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A designed DNA binding motif that recognizes extended sites and spans two adjacent major grooves†

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Abstract

We report the rational design of a DNA-binding peptide construct composed of the DNA-contacting regions of two transcription factors (GCN4 and GAGA) linked through an AT-hook DNA anchor. The resulting chimera, which represents a new, non-natural DNA binding motif, binds with high affinity and selectivity to a long composite sequence of 13 base pairs (TCAT-AATT-GAGAG).

Transcription Factors (TFs) are specialized proteins that bind to specific DNA regulatory sequences,1 and thereby promote or inhibit the transcription of particular genes.2 The recognition process typically requires the cooperative action of several modules, which are connected either in a covalent or non-covalent way. This natural strategy allows the recognition of relatively large DNA sites, which is fundamental to ensure the selective targeting of specific genes.3 Thus, the bZIP or bHLH families bind DNA in the form of leucine zipper-mediated homo- or heterodimers,4 and the Cys_2His_2 zinc finger TFs present multiple recognition modules that simultaneously interact with consecutive sites along the DNA major groove.5 Other TFs, such as the cro repressor, or the glucocorticoid nuclear receptor protein, interact to DNA as non-covalent dimers, inserting recognition helices in the same face of adjacent major grooves.6

Over the last few decades there have been many efforts to develop miniaturized synthetic DNA binders that reproduce the DNA recognition properties of these natural proteins;7,8 some of them have even shown potential for the artificial control of gene expression.9 Most designed DNA-binding peptides rely on the modification of monomeric DNA binding domains,10 or in the artificial dimerization of bZIP basic regions.11 Our group has demonstrated that appropriate conjugation of monomeric fragments of transcription factors with small DNA-binding agents, such as distamycin or pentamidine derivatives,12 or with short AT-hook peptide motives,13 also leads to high-affinity and selective DNA binders. However, these binary artificial constructs allow the specific recognition of relatively short stretches of DNA (up to of 9 base pairs), far from the typical extended DNA sites covered by

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the natural counterparts.3b This represents a serious limitation for future applications in the selective targeting of specific genes. While the desired targeting of long DNA sites has been successfully achieved by recombinant oligomeric zinc fingers,14 we are not aware of synthetic peptide constructs that address extended sites by using TF-based DNA binding modules. Dervan's polyamides are capable of targeting up to sixteen contiguous base pairs, however they interact to the DNA through the minor groove.15

Inspired by proteins like the cro repressor, we explored the possibility of achieving a selective recognition of relatively long DNA sites by a composite “miniprotein” designed to insert TF recognition fragments into two consecutive major grooves (Fig. 1). Herein we demonstrate that covalent tethering of monomeric TF fragments through a polyglycine linker does not produce effective binders. However, if the linkage is carried out by an AT-hook module, the resulting construct binds with high affinity and specificity to an extended consensus sequence spanning 13 bp (TCAT·AATT·GAGAG). This conjugate represents the first synthetic peptide chimera capable of binding specific DNA sites in a tripartite (major–minor–major groove) manner, and provides a novel DNA binding architecture that lacks natural counterparts.

As constituent TF fragments, we selected the basic region (BR) of the yeast GCN4 bZIP protein, and the zinc finger of the GAGA factor of *Drosophila melanogaster,* which are both unable to bind to their target DNA sites as isolated monomers. In the case of GCN4, we chose the sequence between residues Asp^{226} and Gln^{248}, which has been identified as the smallest peptide that retains specific DNA recognition properties when presented as a dimer, 11a,16 or as a stapled derivative.17 With respect to GAGA, we chose a truncated 31-residue peptide of its zinc finger region (residues Ser^{28} to Phe^{58}) that is non functional as an isolated peptide, but can bind to the DNA when conjugated to small DNA-binding agents.18

Using as reference the X-ray structures of the GCN4–DNA19 and GAGA–DNA complexes, 20 we built a model for the simultaneous interaction of the GCN4 basic region and the GAGA zinc finger in contiguous DNA major grooves covering the same face of the DNA double helix (see the ESI†). Inspection of this coarse model suggested that the Arg^{245} in GCN4, and the Arg^{44} in GAGA could be suitable positions for tethering both DNA binding domains, and that the distance between both domains, of about 17 Å, could be effectively satisfied by a peptidic linker composed of nine glycines. After completing the solid-phase synthesis of the GCN4 basic region, the connector bearing a Gly_9 linker was introduced through an orthogonally-deprotected Lys side chain that replaces the natural Arg^{245}. We then added a bromoacetyl moiety that provides a reactive site for coupling the GAGA peptide fragment (Scheme 1).

