure. ^[7] All obtained analytical data were consistent with those in literature.

Synthesis of ¹⁵N labelled TBTQ-TA 14



¹⁵N labelled Trinitro TBTQ 13



Mp: 256-258°C; ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 8.21-8.18 (m, 3H), 8.18-8.17 (m, 3H), 8.10-8.07 (m, 6H), 7.56 (d, J = 9 Hz, 3H), 7.49 (d, J = 8.4 Hz, 3H), 2.29-2.22 (m, 12H), 1.71 (s, 6H), 1.22-1.15 (m, 12H), 0.99-0.96 (m, 18H); ¹³**C NMR** (151 MHz, CD₂Cl₂): δ = 154.4, 154.19, 154.15, 154.0, 149.2, 149.02, 149.0, 148.97, 148.96, 148.91, 148.89, 148.83, 148.81, 148.8, 148.73, 148.7, 124.93, 124.91, 124.90, 124.88, 124.86, 124.85, 124.10, 124.09, 124.06, 124.05, 123.98, 123.97, 123.94, 123.93, 119.45, 119.44, 119.39, 119.39, 119.36, 119.35, 119.31, 119.30, 73.9, 68.1, 67.72, 67.70, 67.4, 40.7, 40.59, 40.57, 40.45, 20.8, 20.72, 20.68, 15.3, 15.0; FT-IR (neat, ATR): $\tilde{\nu}$ (cm⁻¹) 3070 (w), 2964 (w), 2933 (w), 2873 (w), 1591 (w), 1488 (s), 1315 (s), 1149 (w), 1109 (m), 1072 (w), 1056 (w), 906 (w), 896 (w), 839 (m), 825 (m), 759 (w), 723 (s), 686 (w), 665 (w), 622 (w); **HR-MS (ESI**⁺): *m/z* [M+Na]⁺ calcd. for C₃₂H₃₃¹⁵N₃O₆·Na: 581.2178, found 581.2173. **Elemental analysis**: calcd. for C₃₂H₃₃¹⁵N₃O₆·¹/₈·CH₂Cl₂: C 67.79, H 5.89, N 7.91, found C 67.79, H 5.30, N 7.80.

¹⁵N labelled enantiopure (-)-(P) Triamino TBTQ 14



Mp: 289°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂) $\delta = 6.99$ (d, ³*J* = 8.4 Hz, 3H), 6.52 (t, d, *J* = 1.8 Hz, 3H), 6.46 (dt, *J* = 1.8 Hz, 3H), 3.55 (s, 5H), 3.55 (s, 6H), 2.05-1.96 (m, 6H), 1.52 (s, 3H), 1.20-1.14 (m, 6H), 0.90 (t, *J* = 7.8 Hz, 9H); ¹³**C NMR** (151 MHz, CD₂Cl₂): $\delta = 150.2$, 146.44, 146.37, 138.6, 124.14, 124.13, 114.98, 114.96, 109.47, 109.46, 73.0, 66.6, 41.1, 20.8, 15.4, 15.3; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) ; = 3433 (w), 3379 (w), 3330 (m), 3321 (m), 3190 (w), 3006 (w), 2958 (w), 2929 (w), 2910 (w), 2869 (w), 2852 (w), 1608 (s), 1583 (m), 1492 (s), 1456 (m), 1307 (s), 1263 (s), 1245 (w), 1149 (w), 1132 (w), 1101 (w), 1078 (w), 1055 (w), 929 (w), 850 (m), 802 (s), 763 (s), 750 (s), 686 (s), 628 (m); **HR-MS (ESI**⁺): *m/z* [M+Na]⁺ calcd. for C₃₂H₃₉¹⁵N₃Na: 491.2953, found 491.2948 **Elemental analysis:** calcd. for C₃₂H₃₉¹⁵N₃·H₂O: C 79.30, H 8.53, N 8.87, found C 79.03, H 8.05, N 8.60.

a. Synthesis of cages and catenanes

Synthesis of OH-cube



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-dihydroxy-terephthaldehyde **2** (5.4 mg, 0.033 mmol) in deuterated chloroform (2 mL) in a screw-cap 8 mL glass vial, a catalytic amount of TFA (0.2 μ L, 0.0026 mmol) was added and the reaction mixture was stirred at RT for 18 hours. Afterwards, the reaction mixture was diluted with dichloromethane to a total volume of 30 mL and the solution was stirred with saturated aq. solution of Na₂SO₄ (4 mL) at RT for ten minutes. The organic layer was separated, dried over Na₂SO₄ and concentrated to ~ 0.5 mL by rotary evaporation. Acetonitrile (4 mL) was added and the formed precipitate was separated by centrifugation technique. The precipitate was sonicated in acetonitrile and separated by centrifugation technique to give 12.2 mg (86%) of **OH-cube** as a brown solid.

Mp: 314°C (decomposed); ¹**H NMR** (400 MHz, CD₂Cl₂): $\delta = 12.54$ (s, 24H, *H*-13), 8.62 (s, 24H, *H*-9), 7.46 (d, ³*J* = 8.4 Hz, 48H, *H*-8), 7.29 (d, ³*J* = 1.2 Hz, 24H, *H*-5), 7.20 (dd, ³*J* = 8.4 Hz, ⁴*J* = 1.2 Hz, 24 H, *H*-7), 7.03 (s, 24H, *H*-12), 2.22 (br (t), 48H, *H*-15) 1.69 (s, 24H, *H*-14), 1.28-1.15 (m, 48H, *H*-16), 0.97 (t, ³*J* = 7.2 Hz, 72H, *H*-17); ¹³C **NMR** (151 MHz, CD₂Cl₂): $\delta = 161.3$ (*C*-9), 153.3 (*C*-11), 149.7 (*C*-4), 148.0 (*C*-6), 147.7 (*C*-3), 124.9 (*C*-8), 122.8 (*C*-10), 119.4 (*C*-12), 119.2 (*C*-7), 118.8 (*C*-5), 73.5 (*C*-1), 67.4 (*C*-2), 41.0 (*C*-15), 20.9 (*C*-16), 15.4

(*C*-14), 15.2 (*C*-17); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2957 (m), 2927 (m), 2869 (w), 1664 (w), 1612 (s), 1587 (s), 1481 (s), 1457 (w), 1334 (m), 1313 (m), 1257 (m), 1150 (s), 1099 (w), 937 (w), 873 (w), 765 (m), 695 (w), 668 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 294, 356, 433; **MALDI-TOF** (DCTB): m/z [M+H]⁺ calcd. for C₃₅₂H₃₃₇N₂₄O₂₄: 5286.60, found 5286.92. **Elemental analysis** calcd. for C₃₅₂H₃₃₆N₂₄O₂₄·7CH₂Cl₂: C 73.60, H 6.15, N 5.79, found C 73.55, H 6.04, N 5.90.

Synthesis of ¹⁵N labelled *OH-cube



¹⁵N-labelled ***OH-cube** was prepared from TBTQ **14** (5 mg, 0.022 mmol) and 2,5-dihydroxyterephthaldehyde **2** (2.7 mg, 0.033 mmol) in deuterated chloroform (1 mL) as described above for **OH-cube**. Yield: 5.4 mg (76%) of ¹⁵N-labelled ***OH-cube** as a brown solid.

Mp: >290°C; ¹**H NMR** (700 MHz, CD₂Cl₂): $\delta = 12.56$ (s, 24H, H-13), 8.62 (d, ²*J* = 2.1 Hz, 24H, H-9), 7.46 (d, ³*J* = 8.4 Hz, 24H, H-8), 7.29 (s, 24H, H-5), 7.20 (d, ³*J* = 7.7 Hz, 24 H, H-7), 7.03 (s, 24H, H-12), 2.23 (br (t), 48H, H-15) 1.69 (s, 24H, H-14), 1.26-1.25 (m, 48H, H-16), 0.97 (t, ³*J* = 7.0 Hz, 72H, H-17); ¹³C **NMR** (176 MHz, CD₂Cl₂): $\delta = 161.3$, 161.2, 153.4, 149.7, 148.0, 147.7, 124.9, 122.8, 119.4, 119.2, 118.8, 73.5, 67.4, 41.0, 20.9, 15.5, 15.2; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3386 (w), 2960 (m), 2931(w), 2873 (w), 1606 (w), 1579 (s), 1481(s), 1380 (w), 1332 (m), 1315 (m), 1217(m), 1151 (w), 972 (w), 871 (m), 856 (w), 806 (w), 792 (w), 734 (w), 646 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 296, 356, 433; **TIMS-TOF Flex** (DCTB): *m/z* M⁺ calcd. for C₃₅₂H₃₃₆¹⁵N₂₄O₂₄: 5309.5198, found 5309.5188.





To a solution of TBTQ 1 (20 mg, 0.043 mmol) and 2,5-dimethoxy-terephthaldehyde 3 (12.6 mg, 0.0649 mmol) in deuterated chloroform (4 mL) in a screw-capped 8 mL glass vial, a catalytic amount of TFA (0.4 μ L, 0.0052 mmol) was added and the reaction mixture was stirred at RT for 3 days. Afterward, the crude reaction mixture was washed with aq. K₂CO₃ (0.25 M, 3 × 2 mL) and dried over Na₂SO₄. After removal of the solvent by rotary evaporation, the resulting red color solid was immediately dissolved in dichloromethane and injected into a r-GPC which clearly shows three peaks for **OMe-cube**, (**OMe-cube**)₂ and (**OMe-cube**)₃ (See Figure 390 for the GPC trace).

Synthesis of OMe-cube (improved protocol)



To a solution of TBTQ 1 (5 mg, 0.01 mmol) and 2,5-dimethoxy-terephthaldehyde **3** (3.1 mg, 0.016 mmol) in CHCl₃ (25 mL), a catalytic amount of TFA (0.01 μ L 0.13 μ mol) was added and the reaction mixture was stirred at RT for 4 hours. Afterward, the reaction mixture was diluted with dichloromethane to a total volume of 52 mL and the solution was stirred with aq. solution of K₂CO₃ (0.25 M, 3×18 mL) at RT for five minutes each. The organic layer was dried over Na₂SO₄ and diluted to a total volume of 150 mL with acetonitrile before the resulting solution was concentrated to 100 mL by rotary evaporation. The solution was again diluted to a total volume of 150 mL with the addition of acetonitrile and solvents were completely evaporated under reduced pressure. The obtained yellow solid was suspended in

acetonitrile (5 mL) and ultra-sonicated for 30 seconds. The resulting suspension was transferred into a centrifuge tube and the solid was separated by centrifugation technique. The solid was repeatedly washed with acetonitrile (3×8 mL) to give 6.6 mg (88%) of **OMe-cube** as a yellow solid.

Mp: 335°C (decomposed); ¹**H NMR** (400 MHz, CD₂Cl₂): δ = 8.87 (s, 24H, *H*-9), 7.73 (s, 24H, *H*-12), 7.41 (d, ³*J* = 8 Hz, 24H, *H*-8), 7.22 (s, 24 H, *H*-5), 7.04 (dd, ³*J* = 8 Hz, ⁴*J* = 1.6 Hz, 24 H, *H*-7), 3.92 (s, 72H, *H*-13), 2.23 (br (t), 48H, *H*-15) 1.69 (s, 24H, *H*-14), 1.32-1.23 (m, 48H, *H*-16), 0.97 (t, ³*J* = 7.2 Hz, 72H, *H*-17); **FT-IR** (ATR): $\tilde{\nu}$ (cm⁻¹) = 2959 (m), 2871 (w), 1615 (m), 1592 (m), 1482 (m), 1466 (m), 1410 (s), 1373 (w), 1286 (w), 1212 (s), 1043 (m), 881 (w), 817 (w), 717 (w), 679 (w), 649 (w), 617 (w), 608 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 296, 406; **MALDI-TOF** (DCTB): *m*/*z* [M+H]⁺ calcd. for C₃₇₆H₃₈₅N₂₄O₂₄: 5623.98, found 5623.24; **Elemental analysis** calcd. for C₃₇₆H₃₈₄N₂₄O₂₄·4CH₂Cl₂: C 76.54, H 6.63, N 5.64, found C 76.10, H 6.96, N 5.56.

Please note: A decent ¹³C NMR spectrum was not available, even by recording 20000 scans on a 600 MHz NMR instrument (see Figure 28). Unfortunately, no concentrated sample could be used, because the compound forms a catenane immediately.

Synthesis of OMe-cube in 1,1,2,2 tetrachloroethane (TCE)



To a solution of TBTQ **1** (5.4 mg, 0.012 mmol) and 2,5-dimethoxy-terephthaldehyde **9** (3.1 mg, 0.016 mmol) in 1,1,2,2 tetrachloroethane (anhydrous, 2 mL), a catalytic amount of TFA (0.1 μ L, 0.0013 mmol) was added and the reaction mixture was stirred at RT for 2 hours. A saturated solution of aq. NaHCO₃ (2 mL) was added to the reaction mixture and it was further stirred at RT for 10 minutes. The organic layer was separated from the aqueous phase, washed with water (4 x 3 mL) and precipitated by adding methanol into the organic layer (6 mL) and left at RT for 4 hours. The yellow precipitate was separated by centrifugation technique and

repeatedly washed with methanol (4 x 4 mL) to give 5.7 mg (76%) of **OMe-cube** as a yellow solid.

Analytical data is in accordance to those described above.



Synthesis of ¹⁵N labelled *OMe-cube in 1,1,2,2 tetrachloroethane (TCE)

¹⁵N-labeled ***OMe-cube** was prepared from TBTQ **14** (5.3 mg, 0.0114 mmol) and 2,5dimethoxy-terephthaldehyde **9** (3.1 mg, 0.016 mmol) in 1,1,2,2 tetrachloroethane (anhydrous, 2 mL) as described above. Yield: 5.9 mg (78%) of ¹⁵N labelled ***OMe-cube** as a yellow solid. **Mp**: 330°C (decomposed); ¹H NMR (700 MHz, C₂D₂Cl₄): $\delta = 8.84$ (d, J = 3.5 Hz 24H, *H*-9), 7.70 (s, 24H, *H*-12), 7.37 (d, ³J = 7.7 Hz, 24H, *H*-8), 7.20 (s, 24 H, *H*-5), 7.03 (d, ³J = 7.7 Hz, 24 H, *H*-7), 3.91 (s, 72H, *H*-13), 2.22 (br (t), 48H, *H*-15) 1.67 (s, 24H, *H*-14), 1.26 (br (m), 48H, *H*-16), 0.98 (t, ³J = 6.3 Hz, 72H, *H*-17); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2995 (w), 2956 (w), 2931 (w), 2869 (w), 1579 (m), 1481 (s), 1463 (s), 1406 (s), 1367 (m), 1284 (m), 1247 (w), 1209 (s), 1180 (w), 1166 (w), 1137 (m), 1037 (s), 970 (w), 875 (m), 819 (m), 798 (m), 734 (m), 698 (m), 644 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 297, 407; **TIMS-TOF Flex** (DCTB): m/z [M+H]⁺ calcd. for C₃₇₆H₃₈₅¹⁵N₂₄O₂₄: 5647.91, found 5647.91.

Note: A decent ¹³C NMR spectrum was not available, even by recording 10240 on a 700 MHz NMR instrument (see Figure 41). It was also observed that the cage shows decomposition over time in deuterated 1,1,2,2 tetrachloroethane due to acidic nature of solvent.

Synthesis of (OMe-cube)₂



To a solution of TBTQ **1** (20 mg, 0.043 mmol) and 2,5-dimethoxy-terephthaldehyde **3** (12.6 mg, 0.0649 mmol) in deuterated chloroform (4 mL) in a screw-capped 8 mL glass vial, a catalytic amount of TFA (0.4 μ L, 0.0052 mmol) was added and the reaction mixture was stirred at RT for 3 days. Afterward, the crude reaction mixture was washed with aq. K₂CO₃ solution (0.25 M, 3 × 2 mL) was dried over Na₂SO₄ and concentrated under reduced pressure. The resulting red-colored solid was immediately dissolved in dichloromethane and purified by recycling gel permeation chromatography (rGPC) (DCM, 35°C, 5 mL/min) to give 14 mg (46%) of (**OMe-cube**)₂ as a yellow solid.

Mp: 315°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): $\delta = 8.90$ (s, 6H, HC=N), 8.87 (s, 6H, HC=N), 8.85 (s, 12H, HC=N), 8.82 (s, 6H, HC=N), 8.78 (s, 6H, HC=N), 8.65 (s, 6H, HC=N), 8.38 (s, 6H, HC=N), 8.03 (s, 6H, Ar-H), 7.88 (s, 6H, Ar-H), 7.79 (s, 6H, Ar-H), 7.71 (s, 6H, Ar-*H*), 7.69 (12H, Ar-*H*), 7.53 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-*H*), 7.41 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-*H*), 7.38 (d, ³*J* = 8.4 Hz, 6H, TBTQ Ar-*H*), 7.36-7.32 (m, 6H, TBTQ-Ar-*H* & Ar-H), 7.27 (s, 6H, TBTQ-Ar-H), 7.24 (s, 12H, Ar-H, TBTQ-Ar-H), 7.21 (s, 6H, TBTQ-Ar-*H*), 7.19 (s, 6H, TBTQ Ar-*H*), 7.17-7.14 (m, 6H, TBTQ-Ar-*H*), 7.08 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ-Ar-*H*), 7.03-6.97 (m, 18H, TBTQ-Ar-*H*), 6.89 (s, 6H, TBTQ-Ar-*H*), 6.83 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ-Ar-H), 6.78 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ-Ar-H), 6.77 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ-Ar-*H*), 6.72 (s, 6H, TBTQ-Ar-*H*), 6.59 (d, ${}^{3}J = 8.4$ Hz, 12H, TBTQ-Ar-*H*), 6.26 (d, ${}^{3}J$ = 7.8 Hz, 6H, TBTQ-Ar-H), 4.04 (s, 18H, OCH₃), 3.94 (s, 18H, OCH₃), 3.90 (s, 18H, OCH₃), 3.86 (s, 18H, OCH₃), 3.82 (s, 18H, OCH₃), 3.67 (s, 18H, OCH₃), 3.11 (s, 18H, OCH₃), 2.85 (s, 18H, OCH₃), 2.30-1.74 (m, 96H, -CH₂CH₂CH₃) 1.71 (s, 30H), 1.64 (s, 18H), 1.35-1.09 (m, 84H, -CH₂CH₂CH₃), 1.0-0.90 (m, 126H, -CH₂CH₂CH₃), 0.77-0.72 (m, 12H, -CH₂CH₂CH₃), 0.48 (t, ${}^{3}J = 7.2$ Hz, 18H, $-CH_{2}CH_{2}CH_{3}$); ${}^{13}C$ NMR (151 MHz, $CD_{2}Cl_{2}$): $\delta = 156.5$, 155.7, 155.3, 155.2, 155.0, 154.6, 154.31, 154.26, 154.2, 153.9, 153.8, 153.1, 152.7, 152.5, 152.6, 152.3, 152.2, 150.1, 149.8, 149.64, 149.6, 149.5, 149.4, 149.3, 149.1, 146.7, 146.6, 146.4, 146.1, 146.0, 145.5, 145.4, 129.0, 128.9, 128.6, 128.55, 128.49, 127.8, 125.0, 124.6, 124.42, 124.36, 124.2, 124.0, 119.8, 119.7, 119.4, 119.04, 118.96, 118.8, 118.5, 118.4, 118.1, 117.5, 117.3, 116.1, 116.0, 111.2, 110.2, 109.83, 109.77, 109.63, 109.58, 73.5, 73.2, 73.0, 67.39, 67.35, 67.33, 67.23, 67.18, 66.9, 56.71, 56.68, 56.65, 56.61, 56.6, 56.4, 56.1, 55.4, 41.8, 41.2, 41.1, 41.0, 21.3, 21.1, 21.0, 20.9, 20.8, 20.3, 15.5, 15.4, 15.32, 15.29, 15.0, 14.9; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2999 (w), 2957 (m), 2925 (m), 2870 (m), 2853 (m), 1734 (w), 1616 (m), 1593 (m), 1492 (s), 1482 (s), 1465 (s), 1410 (s), 1373 (m), 1211 (s), 1140 (m), 1043 (s), 974 (w), 882 (m), 821 (m), 701 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 296, 406; **MALDI-TOF** (DCTB): *m/z* [M]⁺ calcd. for C₇₅₂H₇₆₈N₄₈O₄₈: 11245.94, found 11245.57. **Elemental analysis** calcd. for C₇₅₂H₇₆₈N₄₈O₄₈·33CH₂Cl₂: C 67.11, H 5.98, N 4.79, found C 66.94, H 5.91, N 4.86.

