



HHS Public Access

Author manuscript

Acta Paediatr. Author manuscript; available in PMC 2019 August 01.

Published in final edited form as:

Acta Paediatr. 2019 June ; 108(6): 998–1007. doi:10.1111/apa.14702.

Review shows that donor milk does not promote the growth and development of preterm infants as well as maternal milk

Anna-Lena Hård¹, Anders K. Nilsson¹, Anna-My Lund², Ingrid Hansen-Pupp², Lois E. H. Smith³, and Ann Hellström¹

¹Department of Ophthalmology, Institute of Neuroscience and Physiology, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden

²Department of Pediatrics, Institute of Clinical Sciences Lund, Lund University and Skane University Hospital, Lund, Sweden

³Department of Ophthalmology, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA

Abstract

Aim: This nonsystematic review examined differences in the composition of raw maternal breastmilk and pasteurised donor milk and possible health effects on preterm infants.

Methods: We searched PubMed up to July 2018 for studies published in English that focused on four comparisons as follows: raw maternal milk versus donor milk, human milk before and after Holder pasteurisation, milk from mothers who delivered preterm and at term and milk collected during early and late lactation. We also searched for possible effects of the milk components, as well as the effects of maternal and donor milk on preterm infants' health.

Results: Raw maternal milk contained factors involved in antioxidant and anti-inflammatory defence, gut microbiome establishment and the maturation of immune defences, food tolerability and metabolism. Many of these factors were reduced or abolished in processed donor milk. Both maternal milk and donor milk have been associated with a reduced incidence of necrotising enterocolitis. High-dose feeding with maternal milk during the neonatal period reportedly reduced the risk of other morbidities and promoted growth and neurodevelopment.

Conclusion: Many of the components in raw maternal breastmilk were lacking in pasteurised donor milk, which was inferior in promoting the growth and development of very preterm infants.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

Correspondence: A-L Hård, c/o Ann Hellström, The Queen Silvia, Children's Hospital, Vitaminvägen 21, 41685, Gothenburg, Sweden. Tel: +46730460130 | Fax: +4631848952 | annalena.hard@gu.se.

CONFLICTS OF INTEREST

The authors have no conflicts of interest to declare.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Appendix S1 References.

Keywords

Donor milk; Maternal milk; Morbidity; Pasteurisation; Preterm

INTRODUCTION

Despite efforts to provide optimal nutrition, most extremely preterm infants suffer from postnatal growth retardation. This has been associated with retinopathy of prematurity (ROP) (1), bronchopulmonary dysplasia, late-onset sepsis, necrotising enterocolitis (NEC) (2,3) and neurodevelopmental impairment (4). Strategies to reduce prematurity-related complications are urgently needed, and the increased use of maternal milk may have preventative effects (5,6). Therefore, current guidelines recommend early enteral feeding, with expressed maternal milk and pasteurised human donor milk as a second choice.

In Sweden, extremely preterm infants always receive human milk during their hospital stay. Mothers are encouraged to start expressing milk for their infants soon after delivery. If maternal milk production is insufficient, or the mother is unable to express milk, or decides not to, infants are fed donor milk from a milk bank until a postmenstrual age of approximately 34 weeks. After that age, they receive preterm formula based on cows' milk. Both expressed maternal milk and donor milk are regularly analysed for their respective macronutrient and energy contents, and the milk is then individually fortified with bovine-derived fortifiers to meet current nutritional recommendations. Notably, donor milk is only provided until several weeks before term-equivalent age. This means that infants who receive donor milk miss human milk intake during the critical developmental period before term, as well as the six months of exclusive breastfeeding and subsequent partial breastfeeding recommended by the World Health Organization (7). In Sweden, the percentage of extremely preterm infants who were exclusively receiving maternal milk at discharge decreased from 55% in 2004 to 16% in 2013 (Fig. 1) (8). A mother's motivation to express milk and then breastfeed is influenced by the information that she receives. It may be insufficient to tell a mother that her milk is the best choice, without presenting the specific advantages and disadvantages of the alternatives. There is a need for a comprehensive summary of the evidence that maternal milk is superior to donor milk in preventing the complications of prematurity.

During the third trimester, a foetus swallows approximately 200–250 mL of amniotic fluid each day for each kilogram of foetal weight (9). Amniotic fluid, like maternal milk, contains nutrients and bioactive factors that promote gut maturation, protect against infection and facilitate growth (9,10). Very preterm infants miss a large portion of the immunoglobulin G transfer from the mother during late gestation, and their immune system is still immature at birth (11). Extremely preterm newborn infants have five to 10 times more microbial infections than infants born at term (12). Breastmilk contains factors that protect against infection and modulate the infants' immunological development (13). These components can also reduce the infants' future incidence of diseases that involve dysregulated immunity, such as allergies (14), inflammatory bowel disease (15), insulin-dependent diabetes mellitus (16) and some lymphomas (17).

The nutrient content of breastmilk varies greatly between mothers and depends on her diet, the gestational age at delivery, the lactation stage, the time of day, the size of the milk portion and whether it is foremilk or hindmilk (18). During the first three to five days after delivery, colostrum is produced, which is particularly rich in the nutrients and factors that are important for gut maturation and defence against infections. Transitional milk is then produced for the next two weeks' post-partum, after which the milk is considered mature.

Expressed maternal milk is usually consumed raw and sometimes after refrigeration and freezing and thawing. Bank milk is typically mature milk from the mothers of term infants and has usually been frozen and thawed before Holder pasteurisation, which implies that it has been heated to 62.5°C for 30 min. Thereafter, the milk is frozen again and reheated before it is fed to the infant. Studies with different designs have compared maternal milk and donor milk and examined the effects of storage and pasteurisation, with highly variable outcomes. In this review, pasteurisation always refers to Holder pasteurisation, since this is the method currently recommended for human milk banks. Table 1 presents several important bioactive factors that are altered after pasteurisation.

This paper reviews the differences between expressed raw maternal breastmilk and pasteurised donor milk in terms of composition and in relation to growth and morbidities. Our aim was to review the relevant scientific reports, so that adequate information could be provided to healthcare professionals and parents, to promote well-informed decisions about breastmilk expression and later breastfeeding. We also briefly address the long-term effects of maternal milk.

METHODS

PubMed was searched up to July 2018 for papers in English. We initially searched for papers that addressed the composition of human milk and differences in the composition between maternal milk and donor milk and raw and pasteurised milk. This review also explored differences in milk composition after term and preterm delivery, as well as in early and mature human milk. After that, we looked for papers that described functions of milk components, especially bioactive compounds, and how they were affected by Holder pasteurisation. Finally, we searched for comparisons between feeding infants fortified raw maternal breastmilk or fortified pasteurised donor milk with regard to growth, morbidity and neurodevelopment.