The resulting peptide **BR(Gly$_9$)**-Br was isolated in an approximate 20% yield after standard cleavage and HPLC purification. The GAGA fragment was engineered to incorporate a Cys residue in the side chain of Lys^{44} (which replaces the native Arg). The expected peptide **GAGA**-SH was obtained in good yield after the standard deprotection/resin cleavage step (Scheme 2). The key coupling reaction between the two peptide fragments was carried out by dissolving the peptide **GAGA**-SH in phosphate buffer at pH 7.5 in presence of 1.5 equiv. of $ZnSO_4$, and 2 equiv. of **BR(Gly$_9$)**-Br (see the ESI†). After 1 h at room temperature, we

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obtained the desired conjugate **BR(Gly$_9$)GAGA** in an approximate 45% yield. The chemoselectivity of this reaction (only the Cys in the side chain of Lys^{44} is modified, and no alkylation of Cys^{36} and Cys^{39} is observed) stems from the coordination of the Zn cation by these other two Cys that support the zinc finger fold.21

With the conjugate at hand, we analyzed its DNA binding properties using standard non-denaturing EMSA assays in polyacrylamide gel, and double-stranded (ds) oligonucleotides featuring composite sites comprising the GCN4 basic region and the GAGA Zf target sequences separated by four (dsDNA A and B), or five base pairs (dsDNA C). These particular base pair spacers were selected so that the interaction of both binding regions of the conjugate could take place through the same face of the DNA double helix. As shown in Fig. 2, incubation of these dsDNAs with **BR(Gly$_9$)GAGA** failed in all cases to rise stable DNA peptide complexes, and we observed only faint slow-migrating bands that suggest the formation of low affinity complexes.

Although these poor results would advice against further pursuing this approach to achieve the desired bipartite major groove binding, we envisioned that using an AT-hook peptide instead of the (Gly$_9$) connector might allow the formation of more stable DNA complexes. AT-hook motives are cationic short peptides present in HMG-I(Y) eukaryotic nuclear proteins that feature a central Arg–Gly–Arg core capable of deeply inserting in the minor groove of AT-rich sites.22 We reasoned that these peptides, in addition to providing stabilizing contacts with the DNA, might work as minor groove anchors to ensure the correct delivery of the TF fragments to their consensus sites.13 Therefore, using as reference the structure of the AT-hook motif RKPRGRPKK, bound to the PRDII sequence of the IFN-β promoter (see the ESI†), we designed a new conjugate, **BR(Hk) GAGA**, comprising three different DNA binding fragments of natural TFs (GCN4, AT-hook, GAGA). Whereas individually these fragments are not functional, they might cooperate to form a trivalent complex with a target composite DNA.

The construct **BR(Hk)GAGA** was made following the same synthetic scheme as described for **BR(Gly$_9$)GAGA**, involving the independent synthesis of an electrophilic GCN4/AT-hook module **(BR(Hk)**-Br), and its chemoselective coupling with a Cys side chain of the GAGA fragment not involved in the zinc finger complexation (see Scheme 3 below and the ESI†). The desired conjugate was obtained after reverse-phase HPLC purification in a reasonable overall yield of approximately 10%, and identified by ESI-MS.

In contrast with the results obtained with our original oligoglycine design, incubation of **BR(Hk)GAGA** with a ds-oligonucleotide featuring a composite sequence comprising the binding sites for the GCN4, the AT-hook, and the GAGA fragment (dsDNA A), led to clear EMSA slow-migrating bands (Fig. 3 top, panel a). This is fully consistent with the formation of a highly-stable peptide–DNA complex. However, incubation of **BR(Hk)GAGA** with a control DNA that does not contain the consensus GAGA binding site (dsDNA D), shows faint bands that indicate the formation of low-affinity complexes, presumably arising from weak binary interactions involving the GCN4 and the AT-hook modules (Fig. 3 top, panel b). 13 Likewise, incubation with a second control oligonucleotide lacking the GCN4 binding site (dsDNA E), leads also to faint retardation bands, and only at high concentrations of the

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conjugate (Fig. 3 top, panel c). Therefore, these results confirm that the trifunctional construct presents and excellent selectivity for its composite tripartite site over potential bipartite competitors. A control oligonucleotide (dsDNA B), lacking the central A/T-rich, also gave rise to weaker complexes than with the consensus DNA A (Fig. 3 top, panel d), although the binding appears to be better than with DNAs D and E, probably because the highly charged AT-hook presents stabilizing electrostatic interactions with the DNA backbone.

In order to quantify the DNA binding of our construct, we carried out fluorescence anisotropy titrations with selected fluorescently-labeled oligonucleotides. Thus, titrations using a tetramethyl rhodamine (TMR)-labeled ds-oligonucleotide containing the target composite site (TMR-A) confirmed formation of a high affinity complex ($K_D \approx 28$ nM at rt), even in the presence of excess of competing calf thymus DNA (Fig. 3 bottom left). Importantly, titration with the ds-oligonucleotide B (TMR-B), which promoted the appearance of electrophoresis retarded bands at high concentrations, revealed a much weaker interaction (more than 500 times lower affinity, see the ESI†). This confirms that in the presence of excess of non-specific DNA, the designed hybrid shows exquisite selectivity for its target 13 base-pair composite tripartite binding site (TCAT·AATT·GAGAG). In agreement with the results obtained by EMSA, circular dichroism experiments revealed that addition of 1 equiv. of the target oligonucleotide A to a 5 μM solution of **BR(Hk)GAGA** promotes a significant increase in the ellipticity of the negative bands at 208 and 222 nm, consistent with the α-helical folding of the GCN4 BR (Fig. 3, bottom right).23,24

While the relatively large size of the synthetic construct might hinder its cellular internalization, we reasoned that the oligocationic character of its basic region and AT-hook units could be beneficial for the cellular transport.25 Indeed, a preliminary test with mammalian Vero cells using a tetramethylrhodamine (TMR) derivative of **BR(Hk)GAGA** (see the ESI†), led to bright emission inside cells, in a pattern consistent with endosomal localization (Fig. 4).26 This efficient cell internalization opens the door for cellular applications of these peptide chimeras.

