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## Variation in the SERPINA6/SERPINA1 locus alters morning plasma cortisol, hepatic corticosteroid binding globulin expression, gene expression in peripheral tissues, and risk of cardiovascular disease

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## Abstract

The stress hormone cortisol modulates fuel metabolism, cardiovascular homeostasis, mood, inflammation and cognition. The CORtisol NETwork (CORNET) consortium previously identified a single locus associated with morning plasma cortisol. Identifying additional genetic variants that explain more of the variance in cortisol could provide new insights into cortisol biology and provide statistical power to test the causative role of cortisol in common diseases. The CORNET consortium extended its genome-wide association meta-analysis for morning plasma cortisol from 12,597 to 25,314 subjects and from ~2.2M to ~7M SNPs, in 17 population-based cohorts of European ancestries. We confirmed the genetic association with SERPINA6/SERPINA1. This locus contains genes encoding corticosteroid binding globulin (CBG) and α1-antitrypsin. Expression quantitative trait loci (eQTL) analyses undertaken in the STARNET cohort of 600 individuals showed that specific genetic variants within the SERPINA6/SERPINA1 locus influence expression of SERPINA6 rather than SERPINA1 in the liver. Moreover, trans-eQTL analysis demonstrated effects on adipose tissue gene expression, suggesting that variations in CBG levels have an effect on delivery of cortisol to peripheral tissues. Two-sample Mendelian randomization analyses provided evidence that each genetically-determined standard deviation (SD) increase in morning plasma cortisol was associated with increased odds of chronic ischaemic heart disease (0.32, 95% CI 0.06 to 0.59) and myocardial infarction (0.21, 95% CI 0.00 to 0.43) in UK Biobank and similarly in CARDIoGRAMplusC4D. These findings reveal a causative pathway for CBG in determining cortisol action in peripheral tissues and thereby contributing to the aetiology of cardiovascular disease.

**Declaration of Interests** The authors declare no competing interests.

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## Introduction

Cortisol plays a vital role in adaptation to environmental stress, modulating fuel metabolism, cardiovascular homeostasis, mood, memory, and inflammation<sup>1</sup>. Cortisol levels vary throughout the day under the control of the hypothalamic-pituitary-adrenal (HPA) axis. Patients with tumours causing excess cortisol develop Cushing's syndrome, characterised by a host of features including obesity, hypertension, diabetes mellitus, depression, cognitive impairment and osteoporosis with an excess mortality due primarily to cardiovascular disease<sup>2</sup>. Similarly, higher plasma cortisol in the population associates with hypertension, hyperglycaemia, cardiovascular disease, type 2 diabetes, cognitive dysfunction, and depression, while lower cortisol associates with immunological abnormalities and post-traumatic stress disorder<sup>3–10</sup>.

We established the CORtisol NETwork (CORNET) consortium with the initial aim of identifying genetic determinants of inter-individual variation in HPA axis function. A genome-wide association meta-analysis (GWAMA), investigating ~2.2 M SNPs in 12,597 individuals from 11 European cohorts, identified a single locus on chromosome 14 associated with morning plasma cortisol at genome-wide significance<sup>11</sup>. The locus spans *SERPINA6* and *SERPINA6* and *serPINA6*, a protein that binds cortisol in the blood. However, it was unclear if the effect is mediated directly through *SERPINA6* or indirectly through the product of *SERPINA1*, a1-antitrypsin, which is involved in regulating cleavage and inactivation of CBG<sup>12</sup>. Using these genetic variants as a proxy for morning plasma cortisol is a causal risk factor for coronary heart disease, but the odds ratio was not statistically significant (OR: 1.06, 95% CI: 0.98-1.15)<sup>13</sup>. An independent study has confirmed recently that common variants in *SERPINA6* are associated with plasma cortisol and with coronary artery disease<sup>14</sup>.

The genetic variants in the *SERPINA6/A1* locus explain only ~0.5% of the variance in morning plasma cortisol. Moreover, attempts to identify genetic variants associated with an alternative phenotype of salivary cortisol have not been successful<sup>15</sup>. Identifying additional genetic variants that explain more of the variance in cortisol could provide new insights into cortisol biology and statistical power to test the causative role of cortisol in the aetiology of other common diseases. We aimed: to identify additional specific loci influencing cortisol; to refine where in the *SERPINA6/A1* locus there is an influence on cortisol; to establish whether *SERPINA6/A1* variation influences tissue-specific expression of CBG and a1-antitrypsin; and to confirm whether high cortisol is causal in ischaemic heart disease and test if it is causal in other common diseases. To achieve these aims, we undertook an extended GWAMA analysis, with more subjects and more SNPs than the original CORNET GWAMA<sup>11</sup>, and used the results to provide instruments for expression quantitative trait loci (eQTL) and Mendelian randomization analyses.

## Methods

## Genome-wide association meta-analysis study

We performed a meta-analysis of genome-wide association studies of morning plasma cortisol in 25,314 subjects from 17 European population-based cohorts: CROATIA-Vis (n=886), CROATIA-Korcula (n = 897), CROATIA-Split (n = 493), ORCADES (n = 1974), Rotterdam Study (n = 2870), NFBC1966 (n = 1324), Helsinki Birth Cohort Study 1934–44 (n = 399), ALSPAC (n = 1487), PREVEND (n = 1151), PIVUS (n = 919), Raine Study (n = 860), ET2DS (n = 847), MrOS-Sweden (n = 969), KORA (n=1651), TwinsUK (n=5654), SHIP (n=910) and VIKING (n=2073). Characteristics of the study populations are presented in Table S1 and details of each cohort are provided in supplementary material. All individuals were of European ancestries. Exclusion criteria were current glucocorticoid use, pregnant or breast-feeding women, and twins (exclusion of one of each twin pair). Cortisol was measured by immunoassay in blood samples collected from study participants between 0700 and 1100 h in all cohorts except for in TwinsUK which measured cortisol using liquid chromatography-mass spectrometry. All participants provided written informed consent and studies were approved by local Research Ethics Committees and/or Institutional Review Boards.