This was not a systematic review of all the factors present in maternal and donor milk and their possible effects on very preterm infants. No other database than PubMed was used. Our aim was to review information that may be of value for healthcare professionals when they inform parents about the advantages and disadvantages of feeding very preterm infants maternal milk or donor milk.

RESULTS

Human milk composition

Macronutrients—Human milk shows wide variations in fat, protein and carbohydrate contents (19). Donor milk contains significantly less protein, fat and energy than the milk, provided by the mothers of preterm infants (20). Compared to term milk, preterm milk is richer in cholesterol, phospholipids and long chain polyunsaturated fatty acids, which are integral cellular membrane components and essential for brain cell growth and myelin development in newborn infants (21). These components decrease in milk as lactation progresses. A review of the effects of pasteurisation on human milk concluded that Holder pasteurisation did not significantly affect its composition with regard to saccharides, protein, fat, energy or fatty acids (22).

Bioactive components—In addition to macronutrients, breastmilk is rich in bioactive components, such as oligosaccharides, immunoglobulins, lactoferrin, cytokines, enzymes, growth factors, hormones, anti-inflammatory agents, microbial factors and fatty acids, particularly in the first months after birth. These factors appear to play a multitude of overlapping roles in gut and general development (23), although little is presently known about their functions in very preterm neonates.

While milk proteins mainly serve as a source of amino acids for infant growth, peptides may also possess bioactive capacities, either directly or after digestion (24). Bioactive milk proteins can provide protection from infections, enhance nutrient absorption and promote immune system development and neurodevelopment. Some of these proteins are destroyed or inactivated by pasteurisation, leading to different properties in raw maternal milk and pasteurised donor milk (22). In preterm formula, the proteins are derived from bovine milk, which differs from human milk in many respects.

The functions of milk proteins and peptides have mainly been investigated in experimental studies. *In vitro* models have revealed that human milk proteins generate a wide variety of bioactive peptides. Beta-casein (25) is the most abundant casein in milk and the greatest source of bioactive peptides (24). *In vitro* studies have shown that kappa-casein from breastmilk inhibits *Helicobacter pylori* binding to human gastric mucosa (26). Pasteurisation did not appear to affect the bioactive peptides derived from the digestion of major human milk proteins (25).

The most prevalent whey protein in human milk is alpha-lactalbumin, which accounts for 20–25% of total milk protein. It plays several physiological roles during the neonatal period, including providing a balanced supply of essential amino acids. Moreover, alpha-lactalbumin digestion results in the transient formation of peptides with immune-stimulatory and bactericidal properties, which may be protective against infection (24,27). One study reported that pasteurisation did not alter the alpha-lactalbumin and serum albumin concentrations in breastmilk (28).

Lymphocytes migrate from the mother's intestine to the mammary gland, where they are transformed to immunoglobulin-A-producing cells and produce the secretory

immunoglobulin-A (sIgA) found in human milk (29). Through this process, the milk contains antibodies that are directed against microbial antigens present in the mother's gut. The milk sIgA blocks the mucosal adherence of bacterial, parasitic and viral pathogens and is critical for maintaining a diversified microbiotic environment (30,31). Thus, sIgA in breastmilk transfers maternal immunity to infectious agents and other antigens in the mother's and hence the infant's environment to the infant.

One study showed that pasteurisation of previously frozen milk reduced sIgA by 51% (32), while another reported that sIgA was 60% lower in pasteurised term donor milk compared to fresh term milk (28). Research also showed that stool samples from breastfed term infants contained large amounts of intact sIgA, with the highest concentrations during the first weeks of life (33).

The iron-binding protein lactoferrin is a major whey protein in human milk. In the gut, it binds to lactoferrin receptors that are expressed in the small intestine. Human lactoferrin receptors have also been found in monocytes, lymphocytes, platelets, fibroblasts and bone (34). Stool samples of full-term exclusively breastfed infants contain intact lactoferrin and the concentration decreases with age and is naturally associated with the decreasing concentrations in milk (33). It has been suggested that absorption of lactoferrin-bound iron in milk is the main route for iron uptake during the neonatal period (35). However, neonatal lactoferrin knockout mice exhibited no evidence of reduced intestinal iron uptake (36). Lactoferrin has been reported to protect newborn infants from infection by withholding iron from bacteria (37) and by destabilising the bacterial cell surface (38). Lactoferrin has also been found to enter cell nuclei and affect the expression of genes, modulate cell proliferation and differentiation (39) and interact with the immune system (34,40).

Studies have shown that lactoferrin has high structural homology between species and bovine lactoferrin exerts biological effects on human enteral cells (41). In addition, bovine milk-based formulas contain very small amounts of lactoferrin without supplementation (42).

The process of freezing, thawing, pasteurising and freezing and thawing again reportedly decreases the lactoferrin concentration in human milk by 70% (43). Other studies have demonstrated that 91% of lactoferrin was lost after the pasteurisation of previously frozen milk (32) and that pasteurised donor milk had a 44% lower lactoferrin concentration than fresh milk (28). In preterm infants, enteral supplementation with bovine lactoferrin decreased invasive fungal infections (44), late-onset sepsis (45–47) and NEC (48). In some countries, bovine lactoferrin is added to commercial formulas, and the US Food and Drug Administration has classified bovine lactoferrin as generally recognised as safe (49).

A randomised controlled trial investigated enteral administration of human recombinant lactoferrin (talactoferrin) to infants with birthweights of 750–1500 g. It reduced hospital-acquired infections by nearly 50%, along with a significant reduction in coagulase-negative *Staphylococcus* and, or, line infections. However, the authors stated that talactoferrin was no longer available in the United States (50,51). A Cochrane review from 2017 concluded that low-quality evidence suggested that supplementing oral feeds with lactoferrin, bovine or

talactoferrin, with or without probiotics, decreased late-onset sepsis and NEC stage II or III in preterm infants, without adverse effects. Ongoing trials will provide evidence from over 6000 infants (52).

Lysozyme is a host defence protein that is secreted in high concentrations in human milk. It is part of the innate immune system and causes lysis of bacteria via degradation of their outer wall. The presence of lysozyme may stimulate overgrowth of intestinal bifidobacteria, which are apparently resistant to lysozyme (53). Lysozyme is excreted in the faeces of preterm infants with a gestational age of 28–30 weeks (54) but not in healthy term infants (33). Pasteurisation reportedly reduces lysozyme by 59% (32) and pasteurised donor milk showed 60% lower lysozyme activity than raw milk (28). Similarly, concentrations of another host defence protein, lactoperoxidase, were 82% lower in donor milk than in the breastmilk of mothers delivering at term (28).