Synthesis of (OMe-cube)₃ (improved protocol)



To a solution of TBTQ **1** (20 mg, 0.043 mmol) and 2,5-dimethoxy-terephthaldehyde **3** (12.6 mg, 0.0649 mmol) in deuterated dichloromethane (4 mL) in a screw-capped 8 mL glass vial a catalytic TFA (0.4 μ L, 0.005 mmol) was added and the reaction mixture was stirred at 80°C for 3 days. After cooling to room temperature, the crude reaction mixture was washed with aq. K₂CO₃ solution (0.25 M, 3 × 2 mL) and dried over Na₂SO₄. The solution was concentrated under reduced pressure and the resulting red-colored solid was immediately dissolved in dichloromethane and purified by rGPC (DCM, 30°C, 5 mL/min) to give 24 mg 80% of (**OMe-cube**)₃ as a yellow solid.

Mp: 310°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): $\delta = 9.31-9.27$ (m, 3H, *H*C=N), 9.06-8.98 (m, 12H, *H*C=N), 8.91-8.72 (m, 66H, *H*C=N), 8.61-8.60 (m, 12H, *H*C=N), 8.51 (s, 6H, *H*C=N), 8.41-8.30 (m, 26H, *H*C=N), 8.09-8.06 (m, 6H, Ar-*H*), 7.92-7.86 (m, 18H, Ar-*H*),

7.81-7.76 (m, 30H, Ar-*H*), 7.72-7.63 (m, 26H, Ar-*H*), 7.56-7.52 (m, 12H), 7.46-7.36 (m, 48H), 7.32-7.27 (m, 26H), 7.23-7.13 (m, 93H), 7.06-6.94 (m, 90H), 6.87 (m, 18H), 7.79-6.72 (m, 36H), 6.66-6.54 (m, 72H), 6.39-6.37 (m, 18H), 6.23-6.15 (m, 12H), 6.02-5.90 (m, 16H), 4.08-4.0 (m, 72H, OC*H*₃), 3.93-3.63 (m, 192H, OC*H*₃), 3.38-3.15 (m, 96H, OC*H*₃), 2.55-2.01 (m, 246H), 1.75-1.55 (m, 144H), 1.35-1.27 (m, 266H), 1.03-0.84 (m, 320H), 0.71-0.38 (m, 84H), 0.11(m, 3H);

¹**H** NMR (700 MHz, 375 K, Toluene-d₈): δ = 9.36 (s, 6H), 9.30 (s, 6H), 9.26 (s, 6H), 9.09-9.08 (m, 18H), 8.99 (s, 6H), 8.73 (s, 6H), 8.52 (s, 6H), 8.39 (s, 6H), 8.36 (s, 6H), 8.33-8.25 (m, 12H), 8.09 (s, 6H), 8.06 (s, 6H), 8.02-7.98 (m, 24H), 7.90 (s, 6H), 7.75 (br (s), 6H), 7.69 (s, 6H), 7.58-7.42 (m, 54H), 7.28-7.13 (66H), 7.03-7.01 (m, 12H), 6.99-6.98 (m, 12H), 6.92-9.91 (m, 30H), 6.78-6.70 (m, 30H), 6.58-6.55 (m, 12H), 6.41 (br (s) 6H), 6.23 (br (s), 6H), 3.98 (s, 18H), 3.69-3.48 (m, 144H), 3.04-2.84 (m, 54H), 2.63-2.16 (m, 128H), 1.94 (br (s), 18H), 1.86-1.82 (m, 30H), 1.72-1.30 (m, 154H), 1.18 (t, *J* = 7 Hz, 18H), 1.15 (t, *J* = 7 Hz, 18H), 1.08 (t, *J* = 7 Hz, 18H), 1.01 (t, *J* = 7 Hz, 18H), 0.94 (t, *J* = 7 Hz, 18H), 0.93-0.87 (m, 102H), 0.83 (t, *J* = 7 Hz, 18H), 0.74 (t, *J* = 7 Hz, 18H), 0.69 (s, 6H), 0.66 (br (s), 12H), 0.42 (s, 6H), 0.38 (t, *J* = 7 Hz, 18H); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3002 (m), 2959 (m), 2936 (m), 2872 (m), 1614 (s), 1591 (s), 1491 (s), 1464 (s), 1372 (s), 1285 (m), 1140 (m), 1043 (s), 975 (w), 880 (m), 803 (m), 710 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 294, 405; **MALDI-TOF** (DCTB): m/z [M]⁺ calcd. for C₁₁₂₈H₁₁₅₂N₇₂O₇₂: 16868.91, found 16868.69. **Elemental analysis** calcd. for C₁₁₂₈H₁₁₅₂N₇₂O₇₂ 19CH₂Cl₂: C 74.53, H 6.49, N 5.46, found C 74.15, H 6.64, N 5.42.

Please note: No decent ¹³C NMR spectrum despite of recording 12000 scans on a 600 MHz NMR instrument was achieved (see Figure 73).

Synthesis of H-cube



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and terephthaldehyde **4** (4.4 mg, 0.033 mmol) in THF (6 mL) a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction 14

mixture was stirred at RT for 4 hrs. The reaction mixture was diluted to 50 mL with dichloromethane. An aqueous solution of K_2CO_3 (0.25 M, 6 mL) was added to the reaction mixture and stirred for five minutes at RT. The aqueous layer was separated and the organic layer was further washed with water (4 × 20 mL). The organic layer was collected, dried over Na₂SO₄ and concentrated at 5 mL on a rotary evaporator. The resulting solution was again diluted to total volume of 50 mL with methanol and then concentrated on a rotary evaporator. The resulting yellow solid was dissolved in a small amount of dichloromethane and precipitated by the addition of methanol (3 mL). The resulting mixture was ultrasonicated and then the precipitate was separated by centrifugation technique. The yellow solid was dried and further dissolved in a small amount of dichloromethane and reprecipitated by the addition of methanol to give 12 mg (89%) of **H-cube** as a yellow solid.

Mp: 335°C (decomposed); ¹**H NMR** (400 MHz, CD₂Cl₂): δ = 8.46 (s, 24H, *H*-9), 7.96 (s, 48H, *H*-11), 7.41 (d, ³*J* = 8.4 Hz, 24H, *H*-8), 7.20 (d, ⁴*J* = 1.6 Hz, 24H, *H*-5), 7.07 (dd, ³*J* = 8.4 Hz, ⁴*J* = 1.6 Hz, 24 H, *H*-7), 2.25 (br (t), 48H, *H*-13) 1.82 (s, 24H, *H*-12), 1.30-1.28 (m, 48H, *H*-14), 0.98 (t, ³*J* = 6.8 Hz, 72H, *H*-15); ¹³C **NMR** (151 MHz, CD₂Cl₂): δ = 159.0 (*C*-9), 151.7 (*C*-6), 149.6 (*C*-4), 146.7 (*C*-3), 139.1 (*C*-10), 129.3 (*C*-11), 124.5 (*C*-8), 120.3 (*C*-7), 116.7 (*C*-5), 73.4 (*C*-1), 67.3 (*C*-2), 41.0 (*C*-13), 20.9 (*C*-14), 15.5 (*C*-12), 15.3 (*C*-15); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3023 (w), 2956 (m), 2929 (w), 2869 (w), 1653 (w), 1622 (m), 1577 (w), 1480 (m), 1414 (w), 1295 (w), 1256 (m), 1204 (w), 1155 (w), 955 (s), 889 (w), 829 (m), 757 (m), 715 (s), 678 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 293, 363; **MALDI-TOF** (DCTB): *m/z* [M+H]⁺ calcd. for C₃₅₂H₃₃₇N₂₄: 4902.72, found 4902.19.

Synthesis of Me-cube



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-dimethyl-terephthaldehyde **5** (5.2 mg, 0.032 mmol) in deuterated chloroform (4 mL) in a screw-cap 8 mL glass vial, a catalytic

amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred at RT for 18 hours. The reaction mixture was concentrated to 1 mL by rotary evaporation and methanol (16 mL) was added. The precipitate was separated by the centrifugation technique. The precipitate was repeatedly dried and dissolved in dichloromethane (3 x 1 mL) and reprecipitated by the addition of methanol to give 11.8 mg (84%) of **Me-cube** as a yellow solid.

Mp: 355°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 8.66 (s, 24H, H-9), 7.89 (s, 24H, H-12), 7.39 (d, ${}^{3}J$ = 7.8 Hz, 24H, H-8), 7.17 (d, ${}^{3}J$ = 1.2 Hz, 24H, H-5), 7.00 (dd, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 1.8 Hz, 24 H, H-7), 2.54 (s, 24H, H-13), 2.23 (br (t), 48H, H-15) 1.70 (s, 24H, H-14), 1.29-1.27 (m, 48H, H-16), 0.97 (t, ${}^{3}J$ = 7.2 Hz, 72H, H-17); ¹³C NMR (151 MHz, CD₂Cl₂): δ = 158.1, 152.5, 149.6, 146.3, 136.7, 136.5, 130.1, 124.5, 119.5, 117.2, 73.3, 67.3, 41.1, 20.9, 18.9, 15.5, 15.3; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2956 (s), 2925 (m), 2869 (m), 1728 (w), 1697 (m), 1618 (s), 1593 (s), 1481(s), 1456 (w), 1402 (w), 1375 (w), 1272 (m), 1186 (m), 1149 (w), 970 (w), 883 (m), 802 (m), 734 (m), 702 (w), 648 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 297, 366; **MALDI-TOF** (DCTB): m/z [M]⁺ calcd. for C₃₇₆H₃₈₄N₂₄: 5239.09, found 5239.96.

Synthesis of Et-cube



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-diethyl-terephthaldehyde **6** (6.1 mg, 0.032 mmol) in deuterated chloroform (4 mL) a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred at RT for 24 hours. The reaction mixture was diluted to a total volume of 20 mL with acetonitrile and the resulting solution was concentrated to 5 mL under reduced pressure. The yellow precipitate was separated by centrifugation technique. The collected yellow precipitate was repeatedly suspended in acetonitrile (3 x 5 mL) sonicated (20 seconds) and separated by the centrifuge technique afforded 11.2 mg (75%) of **Et-cube** as a yellow solid.

Mp: >350°C; ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 8.68 (s, 24H, *H*-9), 7.96 (s, 24H, *H*-12), 7.40 (d, ${}^{3}J$ = 7.8 Hz, 48H, *H*-8), 7.18 (d, ${}^{3}J$ = 1.2 Hz, 24H, *H*-5), 6.99 (dd, ${}^{3}J$ = 7.2 Hz, ${}^{4}J$ = 1.2 Hz, 24 H, *H*-7), 2.92 (q, ${}^{3}J$ = 7.2 Hz, 48H, *H*-13), 2.23 (br (t), 48H, *H*-16), 1.70 (s, 24H, *H*-15), 1.29-1.27 (m, 48H, *H*-17), 1.23 (t, ${}^{3}J$ = 7.2 Hz, 72H, *H*-14);), 0.98 (t, ${}^{3}J$ = 7.2 Hz, 72H, *H*-18); ¹³C **NMR** (151 MHz, CD₂Cl₂): 157.9, 152.7, 149.9, 146.3, 143.2, 136.0, 128.9, 124.5, 119.3, 117.4, 73.4, 67.4, 41.1, 25.8, 20.9, 16.8, 15.5, 15.3; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2960 (s), 2931 (m), 2871 (m), 1699 (w), 1618 (s), 1593 (s), 1548 (w), 1481(s), 1454 (m), 1411 (w), 1375 (m), 1299 (w), 1271 (w), 1184 (m), 1151 (m), 1060 (w), 970 (m), 906 (s), 898 (s), 817 (s), 802 (s), 734 (w), 638 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 298, 362; **MALDI-TOF** (DCTB): m/z [M]⁺ calcd. for C₄₀₀H₄₃₂N₂₄: 5576.48, found 5576.76.

Synthesis of (Et-cube)2



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-diethyl-terephthaldehyde **6** (6.1 mg, 0.032 mmol) in THF (2 mL), a catalytic amount of TFA (0.2 μ L, 0.0026 mmol) was added and the reaction mixture was stirred at RT for 4½ days. Afterward, the crude reaction mixture was concentrated under reduced pressure and the resulting solid was immediately dissolved in chloroform and purified by rGPC (CHCl₃, 40°C, 3.5 mL/min) to give 5.2 mg (35%) of (**Etcube**)₂ as a yellow solid.

Mp: 240°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): $\delta = 8.77$ (s, 6H, *H*C=N), 8.69 (s, 6H, HC=N), 8.67 (s, 12H, HC=N), 8.62 (s, 6H, HC=N), 8.59 (s, 6H, HC=N), 8.33 (s, 6H, HC=N), 8.25 (s, 6H, HC=N), 8.18-8.17 (12H, Ar-H), 7.97 (s, 6H, Ar-H), 7.96 (s, 6H, Ar-H), 7.94 (s, 6H, Ar-H), 7.93 (s, 6H, Ar-H), 7.72 (s, 6H, Ar-H), 7.48 (s, 6H, Ar-H), 7.44-7.41 (12H, TBTQ-Ar-H), 7.39-7.32 (18H, TBTQ-Ar-H), 7.23 (s, 6H, TBTQ-Ar-H), 7.22 (s, 6H, TBTQ-Ar-H), 7.20 (s, 6H, TBTQ-Ar-H), 7.18 (s, 6H, TBTQ-Ar-H), 7.15 (s, 6H, TBTQ-Ar-*H*), 7.10 (s, 6H, TBTQ-Ar-*H*), 7.02-7.00 (18H, TBTQ-Ar-*H*), 6.96 (d, ${}^{3}J$ = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.92 (s, 6H, TBTQ-Ar-*H*), 6.89 (d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ Ar-*H*), 6.88 (s, 6H, TBTQ-Ar-*H*), 6.79 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-*H*), 6.67 (d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ Ar-*H*), 6.62 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-H), 6.49 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-H), 6.33 (d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ Ar-*H*), 5.98 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 3.10-2.81 (m, 82H, -CH₂CH₃), 2.54-242 (m, 14H, -CH2CH3), 2.36-2.04 (m, 84H, -CH2CH2CH3), 1.84-1.66 (m, 48H), 1.60 (q, ${}^{3}J = 7.2$ Hz, 12H, -CH₂CH₂CH₃), 1.32 (t, ${}^{3}J = 7.8$ Hz, 18H, -CH₂CH₂CH₃), 1.29-1.24 (m, 114H), 1.18 (t, ${}^{3}J = 7.2$ Hz, 18H, -CH₂CH₃), 1.14 (t, ${}^{3}J = 7.2$ Hz, 18H, -CH₂CH₃), 1.05 (t, {}^{3}J = 7.2 7.2 Hz, 18H, $-CH_2CH_3$, 1.02-0.94 (m, 108H), 0.89-0.81 (m, 36H), 0.74 (t, ${}^{3}J = 7.8$ Hz, 36H, -CH₂CH₃), 0.49 (t, ${}^{3}J = 7.2$ Hz, 18H), -0.27 (t, ${}^{3}J = 7.8$ Hz, 18H, -CH₂CH₂CH₃); ${}^{13}C$ NMR (151 MHz, CDCl₃): 158.6, 158.3, 157.9, 157.8, 157.7, 157.6, 157.3, 154.1, 153.6, 153.0, 152.6, 152.6, 152.6, 152.5, 152.3, 152.3, 150.5, 149.8, 149.7, 149.7, 149.5, 149.4, 146.5, 146.4, 146.0, 146.0, 145.8, 145.6, 144.7, 143.3, 143.2, 143.0, 142.9, 142.7, 136.9, 136.1, 136.0, 135.9, 135.7, 135.6, 129.0, 128.8, 128.5, 127.9, 124.4, 124.2, 124.2, 119.2, 119.0, 118.1, 117.8, 117.5, 117.3, 116.7, 115.8, 73.5, 73.2, 73.1, 67.5, 67.4, 67.3, 67.3, 67.2, 67.0, 66.9, 41.7, 41.5, 41.5, 41.5, 41.1, 41.0, 40.9, 26.0, 26.0, 25.8, 25.7, 25.6, 24.8, 21.4, 21.1, 21.0, 20.9, 20.4, 16.9, 16.8, 16.7, 16.7, 16.4, 16.3, 16.3, 15.5, 15.4, 15.3, 15.3, 15.2, 15.1, 15.0; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2961 (s), 2930 (m), 2872 (m), 1618 (s), 1593 (s), 1549 (w), 1481 (s), 1456 (m), 1414 (m), 1375 (m), 1296 (w), 1271 (w), 1240 (w), 1184 (m), 1150 (w), 1111 (w), 1101(w), 1061 (w), 970 (m), 953 (w), 910 (m), 878 (m), 820 (m), 802 (m), 781 (w), 712 (w), 677 (w), 642 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 297, 361; **MALDI-TOF** (DCTB): m/z [M+H]⁺ calcd. for C₈₀₀H₈₆₅N₄₈: 11151.94, found 11152.45.

Synthesis of OEt-cube



To a solution of TBTQ **1** (5 mg, 0.01 mmol) and 2,5-diethoxy-terephthaldehyde **7** (3.6 mg, 0.016 mmol) in deuterated chloroform (5 mL), a catalytic amount of TFA (0.1 μ L, 0.001 mmol) was added and the reaction mixture was stirred for 4 hours at RT. Afterward, the crude reaction mixture was purified by r-GPC (CHCl₃) and then the collected fractions (25 mL) were diluted to a total volume of 50 mL with acetonitrile. The solvents were removed under reduced pressure to a volume of 10 mL and diluted again to 50 mL with acetonitrile before solvents were completely removed under vacuum to give 6.7 mg (84%) of **OEt-cube** as a yellow solid.

Mp: 345°C (decomposed); ¹**H NMR** (600 MHz, CDCl₃): $\delta = 8.83$ (s, 24H), 7.66 (s, 24H), 7.37 (d, ³*J* = 7.8 Hz, 24H), 7.19 (s, 24H), 6.98 (d, ³*J* = 7.8 Hz, 24 H), 4.14 (m, 48H), 2.21 (br (t), 48H), 1.67 (s, 24H), 1.38 (t, ³*J* = 7.2 Hz, 72H), 1.25 (br (m), 48H), 0.95 (t, ³*J* = 7.2 Hz, 72H) ; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2956 (m), 2923 m), 2869 (w), 1647 (w), 1616 (w), 1558 (w), 1541 (m), 1529 (m), 1508 (m), 1488 (s), 1473 (s), 1456 (m), 1427 (s), 1396 (w), 1361 (w), 1286 (w), 1203 (s), 1110 (w), 1045 (s), 974 (w), 881 (w); **UV/Vis** (CHCl₃): λ_{max} 19 (nm) = 296, 407; **MALDI-TOF** (DCTB): m/z [M+H]⁺ calcd. for C₄₀₀H₄₃₂N₂₄O₂₄: 5959.35, found 5959.77.1

Please note: OEt-cube is also forming catenane at higher concentration like **OMe-cube**. Therefore, no ¹³C NMR spectrum was recorded.

Synthesis of (OEt-cube)₂



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-diethoxy-terephthaldehyde **7** (7.2 mg, 0.032 mmol) in deuterated chloroform (2 mL), a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred at RT for 7 days. Afterward, the crude reaction mixture was concentrated to 1 mL and purified by r-GPC (CHCl₃, 40°C, 5 mL/min) to give 4 mg (25%) of (**OEt-cube**)₂ as yellow solid.

Mp: 320°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): $\delta = 8.95$ (s, 6H, *H*C=N), 8.92 (s, 6H, *H*C=N), 8.85 (s, 6H, *H*C=N), 8.81 (12H, *H*C=N), 8.78 (s, 6H, *H*C=N), 8.68 (s, 6H, *H*C=N), 8.41 (s, 6H, *H*C=N), 8.17 (s, 6H, Ar-*H*), 7.84 (s, 6H, Ar-*H*), 7.72 (s, 6H, Ar-*H*), 7.70 (s, 6H, Ar-*H*), 7.65 (s, 6H, Ar-*H*), 7.64 (s, 6H, Ar-*H*), 7.62 (s, 6H, Ar-*H*), 7.52 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 7.46 (s, 6H, TBTQ-Ar-*H*), 7.39-7.37 (12H, Ar-*H*), 7.33 (d, ³*J* = 8.4 Hz, 6H, TBTQ Ar-*H*), 7.30 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 7.02-6.93 (30H, TBTQ-Ar-*H*), 6.87 (s, 6H, TBTQ-Ar-*H*), 6.80 (d, ³*J* = 8.4 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, TBTQ-Ar-*H*), 6.57-6.55 (m, 12H, TBTQ-Ar-*H*), 6.47 (d, ³*J* = 7.8 Hz, 6H, TBTQ Ar-*H*), 6.71 (s, 6H, OCH₂CH₃), 2.23-2.12 (m, 72H, -CH₂CH₂CH₃), 1.95 (br (t), 12H, -CH₂CH₂CH₃), 1.77-1.64 (m, 60H, -CH₂CH₂CH₃ & TBTQ-CH₃), 1.51 (t, ³*J* = 7.2 Hz, 18H, -OCH₂CH₃), 1.39-1.26 (m, 138H, -OCH₂CH₃ & -CH₂CH₂CH₃), 0.76-0.74 (m, 30H, -CH₂CH₂CH₃), 0.76-0.74 (m, 30

OCH₂*CH*₃ & -CH₂C*H*₂CH₃), 0.49 (t, ${}^{3}J$ = 7.2 Hz,18H, -CH₂CH₂C*H*₃); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2956 (s), 2923 (s), 2854 (m), 1728 (w), 1616 (m), 1593 (m), 1483 (s), 1427 (s), 1393 (m), 1373 (m), 1280 (m), 1203 (s), 1141 (w), 1110 (m), 1047 (s), 972 (w), 883 (w), 804 (w), 707 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 297, 400; **MALDI-TOF** (DCTB): m/z [M+H]⁺ calcd. for C₈₀₀H₈₆₅N₄₈O₄₈: 11920.70, found 11920.88.