Each study performed linear regression on z-scores of log-transformed morning plasma cortisol (additive genetic effects), adjusted for sex, age and cohort-specific genetic ancestry. Additional models also adjusted for smoking and body mass index. Imputation of the genechip results used the 1000 Genomes European population reference panel. Details of the genotyping, imputation and cohort-specific adjustment for genetic ancestry are provided in Table S2 and Supplementary Material.

Quality control was carried out on the imputed genome-wide data for all 17 studies prior to meta-analysis; this excluded all SNPs with a minor allele frequency (MAF) <0.5%, call rate <95%, and poor imputation quality (MACH R2\_HAT<0.30, IMPUTE PROPER\_INFO<0.60, BEAGLE INFO<0.30, as appropriate). Furthermore, only SNPs with estimates from at least four studies were included, resulting in a final number of 8,452,427 SNPs. Quantile-quantile (QQ) plots and genomic control (lambda) were used to assess evidence for population structure that was not accounted for in association analyses. Sex chromosomes were not analysed. Quality control at the study-level and meta-level was performed using EasyQC software<sup>16</sup>.

The results from all cohorts were combined into a fixed-effects meta-analysis using Stouffer's method with weighting of Z-scores proportional to the square-root of the number of individuals in each sample, using METAL software<sup>17</sup>. Manhattan and QQ plots visualised the results using EasyStrata software<sup>18</sup>.

## **SNP-based heritability**

Linkage disequilibrium (LD) score regression exploits the relationship between SNPphenotype association strengths and LD patterns<sup>19</sup>. Assuming true causal effects, the SNPs which are in higher LD with nearby SNPs are expected to have more inflated test statistics, because they are more likely to tag causal variants with stronger effects. The SNP

heritability was estimated using LD score regression v1.9.0<sup>19</sup>. Since imputation quality can confound LD score regression results, we restricted the analysis to a list of well-imputed SNPs, as recommended by the software authors. After applying default quality control settings, the final SNP number was 1,028,327. The genome-wide summary statistics were partitioned into functional categories using the method described by Finucane et al.<sup>20</sup>. More details of the partitioned heritability method can be found in the Supplementary Materials.

## **Genetic correlations**

Genetic correlations between morning plasma cortisol and selected diseases and traits from UK Biobank were estimated using bivariate LD score regression. This technique examines the correlation structure of genetic effects of SNPs across the genome. The data processing pipeline devised by Bulik-Sullivan *et al.*<sup>21</sup> was followed using LD Hub v1.9.0 software<sup>22</sup>.

## Gene- and pathway-based association analysis

Gene- and pathway-based associations, which assign SNPs to genes and biological pathways respectively, were performed using MAGMA and FUMA software<sup>23,24</sup>. SNPs were assigned to 18 062 genes using the National Center for Biotechnology Information build 37.3. The gene boundary was defined as the start and stop site of each gene. The European panel of the 1000 Genomes data (phase 1, release 3) was used as a reference panel to account for linkage disequilibrium (LD) between the SNPs. A Bonferroni correction was used to control for 18 062 tests ( $\alpha$ =0.05/18 062; P<2.768 × 10<sup>-6</sup>).

# eQTL analysis in the Stockholm Tartu Atherosclerosis Reverse Networks Engineering Task (STARNET) study

Expression Quantitative Trait Loci (eQTL) analyses were undertaken in the STARNET study which is composed of Caucasian individuals of Eastern European origin (30% female), with a confirmed diagnosis of coronary artery disease. Of these individuals 27% had diabetes, 77% had hypertension and 37% had suffered a myocardial infarction before the age of 60 years. Genotyping and RNA sequencing of seven vascular and metabolic tissue samples from 600 patients undergoing coronary artery bypass surgery was performed<sup>25</sup>. The tissue sets available in STARNET include; whole blood, atherosclerotic-lesion free internal mammary artery, atherosclerotic aortic root, subcutaneous fat, visceral abdominal fat, skeletal muscle, and liver. Whole blood samples were taken pre-operatively and the remaining tissue biopsies were obtained during surgery.

For the analysis, we included 580 SNPs within *SERPINA6/SERPINA1* locus on chromosome 14 identified from the CORNET plasma cortisol GWAMA, defined as within 100 Kb of *SERPINA6* ( $\pm$  of the transcription start/end point), with no missing data, and Minor Allele Frequency (MAF) > 5%. All transcripts were annotated using the Human Genome Reference Consortium Human Build 37 (GRCh37).

Transcriptome-wide eQTL associations between selected SNPs and all genes expressed in the STARNET tissues were tested using the Kruskal Wallis test, a non-parametric ANOVA used to determine whether sample groups originate from the same distribution that has been used previously in genetic association studies<sup>26</sup>. Kruskal Wallis test statistics and p-values

were computed using the kruX algorithm<sup>27</sup>, using available pre-processed and normalized genotype and expression data matrices<sup>25</sup> as input. Multiple testing correction was performed for all SNP associations with every STARNET gene in each tissue separately by calculating q-values<sup>28</sup> using the qvalue package in Python. eQTL associations with *SERPINA6* were visualised using the GWAS visualisation tool LocusZoom<sup>29</sup>. The allelic effect of individual SNPs on *SERPINA6* was visualised by constructing boxplots for each allele.

The global tissue-specific effect on gene expression for SNPs associated with plasma cortisol at a genome-wide level of significance was depicted using Q-Q plots showing the observed transcriptome-wide SNP-gene associations against the expected uniform distribution. Deviation from the uniform distribution was tested using the Kolmogorov Smirnov test statistic and p-value for each SNP.