Osteopontin is a multifunctional protein that is present in most tissues and body fluids. It is found in high concentrations in human milk (55,56) and the highest levels have been found three to seven days' post-partum (57). Experimental studies have revealed that milk osteopontin plays essential roles in immunological, intestinal and early cognitive development (55). It can bind to various receptors of enterocytes or be absorbed into systemic circulation. Osteopontin concentrations have been found at lower levels in bovine milk than in human milk and even lower levels have been found in bovine milk-based formula (42,56). Infants born at term who received formula with added bovine osteopontin had lower tumour necrosis factor-alpha serum concentrations than infants on nonsupplemented formula and similar concentrations to breastfed infants. They also had fewer fevers, increased proportions of T cells and upregulated pathways of cell proliferation and cell to cell adhesion. The effects of osteopontin supplements appeared to be dose dependent in one study, with some functions promoted by lower osteopontin concentrations and others by higher concentrations (58). Osteopontin has been found to have high affinity for lactoferrin and may act as a carrier protein for lactoferrin in milk and as a modulator of lactoferrin's anti-microbial and immune-stimulatory activities (59). Little is known about the biological functions of milk osteopontin in preterm infants and we were unable to find any studies on the effect of pasteurisation on osteopontin concentrations. In a preterm animal model that used piglets, formula supplemented with osteopontin reduced the severity, but not the incidence, of NEC (60).

Bile salt-stimulated lipase in milk facilitates lipid digestion and absorption, especially in immature infants with low endogenous capacity to digest fat early in life (S61 in Appendix S1). Bile salt-stimulated lipase is inactivated by pasteurisation (S62) and one study showed that feeding infants with pasteurised maternal milk reduced fat absorption and growth (S63). When recombinant human bile salt-stimulated lipase was used to supplement pasteurised human milk or formula, the absorption of docosahexaenoic acid and arachidonic acid was increased, but not the total fat absorption (S64). Treatment with recombinant human bile salt-stimulated lipase significantly improved growth in infants born small for gestational age, but not in infants born appropriate for gestational age and was associated with increased risk of infections and gastrointestinal intolerance (S65).

Milk fat globules comprise a core of mainly triglycerides, surrounded by a membrane composed of phospholipids, cholesterol, proteins and glycoproteins, including several proteins with known or suggested bio-activities. Free fatty acids and monoglycerides, which are digestion products of the milk fat globule triglyceride core, have been shown to act as detergents and have lytic functions against viruses, bacteria and protozoa (S66). Human milk fat globule membranes contain a multitude of proteins: in one study, 20% were shown to be related to immune responses and 19% to cell communication and signal transduction (S67). In addition, milk fat globule membranes also contain compounds involved in central nervous system development, such as choline and sphingomyelin. Filaments on the surface of human milk fat globules have been reported to contain mucin-1, which inhibits bacterial adhesion to other cells (S68), and lactadherin, which protects against the rotavirus infection (S69). Both mucin-1 and lactadherin survive and maintain their integrity in the stomachs of preterm infants. Their concentrations in human milk have been found to be highest during the first weeks of lactation and to decline thereafter (S70). The average milk fat globule size in mature milk increases with the lactation progress, and thereby, the proportion of milk fat globule membrane decreases. One study showed that, human milk fat globules significantly decreased in size during pasteurisation and showed a greater globule size distribution (S71). Another study reported that storage and cooling affected the stability and membrane composition of bovine milk fat globules and that pasteurisation drastically modified milk fat globule membrane composition and functionality (S72).

Choline is an essential nutrient in breastmilk and it is critical for nervous system development. It is a precursor of membrane constituents such as phosphatidylcholine and sphingomyelin and of the neurotransmitter acetylcholine. A study in rats showed that brain function was positively impacted by oral choline supplementation during neurogenesis and synaptogenesis, corresponding to a period from gestation to four years of age in humans (S73). Notably, phosphatidylcholine is a main component of pulmonary surfactant and the major transporter for arachidonic acid and docosahexaenoic acid (S74). During pregnancy, choline has been shown to selectively transfer from the mother to the foetus and plasma choline concentrations after preterm birth have been reported to decrease to approximately 50% of those in cord blood within 48 h. It has been suggested that choline undernourishment may contribute to adverse neurodevelopmental outcomes after preterm birth (S75). While some authors have reported that preterm human milk contained less choline than term milk during the first weeks of life (S76), others found more phosphocholine in preterm than term milk (S77). Bovine and human milk have a nearly identical phospholipid content, which was not altered by the pasteurisation of bovine milk (S78). An ongoing trial is investigating the effects of choline and docosahexaenoic acid supplementation in preterm infants (NCT02509728).

Human milk oligosaccharides (HMOs) are a group of complex sugars that are highly abundant in human milk. More than 120 different HMO structures have been identified, and it has been shown that each mother produces a unique composition (S79,S80). Most HMOs contain the sugar fucose and a mother's ability to fucosylate HMOs is genetically determined (S81). Some fucosylated HMOs prevent adhesion of enteric pathogens to the intestinal mucosa (S81). *In vitro* digestion studies have demonstrated that HMOs were relatively resistant to human digestion (S82,S83), with a small fraction absorbed into the

infants' systemic circulation and excreted intact in the urine (S84). Another study reported that most HMOs were either metabolised by intestinal microorganisms or excreted with faeces (S85). Within the gastrointestinal tract, different HMO categories apparently serve as metabolic substrates for specific bacteria, selectively promoting bifidobacterial growth in particular (S86,S87).

Experimental studies have shown that HMOs prevented adhesion to epithelial cells of pathogens, such as bacteria (S88), viruses (S89) and parasites (S90). In addition, HMOs inhibited proliferation of Group B *Streptococcus*, a leading cause of invasive infection in newborn infants (S91) and dose-dependently reduced *Candida albicans* invasion of human premature intestinal cells (S92). A study of formula-fed piglets reported that HMOs increased the abundance of various immune cells in both infected and noninfected animals (S93). Interestingly, some HMOs made urogenital tract epithelial cells more resistant to uropathogenic *Escherichia coli* (S94). Human milk oligosaccharides contain large amounts of sialic acid. A review summarised the evidence that supported a role for dietary sialic acid as an essential nutrient for optimal brain development (S95).