To gain some structural insight in the complex between the conjugate **BR(Hk)GAGA** and the target DNA, we carried out a computational study using molecular mechanics (MM) calculations with the obminimize utility script of OpenBabel 2.3.1,27 and the UFF force field.28 Building on the structural data available for the DNA interaction of the parent GCN4 and GAGA proteins, we obtained a model for the interaction of the conjugate with the target sequence: TCAT·AATT·GAGAG. The resulting model is fully consistent with a tripartite major–minor–major groove interaction that involves a relatively large binding surface covering one face of the DNA (see Fig. 5). This type of non-natural DNA binding has not been previously described, and its discovery should open new and important opportunities in the field.

Conclusions

In summary, we have developed a novel DNA binding motif consisting of two DNA binding fragments of natural TFs connected *via* an AT-hook linker, which allows the selective

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recognition of designed, extended DNA sequences (up to 13 bp). The success of this design relies on the ability of the AT-hook moiety to act as a bidentate minor groove-anchoring device that delivers the DNA binding TF fragments to appropriate positions for insertion in their respective major grooves. The peptidic nature of the AT-hook allowed an easy installation of each of the DNA binding peptides at the C- and N-terminus of the anchor.

The construct represents the first demonstration of an engineered synthetic DNA binder that reaches two consecutive major grooves across the minor groove. The tripartite (major–minor–major groove) recognition introduces a novel DNA binding motif that lacks a natural counterpart. This approach promises to be applicable to other DNA binding TF fragments addressing different sites, and introduces a novel way of targeting specific and long DNA sequences.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

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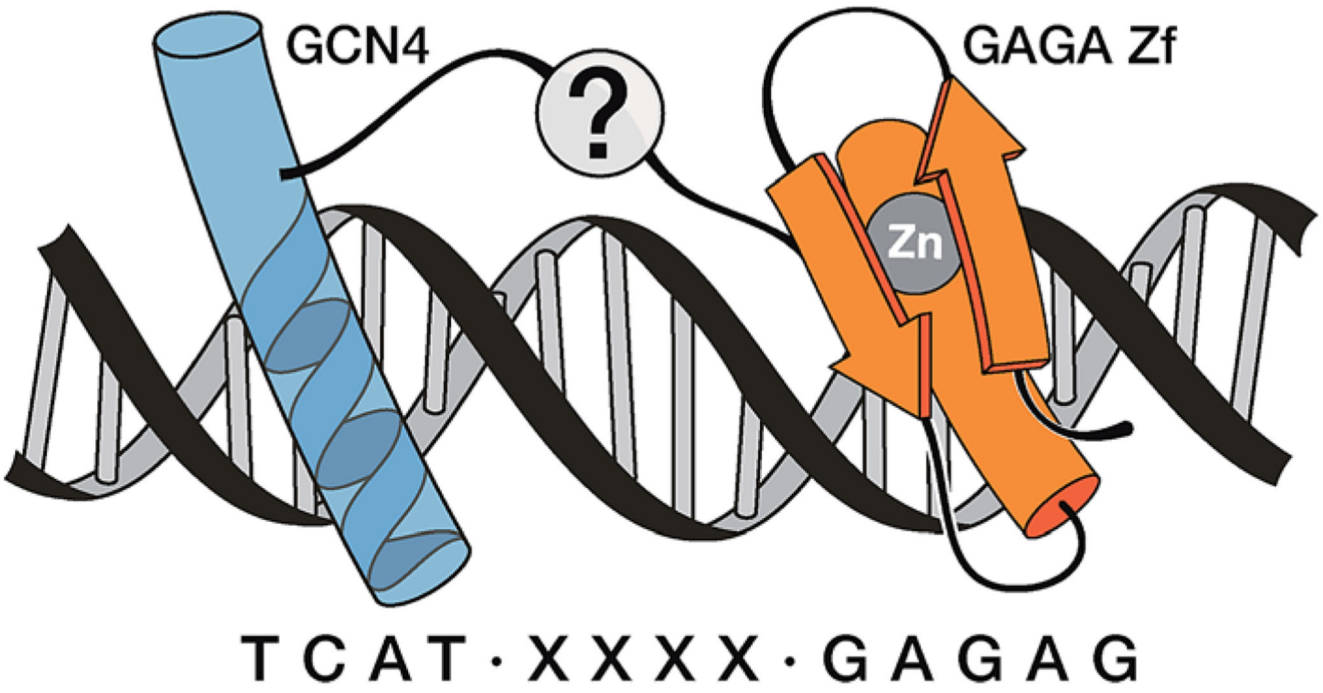

Fig. 1.

Cartoon representing the goal of this research, namely the recognition of extended DNA sites by inserting the basic region (BR) of GCN4 and the GAGA Zf in adjacent major grooves, and along one face of the double helix. The question mark intends to indicate the unknown nature of the connection that could allow the desired recognition.