Bayes factor colocalisation analysis was performed in R using the package Coloc<sup>30</sup>. eQTL analysis was repeated using linear regression with the R package MatrixEQTL<sup>31</sup> to obtain beta values required for colocalisation. linear regression and Kruskal-Wallis p-values were consistent (Figure S7). For visualisation of the colocalisation event, Linkage Disequilibrium with the lead SNP was calculated using the package LDlinkR<sup>32</sup>.

### **Two-sample Mendelian randomization**

The instrument for morning plasma cortisol consisted of independent ( $r^2 < 0.3$ ) genetic variants that reached a genome-wide level of significance ( $P < 5 \times 10^{-8}$ ). To detect independent top SNPs we used the clumping function as implemented in PLINK. The European samples from the 1000 Genomes Project were used to estimate LD between SNPs. Amongst those SNPs within 1000kb and  $r^2 < 0.3$ , only the SNP with the lowest P-value was retained.

Two-sample Mendelian randomization<sup>33</sup> was used to estimate the causal effect of morning plasma cortisol on hypothetically cortisol-related diseases and traits (chronic ischaemic heart disease, myocardial infarction, diabetes mellitus, body mass index and osteoporosis) available in UK Biobank and publicly available GWAS consortia in the MR Base platform<sup>34</sup>.

Genetic instruments for various traits/conditions were also constructed to estimate the causal effect these had on morning plasma cortisol (bidirectional Mendelian randomization). Details of the selected disease and traits, the population, consortia details and genetic instruments are provided in Table S3. The SNP-cortisol estimate was divided by the SNP-outcome estimate (Wald ratio method<sup>35</sup>) and then combined using inverse variance weighting. Additional analyses described in the Supplementary Material were performed to investigate the robustness of this causal estimate and any potential pleiotropic effects.

## Results

## Genome-wide association meta-analysis

The genome-wide association meta-analysis of morning plasma cortisol levels in 25,314 identified a single locus on chromosome 14 reaching a genome-wide level of significance (p  $< 5 \times 10^{-8}$ ) (Figure 1). This is the same locus as the CORNET consortium previously

identified<sup>11</sup> and includes the *SERPINA6* gene, encoding corticosteroid binding globulin (CBG), and the *SERPINA1* gene, encoding  $\alpha$ 1-antitrypsin, an inhibitor of neutrophil elastase which cleaves and inactivates CBG. In an additive genetic model, the top SNP rs9989237 reported a per minor allele effect of 0.11 cortisol z-score (p =  $2.2 \times 10^{-19}$ ). The effect allele frequency was 0.22 and this variation explained 0.13% of the morning plasma cortisol variance. Within this locus we identified four blocks of SNPs in low LD (r<sup>2</sup> < 0.3) visualised in Figure 1 and in Figure S1.

## SNP-based heritability

Using LD score regression, common SNPs across the genome were found to explain 4.2% (s.e. 1.9%) of the phenotypic variation of morning plasma cortisol. There was no evidence of enrichment in a particular cell type, and particularly not in the adrenal/pancreas or liver cell type groups of SNPs, when partitioning the heritability into functional cell types (all P-values > 0.5, Table S4).

## **Genetic correlations**

LD score regression was used to test whether genetic variants associated with morning plasma cortisol also contribute to health-related traits. Estimated genetic correlations are presented in Figure S2. There was evidence of a positive genetic correlation between morning plasma cortisol and acute myocardial infarction ( $r_g = 0.50, 95\%$  CI 0.04 to 0.97) and a negative genetic correlation with BMI ( $r_g = -0.32, 95\%$  CI -0.54 to -0.10).

## Gene-based and pathway-based association analyses

Gene-based association analysis identified three genes, *SERPINA6*, *SERPINA1* and *SERPINA10*, all located on chromosome 14, that attained genome-wide significance following correction for multiple comparisons (Figure S3). Pathway-based association analysis identified the top pathway as metabolism of lipids and lipoproteins ( $P = 1.6 \times 10^{-5}$ ) (Table S5).

## **Tissue-specific eQTL analyses**

The statistical effects of 580 SNPs in the *SERPINA6* region on gene expression were obtained using individual-level genotype and RNA-sequencing data from seven vascular and metabolic tissues from the STARNET study<sup>25</sup>. Following multiple testing correction (q 0.05), 32 cis-eQTLs for *SERPINA6* were identified in the liver, the only tissue where *SERPINA6* is highly expressed (Figure 2A). Of these 32 cis-eQTLs, 21 were also at genome-wide significance in CORNET (p  $5 \times 10^{-8}$ ), and therefore also associated with variation for plasma cortisol (Table S6).

The global effect on tissue-specific gene expression of representative SNPs from each LD block was assessed using the distribution of transcriptome-wide eQTL p-values (Figure S4 and Figure S5). LD block 2, represented by the SNP rs2736898, with the alternate allele C, exerted a negative effect on *SERPINA6* expression in liver (q = 0.00015) (Figure 2B) and showed the strongest tissue-specific effects, particularly in visceral abdominal fat, subcutaneous fat and liver (Figure 2C). In all cases the allele associated with higher plasma

cortisol in the GWAMA was the allele associated with higher *SERPINA6* expression in STARNET (Figure S6).

To determine if the signal identified for *SERPINA6* cis-eQTLs in liver and SNPs associated with plasma cortisol are driven by the same causal variant, Bayes factor colocalisation analysis was performed while accounting for allelic heterogeneity<sup>30</sup> (Figure 3). The probability of both traits sharing a causal variant was low (40.6%) when examining all SNPs within 100Kb of *SERPINA6*. However, when examining each LD block individually, the block represented by rs2736898 returns a 99.2% probability of shared causal variant in this region (Table S7).