Please note: No decent ¹³C NMR spectrum despite of recording 4096 scans on a 600 MHz NMR instrument was achieved.

Synthesis of OⁱPr-cube



To a solution of TBTQ **1** (10 mg, 0.0215 mmol) and 2,5-isopropoxy-terephthaldehyde **8** (8.1 mg, 0.032 mmol) in chloroform (deuterated, 2 mL), a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred at RT for 24 hours. The reaction mixture was concentrated to 1 mL under reduced pressure and then purified by r-GPC (chloroform, 40°C, 3.5 mL/min) to give 16.1 mg (95%) of **OⁱPr-cube** as yellow solid.

Mp: 328°C (decomposed); ¹**H NMR** (600 MHz, CDCl₃): δ = 8.80 (s, 24H, *H*-9), 7.69 (s, 24H, *H*-12), 7.37 (d, ³*J* = 8.4 Hz, 24H, *H*-8), 7.18 (s, 24H, *H*-5), 6.97 (dd, ³*J* = 7.8 Hz, ⁴*J* = 1.2 Hz, 24 H, *H*-7), 4.67 (m, 24, *H*-13), 2.21 (br (t), 48H, *H*-17), 1.67 (s, 24H, *H*-16), 1.31 (d, ³*J* = 6 Hz, 72H, *H*-14), 1.29 (d, ³*J* = 6 Hz, 72H, *H*-15), 1.26 (br (m), 48H, *H*-18), 0.96 (t, ³*J* = 7.2 Hz, 72H, *H*-19); ¹³**C NMR** (151 MHz, CDCl₃): 155.8, 152.4, 152.1, 149.2, 146.1, 129.8, 124.2, 119.2, 117.5, 113.0, 73.1, 72.0, 67.0, 41.1, 22.33, 22.26, 20.7, 15.4, 15.4; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2972 (s), 2961 (s), 2924 (s), 2872 (m), 2818 (w), 2359 (w), 2341(w), 2330 (w), 1616 (m), 1591 (m), 1489 (s), 1479 (s), 1423 (s), 1373 (m), 1335(w), 1277 (m), 1196 (m), 1134 (m), 1109 (s), 1074 (m), 1067 (m), 1040 (w), 972 (m), 935 (m), 910 (m), 891 (w), 872 (w), 843 (w), 824 (w), 802(w),793 (w), 735 (m), 650 (w), 608 (m), 607.55 (m); **UV/Vis** (CHCl₃): λ_{max} (nm) = 297, 351, 405; **MALDI-TOF** (DCTB): *m*/*z* [M+H]⁺ calcd. for C₄₂₄H₄₈₁N₂₄O₂₄: 6296.73, found 6297.58.

Synthesis of SMe-cube



To a solution of TBTQ **1** (5.3 mg, 0.011 mmol) and 2,5-bis(methylthio)terephthalaldehyde) **9** (3.6 mg, 0.016 mmol) in 1,1,2,2 tetrachloroethane (anhydrous, 2 mL), a catalytic amount of TFA (0.1 μ L, 0.001 mmol) was added and the reaction mixture was stirred at RT for 2 hours. The saturated solution of aq. NaHCO₃ (2 mL) was added to the reaction mixture and it was further stirred at RT for 10 minutes. The organic layer was separated, washed with water (4 x 3 mL) and precipitated by adding methanol into the organic layer (6 mL), and left at RT for 4 hours. The yellow precipitate was separated by centrifugation technique, repeatedly washed with methanol (4 x 4 mL) and dried to give 6.4 mg (80%) of **SMe-cube** as a yellow solid.

Mp: 347°C (decomposed); ¹**H NMR** (700 MHz, Cl₂CDCDCl₂): $\delta = 8.87$ (s, 24H), 8.01 (s, 24H), 7.40 (d, J = 7.0 Hz, 24H), 7.23 (s, 24 H), 7.08 (d, J = 7.7 Hz, 24 H), 2.53 (s, 72H), 2.24 (br (t), 48H) 1.69 (s, 24H), 1.31 (br (m), 48H), 0.99 (t, J = 6.3 Hz, 72H; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2959 (m), 2922 (w), 2869 (w), 1616 (m), 1591 (m), 1481 (s), 1433 (m), 1359 (m), 1315 (w), 1267 (w), 1201 (w), 1143 (w), 1099 (m), 1016 (w), 968 (m), 879 (m), 817 (w), 790 (s), 752 (w), 736 (s), 713 (w), 644 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 274, 369; **TIMS-TOF Flex** (DCTB): m/z [M+Na+4(O)]⁺ calcd. for C₃₇₆H₃₈₄N₂₄S₂₄O₄Na: 6095.39, found 6095.41. *Note:*

1. A decent ¹³C NMR spectrum was not available, even by recording 8192 scans on a 700 MHz NMR instrument (see Figure 185). It was also observed that the cage shows decomposition over time in deuterated 1,1,2,2 tetrachloroethane due to the acidic nature of solvent.

2. The mass spectrum shows several additional peaks due to oxidation of sulphur from the cage (See Figure 338).

Synthesis of (SMe-cube)₂



To a solution of TBTQ **1** (10 mg, 0.022 mmol) and 2,5-bis(methylthio)terephthalaldehyde **9** (7.3 mg, 0.032 mmol) in deuterated chloroform (2 mL), a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred at RT for 3 days. Afterward, the crude reaction mixture was concentrated under reduced pressure and the resulting solid was immediately dissolved in chloroform and purified by r-GPC (CHCl₃, 40°C, 3.5 mL/min) to give 10.7 mg (66%) of (**SMe-cube**)₂ as a yellow solid.

Mp: 332°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): $\delta = 9.02$ (s, 6H, *H*C=N), 8.85-8.84 (24H, HC=N), 8.80 (s, 6H, HC=N), 8.59 (s, 6H, HC=N), 8.46 (s, 6H, HC=N), 8.25 (s, 6H, Ar-H), 8.20 (s, 6H, Ar-H), 8.10 (s, 6H, Ar-H), 8.07 (s, 6H, Ar-H), 8.02 (s, 6H, Ar-H), 7.98 (s, 6H, Ar-H), 7.80 (s, 6H, Ar-H), 7.62 (s, 6H, Ar-H), 7.56 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ Ar-H), 7.54 (s, 6H, TBTQ Ar-H), 7.41-7.36 (24H, TBTQ Ar-H), 7.27 (s, 6H, TBTQ-Ar-H), 7.23 (s, 6H, TBTQ-Ar-H), 7.20 (s, 6H, TBTQ-Ar-H), 7.12 (s, 12H, TBTQ-Ar-H), 7.05-7.00 (m, 36H, TBTQ-Ar-*H*), 6.93 (d, ${}^{3}J = 8.4$ Hz, 6H, TBTQ-Ar-*H*), 6.81 (s, 6H, TBTQ-Ar-*H*), 6.66 (d, ${}^{3}J =$ 7.2 Hz, 6H, TBTQ-Ar-H), 6.61 (d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ-Ar-H), 6.55(d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ-Ar-H), 6.33 (brs, 6H, TBTQ-Ar-H), 6.28 (d, ${}^{3}J = 7.8$ Hz, 6H, TBTQ-Ar-H), 2.62 (s, 18H, SCH₃), 2.51 (s, 18H, SCH₃), 2.46 (s, 18H, SCH₃), 2.42 (s, 18H, SCH₃), 2.38 (s, 18H, SCH₃), 2.27-2.10 (m, 102H, SCH₃ & -CH₂CH₂CH₃), 1.96 (s, 18H, SCH₃), 1.76-1.67 (m, 60H, -CH2CH2CH3 & -CH3), 1.30-1.26 (m, 72H, -CH2CH2CH3), 1.12-1.10 (m, 30H, SCH3 & - $CH_2CH_2CH_3$), 0.99-0.95 (m, 108H, -CH₂CH₂CH₃), 0.84 (t, ${}^{3}J = 7.2$ Hz, 9H, -CH₂CH₂CH₃), 0.72-0.69 (m, 12H, -CH₂CH₂CH₃), 0.58 (t, ${}^{3}J = 7.2$ Hz, 9H, -CH₂CH₂CH₃); ${}^{13}C$ NMR (151 MHz, CDCl₃): 158.0, 157.5, 157.3, 156.8, 156.6, 156.5, 156.2, 153.3, 152.8, 152.1, 151.7, 151.6, 150.8, 149.8, 149.7, 149.6, 149.6, 149.4, 146.9, 146.8, 146.5, 146.1, 146.0, 145.8, 145.7, 138.9, 138.8, 138.3, 138.1, 138.0, 137.9, 137.8, 137.6137.5, 136.9, 136.8, 136.7, 136.5,

136.2, 136.1, 129.5, 128.0, 127.9, 127.8, 127.4, 127.3, 126.4, 126.3, 125.0, 124.9, 124.5, 124.4, 124.3, 124.2, 124.1, 120.4, 120.3, 119.7, 118.8, 188.8, 118.6, 118.5, 118.4, 117.5, 117.5, 117.2, 73.4, 73.2, 67.6, 67.5, 67.4, 67.4, 67.3, 67.1, 67.1, 67.0, 41.9, 41.8, 41.6, 41.4, 41.4, 41.3, 41.2, 41.2, 41.2, 41.1, 41.1, 41.0, 40.9, 40.6, 40.5, 21.5, 21.2, 21.0, 20.9, 20.8, 20.3, 17.8, 17.5, 17.4, 17.1, 17.0, 16.7, 16.5, 16.5, 15.5, 15.4, 15.3, 15.3, 15.3, 15.2, 15.1; FT-**IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3684 (w), 3674 (w), 2988 (m), 2972 (m), 2901 (m), 1452 (w), 1406 (w), 1394 (w), 1383 (w), 1259 (w), 1242 (w), 1215 (m), 1074 (s), 1067 (s), 1055 (s), 1028 (m), 928 (w), 893 (w), 879 (w), 870 (w), 748 (s), 669 (m), 629 (w), 611 (w); UV/Vis (CH₂Cl₂): λ_{max} (nm) = 277, 369; MALDI-TOF (DCTB): m/z [M+H]⁺ calcd. for calcd. C752H769N48S48: 12017.84, found 12017.64. Elemental analysis for C₇₈₁H₈₂₆N₄₈S48·29CH₂Cl₂: C 64.78, H 5.75, N 4.64, found C 64.49, H 5.64, N 4.40.

Synthesis of (SMe-cube)₃



To a solution of TBTQ **1** (15.4 mg, 0.0331 mmol) and 2,5-bis(methylthio)terephthalaldehyde) (10.9 mg, 0.0482 mmol) in deuterated dichloromethane (3 mL) in a screw-capped 8 mL glass vial a catalytic TFA (0.3 μ L, 0.0038 mmol) was added and the reaction mixture was stirred at 80°C for 4 days. After cooling down to room temperature, the crude reaction mixture was stirred with aq. K₂CO₃ solution (0.25 M, 3 × 2 mL) for 10 minutes, the organic layer was separated and dried over Na₂SO₄. The solution was concentrated under reduced pressure and the resulting yellow solid was immediately dissolved in dichloromethane and purified by rGPC (THF, 40°C, 3 mL/min) to give 14 mg (58%) of (**SMe-cube**)₃ as a yellow solid. (Note: BHT from the cage was removed by pentane wash).

Mp: 299°C (decomposed); ¹**H NMR** (700 MHz, 295 K, CD₂Cl₂): $\delta = 9.91-8.76$ (m, 48H), 8.46-7.99 (m, 72H), 7.71-7.21 (m, 90H), 7.11-6.26 (m, 150H), 2.69-2.11 (m, 264H), 1.76-

0.28 (m, 456H); ¹³C NMR (176 MHz, 295 K, CD₂Cl₂): $\delta = 158.2$, 157.9, 157.5, 157.0, 156.6, 156.3, 156.0, 153.4, 152.6, 152.0, 151.5, 150.1, 149.9, 149.8, 149.5, 149.1, 147.2, 146.8, 145.9, 145.6, 145.3, 144.8, 139.8, 138.2, 138.1, 138.0, 137.4, 137.0, 136.8, 136.6, 136.5, 135.8, 130.5, 128.0, 127.5, 127.3, 126.4, 125.6, 125.0, 124.8, 124.6, 124.4, 124.2, 123.9, 120.5, 120.0, 119.6, 118.7, 118.1, 117.0, 115.9, 115.0, 73.31, 73.26, 73.2, 72.83, 72.79, 68.1, 67.6, 67.5, 67.4, 67.3, 67.2, 67.1, 66.8, 42.1, 41.6, 41.4, 41.1, 40.9, 40.1, 21.5, 21.3, 21.0, 20.95, 20.90, 20.8, 20.7, 20.2, 17,9, 17.5, 17.4, 17.2, 17.1, 16.4, 15.54, 15.52, 15.39, 15.36, 15.33, 15.30, 15.27, 15.2, 15.03, 15.01;

¹**H** NMR (700 MHz, 410 K, $C_2D_2Cl_4$): $\delta = 9.11-9.09$ (m, 18H), 9.03-9.01 (m, 12H), 8.96 (s, 6H), 8.90 (s, 6H), 8.88 (s, 6H), 8.53 (s, 6H), 8.50 (s, 6H), 8.40 (s, 6H), 8.37-8.36 (m, 12H), 8.31 (s, 6H), 8.29 (s, 6H), 8.282 (s, 6H), 8.275 (s, 6H), 8.20 (s, 6H), 8.17 (s, 6H), 8.16 (s, 6H), 7.79 (s, 6H), 7.74 (br (s), 6H), 7.71 (s, 6H), 7.61 (d, J = 8.4 Hz, 6 H), 7.59-7.57 (m, 12H), 7.47 (d, J = 8.4 Hz, 6 H), 7.42 (d, J = 7.7 Hz, 6 H), 7.40-7.38 (m, 18H), 7.34 (s, 6H), 7.31-7.30 (m, 12H), 7.26 (s, 6H), 7.18 (s, 6H), 7.15-7.13 (m, 12H), 7.10 (d, J = 8.4 Hz, 6 H), 7.04 (br (s), 18H), 7.00 (d, J = 7.7 Hz, 6 H), 6.96-6.89 (m, 30H), 6.82-6.78 (m, 36H), 6.64 (d, J =7.7 Hz, 6 H), 6.52-6.31 (m, 24H), 6.22 (d, J = 8.4 Hz, 6 H), 2.79 (s, 18H), 2.73 (s, 18H), 2.64 (br (s) 12H), 2.60 (s, 18H), 2.55 (s, 18H), 2.49 (s, 18H), 2.48 (s, 18H), 2.39-2.19 (m, 132H), 1.99-1.96 (m, 24H), 1.81-1.74 (m, 86H), 1.52-1.25 (m, 160H), 1.15 (t, J = 7.7 Hz, 18H), 1.11-1.06 (m, 108H), 0.96 (t, J = 7.7 Hz, 18H), 0.92 (t, J = 7.0 Hz, 18H), 0.89-0.87 (m, 30H), 0.72 (t, J = 7.7 Hz, 18H), 0.69-0.59 (m, 36H), 0.42 (s, 6H), 0.34 (t, J = 7.0 Hz, 18H); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2956 (m), 2922 (w), 2869 (w), 1616 (m), 1591 (m), 1483 (m), 1433 (m), 1363 (w), 1263 (m), 1143 (w), 1099 (m), 968 (m), 893 (m), 802 (m), 734 (s), 698 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 276, 366; **MALDI-TOF** (DCTB): m/z [M+Na+O]⁺ calcd. for C₁₁₂₈H₁₁₅₂N₇₂S₇₂ONa: 18064.24, found 18063.63.

Synthesis of ¹⁵N labelled *(SMe-cube)₃



To a solution of TBTQ **14** (10.3 mg, 0.022 mmol) and 2,5-bis(methylthio)terephthalaldehyde) (7.2 mg, 0.032 mmol) in dichloromethane (deuterated, 2 mL) in a screw-capped 8 mL glass vial a catalytic TFA (0.2 μ L, 0.003 mmol,) was added and the reaction mixture was stirred at 80°C for 4 days. After cooling down to room temperature, the crude reaction mixture was stirred with aq. K₂CO₃ solution (0.25 M, 3 × 2 mL) for 10 minutes. The organic layer was separated, dried over Na₂SO₄ and concentrated under reduced pressure. The resulting yellow solid was immediately dissolved in dichloromethane and purified by rGPC (THF, 40°C, 3 mL/min) to give 8.8 mg (55%) of *(**SMe-cube**)₃ as a yellow solid. (Note: BHT from the cage was removed by pentane wash).

Mp: 332 °C (decomposed); ¹**H NMR** (700 MHz, 380K Toluene-d₈): 9.50 (br (d), 12H), 9.37 (br (d), 6H), 9.23 (br (d), 6H), 9.18 (d, J = 2.8 Hz, 6H), 9.15 (d, J = 2.8 Hz, 6H), 8.99 (br (d), 6H), 8.95 (br (d), 6H), 8.75-8.73 (m, 12H), 8.54-8.48 (m, 42H), 8.40 (s, 6H), 8.37 (s, 6H), 8.12 (s, 6H), 8.06 (d, J = 7.7 Hz, 6H), 8.02 (s, 6H), 7.92-7.88 (m, 18H), 7.77 (s, 6H), 7.69 (s, 6H), 7.57 (s, 6H), 7.55 (s, 6H), 7.42 (s, 6H), 7.40-7.20 (m, 54H), 7.15-7.09 (m, 24H), 7.06-6.99 (m, 30H), 6.93-6.85 (m, 24H), 6.76-6.69 (m, 24H), 6.53-6.44 (m, 18H), 2.96-2.72 (m, 262H), 2.00-1.30 (m, 270H), 1.23 (t, J = 6.3 Hz, 18H), 1.18 (t, J = 7 Hz, 18H), 1.15 (t, J = 7 Hz, 18H), 1.04-0.92 (m, 140H), 0.82-0.58 (m, 48H), 0.43 (t, J = 5.6 Hz, 18H); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2956 (m), 2920 (m), 2869 (w), 1579 (m), 1481 (s), 1433 (m), 1359 (m), 1315 (w), 1143 (w), 1099 (m), 968 (m), 879 (m), 802 (s), 733 (m), 653 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 273, 368; **MALDI-TOF** (DCTB): [M+H+8(O)]⁺ calcd. for C₁₁₂₈H₁₁₅₃¹⁵N₇₂S₇₂O₈: 18226.01 found 18226.83.

Synthesis of S^tBu-cube



To a solution of TBTQ 1 (10 mg, 0.022 mmol) and 2,5-bis(tert-butylthio)terephthalaldehyde 10 (10 mg, 0.032 mmol) in chloroform (2 mL), a catalytic amount of TFA (0.2 μ L, 0.003 mmol) was added and the reaction mixture was stirred for 18 hours at RT. The solvent was evaporated and the resulting solid was dissolved in DCM (0.3 mL). Methanol (4 mL) was added and the resulting yellow precipitate was separated by centrifugation technique. The precipitate was repeatedly dried and dissolved in a small amount of dichloromethane (3 x 0.3 mL) and reprecipitated by the addition of methanol to give 14 mg (75%) of S^tBu-cube as a yellow solid.