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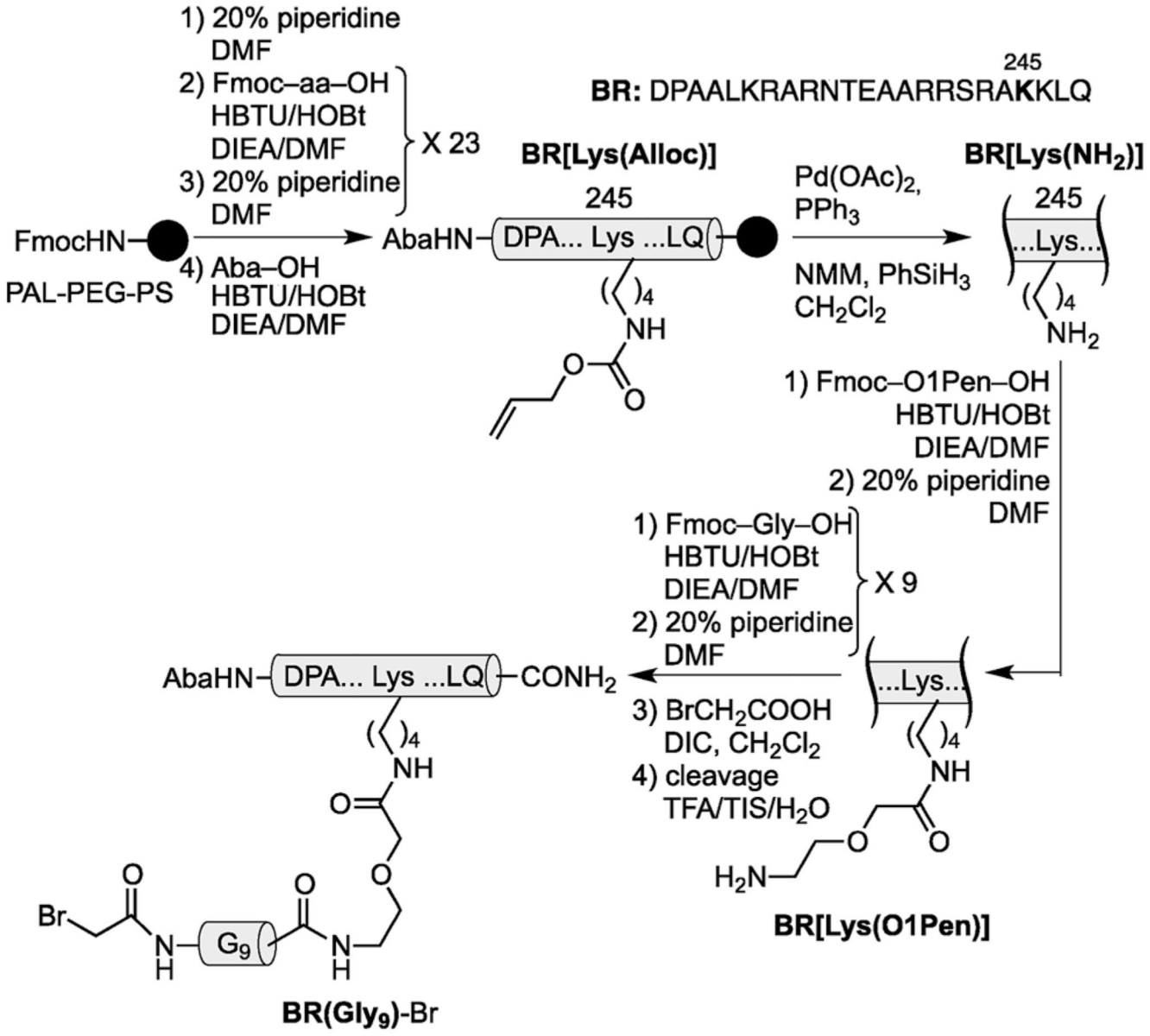
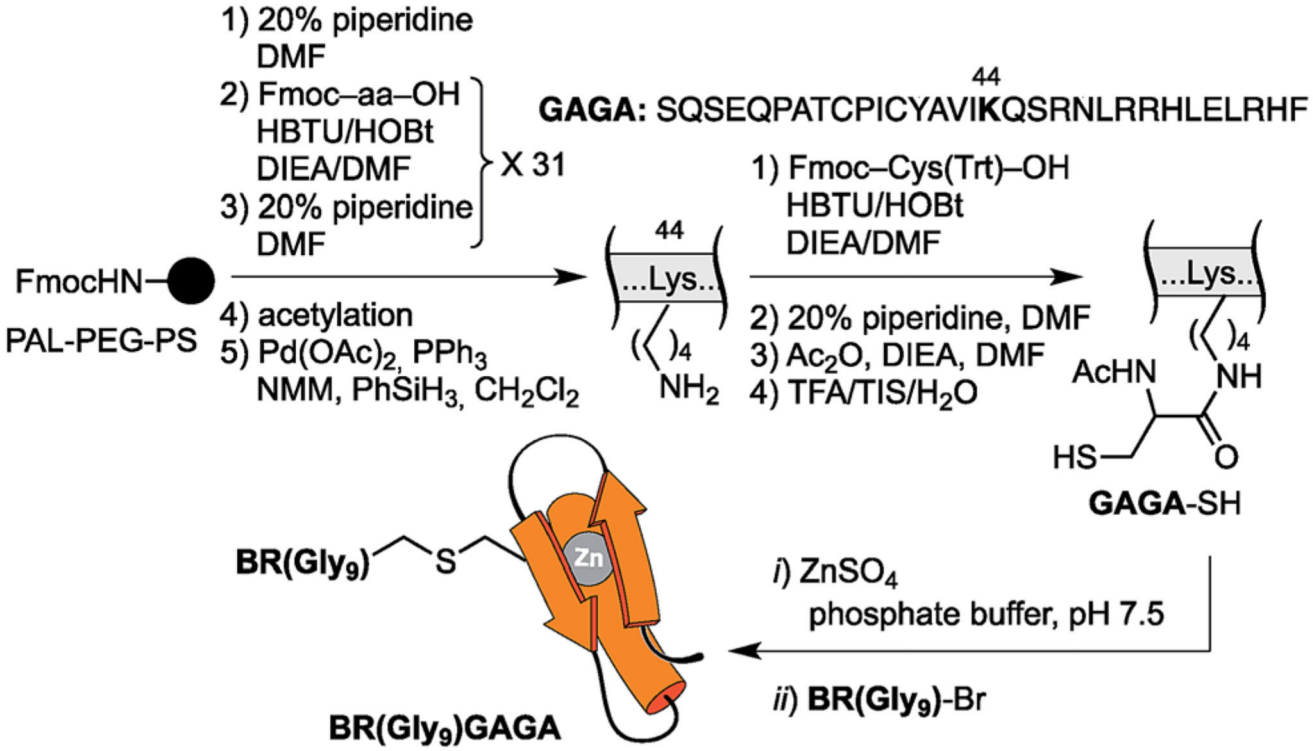