## **Two-sample Mendelian randomization**

The clumping procedure identified four SNPs (rs9989237, rs2736898, rs11620763, rs7146221) as markers representing genome-wide significant signals in this region. Twosample Mendelian randomization analyses provided evidence that each genetically determined standard deviation (SD) increase in morning plasma cortisol was associated with an increased risk of chronic ischaemic heart disease (0.32, 95% CI 0.06 to 0.59) and myocardial infarction (0.21, 95% CI 0.00 to 0.43) in UK Biobank (Figure 4a). Similar estimates were observed for these disease outcomes in non-UK Biobank cohorts (CARDIoGRAM plus C4D, Figure 4b).

The bidirectional Mendelian Randomization analyses, estimating the genetically predicted effect of various traits or disease on plasma cortisol, did not support any causal associations (Figure 5).

## Discussion

These results confirm that genetic variation in the *SERPINA6/A1* locus on chromosome 14 is associated with morning plasma cortisol. Despite doubling the sample size and trebling the SNP density from the previous genome-wide association study<sup>11</sup> no new genetic loci were identified. The improved imputation of genetic markers used in this analysis allowed identification of new SNPs within the *SERPINA6/A1* locus and strengthened the instrument used in two-sample Mendelian randomization analyses. This additional information coupled with tissue-specific gene expression data suggests that genetic variation within this locus influences expression of *SERPINA6* rather than *SERPINA1* in the liver. Furthermore, it appears that the eQTL and GWAMA signals colocalise within the block of SNPs in LD, represented by rs2736898, suggesting this block is primarily responsible for driving CBG-mediated variation for cortisol. Moreover, it demonstrates effects on adipose tissue gene expression, suggesting that resulting variations in CBG levels in turn influence the delivery of cortisol to peripheral tissues.

Given previous evidence of heritability of plasma cortisol it is surprising that the increase in sample size and SNP coverage from our earlier GWAMA did not identify any new loci associated with morning plasma cortisol. Twin studies have estimated the heritability of plasma cortisol ranging from 14% to  $45\%^{36-38}$ . However, SNP heritability estimates of plasma cortisol are considerably lower at  $6\%^{15}$ , and were confirmed in this sample at 4%.

Nevertheless, despite poor prediction of plasma cortisol by common autosomal SNPs, our interrogation of the *SERPINA6/SERPINA1* locus demonstrates important insights.

To test the functional significance of *SERPINA6/A1* variants we used eQTL analyses in the STARNET cohort. A hypothesis-free approach that investigated the effect of all 580 SNPs within the *SERPINA6/SERPINA1* locus on expression of all STARNET genes in each of the seven tissue types identified cis-eQTLs for SERPINA6 expression in the liver, the only tissue where SERPINA6 is highly expressed. This effect was refined to one of the LD blocks, represented by rs2736898. Importantly, genetic variation in *SERPINA6/SERPINA1* was also associated in trans-eQTL analyses with gene expression in visceral abdominal fat and subcutaneous fat as well as liver. These results suggest that not only does cortisol-associated genetic variation influence CBG expression in the liver, but it also influences cortisol signalling in peripheral tissues, an effect that is likely to be mediated by CBG.

CBG binds approximately 90% of cortisol in plasma but it is usually thought that only the free cortisol can access tissues and have biological effects. Patients with mutations in *SERPINA6*<sup>39</sup> and animals with deletion of CBG, however, exhibit features consistent with cortisol deficiency despite biochemical changes in the CBG-bound rather than free cortisol pool. Our data support the hitherto speculative evidence that CBG is involved actively in delivery of cortisol to peripheral tissues. We and others had proposed that this is mediated by altered cleavage of CBG by neutrophil elastase in tissues<sup>11,12</sup>, and evidence from immunoassays suggested that variants in *SERPINA6/SERPINA1* might mediate their effect through altered inhibition of neutrophil elastase by  $\alpha$ 1-antitrypsin. However, the more detailed analyses facilitated by the expanded GWAMA presented here do not support this interpretation since we did not identify eQTLs for  $\alpha$ 1-antitrypsin expression. Moreover, the reliability of immunoassays to determine CBG cleavage has since been called into question<sup>40</sup>.

Epidemiological analysis of cortisol has demonstrated associations with a large number of diseases and traits<sup>2–9</sup>, but the direction of causality, if any, has not been established. The improved imputation of genetic markers used in this analysis allowed identification of new SNPs within the SERPINA6/A1 locus and strengthened the instrument used in the twosample Mendelian randomization analyses. We tested whether genetically-elevated cortisol is causally associated with cortisol-related outcomes selected from the features of Cushing's syndrome – a rare condition caused by tumours secreting ACTH or cortisol - that are highly prevalent in the general population. Consistent with previous reports based on a less refined genetic instrument<sup>13,14</sup> we found evidence that cortisol causally increases the risk of heart disease. However, we did not find conclusive evidence that elevated cortisol causes type 2 diabetes, osteoporosis or obesity; given the relatively small variance in cortisol accounted for by the genetic instrument, it is possible that causal associations with additional diseases and traits would be revealed by analysis of larger sample sizes. We also tested whether common diseases and traits underlie elevated plasma cortisol, using bidirectional Mendelian randomization analyses. The strongest evidence suggested that higher BMI reduces cortisol levels, consistent with prior epidemiological and experimental evidence that obesity enhances clearance of cortisol from the circulation<sup>1</sup>.

It is arguable that morning plasma cortisol or salivary cortisol are poor surrogates for overall cortisol exposure given their diurnal and ultradian fluctuations and variation with acute stress. A more robust assessment of cortisol phenotype, perhaps from hair cortisol<sup>41</sup> or ambulatory sampling of interstitial fluid over 24 hours, may reveal stronger epidemiological and genetic associations.