Similar HMO concentrations have been found in preterm and term milk (S96,S97), but preterm milk contained a much wider variation in the percentage of HMOs that contained fucose or sialic acid than term milk. This may have been due to the immaturity of the fucosylation process, which may influence the developing microbiota (S98). It has been suggested that preterm infants may benefit from supplementation with fucosylated HMOs, for example, from pooled donor human milk (S99). Human milk has been shown to contain 100-fold to 1,000-fold higher concentrations of oligosaccharides than bovine milk (S100). Pasteurisation or freeze drying of donor milk reportedly did not influence HMO content (S101).

Glycosaminoglycans are long un-branched polysaccharides that are found on cell surfaces and cell–extracellular matrix interfaces. They regulate many processes, including cell growth and differentiation, cell to cell and cell to matrix interactions and anti-infective and anti-inflammatory processes (S102). Studies have found that, approximately 55% of the glycosaminoglycans in human milk are chondroitin sulphate and 40% are heparin. The concentrations of these glycosaminoglycans were several times higher in human milk than in bovine milk, which showed a different glycosaminoglycan profile. During the first month of lactation, preterm milk was reported to contain significantly more glycosaminoglycans than term milk (S103) and they were not affected by pasteurisation (S104).

Growth factors, adipokines, hormones and cytokines—Insulin in human milk is thought to play important roles in the development of gut function and general metabolism in infants (S105,S106). Insulin concentrations have been reported to be higher in milk ($60\pm 41 \mu\text{U/mL}$) than in maternal serum. Insulin levels in mature breastmilk were similar after term and preterm deliveries and remained unchanged after more than one year of lactation (S107–S109). Pasteurisation has been reported to reduce milk insulin by 46.1% (S110).

Serum adiponectin improves insulin sensitivity and lipid metabolism (S111). Higher levels of adiponectin have been found in preterm than term breastmilk (S112). One study demonstrated that pasteurisation reduced donor human milk adiponectin by 32.8% (S110).

Insulin-like growth factors (IGFs) possess growth-promoting and anti-apoptotic effects, and their presence in milk is thought to stimulate gut maturation and growth. One study of early human milk found that IGF-2, and its binding protein IGF-binding protein 2, was present in higher concentrations than IGF-1 and IGF-binding protein 3. Preterm milk contained higher IGF-binding protein 2 and IGF-binding protein 3 concentrations than term milk and IGF-binding protein 3 levels declined with time (S113). Enteral administration of IGF-1 to preterm infants supplemented with formula reduced gut permeability, but did not otherwise improve intestinal maturation (S114). Another study found that pasteurisation of human milk significantly reduced IGF-1 by 39.4% (S115).

Epidermal growth factor (EGF) plays important roles in intestinal development, starting during gestation when the foetus swallows amniotic fluid, which contains concentrations of EGF that are correlated with gestational age (S116). After birth, intestinal EGF is mainly obtained from breast-milk and the highest EGF concentrations have been found in colostrum (S117). One study found that EGF levels were 50–80% higher in preterm milk than term milk (S118). Another reported that Holder pasteurisation did not affect EGF concentrations (S115). Infant formulas do not contain EGF. In a rat model, supplementing bovine-based formula with EGF dramatically reduced the occurrence of NEC (S119).

Brain-derived neurotrophic factor (BDNF) has been shown to be involved in neuron growth and differentiation, synaptic plasticity and memory formation (S120). Variants of the BDNF gene have been associated with severe ROP (S121) and infants who developed severe ROP had lower serum BDNF levels at 10–14 days after birth than infants who did not develop ROP (S122). BDNF serum levels were higher at four to six months of age in breastfed term infants than in term infants fed formula (S123). Human milk contains BDNF, which is thought to be mainly produced by BDNF-expressing milk leucocytes (S124). Since cellular milk components are inactivated by pasteurisation, it can be speculated that BDNF concentrations would be lower in donor milk than maternal milk.

Cytokines are pluripotent polypeptides, with multiple and often overlapping effects, which act as messengers among cells and contribute to immune system development and function. Human milk contains various cytokines that can cross the intestinal barrier and the cytokine profile in breastmilk has been shown to vary with gestational age at delivery (S125). Milk cytokines can be divided into two categories: those that enhance inflammation or defend against infection and those that reduce inflammation. One study found that banked pasteurised donor milk contained cytokine concentrations that were similar to those in maternal milk after six weeks of lactation. The concentrations were lower than in milk that had been produced earlier. This provides evidence that donor milk provides less of these factors during the first weeks of life (S126).

The most abundant cytokines in human milk are from the transforming growth factor-beta (TGF-beta) family (S127). These cytokines have immune-regulatory properties and seem to

osteoblasts, chondrocytes, adipocytes, hepatocytes and pancreatic beta cells (S143). However, the roles of breastmilk stem cells for the breastfed infant remain unknown. The long-term impact of breastfeeding has been illustrated by better one-year graft function rates among breastfed recipients (82%) than among nonbreastfed recipients (57%) who received kidney transplants from their mothers later in life. (S144).

Micro ribonucleic acids (micro RNAs) are small noncoding RNA molecules that act on messenger RNA and serve as crucial regulators of gene expression in many biological processes. Human milk cells are one of the richest sources of micro RNAs. The majority of the most abundant micro RNAs are involved in immune responses, development, growth, metabolism, reproduction and enzyme regulatory activity (S145). Interestingly, most of the micro RNAs that are highly expressed in human milk cells regulate the IGF-1 receptor and some play critical roles in insulin receptor signalling (S145). Food-derived micro RNAs can survive the gastrointestinal tract, enter the systemic circulation and be transferred to various organs, where they regulate gene expression (S146,S147). Preterm delivery results in a unique profile of breast milk micro RNAs with metabolic targets (S148), but the roles of these milk micro RNAs remain unexplored in preterm infants. One study reported that pasteurising human milk did not alter the distribution or expression of four micro RNAs involved in the immune response (S149).

Human milk also contains bacteria, viruses and fungi. These include both beneficial bacteria, which develop into healthy gut flora, and pathogens such as the cytomegalovirus and human immunodeficiency virus that may be transferred to the infant. Breastmilk is a natural lactic acid bacterial source for colonisation of the infant's intestine (S150), and the transfer of maternal gut bacteria to the breast has been demonstrated (S151). Studies have shown that preterm infants who were fed maternal milk had increased gut microbial diversity (S152) and different gut microbial profiles, with a higher abundance of Bifidobacteriaceae, than those fed donor milk (S153). Pasteurisation has been demonstrated to inactivate the cellular components of milk (S154) and reduce the bactericidal effects of human breastmilk (S155). Pasteurised human milk showed increased bacterial growth after incubation compared to raw human milk (32).