Scheme 1.

Strategy used for the synthesis of the GCN4/Gly$_9$ chimera **BR(Gly$_9$)**-Br. The 4-acetamidobenzoic acid (Aba) chromophore is introduced at the N-terminus of the GCN4 basic region as a spectroscopic reporter. O1Pen: 5-amino-3-oxapentanoic acid.



Scheme 2.

Strategy used for the synthesis of the GCN4/Gly_9/GAGA chimera, **BR(Gly_9)GAGA**, by chemoselective modification of the GAGA Cys_2His_2 peptide in solution.

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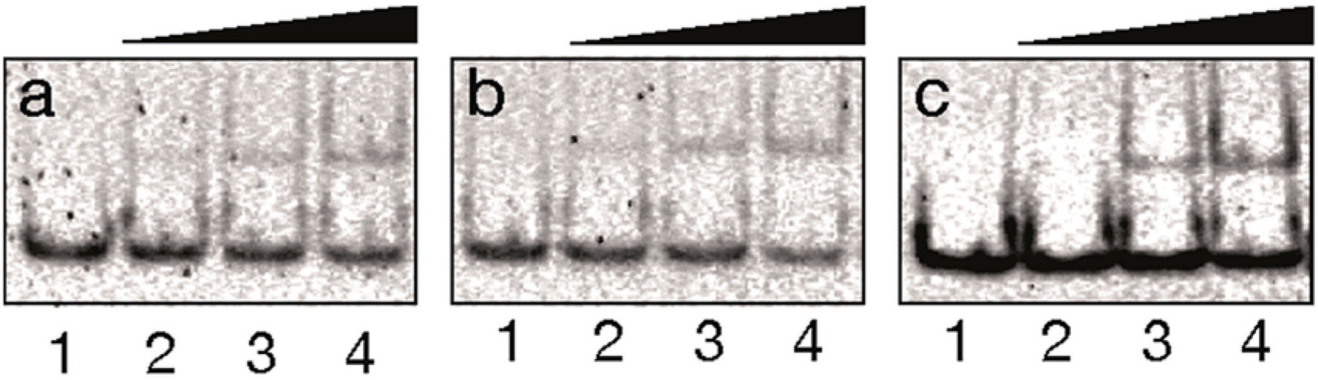

Fig. 2.

DNA binding studies of **BR(Gly$_9$)GAGA** by EMSA. In all cases, lanes 1-4: [**BR(Gly$_9$)GAGA**] = 0, 400, 600, 800 nM with (a) 75 nM of dsDNA A. (b) with 75 nM of dsDNA B. (c) with 75 nM of dsDNA C. Oligonucleotide sequences (only one strand shown): A 5′-CGCG TCAT*AATT*GAGAG CGC-3′; B 5′-CGCG TCAT*CAGC*GAGAG CGC-3′; C 5′-CGCG TCAT*AAATT*GAGAG CGC-3′. Experiment was resolved by PAGE on a 10% nondenaturing polyacrylamide gel and 0.5× TBE buffer over 40 min at rt, and analyzed by staining with SyBrGold (Molecular Probes: 5 μL in 50 mL of 1× TBE) for 10 min, followed by fluorescence visualization.