In summary, this large GWAMA has revealed the limited magnitude and range of genetic effects on plasma cortisol, but has identified a pathway from variation in the *SERPINA6/ SERPINA1* locus through variation in liver *SERPINA6* expression to variation in CBG-mediated gene transcription including in adipose tissue that is causally associated with cardiovascular disease.

## **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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Ethical approval for the study was obtained from the ALSPAC Ethics and Law Committee and the Local Research Ethics Committees. Consent for biological samples has been collected in accordance with the Human Tissue Act (2004). Informed consent for the use of data collected via questionnaires and clinics was obtained from participants following the recommendations of the ALSPAC Ethics and Law Committee at the time. We are extremely grateful to all the families who took part in this study, the midwives for their help in recruiting them, and the whole ALSPAC team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists and nurses.

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## References

- Walker BR. Glucocorticoids and Cardiovascular Disease. European Journal of Endocrinology. 2007; 157:545–559. [PubMed: 17984234]
- Ragnarsson O, Olsson DS, Papakokkinou E, Chantzichristos D, Dahlqvist P, Segerstedt E, Olsson T, Petersson M, Berinder K, Bensing S, et al. Overall and Disease-Specific Mortality in Patients With Cushing Disease: A Swedish Nationwide Study. J Clin Endocrinol Metab. 2019; 104:2375–2384. [PubMed: 30715394]
- Phillips DIW, Barker DJP, Fall CHD, Seckl JR, Whorwood CB, Wood PJ, Walker BR. Elevated Plasma Cortisol Concentrations: A Link between Low Birth Weight and the Insulin Resistance Syndrome? J Clin Endocrinol Metab. 1998; 83:757–760. [PubMed: 9506721]
- 4. Filipovsky J, Ducimetiere P, Eschwege E, Richard JL, Rosselin G, Claude JR. The relationship of blood pressure with glucose, insulin, heart rate, free fatty acids and plasma cortisol levels according

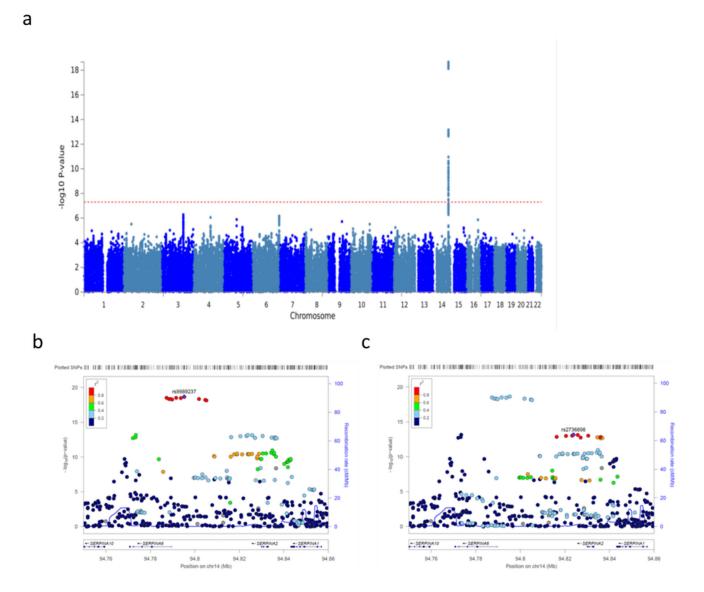
to degree of obesity in middle-aged men. Journal of Hypertension. 1996; 14:229–235. [PubMed: 8728301]