Human milk has antioxidant properties and contains vitamin C, vitamin E, enzymes and other factors that protect against oxidative stress (S156,S157). Extremely preterm infants have low antioxidant capacity and are especially vulnerable to the oxidative stress associated with infections, mechanical ventilation, parenteral nutrition and blood transfusions (S158). It has been suggested that bronchopulmonary dysplasia, ROP, NEC and patent ductus arteriosus are all facets of an oxygen radical disease (S159). Pasteurisation has been shown to significantly reduce the total antioxidant capacity of breastmilk (S160).

Effects of maternal milk versus donor milk on growth and morbidities

Ethical concerns have prevented randomised controlled trials comparing exclusive feeding with maternal milk and donor milk. However, one study compared fortified maternal milk with fortified donor milk in infants with a gestational age of less than 32 weeks. This covered a period of exclusive feeding with human milk, from the discontinuation of parenteral nutrition until the transition to formula. Infants who received at least 80%

maternal milk gained weight more rapidly than those receiving less than 20% maternal milk and weight gain correlated with the proportion of maternal milk intake (S161). In a study of 243 infants with a gestational age of less than 30 weeks, infants who were exclusively fed maternal milk (maternal milk group) were compared to infants who received maternal milk plus pasteurised donor milk from women who had delivered preterm infants (donor milk group) or maternal milk plus preterm formula (formula group) (S162). A human milk fortifier was added when the milk intake reached 100 mL/kg/d. In the donor milk group, 17% of infants were switched to preterm formula due to poor growth, compared to none in the maternal milk group. Proliferative ROP stage 3 was found in 5.6% in the maternal milk group compared to 19% in the donor milk group ($p=0.05$). The maternal milk group also showed fewer episodes of late-onset sepsis and, or, NEC and fewer infection-related events than the other groups. These morbidities were negatively correlated with the cumulative maternal milk intake, indicating dose-dependent effects. Moreover, infants in the maternal milk group stayed in hospital one week less than the infants in the other groups. The authors concluded that donor milk offered no short-term advantages over preterm formula as a substitute for maternal milk.

Two studies since 2017 have focused on reduced weight gain in preterm infants supplemented with fortified donor milk, compared to infants exclusively fed with fortified maternal milk. The first reported that infants with birth-weights of less than 1000 g had a slower rate of weight gain during the first month of life and lower cognitive scores at one and two years of age if they were supplemented with more than 50% donor milk compared to infants who were exclusively fed maternal milk (S163). The second study found a dose-response relationship between maternal milk and growth in infants with a gestational age of 32 weeks or less or a birthweight of 1800 g or less. The mean weight gain velocity before 36 weeks of postmenstrual age, or discharge from the neonatal intensive care unit, decreased by 0.17 g per kg per day and the mean head circumference velocity by 0.01 cm per week for every 10% increase in donor milk intake compared to maternal milk (S164).

DISCUSSION

This paper presents a selection of the most studied elements of breastmilk. We still have very limited knowledge about the roles of these compounds in very preterm infants, despite the wealth of information about various milk components, their effects and their interactions with each other and other factors. Raw maternal milk contains bioactive factors that are involved in antioxidant and anti-inflammatory defence, gut microbiome establishment and the maturation of functions, such as immune defence, food tolerability and metabolism. Exposure to the protective immunological factors and microbiome in maternal milk (S125) counteracts the deficiencies of the infant's innate immune system and supports overall immune function.

It is clear that many of the elements that are present in the raw breastmilk of mothers who have delivered preterm infants are reduced or abolished in processed mature banked milk from mothers of term-born infants and absent or found in other forms in bovine-derived formula. It has been pointed out that the most profound difference between maternal milk and donor milk after preterm deliveries was observed when donor milk was substituted for

maternal milk during the early critical postbirth window. It is thought that components in maternal milk promote nutritional and immunologic programming and the growth of certain organs during this period (S165).

Several studies have shown a dose-dependent relationship between maternal milk intake during the neonatal period and a reduced frequency of ROP (S166,S167), NEC (S168,S169), infections (S170,S171), bronchopulmonary dysplasia (S172) and improved neurodevelopment (S173). Compared to preterm formula, donor milk was found to prevent NEC (S174), which has been reported to affect 5% to 22% of infants with a birthweight of less than 1000 g in high-income countries (S176). However, donor milk has also been reported to result in lower rates of weight gain, linear growth and head growth (S174). Other authors stated that no evidence had been put forward to indicate that donor milk reduced ROP or other morbidities or promoted neurodevelopment (S175).

The main limitation of this review was that it was not a systematic review of all the factors present in maternal and donor milk and their possible effects on very preterm infants. No other database than PubMed was used. We set out to review information of importance for healthcare professionals and parents about advantages and disadvantages of feeding very preterm infants maternal milk or donor milk.

It is well known that maternal milk is the best choice for all infants. The World Health Organization recommends exclusive breastfeeding up to six months of age and continued breastfeeding, together with other foods, up to two years of age and beyond (7). In 2018, a European report on 6592 infants born with a gestational age of less than 32 weeks from 11 countries reported large regional variations in breastmilk feeding. At hospital discharge, the rate of any breastmilk feeding ranged from 36% to 80% (S177). Intake of maternal milk was more important during the first weeks of life than later, with regard to preventing ROP and other prematurity-related morbidities (6). In the Extremely Preterm Infants in Sweden study of infants born before a gestational age of 27 weeks from April 2002 to March 2007, 70% of infants were fed exclusively with maternal milk at four weeks of life. At discharge, 50% received maternal milk, partly or exclusively (20). No later reports of maternal milk intake in the neonatal period of extremely preterm infants in Sweden have been published, but from 2004 to 2013, the rates of exclusively feeding maternal milk to extremely preterm infants at discharge declined from 55% to 16% in Sweden (8).

The World Health Organization and the United Nations Children's Fund launched the Baby-friendly Hospital Initiative to implement practices that promote breastfeeding and defined 10 steps to successful breastfeeding (S178). In 2012, an expanded version was published to meet the needs of preterm and sick infants, emphasising the need for antenatal information that was specifically designed for neonatal intensive care (S179). According to the parents of very preterm infants in one study, the main factors that facilitated maternal milk feeding were the contribution of maternal milk to infant growth and well-being and the parents' knowledge of the benefits of breastfeeding (S180). Other essential factors were the availability of education about breastfeeding and support to mothers who were motivated to provide their own milk.