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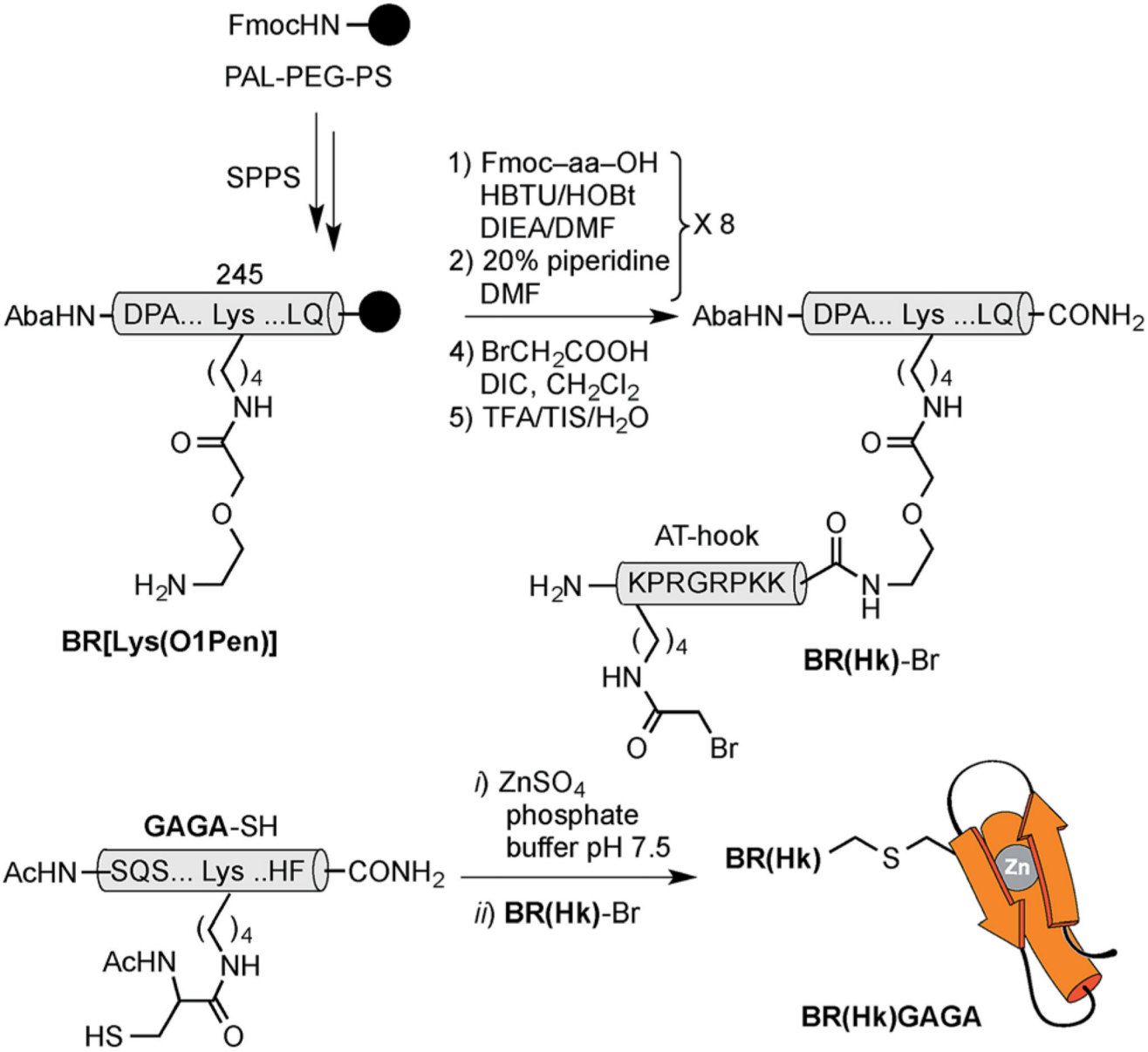

Scheme 3.

Top: synthesis of the GCN4/AT-hook chimera **BR(Hk)**-Br. See Scheme 1 for solid phase peptide synthesis of **BR[Lys(O1Pen)]**. Also note that the bromoacetyl unit was introduced in the side chain of the N-terminal lysine of the AT-hook, using Boc-Lys(Fmoc)-OH as orthogonal amino acid. Bottom: chemoselective modification of the GAGA Cys_2His_2 peptide in solution.