- Fraser, Robert; Ingram, Mary C; Anderson, Niall H; Morrison, Caroline; Davies, Eleanor; Connell, John MC. Cortisol Effects on Body Mass, Blood Pressure, and Cholesterol in the General Population. Hypertension. 1999; 33:1364–1368. [PubMed: 10373217]
- Reynolds RM, Walker BR, Syddall HE, Andrew R, Wood PJ, Whorwood CB, Phillips DIW. Altered Control of Cortisol Secretion in Adult Men with Low Birth Weight and Cardiovascular Risk Factors. J Clin Endocrinol Metab. 2001; 86:245–250. [PubMed: 11232008]
- Lupien SJ, de Leon M, de Santi S, Convit A, Tarshish C, Nair NPV, Thakur M, McEwen BS, Hauger RL, Meaney MJ. Cortisol levels during human aging predict hippocampal atrophy and memory deficits. Nature Neuroscience. 1998; 1:69–73. [PubMed: 10195112]
- Holsboer F. Stress, hypercortisolism and corticosteroid receptors in depression: implicatons for therapy. Journal of Affective Disorders. 2001; 62:77–91. [PubMed: 11172875]
- Seckl JR, Meaney MJ. Glucocorticoid "Programming" and PTSD Risk. Annals of the New York Academy of Sciences. 2006; 1071:351–378. [PubMed: 16891583]
- Ball TM. Cortisol Circadian Rhythms and Stress Responses in Infants at Risk of Allergic Disease. NIM. 2006; 13:294–300.
- Bolton JL, Hayward C, Direk N, Lewis JG, Hammond GL, Hill LA, Anderson A, Huffman J, Wilson JF, Campbell H, et al. Genome Wide Association Identifies Common Variants at the SERPINA6/SERPINA1 Locus Influencing Plasma Cortisol and Corticosteroid Binding Globulin. PLOS Genetics. 2014; 10:e1004474. [PubMed: 25010111]
- Hammond GL, Smith CL, Paterson NaM, Sibbald WJ. A Role for Corticosteroid-Binding Globulin in Delivery of Cortisol to Activated Neutrophils. J Clin Endocrinol Metab. 1990; 71:34–39. [PubMed: 2370299]
- Crawford AA, Soderberg S, Kirschbaum C, Murphy L, Eliasson M, Ebrahim S, Smith GD, Olsson T, Sattar N, Lawlor DA, et al. Morning plasma cortisol as a cardiovascular risk factor: findings from prospective cohort and Mendelian randomization studies. European Journal of Endocrinology. 2019; 181:429–438. [PubMed: 31325907]
- Pott J, Bae YJ, Horn K, Teren A, Kühnapfel A, Kirsten H, Ceglarek U, Loeffler M, Thiery J, Kratzsch J, et al. Genetic Association Study of Eight Steroid Hormones and Implications for Sexual Dimorphism of Coronary Artery Disease. J Clin Endocrinol Metab. 2019; 104:5008–5023. [PubMed: 31169883]
- Neumann A, Direk N, Crawford AA, Mirza S, Adams H, Bolton J, Hayward C, Strachan DP, Payne EK, Smith JA, et al. The low single nucleotide polymorphism heritability of plasma and saliva cortisol levels. Psychoneuroendocrinology. 2017; 85:88–95. [PubMed: 28843169]
- Winkler TW, Day FR, Croteau-Chonka DC, Wood AR, Locke AE, Mägi R, Ferreira T, Fall T, Graff M, Justice AE, et al. Quality control and conduct of genome-wide association meta-analyses. Nature Protocols. 2014; 9:1192–1212. [PubMed: 24762786]
- Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics. 2010; 26:2190–2191. [PubMed: 20616382]
- Winkler TW, Kutalik Z, Gorski M, Lottaz C, Kronenberg F, Heid IM. EasyStrata: evaluation and visualization of stratified genome-wide association meta-analysis data. Bioinformatics. 2015; 31:259–261. [PubMed: 25260699]
- Bulik-Sullivan BK, Loh P-R, Finucane HK, Ripke S, Yang J, Patterson N, Daly MJ, Price AL, Neale BM. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. Nature Genetics. 2015; 47:291–295. [PubMed: 25642630]
- Finucane HK, Bulik-Sullivan B, Gusev A, Trynka G, Reshef Y, Loh P-R, Anttila V, Xu H, Zang C, Farh K, et al. Partitioning heritability by functional annotation using genome-wide association summary statistics. Nature Genetics. 2015; 47:1228–1235. [PubMed: 26414678]
- Bulik-Sullivan B, Finucane HK, Anttila V, Gusev A, Day FR, Loh P-R, Duncan L, Perry JRB, Patterson N, Robinson EB, et al. An atlas of genetic correlations across human diseases and traits. Nature Genetics. 2015; 47:1236–1241. [PubMed: 26414676]
- 22. Zheng J, Erzurumluoglu AM, Elsworth BL, Kemp JP, Howe L, Haycock PC, Hemani G, Tansey K, Laurin C, Pourcain BS, et al. LD Hub: a centralized database and web interface to perform LD

score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. Bioinformatics. 2017; 33:272–279. [PubMed: 27663502]

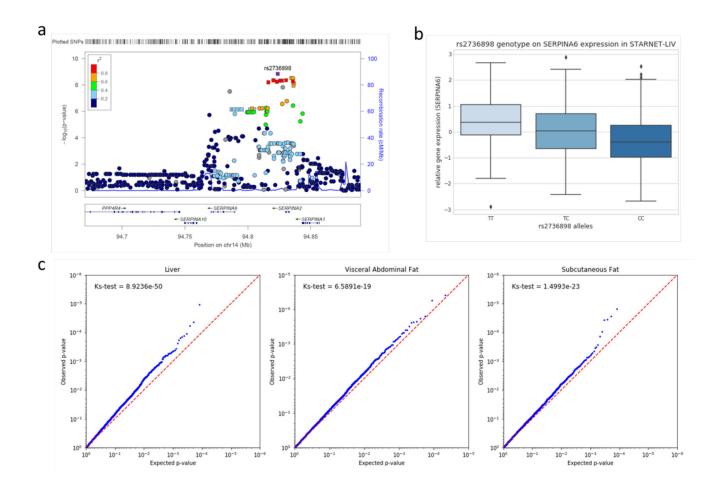
- 23. Leeuw CA, de Mooij JM, Heskes T, Posthuma D. MAGMA: Generalized Gene-Set Analysis of GWAS Data. PLOS Computational Biology. 2015; 11:e1004219. [PubMed: 25885710]
- 24. Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. Nat Commun. 2017; 8:1–11. [PubMed: 28232747]
- 25. Franzén O, Ermel R, Cohain A, Akers NK, Narzo AD, Talukdar HA, Foroughi-Asl H, Giambartolomei C, Fullard JF, Sukhavasi K, et al. Cardiometabolic risk loci share downstream cisand trans-gene regulation across tissues and diseases. Science. 2016; 353:827–830. [PubMed: 27540175]
- Schadt EE, Molony C, Chudin E, Hao K, Yang X, Lum PY, Kasarskis A, Zhang B, Wang S, Suver C, et al. Mapping the Genetic Architecture of Gene Expression in Human Liver. PLOS Biology. 2008; 6:e107. [PubMed: 18462017]
- Qi J, Asl HF, Björkegren J, Michoel T. kruX: matrix-based non-parametric eQTL discovery. BMC Bioinformatics. 2014; 15:11. [PubMed: 24423115]
- Storey JD, Tibshirani R. Statistical significance for genomewide studies. PNAS. 2003; 100:9440– 9445. [PubMed: 12883005]
- Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, Boehnke M, Abecasis GR, Willer CJ. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics. 2010; 26:2336–2337. [PubMed: 20634204]
- Giambartolomei C, Vukcevic D, Schadt EE, Franke L, Hingorani AD, Wallace C, Plagnol V. Bayesian Test for Colocalisation between Pairs of Genetic Association Studies Using Summary Statistics. PLOS Genetics. 2014; 10:e1004383. [PubMed: 24830394]
- Shabalin AA. Matrix eQTL: ultra fast eQTL analysis via large matrix operations. Bioinformatics. 2012; 28:1353–1358. [PubMed: 22492648]
- Machiela MJ, Chanock SJ. LDlink: a web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants. Bioinformatics. 2015; 31:3555–3557. [PubMed: 26139635]
- Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG, EPIC-InterAct Consortium. Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. Eur J Epidemiol. 2015; 30:543–552. [PubMed: 25773750]
- Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, Laurin C, Burgess S, Bowden J, Langdon R, et al. The MR-Base platform supports systematic causal inference across the human phenome. ELife. 2018; 7:e34408. [PubMed: 29846171]
- Halperin M. Fitting of Straight Lines and Prediction when both Variables are Subject to Error. Journal of the American Statistical Association. 1961; 56:657–669.
- 36. Inglis GC, Ingram MC, Holloway CD, Swan L, Birnie D, Hillis WS, Davies E, Fraser R, Connell JMC. Familial Pattern of Corticosteroids and Their Metabolism in Adult Human Subjects the Scottish Adult Twin Study. J Clin Endocrinol Metab. 1999; 84:4132–4137. [PubMed: 10566661]
- Meikle AW, Stringham JD, Woodward MG, Bishop DT. Heritability of variation of plasma cortisol levels. Metabolism - Clinical and Experimental. 1988; 37:514–517. [PubMed: 2967419]
- Froehlich JC, Zink RW, Li T-K, Christian JC. Analysis of Heritability of Hormonal Responses to Alcohol in Twins: Beta-Endorphin as a Potential Biomarker of Genetic Risk for Alcoholism. Alcoholism: Clinical and Experimental Research. 2000; 24:265–277.
- Meyer EJ, Nenke MA, Rankin W, Lewis JG, Torpy DJ. Corticosteroid-Binding Globulin: A Review of Basic and Clinical Advances. Horm Metab Res. 2016; 48:359–371. [PubMed: 27214312]
- Hill LA, Vassiliadi DA, Dimopoulou I, Anderson AJ, Boyle LD, Kilgour AHM, Stimson RH, Machado Y, Overall CM, Walker BR, et al. Neutrophil elastase-cleaved corticosteroid-binding globulin is absent in human plasma. Journal of Endocrinology. 2019; 240:27–39.
- Wester VL, van Rossum EFC. Clinical applications of cortisol measurements in hair. European Journal of Endocrinology. 2015; 173:M1–M10. [PubMed: 25924811]