CONCLUSION

Maternal milk contains many bioactive factors which are adjusted for the specific needs of her very preterm infant. Pasteurised donor milk contains less nutrients and bioactive factors than maternal milk and does not promote growth, health and development as well. Providing maternal milk can be viewed as an example of personalised medicine. Healthcare professionals and parents need comprehensive and up-to-date information about the benefits of maternal milk for very and extremely preterm infants. We believe that this will facilitate well-informed decisions about maternal milk feeding that may improve outcomes.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Abbreviations

BDNF	Brain-derived neurotrophic factor
EGF	Epidermal growth factor
HMO	Human milk oligosaccharides
IGF	Insulin-like growth factor
NEC	Necrotising enterocolitis
ROP	Retinopathy of prematurity
sIgA	Secretory immunoglobulin-A
TGF	Transforming growth factor

References

- Hellstrom A, Ley D, Hansen-Pupp I, Niklasson A, Smith L, Lofqvist C, et al. New insights into the development of retinopathy of prematurity—importance of early weight gain. *Acta Paediatr* 2010; 99: 502–8. [PubMed: 19878131]
- Klevebro S, Lundgren P, Hammar U, Smith LE, Bottai M, Domellof M, et al. Cohort study of growth patterns by gestational age in preterm infants developing morbidity. *BMJ Open* 2016; 6: e012872.
- Martin CR. Preventing bioenergetic failure in the preterm infant. *Arch Dis Child Fetal Neonatal Ed* 2016; 101: F99–101. [PubMed: 26253165]
- Ehrenkranz RA, Dusick AM, Vohr BR, Wright LL, Wrage LA, Poole WK. Growth in the neonatal intensive care unit influences neurodevelopmental and growth outcomes of extremely low birth weight infants. *Pediatrics* 2006; 117: 1253–61. [PubMed: 16585322]
- Cortez J, Makker K, Kraemer DF, Neu J, Sharma R, Hudak ML. Maternal milk feedings reduce sepsis, necrotizing enterocolitis and improve outcomes of premature infants. *J Perinatol* 2018; 38: 71–4. [PubMed: 29048409]
- Corpeleijn WE, Kouwenhoven SM, Paap MC, van Vliet I, Scheerder I, Muizer Y, et al. Intake of own mother's milk during the first days of life is associated with decreased morbidity and mortality in very low birth weight infants during the first 60 days of life. *Neonatology* 2012; 102: 276–81. [PubMed: 22922675]

7. World Health Organization. Global strategy for infant and young child feeding. Geneva, Switzerland: World Health Organization, 2003.
8. Ericson J, Flacking R, Hellstrom-Westas L, Eriksson M. Changes in the prevalence of breast feeding in preterm infants discharged from neonatal units: a register study over 10 years. *BMJ Open* 2016; 6: e012900.
9. Underwood MA, Gilbert WM, Sherman MP. Amniotic fluid: not just fetal urine anymore. *J Perinatol* 2005; 25: 341–8. [PubMed: 15861199]
10. Dasgupta S, Arya S, Choudhary S, Jain SK. Amniotic fluid: Source of trophic factors for the developing intestine. *World J Gastrointest Pathophysiol* 2016; 7: 38–47. [PubMed: 26909227]
11. Englund JA. The influence of maternal immunization on infant immune responses. *J Comp Pathol* 2007; 137(Suppl 1): S16–9. [PubMed: 17553516]
12. Clapp DW, Kliegman RM, Baley JE, Shenker N, Kyllonen K, Fanaroff AA, et al. Use of intravenously administered immune globulin to prevent nosocomial sepsis in low birth weight infants: report of a pilot study. *J Pediatr* 1989; 115: 973–8. [PubMed: 2585237]
13. Lewis ED, Richard C, Larsen BM, Field CJ. The importance of human milk for immunity in preterm infants. *Clin Perinatol* 2017; 44: 23–47. [PubMed: 28159208]
14. Saarinen UM, Kajosaari M. Breastfeeding as prophylaxis against atopic disease: prospective follow-up study until 17 years old. *Lancet* 1995; 346: 1065–9. [PubMed: 7564787]
15. Koletzko S, Sherman P, Corey M, Griffiths A, Smith C. Role of infant feeding practices in development of Crohn's disease in childhood. *BMJ* 1989; 298: 1617–8. [PubMed: 2503151]
16. Mayer EJ, Hamman RF, Gay EC, Lezotte DC, Savitz DA, Klingensmith GJ. Reduced risk of IDDM among breast-fed children. The Colorado IDDM Registry. *Diabetes* 1988; 37: 1625–32. [PubMed: 3192037]
17. Davis MK, Savitz DA, Graubard BI. Infant feeding and childhood cancer. *Lancet* 1988; 2: 365–8. [PubMed: 2899774]
18. Neville MC, Keller RP, Seacat J, Casey CE, Allen JC, Archer P. Studies on human lactation I. Within-feed and between-breast variation in selected components of human milk. *Am J Clin Nutr* 1984; 40: 635–46. [PubMed: 6475828]
19. Weber A, Loui A, Jochum F, Buhner C, Obladen M. Breast milk from mothers of very low birthweight infants: variability in fat and protein content. *Acta Paediatr* 2001; 90: 772–5. [PubMed: 11519980]
20. Stoltz Sjöström E, Öhlund I, Tornevi A, Domellöf M. Intake and macronutrient content of human milk given to extremely preterm infants. *J Hum Lact* 2014; 30: 442–9. [PubMed: 25117506]
21. Bitman J, Wood L, Hamosh M, Hamosh P, Mehta NR. Comparison of the lipid composition of breast milk from mothers of term and preterm infants. *Am J Clin Nutr* 1983; 38: 300–12. [PubMed: 6881084]
22. Peila C, Moro GE, Bertino E, Cavallarin L, Giribaldi M, Giuliani F, et al. The effect of Holder pasteurization on nutrients and biologically-active components in donor human milk: A review. *Nutrients* 2016; 8: 477.
23. Lonnerdal B. Bioactive proteins in human milk: mechanisms of action. *J Pediatr* 2010; 156(Suppl 2): S26–30. [PubMed: 20105661]
24. Wada Y, Lonnerdal B. Bioactive peptides derived from human milk proteins—mechanisms of action. *J Nutr Biochem* 2014; 25: 503–14. [PubMed: 24411973]
25. Wada Y, Lonnerdal B. Bioactive peptides released from in vitro digestion of human milk with or without pasteurization. *Pediatr Res* 2015; 77: 546–53. [PubMed: 25580741]
26. Stromqvist M, Falk P, Bergstrom S, Hansson L, Lonnerdal B, Normark S, et al. Human milk kappa-casein and inhibition of *Helicobacter pylori* adhesion to human gastric mucosa. *J Pediatr Gastroenterol Nutr* 1995; 21: 288–96. [PubMed: 8523212]
27. Lonnerdal B, Lien EL. Nutritional and physiologic significance of alpha-lactalbumin in infants. *Nutr Rev* 2003; 61: 295–305. [PubMed: 14552064]
28. Akinbi H, Meinzen-Derr J, Auer C, Ma Y, Pullum D, Kusano R, et al. Alterations in the host defense properties of human milk following prolonged storage or pasteurization. *J Pediatr Gastroenterol Nutr* 2010; 51: 347–52. [PubMed: 20639776]