Mp: 322°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 9.27 (s, 24H, *H*-9), 8.47 (s, 24H, *H*-12), 7.44 (d, ³*J* = 8.4 Hz, 24H, *H*-8), 7.25 (d, ³*J* = 1.2 Hz, 24H, *H*-5), 7.09 (dd, ³*J* = 7.8 Hz, ⁴*J* = 1.2 Hz, 24 H, *H*-7), 2.25-2.24 (m, 48H, *H*-15), 1.71 (s, 24H, *H*-14), 1.31-1.24 (m, 264H, *H*-13 & *H*-16), 0.98 (t, ³*J* = 7.2 Hz, 72H, *H*-17); ¹³**C NMR** (151 MHz, CD₂Cl₂): δ = 158.9 (*C*-9), 151.8 (*C*-11), 149.6 (*C*-4), 146.8 (*C*-6), 142.2 (*C*-3), 138.5 (*C*-8), 136.2 (*C*-10), 124.5 (*C*-12), 119.8 (*C*-7), 117.9 (*C*-5), 73.4 (*C*-1), 67.3 (*C*-2), 48.3 (*C*-13a), 41.1 (*C*-15), 31.3 (*C*-13), 20.9 (*C*-18), 15.5 (*C*-14), 15.3 (*C*-17); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3057 (w), 3024 (w), 2959 (m), 2926 (m), 2870 (w), 1628 (w) 1591 (s), 1578 (m), 1553 (m), 1526 (w), 1485 (m), 1462 (m), 1441 (m), 1391 (m), 1364 (m), 1348 (w), 1308 (s), 1261 (m), 1238 (m), 1217 (m), 1163 (s), 1150 (s), 1132 (s), 1101 (s),1063 (s), 1030 (m), 1001 (m), 986 (m), 978 (m), 922 (m), 903 (s), 876 (s), 833 (s), 825 (s), 798 (s), 787 (m), 760 (s), 735 (s), 698 (s), 687 (s), 665 (s), 631 (s), 608 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 265, 295, 363; **MALDI-TOF** (DCTB): *m*/*z* [M-C₄H₉]⁺ calcd. for C₄₄₄H₅₁₉N₂₄S₂₄: 6960.48, found 6960.22.

Synthesis of Br-cube



To a solution of TBTQ **1** (5 mg, 0.01 mmol) and 2,5-bromo-terephthaldehyde **11** (4.7 mg, 0.016 mmol) in chloroform (1 mL), a catalytic amount of TFA (0.1 μ L, 0.001 mmol) was added and the reaction mixture was stirred at RT for 18 hours. The reaction mixture was concentrated to ~0.2 mL and acetonitrile (4 mL) was added. The resulting precipitate was separated by the centrifugation technique. The precipitate was repeatedly dried and dissolved in a small amount of dichloromethane (3 x 0.2 mL) and reprecipitated by the addition of acetonitrile afforded 8.4 mg (92%) of **Br-cube** as a yellow solid.

Mp: 332°C (decomposed); ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 8.76 (s, 24H, *H*-9), 8.45 (s, 24H, *H*-12), 7.45 (d, ³*J* = 8.4 Hz, 48H, *H*-8), 7.28 (d, ³*J* = 0.6 Hz, 24H, *H*-5), 7.11 (dd, ³*J* = 8.4 Hz, ⁴*J* = 1.2 Hz, 24 H, *H*-7), 2.24 (br (t), 48H, *H*-14) 1.70 (s, 24H, *H*-13), 1.31-1.25 (m, 48H, *H*-15), 0.98 (t, ³*J* = 7.2 Hz, 72H, *H*-16); ¹³C **NMR** (151 MHz, CD₂Cl₂): δ = 156.6 (*C*-9), 150.8 (*C*-6), 149.6 (*C*-4), 147.5 (*C*-3), 138.0 (*C*-11), 133.3 (*C*-12), 125.0 (*C*-10), 124.7 (*C*-8), 119.1 (*C*-7), 118.9 (*C*-5), 73.4 (*C*-1), 67.4 (*C*-2), 41.1 (*C*-14), 20.9 (*C*-15), 15.5 (*C*-13), 15.2 (*C*-16); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 3077 (w), 3020 (w), 2956 (s), 2928 (m), 2869 (m), 1734 (w), 1696 (w), 1616 (m), 1593 (m), 1482 (s), 1456 (m), 1359 (s), 1260 (m), 1145 (w), 903 (w), 806 (w), 764 (s), 750 (s), 699 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 269, 300, 378; **MALDI-TOF** (DCTB): *m*/*z* [M]⁺ calcd. for C₃₅₂H₃₁₂N₂₄Br₂₄: 6795.54, found 6795.56.

Synthesis H/OMe-cube



To a solution of TBTQ **1** (5.4 mg, 0.012 mmol) and 2-methoxy-terephthaldehyde **12** (2.6 mg, 0.016 mmol) in deuterated chloroform (1 mL) in a screw-capped 8 mL glass vial, a catalytic amount of TFA (0.1 μ L, 0.001 mmol) was added and the reaction mixture was stirred at RT for 3 hours. Solid NaHCO₃ (200 mg) was added and the reaction mixture was further stirred at RT for 10 minutes. The organic layer was separated by decantation and concentrated to a total volume of 1 mL and then purified by recycling gel permeation chromatography (rGPC) (THF, 40°C, 5 mL/min) to give 2.4 mg (35%) of **Mono-OMe-cube** as a yellow solid.

Mp: >350°C; ¹**H NMR** (600 MHz, C₂D₂Cl₄): δ = 8.85-8.84 (m, 12H), 8.40-8.39 (m, 12H), 8.16-8.15 (m, 12H), 7.52 (br (s), 12H), 7.46-7.44 (m, 12H), 7.40-7.37 (m, 24H), 7.20-7.18 (m, 24H), 7.05-7.04 (m, 24H), 3.94-3.92 (m, 36H), 2.22 (br (s), 4H), 1.67 (br (s). 24H), 1.30-1.26 (m, 48H), 0.99-0.96 (m, 72H); ¹³**C NMR** (151 MHz, C₂D₂Cl₄): δ =159.4, 158.71, 158.69, 155.12, 155.11, 155.08, 151.60, 151.59, 150.81, 150.78, 149.32, 149.30, 149.27, 149.23, 149.20, 149.16, 149.14, 146.04, 146.02, 145.95, 145.81, 145.78, 139.87, 139.84, 139.81, 139.78, 127.5, 127.2, 124.1, 124.0, 121.96, 121.92, 119.4, 119.33, 119.28, 116.9, 116.4, 109.86, 109.83, 72.8, 66.72, 66.69, 66.67, 55.8, 40.6, 40.5, 20.38, 15.17; **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2858 (m), 2931 (w), 2914 (w), 2871 (w), 1618 (m), 1595 (m), 1564 (w), 1481 (s), 1461 (m), 1415 (w), 1375 (w), 1303 (w), 1286 (w), 1259 (m), 1193 (w), 1153 (w), 1112 (w), 1076 (w), 1035 (m), 970 (w), 941 (w), 875 (m), 823 (m), 775 (w), 767 (w), 732 (w), 698 (w), 673 (w), 648 (w), 617 (w); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 295, 375; **MALDI-TOF** (DCTB): m/z [M]⁺ calcd. for C₃₆₄H₃₆₀N₂₄O₁₂: 5262.84, found 5262.84.

Synthesis (H/OMe-cube)₂



To a solution of TBTQ 1 (5.3 mg, 0.011 mmol) and 2-methoxy-terephthaldehyde 12 (2.6 mg, 0.016 mmol) in deuterated dichloromethane (1 mL) in a screw-capped 8 mL glass vial, a catalytic amount of TFA (0.1 μ L, 0.001 mmol) was added and the reaction mixture was stirred at RT for 4 days. Afterward, the crude reaction mixture was stirred with aq. K₂CO₃ solution (0.25 M, 3 × 2 mL) for 10 minutes, the organic layer was separated, dried over Na₂SO₄ and concentrated under reduced pressure. The resulting yellow solid was dissolved immediately in dichloromethane and purified by recycling gel permeation chromatography (rGPC) (THF, 40°C, 3 mL/min) to give 2.9 mg (42%) of (Mono-OMe-cube)₂ as a yellow solid.

Mp: decomposed above 290°C; ¹**H NMR** (600 MHz, CD₂Cl₂): δ = 8.96-8.69 (m, 18H), 8.53-8.15 (m, 44H), 8.01-6.17 (m, 202H), 4.03-3.92 (m, 46H), 3.69-3.42 (m, 12 H), 2.94-2.48 (m, 14H), 2.24-2.00 (m, 88H), 1.72-1.65 (m, 52H), 1.30-0.38 (m, 244H); **FT-IR** (neat, ATR): $\tilde{\nu}$ (cm⁻¹) = 2960 (m), 2932 (w), 2913 (w), 2873 (w), 1618 (m), 1595 (m), 1564 (w), 1481 (m), 1461 (w), 1415 (m), 1371 (w), 1307 (w), 1263 (w), 1193 (w), 1151 (w), 1112 (w), 1035 (w), 952 (s), 865 (w), 825 (w), 711 (s), 676 (m); **UV/Vis** (CH₂Cl₂): λ_{max} (nm) = 294, 370; **TIMS-TOF Flex** (DCTB): m/z [M+H]⁺ calcd. for C₃₅₂H₃₃₇N₂₄O₂₄: 10526.69, found 10526.70.

Note: A decent ¹³C NMR spectrum was not available, even by recording 6144 on a 600 MHz NMR instrument (see Figure 296).

3. 1D and 2D NMR spectra



3:1) 13.



Figure 3. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of enantiopure ¹⁵N labelled (-)-(*P*)-TA-TBTQ 14.



TBTQ 14.



Figure 6. ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of OH-cube. * H-grease



Figure 7. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of OH-cube.



Figure 8. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of OH-cube.



Figure 9. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of OH-cube.



cube.



Figure 11. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of OH-cube.



Figure 12. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of OH-cube.


Figure 13. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of OH-cube.



cube.



Figure 15. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of OH-cube.



Figure 16. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of OH-cube.





grease.



Figure 19. ¹H-¹H COSY NMR spectrum (700 MHz, CD_2Cl_2) of ¹⁵N labelled *OH-cube.



Figure 20. Partial ¹H-¹H COSY NMR spectrum (700 MHz, CD₂Cl₂) of ¹⁵N labelled ***OH-cube**.



Figure 21. ¹H-¹³C HSQC NMR (700 MHz and 151 MHz, CD₂Cl₂) spectrum of ¹⁵N labelled ***OH-cube**.



Figure 22. Partial ¹H-¹³C HSQC NMR (700 MHz and 151 MHz, CD₂Cl₂) spectrum of ¹⁵N labelled ***OH-cube**.



Figure 23. $^{1}H^{-13}C$ HMBC NMR (700 MHz and 176 MHz, $CD_{2}Cl_{2}$) spectrum of ^{15}N labelled *OH-cube.



Figure 24. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of ¹⁵N labelled ***OH-cube**.



13.0 12.0 11.0 10.0 9.0 8.0 7.0 6.0 5.0 4.0 3.0 2.0 1.0 0.0 ppm Figure 25. ${}^{1}H{}^{-1}H$ NOESY NMR (700 MHz, CD₂Cl₂) spectrum of ${}^{15}N$ labelled *OH-cube.



cube.



Figure 27. ¹H NMR (400 MHz, CD₂Cl₂) spectrum of **OMe-cube**. (*silicone grease).



Figure 28. ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of **OMe-cube**. (*H-grease. +silicon grease).



Figure 29. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of OMe-cube.



Figure 30. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of **OMe-cube**.



Figure 32. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **OMe-cube**.



Figure 33. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of OMe-cube.



Figure 34. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of **OMe-cube**.



Figure 36. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **OMe-cube**.







Figure 38. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of OMe-cube.



Figure 39. ¹H NMR (300 MHz, C₂D₂Cl₄) spectrum of OMe-cube synthesized in TCE.



Figure 40. ¹H NMR (700 MHz, C₂D₂Cl₄) spectrum of ¹⁵N labelled *OMe-cube. (⁺CHCl₃, *DCM).



Figure 41.¹³C NMR (176 MHz, C₂D₂Cl₄) spectrum of ¹⁵N labelled *OMe-cube. *C₂Cl₄, **CCl₄, ⁺CDCl₃)



Figure 42. ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄) of ¹⁵N labelled *OMe-cube.



Figure 43. Partial ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄) of ¹⁵N labelled *OMecube.



Figure 44. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of ¹⁵N labelled ***OMe-cube**.



Figure 45. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of ¹⁵N **labelled *OMe-cube**.



Figure 46. ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of ¹⁵N labelled ***OMe-cube**.



Figure 47. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of ¹⁵N labelled ***OMe-cube**.



Figure 48. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



grease, ⁺silicone grease).



Figure 50. Expanded ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 51. Expanded ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 52. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (OMe-cube)₂.



Figure 53. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (OMe-cube)₂.



Figure 54. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 55. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂.



Figure 56. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂.





Figure 58. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 59. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂.



Figure 60. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂.



cube)2.



Figure 62. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂.



63



Figure 64. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 65. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 66. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 67. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 68. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 69. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 70. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 71. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 72. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.





Figure 74. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (OMe-cube)₃.



Figure 75. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (OMe-cube)₃.



Figure 76. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 77. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₃.



Figure 78. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 79. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 80.¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



(OMe-cube)₃.


Figure 82. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 83. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 84. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 85. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 86. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 87. ¹H NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (**OMe-cube**)₃.*unknown impurity, ^{+}EA , $^{\Delta}silicone$ grease



Figure 88. ¹H NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (OMe-cube)₃.



Figure 89. ¹H NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (OMe-cube)₃.



Figure 90. Partial ¹H NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (OMe-cube)₃.



Figure 91. Partial ¹H NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (OMe-cube)₃.



Figure 92. ¹H-¹H COSY NMR spectrum (700 MHz, 375 K, Toluene-d₈) of (OMe-cube)₃.



Figure 93. Partial ¹H-¹H COSY NMR spectrum (700 MHz, 375 K, Toluene-d₈) of (**OMe-cube**)₃.



Figure 94. Partial ¹H-¹H COSY NMR spectrum (700 MHz, 375 K, Toluene-d₈) of (**OMe-cube**)₃.



Figure 95. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) CD₂Cl₂) spectrum of (OMe-cube)₃.



Figure 96. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of **(OMe-cube)**3.



Figure 97. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of (**OMe-cube**)₃.



Figure 98. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of **(OMe-cube)**₃.



Figure 99.¹H-¹³C HMBC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of **(OMe-cube)**₃.



Figure 100. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of (**OMe-cube**)₃.



Figure 101. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, 375 K, Toluene-d₈) spectrum of (**OMe-cube**)₃.



Figure 102. ¹H-¹H NOESY NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (OMe-cube)₃.



Figure 103. Partial ¹H-¹H NOESY NMR (700 MHz, 375 K, Toluene-d₈) spectrum of (**OMe-cube**)*3*.



Figure 104. ¹H NMR (400 MHz, CD₂Cl₂) spectrum of H-cube. (*THF).





Figure 106. $^{1}H^{-1}H$ COSY NMR (600 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 107. Partial ¹H-¹H COSY NMR (600 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 108. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 109. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **H**-cube.



Figure 110. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of **H-cube**.



Figure 111. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 112. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 113. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **H**-cube.



Figure 114. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of **H-cube**.



Figure 115. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of H-cube.



Figure 116. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of Me-cube. (*CHCl₃).



Figure 117. ¹³C NMR spectrum (151 MHz, CD₂Cl₂) of Me-cube.



Figure 118. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of **Me-cube**.



Figure 119. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of **Me-cube**.



Figure 120. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of Me-cube.



Figure 121. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Me-cube**.



Figure 122. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of Me-cube.



Figure 123. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of Me-cube.







Figure 126. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of **Et-cube**.



Figure 127. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of Et-cube.



Figure 128. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 129. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Etcube**.



Figure 130. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 131. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 132. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 133. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Et-cube**.



Figure 134. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 135. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of Et-cube.



Figure 136. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂. (* silicon grease)



Figure 137. ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 138. Expanded ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 139. Expanded ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 140. Expanded ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 141. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (**Et-cube**)₂.



Figure 142. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (Et-cube)₂.



Figure 143. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 144. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**Et-cube**)₂.



Figure 145. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**Etcube**)₂.



Figure 146. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.


Figure 147. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 148. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 149. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**Et-cube**)₂.



Figure 150. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 151. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 152. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 153. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 154. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (Et-cube)₂.



Figure 155. ¹H NMR (600 MHz, CDCl₃) spectrum of OEt-cube. (*DCM, + silicone grease)



Figure 156. ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃) of OEt-cube.



Figure 157. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃) of OEt-cube.



Figure 158. ¹H-¹H NOESY NMR (600 MHz, CDCl₃) spectrum of OEt-cube



Figure 159. Partial ¹H-¹H NOESY NMR (600 MHz, CDCl₃) spectrum of OEt-cube.





Figure 162 ¹³C NMR of **OEt-cube**)₂. Expanded ¹H NMR (600 MHz, CD₂Cl₂) spectrum of **(OEt-cube**)₂.







Figure 164. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (OEt-cube)₂.



Figure 165. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 166. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 167. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 168. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 169. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 170. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.



Figure 171. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (OEt-cube)₂.







Figure 174. ¹H-¹H COSY NMR (600 MHz, CDCl₃) spectrum OⁱPr-cube.



Figure 175. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃) of **OⁱPr-cube**.



Figure 176. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CDCl₃) spectrum of OⁱPr-cube.



Figure 177. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CDCl₃) spectrum of **OⁱPr-cube**.



Figure 178. ¹H-¹H TOCSY NMR spectrum (600 MHz, CDCl₃) of OⁱPr-cube.



Figure 179. Partial ¹H-¹H TOCSY NMR (600 MHz, CDCl₃) spectrum of OⁱPr-cube.



Figure 180. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CDCl₃) spectrum of OⁱPr-cube.



Figure 181. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CDCl₃) spectrum of **O**ⁱ**Pr-cube**.



Figure 182. ¹H-¹H NOESY NMR (600 MHz, CDCl₃) spectrum of OⁱPr-cube.



Figure 183. Partial ¹H-¹H NOESY NMR (600 MHz, CDCl₃) spectrum of OⁱPr-cube.



Figure 184. ¹H NMR (700 MHz, C₂D₂Cl₄) spectrum of SMe-cube.





Figure 186. ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄) of SMe-cube.



Figure 187. Partial ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄) of **SMe-cube**.



Figure 188. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of SMe-cube.



Figure 189. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of SMe-cube.



Figure 190. ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of SMe-cube.



Figure 191. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄) spectrum of SMe-cube.



Figure 192. ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of SMe-cube.



Figure 193. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of SMe-cube.



Figure 194. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂. (+CHCl₃, *silicone grease).



Figure 195. ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 196. Expanded ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 197. Expanded ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 198. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (SMe-cube)₂.



Figure 199. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (SMe-cube)₂.



Figure 200. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂.



Figure 201. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **(SMe-cube)**₂.



Figure 202. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **(SMe-cube)**₂.



Figure 203. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 204. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 205. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 206. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂.



Figure 207. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 208. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 209. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 210. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 211. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (SMe-cube)₂.



Figure 212. ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃. (*THF)



Figure 213. Expanded ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃.






Figure 216. ¹H-¹H COSY NMR spectrum (700 MHz, CD₂Cl₂) of (SMe-cube)₃.



Figure 217. Partial ¹H-¹H COSY NMR spectrum (700 MHz, CD₂Cl₂) of (SMe-cube)₃.



Figure 218. Partial ¹H-¹H COSY NMR spectrum (700 MHz, CD₂Cl₂) of (SMe-cube)₃.



Figure 219. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (SMecube)₃.



Figure 220. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃.



Figure 221. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₃.



Figure 222. ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₃.



Figure 223. ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃.



Figure 224. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₃.





Figure 226. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃.



Figure 227. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃



Figure 228. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of (SMe-cube)₃



Figure 229. ¹H NMR (700 MHz, 410 K, C₂D₂Cl₄) spectrum of (SMe-cube)₃, .*THF



Figure 230. Expanded ¹H NMR (700 MHz, 410 K, C₂D₂Cl₄) spectrum of (SMe-cube)₃.



Figure 231. ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄, 410 K) of (SMe-cube)₃.



Figure 232. Partial ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄, 410 K) of (**SMe-cube**)₃.



Figure 233. Partial ¹H-¹H COSY NMR spectrum (700 MHz, C₂D₂Cl₄, 410 K) of (SMe-cube)₃.



Figure 234. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 235. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 236. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 237. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 238. 1 H- 13 C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (SMe-cube)₃.



Figure 239. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 240. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 241. ¹H-¹H NOESY NMR (700 MHz, C₂D₂Cl₄, 410 K) spectrum of (SMe-cube)₃.



Figure 242. Partial ¹H-¹H NOESY NMR (700 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



Figure 243. Partial ¹H-¹H NOESY NMR (700 MHz, C₂D₂Cl₄, 410 K) spectrum of (**SMe-cube**)₃.



cube)3



Figure 245. ¹H NMR (700 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(**SMe-cube**)₃. **DCM, *THF. ⁺MeOH.



*(SMe-cube)₃.



Figure 247. ¹H-¹H COSY NMR spectrum (700 MHz, Toluene-d₈, 380 K) of ¹⁵N labelled *(SMe-cube)₃..



Figure 248. Partial ¹H-¹H COSY NMR spectrum (700 MHz, Toluene-d₈, 380 K) of ¹⁵N labelled *(SMe-cube)₃.



Figure 249. Partial ¹H-¹H COSY NMR spectrum (700 MHz, Toluene-d₈, 380 K) of ¹⁵N labelled *(SMe-cube)₃.



Figure 250. ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 251. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 252. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 253. Partial ¹H-¹³C HSQC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 254. ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 255. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 256. Partial ¹H-¹³C HMBC NMR (700 MHz and 176 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 257. ¹H-¹H NOESY NMR (700 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 258. Partial ¹H-¹H NOESY NMR (700 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 259. Partial ¹H-¹H NOESY NMR (700 MHz, Toluene-d₈, 380 K) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 260. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube. (^{*}silicone grease).



Figure 261. ¹³C NMR spectrum (151 MHz, CD₂Cl₂) of **SC(CH₃)₃-cube**. (⁺H grease, *silicone grease).



Figure 262. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of SC(CH₃)₃-cube.



Figure 263. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of SC(CH₃)₃-cube.



Figure 264. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.



Figure 265. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **SC(CH₃)₃-cube**.



Figure 266. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.



Figure 267. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.



Figure 268. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **SC(CH₃)3-cube**.



Figure 269. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **SC(CH₃)₃-cube**.



Figure 270. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.



Figure 271. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.





Figure 273. ¹³C NMR (151 MHz, CD₂Cl₂) spectrum of **Br-cube**. (*ethyl acetate).



Figure 274. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of Br-cube.



Figure 275. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of **Br-cube**.



Figure 276. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Br-cube**.



Figure 277. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Br-cube**.


Figure 278. ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of Br-cube



Figure 279. Partial ¹H-¹H TOCSY NMR (600 MHz, CD₂Cl₂) spectrum of **Br-cube**.



Figure 280. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of Br-cube.



Figure 281. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **Br-cube**.



Figure 282. ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of Br-cube.



Figure 283. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of **Br-cube**.



Figure 284. ¹H NMR (600 MHz, C₂D₂Cl₄) spectrum of H/OMe-cube. (*THF)





Figure 286. ¹H-¹H COSY NMR spectrum (600 MHz, C₂D₂Cl₄) of H/OMe-cube.



Figure 287. Partial ¹H-¹H COSY NMR spectrum (600 MHz, C₂D₂Cl₄) of H/OMe-cube.



Figure 288. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, C₂D₂Cl₄) spectrum of **H/OMe-cube**.



Figure 289. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, C₂D₂Cl₄) spectrum of **H/OMe-cube**.



Figure 290. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, C₂D₂Cl₄) spectrum of H/OMe-cube.



Figure 291. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, C₂D₂Cl₄) spectrum of **H/OMe-cube**.



Figure 292. ¹H-¹H NOESY NMR (600 MHz, C₂D₂Cl₄) spectrum of H/OMe-cube.



Figure 293. Partial ¹H-¹H NOESY NMR (600 MHz, C₂D₂Cl₄) spectrum of H/OMe-cube.



Figure 295. ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (H/OMe-cube)₂.





Figure 297. ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (H/OMe-cube)₂.



Figure 298. Partial ¹H-¹H COSY NMR spectrum (600 MHz, CD₂Cl₂) of (H/OMe-cube)₂.



Figure 299. ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**H/OMe-cube**)₂.



Figure 300. Partial ¹H-¹³C HSQC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (H/OMe-cube)₂.



Figure 301. ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of (**H/OMe-cube**)₂.



Figure 302. Partial ¹H-¹³C HMBC NMR (600 MHz and 151 MHz, CD₂Cl₂) spectrum of **(H/OMe-cube)**₂.





Figure 304. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (H/OMe-cube)₂.



Figure 305. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum of (H/OMe-cube)₂.

4. DOSY experiments

DOSY NMR experiments were calibrated using self-diffusion values for the used solvents (D_{solv}) .^[S8] Tha solvodynamic radii were calculated using the semi-empirical modification of the Stokes-Einstein equation proposed by Chen and Chen. ^[S9] This equation was solved for r_s using values of r_{solv} and η from the literature. ^[S10]



D is the measured diffusion coefficient $(m^2 \cdot s^{-1})$

 $k_{\rm B}$ is Boltzmann constant (1.3806485 · 10 m² · kg · s⁻² · K⁻¹)

T is the temperature (K)

 r_{solv} is the hydrodynamic radius of the solvent (m)

 r_s is the hydrodynamic radius of the analyte (m)

 η viscosity of the solvent at temperature T (Kg·m⁻¹·s⁻¹)



Figure 307. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum ¹⁵N labelled *OH-cube.



Figure 309. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of (OMe-cube)₂.



Figure 311. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of H-cube.



Figure 313. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of Et-cube.



Figure 315. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of (OEt-cube)₂



Figure 317. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of (SMe-cube)₂.





Figure 319. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of SC(CH₃)₃-cube.



Figure 321. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of (H/OMe-cube)₂.



Figure 322. DOSY NMR (400 MHz, 295 K, CDCl₃) spectrum of OEt-cube.



Figure 323. DOSY NMR (400 MHz, 295 K, CD₂Cl₂) spectrum of (H/OMe-cube)₂



Figure 324. DOSY NMR (400 MHz, 295 K, C2D₂Cl₄) spectrum of SMe-cube.

5. Mass Spectra



Figure 325. MALDI-TOF mass spectrum of OH-cube.



Figure 326. TIMS-TOF mass spectrum of ¹⁵N labelled *OH-cube.



Figure 327. MALDI-TOF mass spectrum of OMe-cube.



Figure 328. MALDI-TOF mass spectrum of ¹⁵N labelled *OMe-cube.



Figure 329. MALDI-TOF mass spectrum of (OMe-cube)2.



Figure 330. MALDI-TOF (DCTB matrix) mass spectrum of (**OMe-cube**)₃. Inset: low *m*/*z* region.



Figure 331. MALDI-TOF mass spectrum of H-cube.



Figure 332. MALDI-TOF (DCTB matrix) mass spectrum of Me-cube.



Figure 333. MALDI-TOF mass spectrum of Et-cube.



Figure 334. MALDI-TOF (DCTB matrix) mass spectrum of (Et-cube)2.



Figure 335. MALDI-TOF (DCTB matrix) mass spectrum of OEt-cube.



Figure 336. MALDI-TOF (DCTB matrix) mass spectrum of (OEt-cube)2



Figure 337. MALDI-TOF (DCTB matrix) mass spectrum of OⁱPr-cube.



Figure 338. TIMS-TOF (DCTB) mass spectrum of SMe-cube.



Figure 339. MALDI-TOF (DCTB matrix) mass spectrum of (SMe-cube)2.



Figure 340. MALDI-TOF (DCTB) mass spectrum of (SMe-cube)3.



Figure 341. MALDI-TOF (DCTB) mass spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 342. MALDI-TOF (DCTB matrix) mass spectrum of SC(CH₃)₃-cube.



Figure 343. MALDI-TOF mass spectrum of Br-cube.



Figure 344. TIMS-TOF (DCTB) mass spectrum of H/OMe-cube.



Figure 345. TIMS-TOF (DCTB) mass spectrum of (H/OMe-cube)₂.




Figure 347. FT-IR (ATR) spectrum of 15N labelled *OH-cube.



Figure 348. FT-IR (ATR) spectrum of OMecube.



Figure 349. FT-IR (ATR) spectrum of ¹⁵N labelled *OMe-cube.



Figure 350. FT-IR (ATR) spectrum of (OMe-cube)₂.



Figure 351. FT-IR spectrum (DCM) of (OMe-cube)₃.



Figure 352. FT-IR (ATR) spectrum of H-cube.



Figure 353. FT-IR (ATR) spectrum of Me-cube.



Figure 354. FT-IR (ATR) spectrum of Et-cube.



Figure 355. FT-IR spectrum (DCM) of (Et-cube)₂.







Figure 357. FT-IR (ATR) spectrum of (OEt-cube)2.



Figure 358. FT-IR (ATR) spectrum of OⁱPr-cube.



Figure 359. FT-IR (ATR) spectrum of SMe-cube.



Figure 360. FT-IR spectrum (DCM) of (SMe-cube)2.



Figure 361. FT-IR (ATR) spectrum of (SMe-cube)3.



Figure 362. FT-IR (ATR) spectrum of ¹⁵N labelled *(SMe-cube)₃.



Figure 363. FT-IR spectrum (DCM) of SC(CH₃)₃-cube.



Figure 364. FT-IR (ATR) spectrum of Br-cube.



Figure 365. FT-IR (ATR) spectrum of Mono-OMe-cube.



Figure 366. FT-IR (ATR) spectrum of (mono-OMe-cube)₂.

7. UV-Vis spectra



Figure 367. UV-Vis spectrum (DCM, conc. 3.2 µM) of OH-cube.



Figure 368. UV-Vis spectrum (DCM, conc. 0.94 µM) of OH-cube.



Figure 369. UV-Vis spectrum (DCM, Conc. 3.6 μ M) and CD spectrum of OMe-cube.



Figure 370. UV-Vis spectrum (DCM, conc. 0.59 µM) of ¹⁵N labelled *OMe-cube.



Figure 371. UV-Vis spectrum (DCM, Conc. 1.1 μ M) and CD (DCM) spectrum of (OMe-cube)₂.



Figure 372. UV-Vis spectrum (DCM, Conc. 1.1 μ M) and CD spectrum (DCM) of (OMe-cube)₃.



Figure 373. UV-Vis spectrum (DCM, Conc. 4.2 $\mu M)$ of H-cube.



Figure 374. UV-Vis spectrum (DCM, Conc. 3.18 µM) of Me-cube.



Figure 375. UV-Vis spectrum (DCM, Conc. 5.1 μ M) of Et-cube.



Figure 376. UV-Vis spectrum (DCM, Conc. 2.1 µM) of (Et-cube)₂.



Figure 377. UV-Vis spectrum (CHCl₃, Conc. 0.012 μ M) of OEt-cube.



Figure 378. UV-Vis spectrum (DCM, Conc. 0.011 µM) of (OEt-cube)₂.



Figure 379. UV-Vis spectrum (CHCl₃, Conc. 5.2 μ M) of OiPr-cube



Figure 380. UV-Vis spectrum (DCM, conc.2.77 μ M) of SMe-cube.



Figure 381. UV-Vis spectrum (DCM, Conc. 2.7 µM) of (SMe-cube)₂.



Figure 382. UV-Vis spectrum (DCM, conc. 0.65 µM) of (SMe-cube)₃.



Figure 383. UV-Vis spectrum (DCM, conc. 0.64 μ M) of ¹⁵N labelled *(SMe-cube)₃.



Figure 384. UV-Vis spectrum (DCM, Conc. 1.9 µM) of SC(CH₃)₃-cube.



Figure 385. UV-Vis spectrum (DCM, Conc. 9.8 μ M) of Br-cube.



Figure 386. UV-Vis spectrum (DCM, conc. 1.27 μ M) of Mono-OMe-cube.



Figure 387. UV-Vis spectrum (DCM, conc. 1.38 µM) of (Mono-OMe-cube)₂

8. GPC data:

OMe-cube:

Sample preparation: To avoid the catenane formation during separation, pure **OMe-cube** in DCM (0.8 mg dissolved in 5 mL of DCM) was injected into the r-GPC to get the trace of the pure **OMe-cube**.



Figure 388. GPC trace of pure **OMe-cube** (solvent: dichloromethane, flow rate: 5mL/min, Temp: 30°C, wavelength: 260 nm, GPC Columns: 3 x 100 Å (20 x 300 mm))

(OMe-cube)₂:

Sample preparation for GPC: The crude reaction mixture (~32 mg) was first passed through a small pre-packed size exclusion column using DCM as a mobile phase and the collected fraction (10 mL) was concentrated to 0.5 mL under reduced pressure and injected into the r-GPC in one portion. Total 14 mg of pure (**OMe-cube**)₂ was collected.



Figure 389. GPC traces of (**OMe-cube**)₂. a) GPC trace of crude reaction mixture of (**OMe-cube**)₂ prepared by improved protocol and b) GPC trace of isolated pure (**OMe-cube**)₂ (solvent: dichloromethane, flow rate: 5mL/min, temp: $30^{\circ}C$, wavelength: 260 nm, GPC Columns: $3 \times 100 \text{ Å} (20 \times 300 \text{ mm})$).

Note: It was found that (*OMe-cube*)₂ shows decomposition after multiple cycles on rGPC column when chloroform solvent was used instead of DCM.

(OMe-cube)3:

Sample preparation for GPC: The crude reaction mixture (~ 16 mg) was first passed through a small pre-packed size exclusion column using DCM as a mobile phase and then the collected fraction (10 mL) was concentrated to 0.5 mL under reduced pressure and injected into the rGPC. Very small amount (< 1mg) of pure (**OMe-cube**)₃ was collected. (**OMe-cube**)₃ was confirmed by MALDI-TOF mass technique.



Figure 390. Separation of (**OMe-cube**)³ from crude reaction mixture (reaction solvent: CHCl₃). (Chloroform: flow rate: 5mL/min, temp: 40°C, wavelength: 260 nm, GPC columns: 3 x 100 Å (20 x 300 mm)).

(OMe-cube)₃ (synthesized by improved protocol):

Sample preparation for GPC: The crude reaction mixture (~32 mg) was first passed through a small pre-packed size exclusion column using DCM as a mobile phase and then the collected fraction (10 mL) was concentrated to 0.5 mL under reduced pressure and injected into the rGPC in one portion. Total 16 mg of pure (**OMe-cube**)₃ was collected.



Figure 391. GPC profile of **(OMe-cube)**₃. a) GPC trace of crude reaction mixture of **(OMe-cube)**₃ prepared by improved protocol and b) GPC trace of pure **(OMe-cube)**₃ (solvent: dichloromethane, flow rate: 5mL/min, temp: 30°C, wavelength: 260 nm, GPC columns: 3 x 100 Å (20 x 300 mm)).

Note: It was found that (*OMe-cube*)₃ shows decomposition after multiple cycles on rGPC column when chloroform solvent was used instead of DCM.

(Et-cube)2: (synthesized by improved protocol)

Sample preparation for GPC: THF was first removed from the crude reaction mixture under reduced pressure. The crude reaction mixture (~17 mg) was redissolved in chloroform and passed through a small pre-packed size exclusion column using chloroform solvent as a mobile and then the collected fraction (6 mL) was concentrated to 0.5 mL under reduced pressure and injected into rGPC in one portion. Total 5.2 mg of pure (**Et-cube**)₂ was collected.



Figure 392. GPC profile of (**Et-cube**)₂. a) GPC trace of crude reaction mixture of (**Et-cube**)₂ prepared by improved protocol and b) GPC trace of pure (**Et-cube**)₂ (solvent: chloroform, flow rate: 3.5mL/min, Temp: 40°C, wavelength: 260 nm, GPC columns: 3 x 100 Å (20 x 300 mm)).

(OEt-cube)2: (synthesized by improved protocol)

Sample preparation for GPC: The rude reaction mixture (~17 mg) was first passed through a small pre-packed size exclusion column using chloroform solvent as a mobile phase and then the collected fraction (6 mL) was concentrated to 0.5 mL under reduced pressure and injected into the rGPC in one portion. Total 4 mg of pure (**OEt-cube**)₂ was collected.



Figure 393. GPC profile of (**OEt-cube**)₂ a) GPC trace of crude reaction mixture of (**OEt-cube**)₂ prepared by improved protocol and b) GPC trace of pure (**OEt-cube**)₂ (solvent: chloroform, flow rate: 5mL/min, Temp: 40°C, wavelength: 254 nm, GPC columns: 3 x 100 Å (20 x 300 mm)).

(SMe-cube)2: (synthesized by improved protocol)

Sample preparation for GPC: The crude reaction mixture (~17 mg) was first passed through a small pre-packed size exclusion column using chloroform as a mobile phase and then the collected fraction (8 mL) was concentrated to 0.5 mL under reduced pressure and injected into rGPC in one portion. Total 10.7 mg of pure (**SMe-cube**)₂ was collected.



Figure 394. GPC profile of (**SMe-cube**)₂ a) GPC trace of crude reaction mixture of (**SMe-cube**)₂ prepared by improved protocol and b) GPC trace of pure (**SMe-cube**)₂ (solvent: chloroform, flow rate: 3.5mL/min, Temp: 40°C, wavelength: 260 nm, GPC columns: 3 x 100 Å (20 x 300 mm)).



Figure 395. GPC profile of (**SMe-cube**)₃. a) GPC trace of crude reaction mixture of (**SMe-cube**)₃ and b) GPC trace of pure (**SMe-cube**)₃. (solvent: THF, flow rate: 3 mL/min, temp: 40°C, wavelength: 260 nm, GPC columns: $(1 \times 500 \text{ Å} (20 \times 300 \text{ mm}) + 3 \times 100 \text{ Å} (20 \times 300 \text{ mm}))$.



Figure 396. GPC trace of crude reaction mixture of ¹⁵N labelled *(SMe-cube)₃. (solvent: THF, flow rate: 3 mL/min, temp: 40°C, wavelength: 260 nm, GPC columns: (1 x 500 Å (20 x 300 mm)) + 3 x 100 Å (20 x 300 mm)).



Figure 397. GPC trace of crude reaction mixture of (**mono-OMe-cube**)₂. (solvent: THF, flow rate: 3 mL/min, temp: 40°C, wavelength: 260 nm, GPC columns: (1 x 500 Å (20 x 300 mm)) + 3 x 100 Å (20 x 300 mm)).

9. Imine condensation between TA-TBTQ and dialdehyde linkers at variable concentrations to study the formation of cage /catenanes

General Procedure: To a solution TBTQ **1** (5 mg, 10.7 μ mol, 1equiv) and various dialdehyde (16.1 μ mol, 1.5 equiv) in CHCl₃ of different concentrations (10.7 mM, 21.4 mM and 42.8 mM), a catalytic TFA (0.1 μ L, 1.3 μ mol, 0.12 equiv) was added and the reaction mixture was stirred at RT. After 3 days, solvent was evaporated and ¹H NMR spectra of the crude reaction mixtures were immediately measured in deuterated dichloromethane (Figure 398-S399) to check the formation of catenane.

2,5-Dihydroxy-terephthaldehyde (2)



Scheme 1. Imine condensation at different concentrations between TBTQ 1 and 2,5dihydroxy-terephthaldehyde 2.

¹H NMR, 300 MHz, CD₂Cl₂



Figure 398. Partial ¹H NMR spectra (300 MHz, CD₂Cl₂) of crude reaction mixture of **OH-cube** at 10.7 mM, 21.4 mM and 42.8 mM concentrations.

2,5-Dimethoxy-terephthaldehyde (3)



Scheme 2. Imine condensation at different concentrations between TBTQ 1 and 2,5dimethoxy-terephthaldehyde 3.



Figure 399. Partial ¹H NMR (300 MHz, CD₂Cl₂) spectra of crude reaction mixture of **OMe-cube**, **(OMe-cube**)₂ and **(OMe-cube**)₃. at 10.7 mM, 21.4 mM and 42.8 mM concentrations.

Entry	TBTQ 1	2,5-dimethoxy-	CHCl ₃	[8+12]	[16+24]	[24+36]
	(mg)	terephthaldehyde	(mL)	cage ^[a]	cage ^[a]	cage ^[a]
		(mg)				
1	5	3.1	0.25	Yes	Yes	Yes
2	5	3.1	0.50	Yes	Yes	Yes
3	5	3.1	1.00	Yes	Yes	Yes
4	5	3.1	2.00	Yes	Yes	No
5	5	3.1	3.00	Yes	Yes	No
6	5	3.1	4.00	Yes	Yes	No
7	5	3.1	6.00	Yes	Yes	No
8	5	3.1	8.00	Yes	Yes	No
9	5	3.1	25.0	Yes	No	No

Table 1: Synthesis of dimethoxy OMe-cube at different concentrations

Reaction time: 24 h, Cat TFA (12 mol%), [a] crude reaction mixture was analyzed by MALDI-TOF mass spectroscopy.

10. Imine condensation between TA-TBTQ and 2,5-dihydroxy dialdehyde linkers at different catalyst concentrations and temperatures to study the formation of cage /catenanes

To a solution TBTQ **1** (5 mg, 10.7 μ mol, 1equiv) and 2,5-dihydroxy dialdehyde (2.7 mg, 16.1 μ mol, 1.5 equiv) in CDCl₃, catalytic TFA was added and the reaction mixture was stirred at RT and at 80 °C After 3 days, ¹H NMR spectra of the crude reaction mixtures were measured (Figure 400) to check the formation of catenane.



Figure 400. Partial ¹H NMR (300 MHz, CDCl₃) spectra of crude reaction mixture of **OH-cube** at diff TFA concentrations (a-c) and at diff. temperatures (a and d).
11. Imine condensation between TA-TBTQ and dialdehyde linkers in different solvents to study the formation of cage /catenanes

General procedure: To a solution of TBTQ **1** (5.6 mg, 12 μ mol, 1equiv) and 2,5-substituted terephthaldehyde (16.1 μ mol) in CDCl₃ or CD₂Cl₂ or THF-d₈ or 1,4 dioxane-d₈ (1 mL), a catalytic amount of TFA (0.1 μ L, 1.3 μ mol) was added and the reaction mixture was stirred at RT for 3 days. ¹H NMR and MALDI-TOF spectra of the crude reaction mixtures were measured after three days (Figure 401-S413) to check the formation of [12+8] cages and its catenanes.

2,5-Dihydroxy-terephthaldehyde (2)



Scheme 3. Imine condensation between TBTQ 1 and 2,5-dihydroxyterephthaldehyde 2 in different solvents.



Figure 401. Partial ¹H NMR (500 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-dihydroxy-terephthaldehyde **2** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

Note: Cage reaction in chloroform solvent was continued for three months and then analyzed by MALDI-TOF MS. The MALDI-TOF spectrum remains unchanged after 3 months for **OH-cube**.

2,5-Dimethoxy-terephthaldehyde (3)



Scheme 4. Imine condensation between TBTQ 1 and 2,5-dimethoxy-terephthaldehyde 3 in different solvents.



Figure 402. Partial ¹H NMR (300 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-dimethoxy-terephthaldehyde **3** in different solvents (CDCl₃, CD₂Cl₂, THF- d_8 and 1,4-dioxane- d_8) after 3 days of reaction.

2,5-Dimethoxy-terephthaldehyde (3)



Scheme 5. Imine condensation between TBTQ 1 and 2,5-dimethoxy-terephthaldehyde 3 in mixed solvents (CD₂Cl₂ and CD₃CN).



Figure 403. a) and b) ¹H NMR (600 MHz) and c) and d) MALDI-TOF mass spectra of reaction between TBTQ 1 and 2,5-dimethoxy-terephthaldehyde 3 in mixed solvents after 3 days of reaction (reaction temp 80 $^{\circ}$ C).

MALDI-TOF



Figure 404. MALDI-TOF (DCTB) mass spectra of reaction between TBTQ 1 and 2,5dimethoxy-terephthaldehyde 3 in mixed solvents after 3 days of reaction (reaction temp 80 $^{\circ}$ C).

Terephthaldehyde (4)



Scheme 6. Imine condensation between TBTQ 1 and terephthaldehyde 4 in different solvents.



Figure 405. Partial ¹H NMR (500 MHz) and MALDI-TOF mass spectra of reaction between -TBTQ 1 and terephthaldehyde 4 in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction. *reaction temp 80 °C.

2,5-Dimethyl-terephthaldehyde (5)



Scheme 7. Imine condensation between TBTQ 1 and 2,5-dimethyl-terephthaldehyde 5 in different solvents.



Figure 406. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra of reaction between TA-TBTQ and 2,5-dimethyl-terephthaldehyde in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.*reaction temp 80 °C.

2,5-Diethyl-terephthaldehyde (6)



Scheme 8. Imine condensation between TBTQ 1 and 2,5-diethyl-terephthaldehyde 6 in different solvents.



Figure 407. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-diethyl-terephthaldehyde **6** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2,5-Diethoxy-terephthaldehyde (7)



Scheme 9. Imine condensation between TBTQ 1 and 2,5-diethoxy-terephthaldehyde 7 in different solvents.



Figure 408. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-diethoxy-terephthaldehyde **7** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2,5-Diisopropoxy-terephthaldehyde (8)



Scheme 10. Imine condensation between TBTQ 1 and 2,5-diisopropoxy-terephthaldehyde 8 in different solvents.



Figure 409. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-diisopropoxy-terephthaldehyde **8** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2,5-Bis(methylthio)terephthaldehyde (9)



Scheme 11. Imine condensation between TBTQ 1 and 2,5-bis(methylthio)terephthaldehyde 9 in different solvents.



Figure 410. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-bis(methylthio)terephthaldehyde **9** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2,5-bis(tert-methylthio)terephthaldehyde (10)



Scheme 12. Imine condensation between TBTQ 1 and 2,5-bis(*tert*-methylthio)terephthaldehyde 10 in different solvents.



Figure 411. Partial ¹H NMR (600 MHz) and MALDI-TOF mass spectra reaction between TBTQ 1 and 2,5-bis(*tert*-methylthio)terephthaldehyde 10 in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2,5-Dibromo-terephthaldehyde (11)



Scheme 13. Imine condensation between TBTQ 1 and 2,5-dibromo-terephthaldehyde 11 in different solvents.



Figure 412. Partial ¹H NMR (300 MHz) and MALDI-TOF mass spectra of reaction between TBTQ **1** and 2,5-dibromo-terephthaldehyde **11** in different solvents (CDCl₃, CD₂Cl₂, THF-d₈ and 1,4-dioxane-d₈) after 3 days of reaction.

2-methoxyterephthaldehyde (12)



Scheme 14. Imine condensation between TBTQ 1 and 2,5-dimethoxy-terephthaldehyde 12 in different solvents.



Figure 413. a) and b) ¹H NMR (600 MHz) and c) and d) MALDI-TOF (DCTB) mass spectra of reaction between TBTQ 1 and 2-methoxyterephthaldehyde 12 in CDCl₃ and CD₂Cl₂ after 3 days of reaction.

12. Thermodynamic data

a) Calculation of ΔG by variable concentration measurements

General procedure: To a solution of TBTQ **1** (5 mg, 10.7 μ mol, 1equiv) and 2,5-substituted terephthaldehydes (1.5 equiv) in CDCl₃, a catalytic amount of TFA (0.1 μ L, 1.3 μ mol) was added and the reaction mixture was stirred at RT at different concentrations (0.43 mM, 0.71 mM, 1.07 mM, 2.14 mM and 10.7 mM (concentration is with respect to TA-TBTQ). After three days, ¹H NMR spectra (600 MHz, RT) were measured.

Equations used to calculate the K_{eq} and ΔG

*K*_{eq} = [Dicatenane]/[Cage]²

where: K_{eq} is the equilibrium constant

[Catenane] is the concentration of [12+8]₂ dicatenane (M)

[Cage] is the concentration of [12+8] monomeric cage (M)

$\Delta \mathbf{G} = -\mathbf{RT} \ln \mathbf{K}_{eq}$

 ΔG is the Gibbs free energy (kJmol⁻¹)

R is the universal gas constant (Jmol⁻¹K⁻¹)

T is the temperature (K)

• Calculation of cage to dicatenane ratio from ¹H NMR spectrum



Integration of cage = (cage+dicatenane peak)-5

Integration of cage = (7.40)-5 = 2.40

cage:dicatenane ratio is 2.40:1

Cage has 24 protons per imine signals and dicatenane has 6 imine protons per imine signals.

Cage:dicatenane ratio is (2.40/24):(1/6) = 0.100:0.167

cage:dicatenane ratio is (0.100/0.100):(0.167/0.100) = 1.00:1.67

a) **OMe-cube** \rightarrow (**OMe-cube**)



¹H NMR, 600 MHz, CDCl₃



Figure 414. Partial ¹H NMR spectra of the reaction (**OMe-cube**) \rightarrow (**OMe-cube**)₂ at various concentrations under thermodynamic equilibrium.

Entry	Conc	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnKeq	Δ G
	(mM)					(kJmol ⁻¹)
1	10.7	295	1:1.16	2851.75	7.955	-19.51
2	2.14	295	1:0.65	5601.06	8.630	-21.17
3	1.07	295	1:0.52	7796.71	8.961	-21.98
4	0.71	295	1:0.34	6485.31	8.777	-21.53
5	0.43	295	1:0.13	3121.88	8.046	-19.73

Table 3. Calculation of equilibrium constant and ΔG for the reaction between OMe-cube \rightarrow (OMe-cube)₂.

b) OEt-cube \rightarrow (OEt-cube)₂



¹H NMR, 600 MHz, CDCl₃



Figure 415. Partial ¹H NMR spectra of the reaction (OEt-cube) \rightarrow (OEt-cube)₂ at various concentrations under thermodynamic equilibrium.

Table 4. Calculation of equilibrium constant and ΔG for the reaction OEt-cube \rightarrow (OEt-cube)_2

Entry	Conc	T (K)	Cage:	Keq (M-1)	lnK _{eq}	$\Delta \mathbf{G}$
	(mM)		Catenane			(kJmol ⁻¹)
1	10.7	295	1:0.22	235.64	5.462	-13.40
2	2.14	295	1:0.17	857.26	6.753	-16.56
3	1.07	295	1:0.09	758.89	6.631	-16.27
4	0.71	295	1:0.07	874.28	6.773	-16.61

c) SMe-cube \rightarrow (SMe-cube)₂





Figure 416. Partial ¹H NMR spectra of reaction (SMe-cube) \rightarrow (SMe-cube)₂ at various concentrations under thermodynamic equilibrium.

Entry	Conc	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnKeq	$\Delta \mathbf{G}$
	(mM)					(kJmol ⁻¹)
2	2.14	295	1:2.06	39376.57	10.581	-25.95
3	1.07	295	1:1.79	61341.00	11.024	-27.04
4	0.71	295	1:1.32	53736.14	10.892	-26.71
5	0.43	295	1:1.03	59081.81	10.986	-26.95

Table 5. Calculation of equilibrium constant and ΔG for the reaction SMe-cube \rightarrow (SMe-cube)₂

b) van't Hoff plots

$(OMe-cube) \rightarrow (OMe-cube)_2.$

Procedure: A vial 'A' was charged with **OMe-cube** (1.9 mg, 0.34 μ mol) and deuterated CHCl₃ (0.5 mL). Catalytic TFA (5 μ L of a 65.3 mM solution in CDCl₃, 0.33 μ mol TFA) was added and reaction mixture was stirred at 295 K for 40 hours. In an exactly similar scale, another six reactions were set up in five different vials B, C, D, E, F and G and stirred at 303 K, 308 K, 313 K, 318 K, 323 K and 328 K respectively. After 40 hours, ¹H NMR spectra of each reaction mixture were measured at the same stirring temperature on 700 MHz NMR instrument.

Enthalpy and entropy of the reaction (OMe-cube) \rightarrow (OMe-cube)₂ were calculated according to the van't Hoff equation,:

$$\ln K_{eq} = -\frac{\Delta H}{RT} + \frac{\Delta S}{R} \qquad \text{..... Equation (2)}$$

where: K_{eq} is the equilibrium constant

 ΔH^{θ} is the difference in enthalpy between (**OMe-cube**) and (**OMe-cube**)₂ catenane (J·mol⁻¹) ΔS^{θ} is the difference in entropy between (**OMe-cube**) and (**OMe-cube**)₂ catenane (J·K⁻¹·mol⁻¹)

T is the temperature (K)

R is the universal gas constant $(J \cdot K^{-1} \cdot mol^{-1})$

• Equation used to find the concentrations (M) of OMe cage and dicatenane in the NMR sample used for thermodynamic and kinetic studies

mass of cage + mass of dicatenane = 0.0019 g

(no. of moles of cage \times mol. wt. of cage) + (no. of moles of dicatenane + mol. wt. of dicatenane) = 0.0019 g

(for example, the ratio of **cage:diatenane** calculated from the ¹H NMR spectra is **1.00:1.67**).

 $(n_{cage} \times 5623.23 \text{ g/mol}) + (n_{cat} \times 11246.47 \text{ g/mol}) = 0.0019 \text{ g}$

(e.g. cage to dicatenane ratio obtained from ${}^{1}H$ NMR is 1:1.67)

 $n_{cage} = A \mod n_{cat} = 1.67 \times A \mod n_{cage}$

 $(A \text{ mol} \times 5623.23 \text{ g/mol}) + (1.67 \times A \text{ mol} \times 11246.47 \text{ g/mol}) = 0.0019 \text{ g}$

5623.23 g/mol \times A mol + 18781.60 g/mol \times A mol = 0.0019 g

A mol = 0.0019 g/24404.83 g/mol

A mol = 0.0000007785 mol = n_{cage}

 $n_{cat} = 1.67 \text{ x } 0.0000007785 \text{ mol} = 0.00000013 \text{ mol}$

[cage] = 0.0000007785 mol/ 0.0005 L = 0.0001557 M,

[dicatenane] = 0.00000013 mol / 0.0005 L = 0.00026 M

Methoxy cage (1st set data)





Figure 417. ¹H NMR spectra of reaction (OMe-cube) \rightarrow (OMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnK _{eq}	1/T (K ⁻¹)
1	295	1:1.0152	4552.542	8.423	0.00339
2	303	1:1.0782	5035.600	8.524	0.00330
3	308	1:1.1204	5373.275	8.589	0.00325
4	313	1:1.1594	5693.895	8.647	0.00319
5	318	1:1.2270	6271.124	8.744	0.00314
6	323	1:1.3333	7234.230	8.887	0.00310
7	328	1:1.3889	7764.010	8.957	0.00305

Table 6. Calculation of Δ H and Δ S for the reaction (**OMe-cube**) \rightarrow (**OMe-cube**)₂.

The slope value (-1592.68 ± 131) was $-\Delta H^{\theta}/R$ and the intercept value (13.78 ± 0.42) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 13.2 \pm 1.1$ kJ/mol, and $\Delta S = 114.6 \pm 3.5$ J/Kmol.



Figure 418. van't Hoff plot of the reaction (OMe-cube) \rightarrow (OMe-cube)₂.



Figure 419. ¹H NMR spectra of reaction (OMe-cube) \rightarrow (OMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnKeq	1/T (K ⁻¹)
1	295	1:1.0256	4630.841	8.440	0.00339
2	303	1:1.0929	5152.032	8.547	0.00330
3	308	1:1.1204	5373.275	8.589	0.00325
4	313	1:1.1940	5986.161	8.697	0.00319
5	318	1:1.2500	6473.814	8.776	0.00314
6	323	1:1.3201	7111.029	8.869	0.00310

Table 7. Calculation of ΔH and ΔS for the reaction (**OMe-cube**) \rightarrow (**OMe-cube**)₂.



Figure 420. van't Hoff plot of the reaction (OMe-cube) \rightarrow (OMe-cube)₂.

The slope value (-1458.52 ± 103.64) was $-\Delta H^{\theta}/R$ and the intercept value (13.36 ± 0.33) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 12.1 \pm 0.9$ kJ/mol, and $\Delta S = 111.1 \pm 2.7$ J/Kmol



Figure 421. ¹H NMR spectra of reaction (OMe-cube) \rightarrow (OMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnK _{eq}	1/T (K ⁻¹)
1	295	1:1.0283	4650.770	8.445	0.00339
2	303	1:1.0899	5128.388	8.543	0.00330
3	308	1:1.1142	5322.758	8.580	0.00325
4	313	1:1.2085	6110.107	8.718	0.00319
5	318	1:1.3115	7030.800	8.858	0.00314
6	323	1:1.3514	7404.049	8.910	0.00310

Table 8. Calculation of ΔH and ΔS for the reaction (**OMe-cube**) \rightarrow (**OMe-cube**)₂.



Figure 422. van't Hoff plot of the reaction (OMe-cube) \rightarrow (OMe-cube)₂.

The slope value (-1682.17 ± 187.97) was $-\Delta H^{\theta}/R$ and the intercept value (14.11 ± 0.61) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 14.0 \pm 0.9$ kJ/mol, and $\Delta S = 117.3 \pm 5.1$ J/Kmol.

Table 9. Δ **H** and Δ **S** values calculated from Van't Hoff plots from three sets of experiments for (OMe-cube) \rightarrow (OMe-cube)₂ reaction.

Entry	Exp No.	ΔΗ	ΔH (kJ/mol)	Exp No.	ΔS	ΔS (J/Kmol)
1	Set 1	13.2		Set 1	114.6	
2	Set 2	12.1	13.1 ± 0.8	Set 2	111.1	114.3 ± 2.5
3	Set 3	14.0		Set 3	117.3	

The average value of three measurements reported as ΔH and ΔS . The standard deviations calculated using STDEV.P function of Microsoft Excel 2016.

$(SMe-cube) \rightarrow (SMe-cube)_2$

Procedure: A vial 'A' was charged with **SMe-cube** (2.0 mg, 0.33 μ mol) and deuterated CHCl₃ (0.5 mL). Catalytic TFA (5 μ L of a 0.653 mM solution in CDCl₃, 0.33 μ mol TFA) was added and reaction mixture was stirred at 295 K for 40 hours. In a exactly similar scale another reactions were set up in five different vials B, C, D, E, F and G and stirred at 303 K, 308 K, 313 K, 318 K, 323 K and 328 K respectively. After 40 hours, ¹H NMR spectra of each reaction mixture were measured at the same stirring temperature on 700 MHz NMR instrument.

Thiomethyl cage (1st set data)





Figure 423. ¹H NMR spectra of reaction (SMe-cube) \rightarrow (SMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnK _{eq}	1/T (K ⁻¹)
1	295	1:3.0075	33361.455	10.415	0.00339
3	308	1:3.6036	43763.755	10.687	0.00325
3	313	1:3.9604	52278.631	10.864	0.00319
4	323	1:4.8780	77639.556	11.259	0.00310
5	328	1:5.4795	96964.036	11.482	0.00305

Table 10. Calculation of ΔH and ΔS for the reaction (SMe-cube) \rightarrow (SMe-cube)₂.



Figure 424. van't Hoff plot of the reaction (SMe-cube) \rightarrow (SMe-cube)₂.

The slope value (-3161.02 ± 376.7) was $-\Delta H^{\theta}/R$ and the intercept value (21 ± 1.21) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 26.3 \pm 3.1$ kJ/mol, and $\Delta S = 174.9 \pm 10.1$ J/Kmol.



Figure 425. ¹H NMR spectra of reaction (SMe-cube) \rightarrow (SMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Table 11. Calculation of ΔH and ΔS for the react	tion (OMe-cube) \rightarrow (OMe-cube) ₂ .
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Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnKeq	1/T (K ⁻¹)
1	308	1:3.5714	43032.934	10.670	0.00325
2	313	1:3.8095	48586.111	10.791	0.00319
3	318	1:4.3011	61112.173	11.020	0.00314
4	323	1:4.6512	70905.346	11.169	0.00310
5	328	1:5.0633	83363.675	11.331	0.00305



Figure 426. van't Hoff plot of the reaction (SMe-cube) \rightarrow (SMe-cube)₂.

The slope value (-3435.08 ± 171.0) was $-\Delta H^{\theta}/R$ and the intercept value (21.80 ± 0.54) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 28.6 \pm 1.4$ kJ/mol, and $\Delta S = 181.2 \pm 4.5$ J/Kmol.



Figure 427. ¹H NMR spectra of reaction (SMe-cube) \rightarrow (SMe-cube)₂ at various temperatures under thermodynamic equilibrium.

Table 12.	Calculation	of ΔH and Δ	S for the reaction	on (SMe-cube) →	• (SMe-cube) ₂ .

Entry	T (K)	Cage:Catenane	Keq (M ⁻¹)	lnK _{eq}	1/T (K ⁻¹)
1	295	1:3.0303	33832.545	10.429	0.00339
2	303	1:3.6364	44514.160	10.704	0.00330
3	308	1:4.3011	61112.173	11.021	0.00319



Figure 428. van't Hoff plot of the reaction (SMe-cube) \rightarrow (SMe-cube)₂.

The slope value (-4023.2 ± 847.7) was $-\Delta H^{\theta}/R$ and the intercept value (21.36 ± 2.8) was $\Delta S^{\theta}/R$. Therefore, $\Delta H^{\theta} = 33.4 \pm 7.0$ kJ/mol, and $\Delta S = 199.9 \pm 23.3$ J/Kmol.

Table 13. Δ **H** and Δ **S** values calculated from Van't Hoff plots from three sets of experiments for (SMe-cube) \rightarrow (SMe-cube)₂ reaction.

Entry	Exp No.	ΔH	ΔH (kJ/mol)	Exp No.	ΔS	ΔS (J/Kmol)
1	Set 1	26.3		Set 1	174.9	
2	Set 2	28.6	29.4 ± 3.0	Set 2	181.2	185.3 ± 10.6
3	Set 3	33.4		Set 3	199.9	

The average value of three measurements reported as ΔH and ΔS . The standard deviations calculated using STDEV.P function of Microsoft Excel 2016.

- 13. ¹H NMR assignment of (OMe-cube)₂ and (SMe-cube)₂
- a) ¹H NMR assignment of (OMe-cube)₂
- ¹H NMR, 700 MHz, CD_2CI_2



Figure 429. ¹H NMR spectrum (700 MHz, CD₂Cl₂) of (**OMe-cube**)₂. (The group of protons were assigned by 1D and 2D NMR spectra).

¹³H NMR, 700 MHz, CD₂Cl₂



Figure 430. ¹³C NMR spectrum (700 MHz, CD₂Cl₂) of (OMe-cube)₂. (*pentane, +THF)



Figure 431. Labelling of protons and carbons to the partial chemical structure of (OMe-cube)₂.




Figure 432. Partial ¹³C NMR spectrum (700 MHz, CD₂Cl₂) of (**OMe-cube**)₂. (All the carbon regions were assigned by HSQC, HMBC, DEPT).

Assignment of eight different signals of imine, linker and methoxy protons to the 3D model of (OMe-cube)₂

1. First imine protons were labelled as A, B, C, D, E, F, G and H from lower chemical shift values to higher chemical shift values as shown in Figure 433.



Figure 433. Partial ¹H NMR (700 MHz, CD_2Cl_2) spectrum of (**OMe-cube**)₂ showing the labelling of imine protons.

2. Four pairs of neighboring imine protons and dialdehyde linker protons as shown in following Figure 434 were identified by multiple HSQC and HMBC interactions (see Figure 435-438, Table 14 and 15).



Figure 434. Labelling of protons and carbons of **(OMe-cube)**₂. (Arrow indicates that the how neighboring linker and imine protons are identified by HMBC and HSQC measurements).



Figure 435. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between H-(A-H) and C-(10a-10h). (See Table 14 for chemical shift values).



Figure 436. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between C-(10a-10h) and H-($\alpha - \theta$). (See Table 14 for chemical shift values).



Figure 437. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between H-(a- θ) and C-(9a-9h). (See Table 15 for chemical shift values).



Figure 438. Partial ¹H-¹³C HSQC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between C-(9a-9h) and H-(A-H). (See Table 15 for chemical shift values).

Table 14.	$^{1}\text{H}-^{13}\text{C}$	HMBC	interactions	between	H-(A-H)	and	C-(10a-10h)	, and	C-(1	0a-10h)
and H- $(\alpha - \theta)$).									

¹ H-	¹ H-	¹³ C	¹³ C	¹³ C	¹³ C	¹ H-	¹ H-
Atom	NMR	Atom	Atom	Atom	Atom	Atom	NMR
	(HMBC)		(HMBC)		(HMBC)		(HMBC)
А	8.39	10a	127.75	10a	127.75	β	7.34
В	8.65	10b	128.90	10b	128.90	α	7.24
С	8.78	10c	128.55	10c	128.55	δ	7.88
D	8.82	10d	128.42	10d	128.42	γ	7.79
Е	8.85	10e	128.46	10e	128.46	ζ	7.71
F	8.85	10f	128.45	10f	128.45	3	7.69
G	8.87	10g	128.51	10g	128.51	θ	8.03
Н	8.90	10h	128.92	10h	128.92	η	7.69

Table 15. HMBC interactions between H-(α - θ) and C-(9a-9h) and HSQC interactions between C-(9a-9h) and H-(A-H).

¹ H-	¹ H-	¹³ C	¹³ C	¹³ C	¹³ C	¹ H-	¹ H-
Atom	NMR	Atom	Atom	Atom	Atom	Atom	NMR
	(HMBC)		(HMBC)		(HMBC)		(HSQC)
β	7.34	9b	155.27	9b	155.27	В	8.65
α	7.24	9a	155.53	9a	155.53	А	8.39
δ	7.88	9d	155.33	9d	155.33	D	8.82
γ	7.79	9c	155.17	9c	155.17	С	8.78
ζ	7.71	9f	155.03	9f	155.03	F	8.85
3	7.69	9e	155.33	9e	155.33	Е	8.85
θ	8.03	9h	156.47	9h	156.47	Н	8.90
η	7.69	9g	155.72	9g	155.72	G	8.87



Figure 439. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing four pairs of neighboring imine protons.



Figure 440. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing four pairs of neighboring linker protons.

3. Four neighboring methoxy protons were assigned by HSQC, HMBC and NOESY interactions as shown in Figure 441 and 442 (Table 16 and 17).



Figure 441. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between H-(α - θ) and C-(11a-11h). (See Table 16 for chemical shift values).



Figure 442. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between C-(11a-11h) and H-(a-h). (See Table 16 for chemical shift values).

¹ H-	¹ H-	¹³ C	¹³ C Atom	¹³ C	¹³ C Atom	¹ H-	¹ H-
Atom	NMR	Atom	(HMBC)	Atom	(HMBC)	Atom	NMR
	(HMBC)						(HMBC)
α	7.24	11a	153.88	11a	153.88	a	2.85
β	7.34	11b	153.75	11b	153.75	b	3.11
γ	7.79	11c	154.28	11c	154.28	с	3.86
δ	7.88	11d	154.56	11d	154.56	d	3.67
3	7.69	11e	154.24	11e	154.24	e/f/g/h	*
ζ	7.71	11f	154.21	11f	154.21	e/f/g/h	*
η	7.69	11g	154.24	11g	154.24	e/f/g/h	*
θ	8.03	11h	154.24	11h	154.24	e/f/g/h	*

Table 16. ¹H-¹H HMBC interactions between H-(α - θ) and C-(11a-11h), and C-(11a-11h) and H-(a-h).

*Could not assigned due to overlapped carbon signals. These signals were assigned from ¹H-¹H NOESY spectrum (See Figure 443).



Figure 443. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between H-(a-h) and H-(A-H). (See Table 17 for chemical shift values).



Table 17. ¹H-¹H NOESY interactions between H-(A-H) and H-(a-h).

Figure 444. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing four pairs of neighboring methoxy protons.

4. The neighboring TBTQ aromatic protons were assigned by HSQC, HMBC and COSY interactions as shown in Figure 445-447.



Figure 445. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ showing cross peaks between H-(A-H) and C-(6a-6h). (See Table 18 for chemical shift values).

Table 18. ¹H-¹H HMBC interactions between H-(A-H) and C-(6a-6h).

¹ H-	¹ H-	¹³ C	¹³ C
Atom	NMR	Atom	Atom
	(HMBC)		(HMBC)
А	8.39	6a	152.48
В	8.65	6b	153.09
С	8.78	6c	152.28
D	8.82	6d	152.66
Е	8.85	6e	152.21
F	8.85	6f	152.21
G	8.87	6g	152.52
Н	8.90	бh	154.21



Figure 446. Partial ${}^{1}\text{H}{}^{-13}\text{C}$ HMBC NMR (700 MHz, CD_2Cl_2) spectrum of (**OMe-cube**)₂ showing cross peaks for pairs 1) C-(6a-6h) and H-(I-VIII) and 2) C-(6a-6h) and H-(i-viii)). (See Table 19 for chemical shift values).

Table 19. ¹ H- ¹ H HMBC interactions	between pairs 1) C-(6a-6h)	and H-(I-VIII)	and 2) C-(6a-
бh) and H-(i-viii).			

¹³ C	¹³ C	¹ H-	¹ H-	¹³ C	¹³ C	¹ H-	¹ H-
Atom	Atom	Atom	NMR	Atom	Atom	Atom	NMR
	(HMBC)		(HMBC)		(HMBC)		(HMBC)
ба	152.48	Ι	6.98	ба	152.48	i	6.72
6b	153.09	II	7.54	6b	153.09	ii	7.36
6с	152.28	III	7.33	6с	152.28	iii	7.27
6d	152.66	IV	6.27	6d	152.66	iv	7.14
бе	152.21	V	7.39/7.42	6e	152.21	v	7.19/7.24
6f	152.21	VI	7.39/7.42	6f	152.21	vi	7.19/7.24
6g	152.52	VII	6.83	6g	152.52	vii	7.21
бh	154.21	VIII	7.35	6h	154.21	viii	6.89



Figure 447. Partial ¹H-¹³C HMBC NMR (700 MHz, CD_2Cl_2) spectrum of (**OMe-cube**)₂ showing cross peaks for pairs 1) H-(i-viii) and C-(4a-4h) and 2) C-(4a-4h) and H-(1-8). (To assign the protons 1,4 and 7 clearly ¹H-¹H COSY spectrum was used). (See Table 20 for chemical shift values).

(1-0).							
¹ H-	¹ H-	¹³ C	¹³ C Atom	¹³ C	¹³ C Atom	¹ H-	¹ H-
Atom	NMR	Atom	(HMBC)	Atom	(HMBC)	Atom	NMR
	(HMBC)						(HSQC)
i	6.72	4a	146.11	4a	146.11	1	6.60
ii	7.36	4b	145.49	4b	145.49	2	7.16
iii	7.27	4c	146.38	4c	146.38	3	7.00
iv	7.14	4d	145.99	4d	145.99	4	6.60

146.56/146.66

146.56/146.66

145.99

145.49

4e

4f

4g

4h

146.56/146.66

146.56/146.66

145.99

145.49

5

6

7

8

7.19/7.24

7.19/7.24

7.21

6.89

4e

4f

4g

4h

v

vi

vii

viii

Table 20. ¹H-¹H HMBC interactions for pairs 1) H-(i-viii) and C-(4a-4h) 2) C-(4a-4h) and H-(1-8).

7.06/7.03

7.06/7.03

6.75

6.79

Assignment of Imine protons A and B to the 3D model of (OMe-cube)2

The first imine signal at $\delta = 8.39$ ppm, the most upfield shifted imine peak (labelled as **A**) assigned to the blue imine protons of the dicatenane (Figure 448 and 449) which are found inside the other cube. This imine proton in the catenane with highest shielding effect of the surrounding rings compared to other seven different imine protons of dicatenane. Based on multiple HMBC and HSQC interactions (Figure 435-438, Table 14 and 15), the neighboring imine proton at $\delta = 8.65$ ppm (labelled as **B**) was identified and assigned to the red imine protons of the dicatenane (Figure 448 and 449).



Figure 448. Partial ¹H NMR (600 MHz, CD_2Cl_2) spectrum of (OMe-cube)₂ and assignment of imine protons A and B.



Figure 449. Cartoon representation of one (OMe-cube)₂ showing the assignment of imine protons **A** and **B**.

Assignment of imine protons D and H to the 3D model of (OMe-cube)2

The imine proton **B** ($\delta = 8.65$ ppm) of one cube of the catenane show NOESY cross peaks with the imine proton at $\delta = 8.90$ (labelled as **H**) (Figure 452) and methoxy peak at $\delta = 3.67$ ppm (labelled as **d**) of another cube (Figure 453). Therefore, the imine peak at $\delta = 8.90$ (labelled as **H**) is assigned to the purple imine proton of the dicatenane (Figure 450 and 451). Furthermore, the methoxy peak at $\delta = 3.67$ (labelled as **d**) shows NOESY cross peaks with the methoxy proton at $\delta = 3.11$ (labelled as **b**) (Figure 454) further support the assignment of imine proton **H**. Based on multiple HSQC and HMBC interactions (Figure 435-438, Table 14 and 15), the neighboring imine proton at $\delta = 8.82$ ppm (labelled as **D**) was identified and assigned to the lime green proton of the dicatenane (Figure 450 and 451).



Figure 450. Partial ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ and assignment of imine protons **D** and **H**.



Figure 451. Cartoon representation of (OMe-cube)₂ showing the assignment of imine protons **D** and **H**.



Figure 452. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum showing cross peaks between imine protons



Figure 453. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum showing cross peaks between methoxy protons H-(a-h) and imine protons H-(A-H).



Figure 454. Partial ¹H-¹H NOESY NMR (600 MHz, CD_2Cl_2) spectrum showing cross peaks between methoxy protons.

Assignment of imine protons C and G to the 3D model of (OMe-cube)2

The triply interlocked model of dicatenane shows that the chemical environment of the pair of purple imine proton **H** ($\delta = 8.90$ ppm) and lime imine proton **D** ($\delta = 8.82$ ppm) is more similar to the pair of yellow and orange imine protons than the pair of grey and dark blue imine protons (Figure 456). Therefore, downfield shifted imine signal at $\delta = 8.87$ ppm (labelled as **G**) is assigned to the yellow imine proton of the dicatenane and upfield shifted imine peak at $\delta = 8.78$ (labelled as **C**) is assigned to the orange imine proton of the dicatenane (Figure 455 and 456). Multiple interactions from HMBC and HSQC (Figure 435-438, Table 14 and 15) also shows that these both protons are the neighboring imine protons from the same pair.



Figure 455. Partial ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (OMe-cube)₂ and assignment of proton G and C.



Figure 456. Cartoon representation of (OMe-cube)₂ showing the assignment of imine protons G and C.

Assignment of imine protons E and F to the 3D model of (OMe-cube)2

For the assignment of protons **E** and **F**, NOE interactions of the highly shielded aliphatic protons of the propyl chains of the TBTQ units that are in direct connection to imine proton **A** were investigated. Figure 459 shows NOESY cross peaks of aliphatic protons of the propyl chain at $\delta = 0.48$ ppm with methoxy protons at $\delta = 3.82$ ppm. Based on this interaction, the methoxy peak at $\delta = 3.82$ (labelled as **e**) assigned to the grey methoxy proton of the dicatenane. The methoxy proton **e** shows a NOESY cross peaks with the imine proton at $\delta =$ 8.85 (labelled as **E**) (See Figure 443). Therefore, the imine proton at $\delta =$ 8.85 (labelled as **E**) is assigned to the grey imine protons (Figure 457 and 458). Based on multiple HMBC and HSQC interactions, the imine peak at $\delta =$ 8.85 ppm (labelled as **F**) is assigned to the dark blue proton of the catenane (see Figure 435-438, Table 14 and 15). This is the last pair of neighbouring imine protons. Figure 460 shows NOESY cross peaks of shielded protons of aliphatic chains with the linker aromatic proton ε at $\delta =$ 7.69 further support the assignment of imine protons **E** and **F**.



Figure 457. Partial ¹H NMR (600 MHz, CD_2Cl_2) spectrum of (OMe-cube)₂ and assignment of protons E and F.



Figure 458. Cartoon representation of (OMe-cube)₂ showing the assignment of linker protons E and F.



Figure 459. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum showing cross peaks between methoxy and aliphatic protons.



Figure 460. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) spectrum showing cross peaks between the aliphatic protons and aldehyde linker protons.

-N=CH	δ (ppm)	OCH3	δ (ppm)	Ar <i>H</i> (linker)	δ (ppm)
A	8.39	a	2.85	α	7.24
В	8.65	b	3.11	β	7.34
С	8.78	с	3.86	γ	7.79
D	8.82	d	3.67	δ	7.88
E	8.85	e	3.82	3	7.69
F	8.85	f	3.90	ζ	7.71
G	8.87	g	4.04	η	7.69
Η	8.90	h	3.94	θ	8.03

 Table 21. Assignment of imine, methoxy and linker protons of (OMe-cube)2

 Table 22. Assignment of TBTQ aromatic protons

TBTQ-	δ (ppm)	TBTQ-	δ (ppm)	TBTQ-	δ (ppm)
ArH _i		ArH ₁		ArH _I	
i	6.72	1	6.60	Ι	6.98
ii	7.36	2	7.16	II	7.54
iii	7.27	3	7.00	Ш	7.33
iv	7.14	4	6.60	IV	6.27
V	7.19/7.24	5	7.06/7.03	V	7.39/7.42
vi	7.19/7.24	6	7.06/7.03	VI	7.39/7.42
vii	7.21	7	6.75	VII	6.83
viii	6.89	8	6.79	VIII	7.35



Figure 461. Partial ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (**OMe-cube**)₂ and assignment of imine, methoxy and linker protons.



Figure 462. Partial ¹H NMR (600 MHz, CD₂Cl₂) spectrum of **(OMe-cube)**² and assignment of TBTQ and aliphatic protons.

b) Assignment of protons and carbons of (SMe-cube)₂

¹H NMR, 700 MHz, CD₂Cl₂



assigned by 1D and 2D NMR spectra).



 $^{13}\mathrm{C}\,\mathrm{NMR}$, 700 MHz, $\mathrm{CD_2Cl_2}$



Figure 465. Labelling of protons and carbons to the partial chemical structure of (**SMe-cube**)₂.





Figure 466. Partial ¹³C NMR spectrum (700 MHz, CD₂Cl₂) of (**SMe-cube**)₂. (All the carbon regions were assigned by HSQC, HMBC, DEPT).

Assignment of eight different signals of dialdehyde protons to the 3D model of (SMecube)₂

1. First, dialdehyde linker protons were labelled as α , β , γ , δ , ϵ , ζ , η and θ from lower chemical shift values to higher chemical shift values (Figure 467).



Figure 467. Partial ¹H NMR (700 MHz, CD_2Cl_2) spectrum of (**SMe-cube**)₂ showing the labelling to dialdehyde linker protons.

2. Four pairs of neighboring dialdehyde linker protons and imine protons as shown in Figure 468 were assigned by multiple HSQC and HMBC interactions (see Figure 469-472, Table 23 and 24).



Figure 468. Labelling of protons and carbons of (SMe-cube)₂. (Arrows indicate how neighboring linker and imine protons are identified by HMBC and HSQC measurements).



Figure 469. Partial ¹H-¹³C HMBC NMR (700 MHz, CD_2Cl_2) showing cross peaks between H-(a- θ) and C-(10a-10h).



Figure 470. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) showing cross peaks between C-(10a-10h) and H-(A-H).



Figure 471. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) showing cross peaks between H-(A-H) and C-(12a-12h).



Figure 472. Partial ¹H-¹³C HSQC NMR (700 MHz, CD₂Cl₂) showing cross peaks between C-(12a-12h) and H-(a- θ).

¹ H-	¹ H-	¹³ C	¹³ C	¹³ C	¹³ C	¹ H-	¹ H-
Atom	NMR	Atom	Atom	Atom	Atom	Atom	NMR
	(HMBC)		(HMBC)		(HMBC)		(HMBC)
α	7.62	10b	137.46	10b	137.46	В	8.59
β	7.80	10a	136.22	10a	136.22	А	8.46
γ	7.98	10d	136.44	10d	136.44	D	8.84
δ	8.02	10c	136.87	10c	136.87	С	8.80
3	8.07	10f	136.80	10f	136.80	F	8.845
ζ	8.10	10e	136.63	10e	136.63	Е	8.84
η	8.20	10h	136.82	10h	136.82	Н	9.02
θ	8.25	10g	136.11	10g	136.11	G	8.85

Table 23. HMBC interactions between H-(α - θ) and C-(10a-10h), and C-(10a-10h) and H-(A-H).

Table 24. HMBC interactions between H-(A-H) and C-(12a-12h), and HSQC interactions between C-(12a-12h) and H-(α - θ).

¹ H-	¹ H-	¹³ C	¹³ C	¹³ C	¹³ C	¹ H-	¹ H-
Atom	NMR	Atom	Atom	Atom	Atom	Atom	NMR
	(HMBC)		(HMBC)		(HMBC)		(HSQC)
В	8.59	12b	126.37	12b	126.37	β	7.80
А	8.46	12a	129.47	12a	129.47	α	7.62
D	8.84	12d	127.94	12d	127.94	δ	8.02
С	8.80	12c	127.24	12c	127.24	γ	7.98
F	8.845	12f	127.84	12f	127.84	ζ	8.10
Е	8.84	12e	127.42	12e	127.42	3	8.07
Н	9.02	12h	127.77	12h	127.77	θ	8.25
G	8.85	12g	126.37	12g	126.37	η	8.20



Figure 473. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂ showing four pairs of neighboring linker protons.



Figure 474. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂ showing four pairs of neighboring imine protons.

3. Four neighboring methoxy protons were assigned by HSQC and HMBC interactions as shown in Figure 475 and 476 (Table 25).



Figure 475. Partial ¹H-¹³C HMBC NMR (700 MHz, CD_2Cl_2) showing cross peaks between H-(a- θ) and C-(11a-11h).



Figure 476. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) showing cross peaks between C-(11a-11h) and H-(a-h).

Table 25. HMBC interactions between H-(α - θ) and C-(11a-11h), and C-(11a-11h) and H-(a-h).

¹ H-	¹ H-	¹³ C	¹³ C Atom	¹³ C	¹³ C Atom	¹ H-	¹ H-
Atom	NMR	Atom	(HMBC)	Atom	(HMBC)	Atom	NMR
	(HMBC)						(HMBC)
α	7.62	11a	138.92	11a	138.92	a	1.96
β	7.80	11b	137.60	11b	137.60	b	1.12
γ	7.98	11c	137.83	11c	137.83	с	2.42
δ	8.02	11d	138.30	11d	138.30	d	2.38
3	8.07	11e	137.89	11e	137.89	e	2.62
ζ	8.10	11f	138.13	11f	138.13	f	2.51
η	8.20	11g	178.78	11g	178.78	g	2.27
θ	8.25	11h	137.98	11h	137.98	h	2.45



Figure 477. Partial ¹H NMR (700 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂ showing four pairs of neighboring methoxy protons.

4. The neighboring TBTQ aromatic protons were assigned by HSQC, HMBC and COSY interactions as shown in Figure 478-481.



Figure 478. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) showing cross peaks between H-(A-H) and C-(6a-6h).







Figure 480. Partial ¹H-¹³C HMBC NMR (700 MHz, CD₂Cl₂) showing cross peaks between C-(6a-6h) and H-(i-viii).



Figure 481. Partial ¹H-¹H COSY NMR (700 MHz, CD₂Cl₂) showing cross peaks between H-(I-VIII) and H-(1-8).

Assignment of dialdehyde linker α and β protons to the 3D model of (SMe-cube)₂

The first aldehyde linker signal at $\delta = 7.62$ ppm, the most upfield shifted peak (labelled as α) is assigned to the blue imine protons of the dicatenane (Figure 482 and 483) which are found inside the other cube. This is the only linker aromatic proton in the catenane which experiences the highest shielding effect of the surrounding rings compared to other seven different imine protons of dicatenane. Based on multiple HMBC and HSQC interactions (Figure 469-472, Table 23), the neighboring dialdehyde linker proton at $\delta = 7.80$ ppm (labelled as β) was identified and assigned to the red aldehyde protons of the dicatenane (Figure 482 and 483).



Figure 482. Partial ¹H NMR (600 MHz, CD_2Cl_2) spectrum of (**SMe-cube**)² and assignment of linker protons α and β .



Figure 483. Cartoon representation [SMe-cube]₂ showing the assignment of linker protons α and β .

Assignment of dialdehyde linker protons η and θ to the 3D model of (SMe-cube)₂

The dialdehyde linker proton β ($\delta = 7.80$ ppm) of one cube of catenane shows a NOESY cross peaks to the imine proton at $\delta = 9.02$ (labelled as **H**) and SMe proton at $\delta = 2.27$ ppm (labelled as **g**) of another cube (Figure 486 and 487). Therefore, the dialdehyde linker peak at $\delta = 8.25$ (labelled as θ) is assigned to the purple linker proton of the dicatenane (Figure 484 and 485). Furthermore, the imine proton at $\delta = 8.25$ (labelled as **B**) shows a NOESY cross peak with dialdehyde linker proton at $\delta = 8.25$ (labelled as **B**) shows a NOESY cross peak with dialdehyde linker proton at $\delta = 8.25$ (labelled as **B**) shows a NOESY cross peak with dialdehyde linker proton at $\delta = 8.25$ (labelled as **B**) and SMe proton at $\delta = 2.27$ ppm (labelled as **g**) (Figure 487), which further supports the assignment of dialdehyde linker proton θ . Based on multiple HSQC and HMBC interactions (Figure 469-472, Table 23 and 24), the neighboring linker peak at $\delta = 8.20$ ppm (labelled as η) was identified and assigned to the lime green proton of the dicatenane (Figure 484 and 485).



Figure 484. Partial ¹H NMR (600 MHz, CD_2Cl_2) spectrum of (SMe-cube)₂ and assignment of Imine protons η and θ .



Figure 485. Cartoon representation of [SMe-cube]₂ showing the assignment of linker protons η and θ .


Figure 486. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) showing cross peaks between dialdehyde linker proton and imine protons.



Figure 487. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) showing cross peaks between dialdehyde linker proton and SMe proton.



Figure 488. Partial ¹H-¹H NOESY NMR (600 MHz, CD₂Cl₂) showing cross peaks between imine protons and SMe protons.

Assignment of imine protons γ and δ to the 3D model of (SMe-cube)₂

For the assignment of protons γ and δ , NOE interactions of the highly shielded aliphatic protons of the propyl chain of the TBTQ units that are in direct connection to dialdehyde linker proton α were investigated. Figure 491 shows the NOESY cross peaks of aliphatic protons of the propyl chain at $\delta = 0.58$ ppm (CH₃) and 0.70 ppm (CH₂) with dialdehyde linker proton at $\delta = 8.02$ ppm (labelled as δ). Based on this interaction, the dialdehyde linker peak at $\delta = 8.02$ (labelled as δ) assigned to the gray linker proton of the dicatenane (Figure 489 and 490). Based on multiple HMBC and HSQC interactions (Figure 469-472, Table 23 and 24) the dialdehyde linker peak at $\delta = 7.98$ ppm (labelled as γ) is assigned to the dark blue proton of the catenane.



Figure 489. Partial ¹H NMR (600 MHz, CD_2Cl_2) spectrum of (SMe-cube)₂ and assignment of protons γ and δ .



Figure 490. Cartoon representation of [SMe-cube]₂ showing the assignment of linker protons γ and δ .



Figure 491. Partial ¹H-¹H NOESY NMR (700 MHz, CD₂Cl₂) showing cross peaks between dialdehyde linker protons and aliphatic protons.

Assignment of imine protons ε and ζ to the 3D model of (SMe-cube)₂

The triply interlocked model of dicatenane shows that the chemical environment of the pair of lime green dialdehyde linker proton η ($\delta = 8.20$ ppm) and purple dialdehyde linker proton θ ($\delta = 8.25$ ppm) is similar to the pair of yellow and orange dialdehyde protons. Therefore, the downfield shifted imine signal at $\delta = 8.10$ ppm (labelled as ζ) is assigned to the yellow dialdehyde linker proton of the dicatenane and upfield shifted imine peak at $\delta = 8.07$ (labelled as ε) is assigned to the orange imine proton of dicatenane (Figure 492 and 493). Multiple interactions from HMBC and HSQC (Figure 469-472, Table 23 and 24) also shows that these both protons are the neighboring dialdehyde linker protons from the same pair.



Figure 492. Partial ¹H NMR (600 MHz, CD₂Cl₂) spectrum of (**SMe-cube**)₂ and assignment of proton ε and ζ .



Figure 493. Cartoon representation of $[SMe-cube]_2$ showing the assignment of linker protons ϵ and ζ .

SMe protons and imine protons were assigned similarly like imine protons and the complete assignment is shown in below.

ArH (linker)	δ (ppm)	-N=CH	δ (ppm)	SCH ₃	δ (ppm)
α	7.62	Α	8.46	a	1.96
β	7.80	В	8.59	b	1.12
γ	7.98	С	8.80	С	2.42
δ	8.02	D	8.84	d	2.38
3	8.07	Ε	8.84	e	2.62
ζ	8.10	F	8.845	f	2.51
η	8.20	G	8.85	g	2.27
θ	8.25	Н	9.02	h	2.45

Table 26.





Figure 495. ¹H NMR (700 MHz, TCE, 410 K) and number of imine and TBTQ aromatic signals of (**SMe-cube**)₃. (24 *TBTQ aromatic protons (protons ortho to each other) are assigned mainly from* ¹H-¹H COSY (Fig. S232) and other 2D spectra. For 2D spectra please see Figs. S229-S244)



Figure 496. Highlighting of 12 magnetically equivalent methoxy groups for a singly interlocked (**OMe-cube**)₂ (not observed).



14. Variable temperature NMR studies with (OMe cube)3 and (SMe cube)3

Figure 497 Temperature dependent ¹H NMR (700 MHz, Toluene-d₈) spectra of (**OMe cube**)₃. *DCM, +silicon grease.



Figure 498. Temperature dependent partial ¹H NMR (700 MHz, Toluene-d₈) spectra of (**OMe cube**)₃.



Figure 499. Temperature dependent partial ¹H NMR (700 MHz, Toluene-d₈) spectra of (**OMe cube**)₃. *DCM, +silicon grease.



Figure 500. Temperature dependent partial ¹H NMR (700 MHz, Toluene-d₈) spectra of (**OMe cube**)₃.



Figure 501. Temperature dependent ¹H NMR (700 MHz, CD₂Cl₂) spectra of (**SMe cube**)₃. ⁺THF, *silicon grease.



Figure 502. Temperature dependent partial ¹H NMR (700 MHz, CD₂Cl₂) spectra of (SMe cube)₃.



Figure 503. Temperature dependent ¹H NMR (700 MHz, $C_2D_2Cl_4$) spectra of (SMe cube)₃. ⁺CHCl₃ *THF.



Figure 504. Temperature dependent partial ¹H NMR (700 MHz, C₂D₂Cl₄) spectra of (**SMe cube**)₃.



Figure 505. Temperature dependent partial ¹H NMR (700 MHz, C₂D₂Cl₄) spectra of (**SMe cube**)₃. *THF.

15. ¹H NMR Kinetic srudies:



The **OMe-cube** \rightarrow (**OMe-cube**)₂ reaction was treated as second order reaction. ¹H NMR measurements were performed on a 700 MHz NMR instrument. The rate constant k_1 was determined from the graphical evaluation of the slope with the following equation (1):



Figure 506. ¹H NMR kinetic data of two measurements. Concentration vs. time plot **OMe**cube \rightarrow (**OMe**-cube)₂ reaction a) and c), and graphical evaluation rate constant k_1 b) and d).

Cage	CDCl ₃	OMe-	TFA	T (K)	<i>k</i> ₁ (x 10 ⁻² M ⁻¹	<i>k</i> ₁ (x 10 ⁻² M ⁻¹
	(µL)	cube (µmol)	(µmol)		s ⁻¹)	s -1)
OMe aube	500	0.676	0.653	295	10.1	10.5 + 0.4
One-cube	500	0.676	0.653	295	10.8	10.3 ± 0.4

 Table 27. Experimental details of ¹H NMR kinetic measurements

The average value of two measurements reported as k_1 . The standard deviations calculated using STDEV.P function of Microsoft Excel 2016.

Calculation of cage to catenane ratio from ¹H NMR spectrum



Integration of cage = (cage+catenane peak)-5

Integration of cage = (7.40)-5 = 2.40

cage:catenane ratio is 2.40:1

Cage has 24 protons per imine signals and catenane has 6 imine protons per imine signals.

Cage:catenane ratio is (2.40/24):(1/6) = 0.100:0.167

cage:catenane ratio is (0.100/0.100):(0.167/0.100) = 1.00:1.67

Equation used to find the concentrations (M) of cage and catenanes in the NMR sample used for kinetic studies

mass of cage + mass of catenane = 0.0038 g

(no. of moles of cage \times mol. wt. of cage) + (no. of moles of catenane + mol. wt. of catenane) = 0.0038 g

(for example, the ratio of **Cage:Catenane** calculated from the ¹H NMR spectra is **1.00:1.67**) (NMR spectra number 112.)

 $(n_{cage} \times 5623.23 \text{ g/mol}) + (n_{cat} \times 11246.47 \text{ g/mol}) = 0.0038 \text{ g}$

(e.g. Cage to catenane ratio obtained from ¹H NMR is 1:1.67)

 $n_{cage}=A mol, n_{cat}=1.67 \times A mol$

 $(A \text{ mol} \times 5623.23 \text{ g/mol}) + (1.67 \times A \text{ mol} \times 11246.47 \text{ g/mol}) = 0.0038 \text{ g}$

 $5623.23 \text{ g/mol} \times \text{A mol} + 18781.60 \text{ g/mol} \times \text{A mol} = 0.0038 \text{ g}$

A mol = 0.0038 g/24404.83 g/mol

A mol = $0.000001557 \text{ mol} = n_{cage}$

 n_{cat} = 1.67 x 0.0000001557 mol = 0.00000026 mol

[Cage] = 0.0000001557 mol/ 0.0005L = 0.0003114 M,

[Catenane] = 0.00000026 mol/ 0.0005 L = 0.00052 M

16 Catenation Test of OMe-cube without acid catalyst trifluoroacetic acid

Procedure: In a 1.5 mL vial **OMe-cube** (4 mg, 0.71 μ mol) was completely dissolved in CDCl₃ (526 μ L) and then reaction mixture was transferred into an NMR tube and analysed by NMR spectroscopy at RT at different time intervals. (CDCl₃ was passed through the basic alumina just before using it for the reaction).



Figure 507. Time dependent ¹H NMR (300 MHz, CDCl₃) spectra monitoring (**OMe-cube**) \rightarrow (**OMe-cube**)₂ reaction without TFA catalyst. *1,1,2,2-tetrachloroethane.

17. Mechanistic Studies of catenane Formation by Mass Spectrometry

1) Mechanistic Studies for OMe-cube and (OMe-cube)₂

Procedure: To a mixture of **OMe-cube** (2 mg, 0.36 μ mol) and ¹⁵N labelled ***OMe-cube** (2 mg, 0.36 μ mol) in a 1:1 ratio in CDCl₃ (0.53 mL), TFA (5.3 μ L of a 0.131M solution in CDCl₃, 0.69 μ mol) was added and the reaction mixture was stirred for 3 days at RT and then analysed by TIMS-TOF mass spectrometry.



Figure 508. Full TIMS-TOF (DCTB) mass spectra of crude reaction mixture where 1:1 mixture of **OMe-cube** and ¹⁵N labelled ***OMe-cube** was used for catenane reaction.



Figure 509. a) and c) partial TIMS-TOF (DCTB) mass spectra of crude reaction mixture where 1:1

mixture of **OMe-cube** and ¹⁵N labelled ***OMe-cube** was used for catenane reaction. c) and d) calculated mass spectra.

2) Mechanistic Studies for OH-cube

Procedure: To a mixture of **OH-cube** (2 mg, 0.38 μ mol) and ¹⁵N labelled ***OH-cube** (2 mg, 0.38 μ mol) in a 1:1 ratio in CD₂Cl₂ (0.53 mL), TFA (5.6 μ L of a 0.131M solution in CDCl₃, 0.73 μ mol TFA) was added and the reaction mixture was stirred for 3 days and then analysed by TIMS-TOF mass spectrometry.



Figure 510. Full TIMS-TOF (DCTB) mass spectra of crude reaction mixture where 1:1 mixture of **OH-cube** and ¹⁵N labelled ***OH-cube** was stirred at a) RT and b) at 80 °C for scrambling studies.



Figure 511. a) TIMS-TOF (DCTB) mass spectra of pure **OH-cube** and ¹⁵N labelled ***OH-cube**. b) and c) TIMS-TOF (DCTB) mass spectra of reaction mixture where 1:1 mixture of **OH-cube** and ¹⁵N labelled ***OH-cube** was used for catenane reaction at RT and at 80 °C respectively. d) calculated mass spectra of **M**⁺ (**OH-cube**), ¹⁴N₁₂:¹⁵N₁₂ (**OH-cube**: ***OH-cube**, **1:1**), **M+Na** (**OH-cube+Na**), ***M**⁺ (***OH-cube**), ¹⁴N₁₂:¹⁵N₁₂+Na (**OH-cube**: ***OH-cube** +Na, **1:1**) and ***M+Na** (***OH-cube+Na**).



Figure 512: DOSY NMR of 1:1 mixture of pure (**OMe-cube**)₂ (6.7 mg, 0.593 mmol) and (**OME-cube**)₃ (10.0 mg, 0.593 μ mol) in CD₂Cl₂ (0.5 mL).

18. Models of Weak Interactions



Figure 513. Comparison of weak interactions. **a** X-ray structure of a quinoxalinophenthrophenazine with methoxy-methoxy interactions $(d_{C\cdots O} = 3.1 \text{ Å})$.^[S11] **b** and **c** Cooper's knot $(d_{C\cdots O} = 3.5 \text{ Å})$.^[S12] **d** and **e** Model of possible conformation of (**OMe-cube**)₂ with three methoxy-methoxy interactions with $d_{C\cdots O} = 3.0 \text{ Å}$. **f** and **g** Model of possible conformation of (**SMe-cube**)₂ with three sulfur-sulfur interactions with $d_{S\cdots S} = 3.8 \text{ Å}$.

19. Computational Details

All quantum-chemical calculations were performed by employing the Gaussian16 program package.^[13] The theoretical approach is based on the semi-empirical PM6 method.^[14] The geometries of the regarded species were fully optimized, and ground states of the monomeric structures were confirmed by using frequency calculations to not exhibit any imaginary frequency. *For detailed computational data, see Supporting Information File 2 (or xyz zip files).* Here, only the results are discussed.

Heat of formations

All heat of formations were calculated for minimum geometries. As

28 shows, conformer 2 is the more stable conformer for all cage compounds. The regarding geometries are shown in Figures 512, 513 and 522. For all cages the propyl chains of the TA-TBTQ units were shortened to methyl groups.

Table 28. PM6 calculated heat of formations for the different conformers of the cage compounds. For structures, see Figures 512, 513 and 522.

Cage Compound		Δ _f H [kJ mol ⁻¹]
OH-cube	Type 1 conformer	742
Oll-cube	Type 2 conformer	728
OMe who	Type 1 conformer	1462
Ome-cube	Type 2 conformer	1438
CM	Type 1 conformer	5346
Sivie-cube	Type 2 conformer	5326





Figure 514. PM6 optimized structures of the **OH-cube**. **a**) Type 1 conformer, **b**) Type 2 conformer. white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen.



Figure 515. PM6 optimized structures of the **OMe-cube**. **a**) conformer 1, **b**) conformer 2. white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen.



Figure 516. PM6 optimized structure of a hypothetically triply interlocked (**OH-cube**)₂ (Type 1 conformer). **a**) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting both single cages in red and blue.



Figure 517. PM6 optimized structure of the triply interlocked (**OMe-cube**)₂ (Type 1 conformer). **a**) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting both single cages in red and blue.



Figure 518. PM6 optimized structure of a hypothetical singly interlocked (**OMe-cube**)₂ (Type 1 conformer). **a**) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting both single cages in red and blue.



Figure 519. PM6 optimized structure of a hypothetical singly interlocked (**OMe-cube**)₂ (Type 2 conformer). **a**) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting both single cages in red and blue.



Figure 520. PM6 optimized structure of triply interlocked **[(OMe-cube)@(OMe-cube)]@** (**OMe-cube**) (Type 1 conformer). **a**) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting three cages in red, blue and green colour. Hydrogen atoms are omitted for clarity.





Figure 521. PM6 optimized structure of linear triply interlocked (OMe-cube)@(OMe-cube)@(OMe-cube) (Type 1 conformer, 1 and 4 position of middle cage is interlocked). a) structure coloured after the elements (white = hydrogen, grey = carbon, blue = nitrogen, red = oxygen) and b) highlighting outer two cages in blue color and middle in e colour. Hydrogen atoms are omitted for clarity.



Figure 522. PM6 optimized structure of triply interlocked trimeric catenane with **syn-distal type** connectivity (middle cage is interlocked in 1 and 3 position) (Type 1 conformer). **a**) structure coloured after the elements (grey = carbon, blue = nitrogen, red = oxygen) and **b**) highlighting three cages in red and blue colour. Hydrogen atoms are omitted for clarity. **a**)



b)



Figure 523. PM6 optimized structure of triply interlocked trimeric catenane with synproximal type connectivity (middle cage is interlocked in 1 and 2 position) (Type 1 conformer). a) structure coloured after the elements (grey = carbon, blue = nitrogen, red = oxygen) and b) highlighting three cages in red and blue colour. Hydrogen atoms are omitted for clarity.



Figure 524. PM6 optimized structures of the **SMe-cube**. **a**) Type 1 conformer, **b**) Type 2 conformer. white = hydrogen, grey = carbon, blue = nitrogen, yellow = sulphur.

Entry	Cage/catenane	Cage/catenane Synthesis	Yield (%)
1	OH cage	1st run	84
		2nd run	85
		3rd run	82
2	OMe cube	1st run	80
		2nd run	76
3	(OMe-cube) ₂	1st run	60
		2nd run	46
		3rd run	47
4	(OMe-cube) 3	1st run	80
		2nd run	79
		3rd run	84
5	H-cube	1st run	89
		2nd run	90
6	Me-cube	1st run	80
		2nd run	84
7	Et-cube	1st run	75
		2nd run	79
8	(Et-cube) 2	1st run	35
		2nd run	38
9	OEt-cube	1st run	84
		2nd run	81
10	(OEt cube) 2	1st run	25
		2nd run	32
11	O-ipr-cube	1st run	95
		2nd run	81
12	SMe-cube	1st run	86
		2nd run	80
13	(Sme-cube) 2	lst run	70
		2nd run	66 70
14	(SMe-cube) 2	1st run	58
1.5		2nd run	54
15	SC(CH ₃) ₃ -cube	1st run	/5
10	Deck	2nd run	81
10	Br-cube	1st run	93
17	II/OMa awk -	2nd run	92
1/	n/Ome-cube	1st run 2nd min	33 26
10	(U/OMo onho):	∠nu run 1st mm	30 42
18	(II/OMe-cube)2	1st run 2nd min	42
		2na run	44

 Table 29. Cage/catenane reactions reproducibility details
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