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## Figure 1.

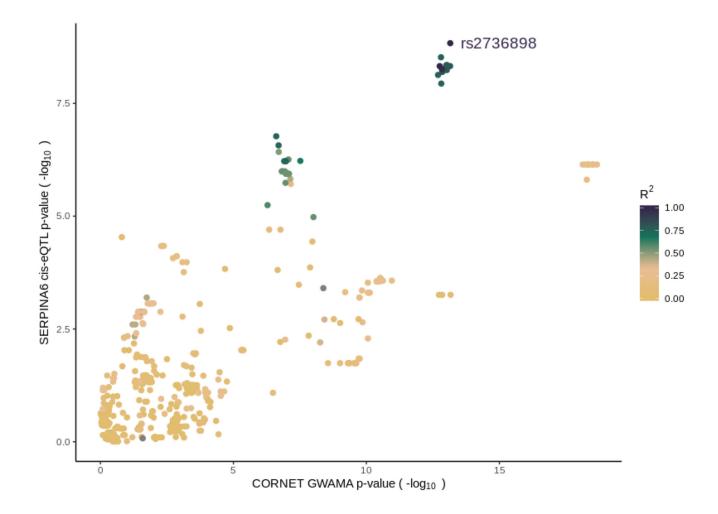
**a)** Manhattan plot of -log10 P values of the SNP-based association analysis of morning plasma cortisol (n=25,314). The locus on chr14 spans *SERPINA6* and *SERPINA1* genes; no other loci reached genome-wide significance. **b)** and **c**) Zoomed in Manhattan plot (LocusZoom plot) of -log10 P values of the SNP-based association analysis of morning plasma cortisol (n=25,314). These show two (of the four) LD blocks ( $r^2 > 0.3$ ) in this locus.



#### Figure 2.

Tissue-specific association of cortisol-related SNPs with gene expression in STARNET. (a) LocusZoom plot showing genomic loci of given SNPs against measure of significance (log10 (p-value)) for an eQTL analysis in liver for all SNPs within 100 Kb of *SERPINA6*. Squares represent the 21 significant cis-eQTLs (q 0.05) that are also at genome wide significance in CORNET (p 5  $10^{-8}$ ). (b) Genotypic effect of representative SNP for LD block 2 (rs2736898) on SERPINA6 gene expression in liver. (c) Global tissue specific effects on gene expression for rs2736898 represented as Q-Q plots for genes in liver, subcutaneous fat and visceral abdominal fat describing observed p-values vs those expected by chance. Deviation from expected uniform distribution described by Kolmogorov-Smirnov test p-value (Ks-test).

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## Figure 3.

Scatterplot showing colocalisation of joint signal from CORNET GWAMA and *SERPINA6* cis-eQTLs from STARNET-liver. Includes all SNPs within 100 Kb of *SERPINA6* that were present in both datasets (n=535). Colour bar indicates degree of LD with rs2736898. Formal colocalisation analysis with Coloc indicates 99.2% probability of the presence of a shared causal variant within LD block 2 mediating GWAMA and *SERPINA6* cis-eQTL signal.

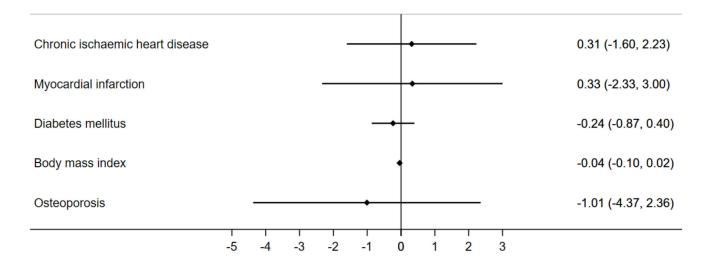
а		b	
Chronic ischaemic heart disease rs11620763 rs2736898 rs7146221 rs9989237 Subtotal (I-squared = 15.2%, p = 0.316)	001 (-0.42, 0.44) 29.85 0.64 (0.13, 1.15) 22.67 0.41 (-0.19, 1.01) 17.06 0.35 (-0.08, 0.78) 30.43 0.32 (0.06, 0.59) 100.00	Coronary heart disease rs11620763 rs2736898 rs7146221 rs9889237 Subtotal (I-squared = 16.0%, p = 0.311)	0.29 (0.00, 0.56) 26.71 0.09 (-0.22, 0.41) 22.93 0.39 (0.00, 0.79) 15.68 0.02 (-0.22, 0.27) 34.67 0.17 (0.00, 0.33) 100.00
Myocardial infarction rs11620763 rs2736898 rs7146221 rs9989237 Subtotal (I-squared = 0.0%, p = 0.586)	0 12 (-0.27, 0.51) 30.58 0.50 (0.04, 0.86) 22.07 0.20 (-0.35, 0.74) 15.94 0.11 (-0.27, 0.50) 31.41 0.21 (-0.00, 0.43) 100.00	Myocardial infarction rs11620763 rs7136898 rs7146221 rs9889237 Subtotal (I-squared = 49.2%, p = 0.116)	■         0.41 (0.09, 0.73)         26.51           -0.05 (-0.40, 0.30)         24.22           0.35 (-0.08, 0.79)         18.59           -0.00 (-0.28, 0.27)         30.69           0.16 (-0.08, 0.40)         100.00
Diabetes melitus rs1162/0763 rs2736896 rs7146221 rs9969237 Subtotal (I-squared = 0.0%, p = 0.688)	-0.15(-0.42, 0.13) 30.58 -0.08(-0.24, 0.41) 22.07 -0.07(-0.32, 0.45) 15.94 0.01(-0.26, 0.28) 31.41 -0.01(-0.17, 0.14) 100.00	Type 2 Diabetes rs11620763 rs2736898 rs9869237 Subtotal (I-squared = 0.0%, p = 0.973)	0.23 (0.28, 0.74) 21.40 0.17 (0.26, 0.60) 31.01 0.23 (0.11, 0.57) 47.60 0.21 (-0.02, 0.45) 100.00
Body mass index rs11620763 rs2736986 rs7144221 rs96989237 Subtotal (I-squared = 0.0%, p = 0.642)	0 00 (-0 05, 0 06) 30.57 0 02 (-0 04, 0 09) 22.08 0 02 (-0 06, 0 10) 15.94 -0 03 (-0 08, 0 03) 31.42 -0 00 (-0 03, 0 03) 100.00	Body mass index rs11620763 rs2736898 rs98802237 Subtotal (I-squared = 0.0%, p = 0.931)	-0.01 (-0.13, 0.11) 30.28 0.02 (-0.11, 0.14) 29.37 0.01 (-0.09, 0.12) 40.34 0.01 (-0.06, 0.07) 100.00
Osteoporosis rs11620763 rs2736698 rs7146221 rs9989237 Subtotal (i-squared = 0.0%, p = 0.820)	0.11 (-0.36, 0.57) 30.58 -0.22 (-0.76, 0.33) 22.07 -0.10 (-0.54, 0.75) 15.94 -0.03 (-0.42, 0.49) 31.42 -0.01 (-0.24, 0.27) 100.00	Lumbar spine bone mineral density rs11620763 rs2736698 rs716221 rs99690237 Subtotal (I-squared = 0.8%, p = 0.388)	-0.13 (-0.42, 0.15) 24.91 0.22 (-0.07, 0.51) 23.84 0.08 (-0.27, 0.43) 16.69 - 0.09 (-0.15, 0.34) 34.56 0.07 (-0.08, 0.21) 100.00
NOTE: Weights are from random effects analysis I -1.15 0	1.15	NOTE: Weights are from random effects analysis	.792

## Figure 4.

Causal estimates of a 1 SD increase in morning plasma cortisol on relevant disease and trait outcomes. Estimates are from two-sample Mendelian randomisation analyses using inverse variance weighting to combine estimates from each genetic variant. Panel a) Outcomes from UK Biobank (chronic ischaemic heart disease, cases = 8755, controls = 328444; myocardial infarction, cases = 7790, controls = 328893; diabetes mellitus, cases = 16183, controls = 320290; body mass index, sample size = 336107; osteoporosis, cases = 5266, controls = 331893); b) Equivalent outcomes from non-UK Biobank sources - CARDIoGRAMplusC4D (coronary heart disease, cases = 60801, controls = 123504; myocardial infarction, cases = 43676, controls = 128199), DIAGRAM (type 2 diabetes, cases=26488, controls=83964), GIANT (BMI, sample size = 339224) and GEFOS (lumber spine mineral density, sample size = 28498).

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## Figure 5.

Bidirectional causal estimates of the effect of the disease or trait on morning plasma cortisol. Estimates are from two-sample Mendelian randomisation analyses using inverse variance weighting to combine estimates from each genetic variant.