29. Goldman AS. The immune system of human milk: antimicrobial, antiinflammatory and immunomodulating properties. *Pediatr Infect Dis J* 1993; 12: 664–71. [PubMed: 8414780]
30. Suzuki K, Nakajima A. New aspects of IgA synthesis in the gut. *Int Immunol* 2014; 26: 489–94. [PubMed: 24872116]
31. Hanson LA, Korotkova M, Teleme E. Breast-feeding, infant formulas, and the immune system. *Ann Allergy Asthma Immunol* 2003; 90(Suppl 3): 59–63. [PubMed: 12839115]
32. Christen L, Lai CT, Hartmann B, Hartmann PE, Geddes DT. The effect of UV-C pasteurization on bacteriostatic properties and immunological proteins of donor human milk. *PLoS ONE* 2013; 8: e85867. [PubMed: 24376898]
33. Davidson LA, Lonnerdal B. Persistence of human milk proteins in the breast-fed infant. *Acta Paediatr Scand* 1987; 6: 733–40.
34. Suzuki YA, Lopez V, Lonnerdal B. Mammalian lactoferrin receptors: structure and function. *Cell Mol Life Sci* 2005; 62: 2560–75. [PubMed: 16261254]
35. Lopez V, Suzuki YA, Lonnerdal B. Ontogenic changes in lactoferrin receptor and DMT1 in mouse small intestine: implications for iron absorption during early life. *Biochem Cell Biol* 2006; 84: 337–44. [PubMed: 16936804]
36. Ward PP, Mendoza-Meneses M, Cunningham GA, Conneely OM. Iron status in mice carrying a targeted disruption of lactoferrin. *Mol Cell Biol* 2003; 23: 178–85. [PubMed: 12482971]
37. Bullen JJ. Iron-binding proteins in milk and resistance to *Escherichia coli* infection in infants. *Proc R Soc Med* 1972; 65: 1086. [PubMed: 4568537]
38. Ellison RT 3rd, Giehl TJ, LaForce FM. Damage of the outer membrane of enteric gram-negative bacteria by lactoferrin and transferrin. *Infect Immun* 1988; 56: 2774–81. [PubMed: 3169987]
39. Liao Y, Jiang R, Lonnerdal B. Biochemical and molecular impacts of lactoferrin on small intestinal growth and development during early life. *Biochem Cell Biol* 2012; 90: 476–84. [PubMed: 22332905]
40. Spadaro M, Caorsi C, Ceruti P, Varadhachary A, Forni G, Pericle F, et al. Lactoferrin, a major defense protein of innate immunity, is a novel maturation factor for human dendritic cells. *FASEB J* 2008; 22: 2747–57. [PubMed: 18364398]
41. Lonnerdal B, Jiang R, Du X. Bovine lactoferrin can be taken up by the human intestinal lactoferrin receptor and exert bioactivities. *J Pediatr Gastroenterol Nutr* 2011; 53: 606–14. [PubMed: 21832946]
42. Chatterton DE, Nguyen DN, Bering SB, Sangild PT. Anti-inflammatory mechanisms of bioactive milk proteins in the intestine of newborns. *Int J Biochem Cell Biol* 2013; 45: 1730–47. [PubMed: 23660296]
43. Arroyo G, Ortiz Barrientos KA, Lange K, Nave F, Miss Mas G, Lam Aguilar P, et al. Effect of the various steps in the processing of human milk in the concentrations of IgA, IgM, and lactoferrin. *Breastfeed Med* 2017; 12: 443–5. [PubMed: 28742378]
44. Manzoni P, Stolfi I, Messner H, Cattani S, Laforgia N, Romeo MG, et al. Bovine lactoferrin prevents invasive fungal infections in very low birth weight infants: a randomized controlled trial. *Pediatrics* 2012; 129: 116–23. [PubMed: 22184648]
45. Manzoni P, Rinaldi M, Cattani S, Pagni L, Romeo MG, Messner H, et al. Bovine lactoferrin supplementation for prevention of late-onset sepsis in very low-birth-weight neonates: a randomized trial. *JAMA* 2009; 302: 1421–8. [PubMed: 19809023]
46. Akin IM, Atasay B, Dogu F, Okulu E, Arsan S, Karatas HD, et al. Oral lactoferrin to prevent nosocomial sepsis and necrotizing enterocolitis of premature neonates and effect on T-regulatory cells. *Am J Perinatol* 2014; 31: 1111–20. [PubMed: 24839144]
47. Ochoa TJ, Zegarra J, Cam L, Llanos R, Pezo A, Cruz K, et al. Randomized controlled trial of lactoferrin for prevention of sepsis in peruvian neonates less than 2500 g. *Pediatr Infect Dis J* 2015; 34: 571–6. [PubMed: 25973934]
48. Manzoni P, Meyer M, Stolfi I, Rinaldi M, Cattani S, Pagni L, et al. Bovine lactoferrin supplementation for prevention of necrotizing enterocolitis in very-low-birth-weight neonates: a randomized clinical trial. *Early Hum Dev* 2014; 90(Suppl 1): S60–5. [PubMed: 24709463]
49. Manzoni P. Clinical benefits of lactoferrin for infants and children. *J Pediatr* 2016; 173(Suppl): S43–52. [PubMed: 27234411]