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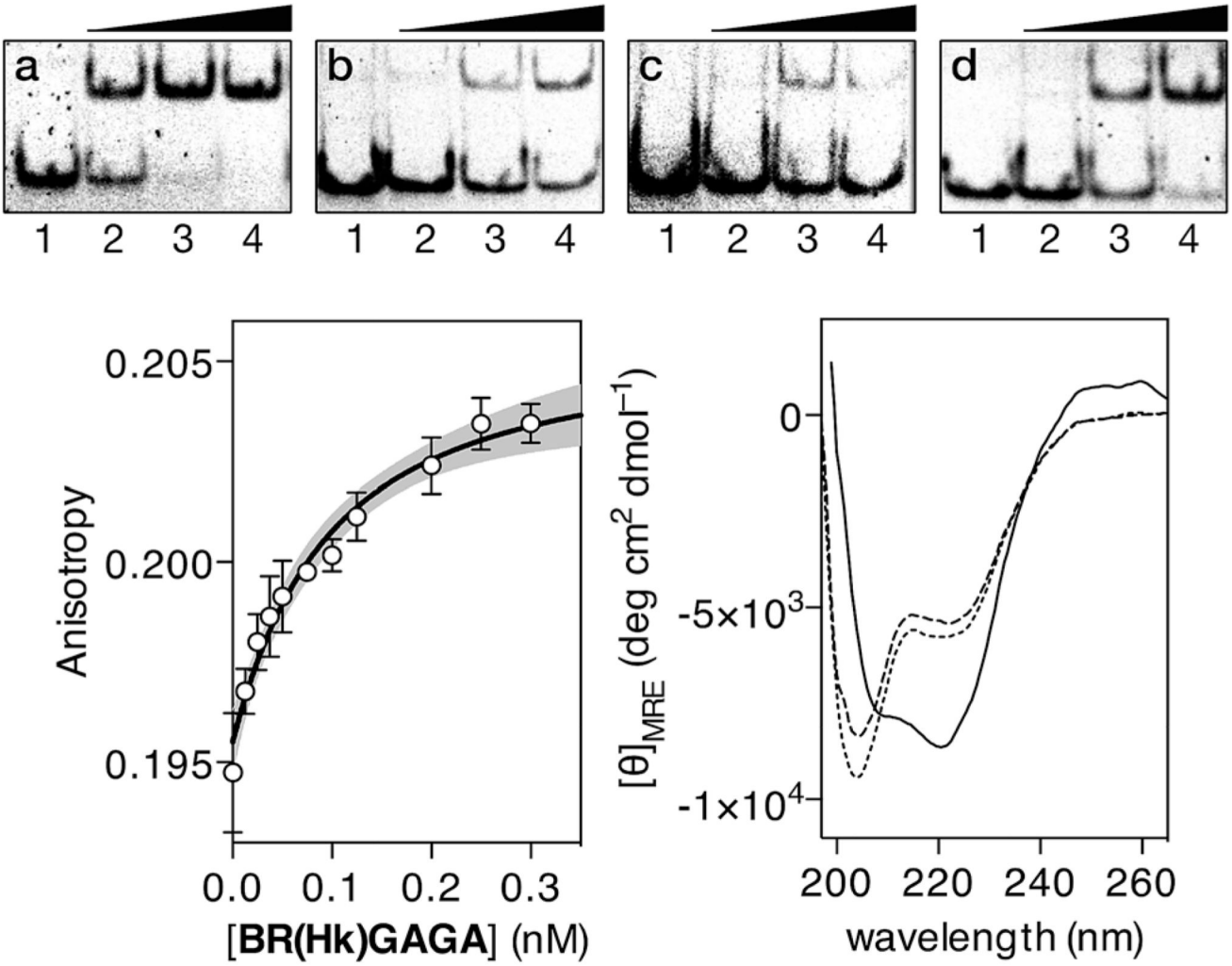

Fig. 3.

EMSA DNA binding studies of **BR(Hk)GAGA**. In all cases, lanes 1–4: **[BR(Hk)GAGA]** = 0,400, 600, 800 nM with (a) 75 nM of dsDNAA; (b) 75 nM of dsDNA D; (c) 75 nM of dsDNA E; (d) 75 nM of dsDNA B. Oligonucleotide sequences (only one strand shown): A: 5′-CGCG TCATAATTGAGAG CGC-3′; D: 5′-CGCG TCATAATTCGCGA CGC-3′; E: 5′-CGCG TGCTAATTGAGAG CGC-3′; B: 5′-CGCG TCATCAGC-GAGAG CGC-3′. Experiments were carried out by PAGE on a 10% non-denaturing gel and 0.5× TBE buffer over 40 min at rt, and analyzed by staining with SyBrGold (Molecular Probes: 5 mL in 50 mLof TBE) for 10 min, followed by fluorescence visualization. Bottom left: fluorescence anisotropy titration of a 25 nM solution of TMR-A in the presence of competing non-specific calf thymus DNA (50 mM) and with increasing concentrations of **BR(Hk)GAGA**. The best fit to a 1: 1 binding model and the 95% confidence band of the fit (in grey) are also shown. Bottom right: circular dichroism of a 5 mM solution of **BR(Hk)GAGA** (dotted line), of the same solution after the addition of 1 equiv. of $ZnSO_4$ (dashed line), and after the subsequent addition of 1 equiv. of the target dsDNA A (10 mM phosphate buffer pH 7.5 and

100 mM of NaCl; the contribution of the DNA to the CD spectrum has been subtracted for clarity).

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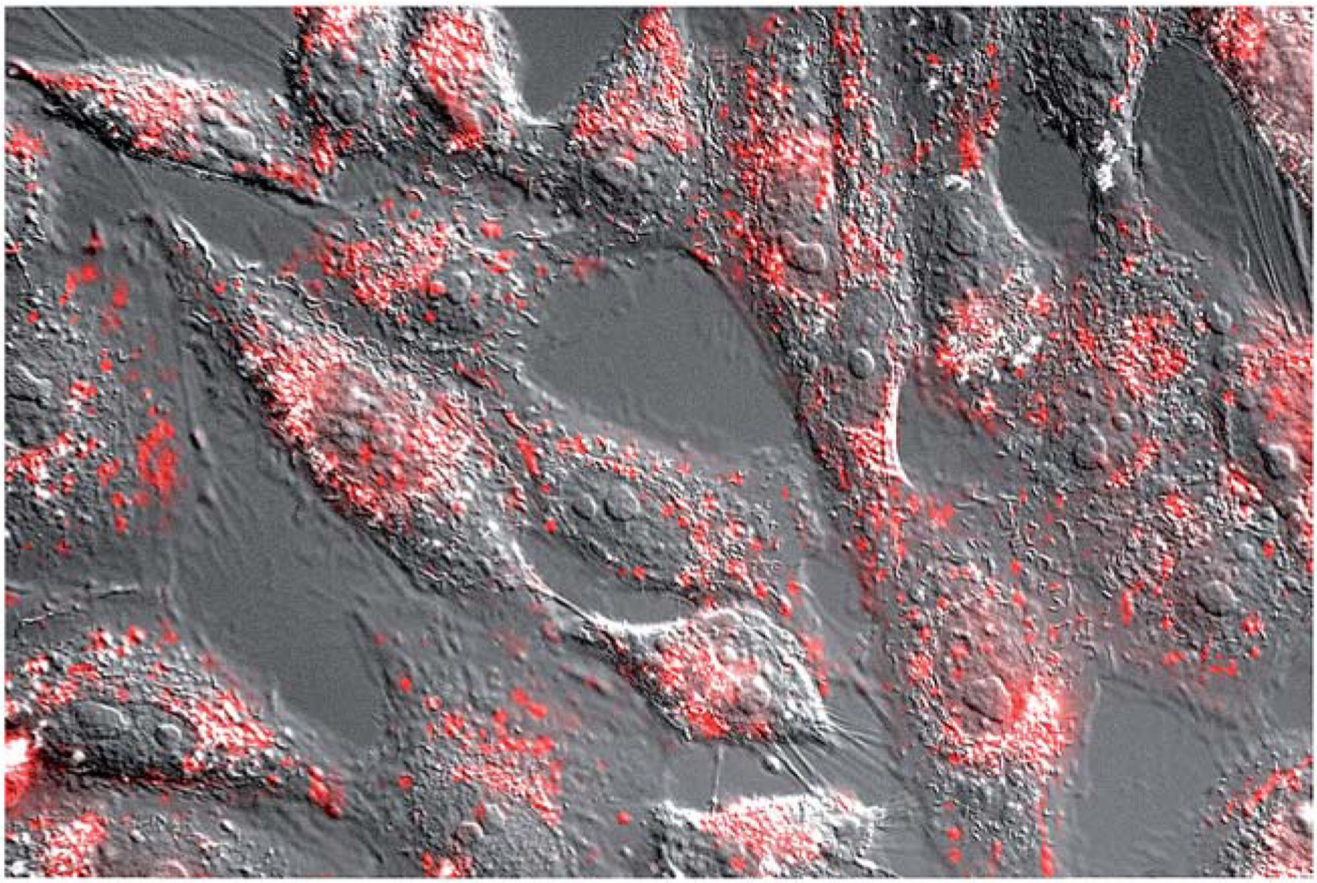

Fig. 4.

Fluorescence micrography of Vero cells. Brightfield images are superimposed to the red emission channel after incubation with 5 μM TMR-**BR(Hk)GAGA** for 30 min at 37 °C.

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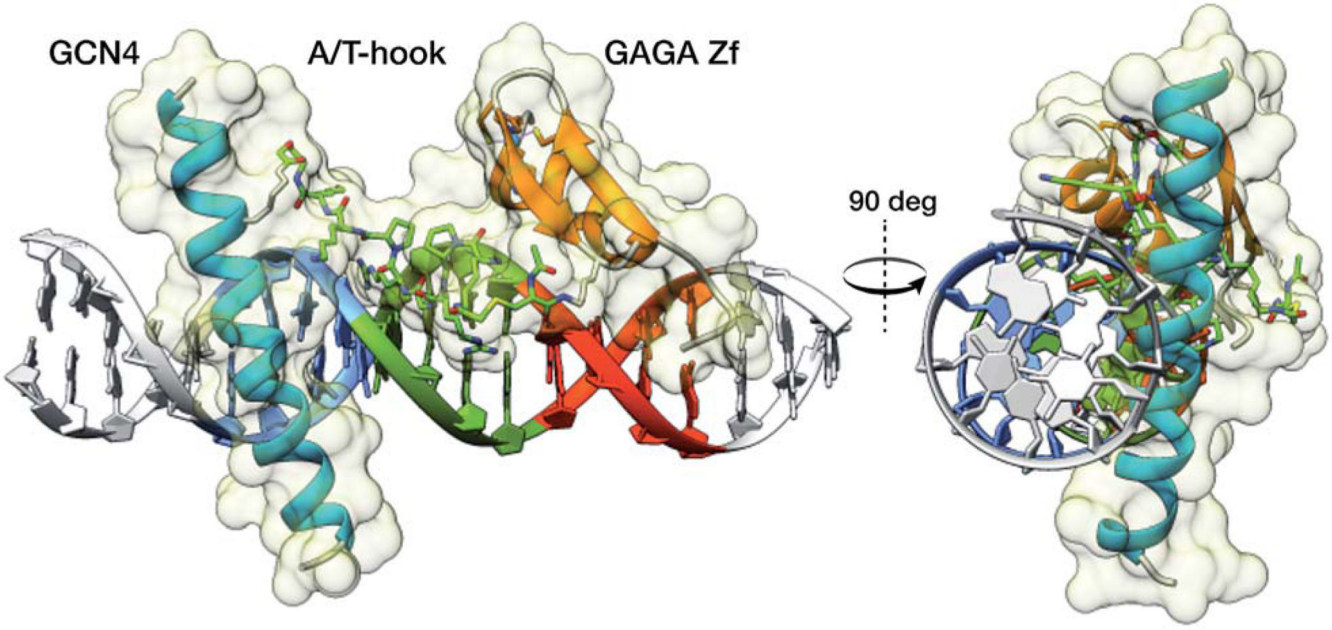

Fig. 5.

Model obtained using MM calculations of the interaction between the tripartite construct and the target composite DNA sequence. The image on the right shows the interaction of the three modules along the DNA axis covering one side of the double helix.