50. Sherman MP, Sherman J, Arcinue R, Niklas V. Randomized control trial of human recombinant lactoferrin: A substudy reveals effects on the fecal microbiome of very low birth weight infants. *J Pediatr* 2016; 173(Suppl): S37–42. [PubMed: 27234409]
51. Sherman MP, Adamkin DH, Niklas V, Radmacher P, Sherman J, Wertheimer F, et al. Randomized controlled trial of Talactoferrin oral solution in preterm infants. *J Pediatr* 2016; 175: 68–73 e3. [PubMed: 27260839]
52. Pammi M, Suresh G. Enteral lactoferrin supplementation for prevention of sepsis and necrotizing enterocolitis in preterm infants. *Cochrane Database Syst Rev* 2017; 6: CD007137. [PubMed: 28658720]
53. Sakurai T, Hashikura N, Minami J, Yamada A, Odamaki T, Xiao JZ. Tolerance mechanisms of human-residential bifidobacteria against lysozyme. *Anaerobe* 2017; 47: 104–10. [PubMed: 28478277]
54. Schanler RJ, Goldblum RM, Garza C, Goldman AS. Enhanced fecal excretion of selected immune factors in very low birth weight infants fed fortified human milk. *Pediatr Res* 1986; 20: 711–5. [PubMed: 3737281]
55. Jiang R, Lonnerdal B. Biological roles of milk osteopontin. *Curr Opin Clin Nutr Metab Care* 2016; 19: 214–9. [PubMed: 27504516]
56. Schack L, Lange A, Kelsen J, Agnholt J, Christensen B, Petersen TE, et al. Considerable variation in the concentration of osteopontin in human milk, bovine milk, and infant formulas. *J Dairy Sci* 2009; 92: 5378–85. [PubMed: 19841198]
57. Nagatomo T, Ohga S, Takada H, Nomura A, Hikino S, Imura M, et al. Microarray analysis of human milk cells: persistent high expression of osteopontin during the lactation period. *Clin Exp Immunol* 2004; 138: 47–53. [PubMed: 15373904]
58. Lonnerdal B, Kvistgaard AS, Peerson JM, Donovan SM, Peng YM. Growth, nutrition, and cytokine response of breast-fed infants and infants fed formula with added bovine osteopontin. *J Pediatr Gastroenterol Nutr* 2016; 62: 650–7. [PubMed: 26465791]
59. Yamniuk AP, Burling H, Vogel HJ. Thermodynamic characterization of the interactions between the immunoregulatory proteins osteopontin and lactoferrin. *Mol Immunol* 2009; 46: 2395–402. [PubMed: 19477017]
60. Moller HK, Thymann T, Fink LN, Frokiaer H, Kvistgaard AS, Sangild PT. Bovine colostrum is superior to enriched formulas in stimulating intestinal function and necrotising enterocolitis resistance in preterm pigs. *Br J Nutr* 2011; 105: 44–53. [PubMed: 20723273]

Key notes

- This review focused on differences in the composition of raw maternal breastmilk and pasteurised donor milk and the possible health effects on preterm infants.
- Maternal milk prevented necrotising enterocolitis and other morbidities and promoted growth and neurodevelopment.
- Donor milk was associated with reduced necrotising enterocolitis compared to preterm formula, but it was inferior to maternal milk in promoting the growth and development of very preterm infants.

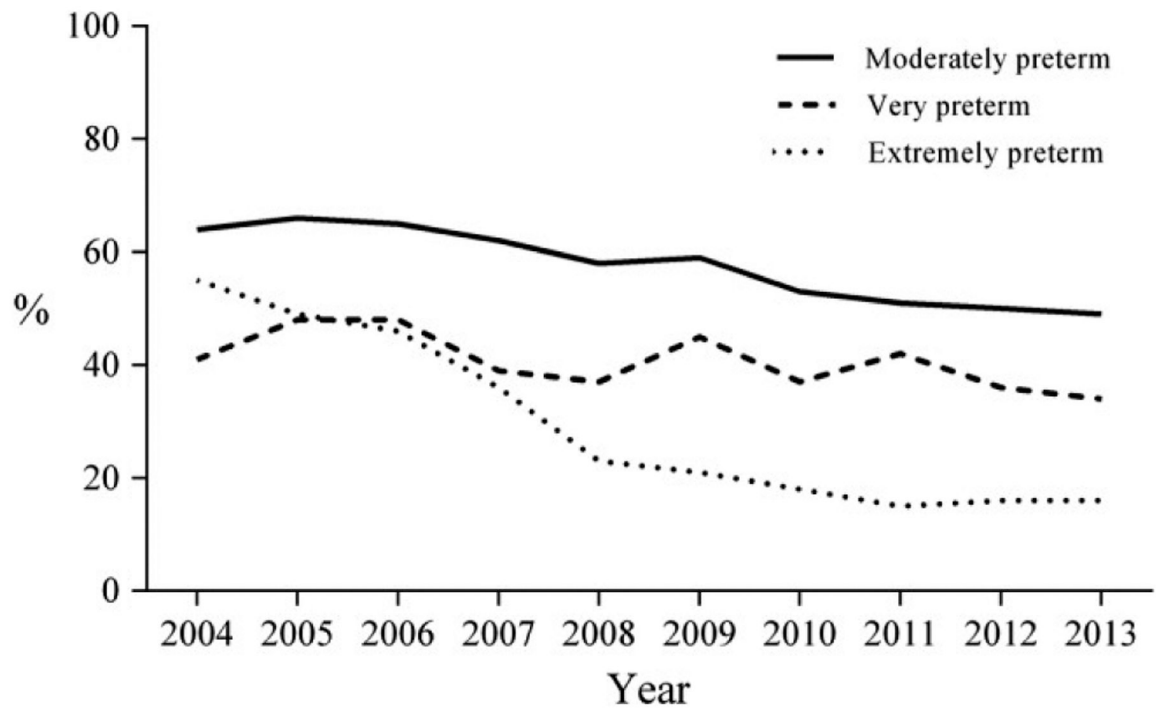


Figure 1.

Exclusive maternal breastmilk feeding at discharge from neonatal units, among infants of three gestational age groups: extremely preterm (n=550), very preterm (n=1848) and moderately preterm (n=12 958). Data from the Swedish Neonatal Quality register (2004–2013) (8).

Table 1

Some human milk bioactive components/functions shown to be affected by Holder pasteurization

Component concentrations/functions	Change with pasteurization	Reference
Secretory IgA	51% lower	Christen et al. (32)
Lactoferrin	91% lower	Christen et al. (32)
Lysozyme	59% lower	Christen et al. (32)
BSSL activity	Inactivated	Baro et al. (S62)
Insulin	46% lower	Ley et al. (S110)
Adiponectin	33% lower	Ley et al. (S110)
IGF-1	39% lower	Goelz et al. (S115)
TGF- β	14% lower	Reeves et al. (S136)
Maternal leucocytes	Destroyed	Goldman (S154)
Antioxidant capacity	67% lower	Silvestre et al. (S160)
Bacterial growth rate	Doubled	Christen et al. (32)

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript