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## Quantitation of TLR4 Internalization in Response to LPS in Thioglycollate Elicited Peritoneal mouse Macrophages by Flow Cytometry.

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### Abstract

Internalization of cell surface Toll Like Receptor 4 (TLR4) is a convenient read-out to measure LPS dependent activation of the TRIF adaptor pathway. We here provide a protocol to quantify the LPS dependent internalization of TLR4 using thioglycollate-elicited peritoneal macrophages by flow cytometry.

### Keywords

TLR4; LPS; TRIF; CD14; Endocytosis; peritoneal macrophages

### [Background]

Toll like receptor 4 (TLR4) is unique among the Toll like receptors in that it can deliver qualitatively different signals from two distinct cellular locations (reviewed in Takeda and Akira, 2004). Cell surface TLR4/MD2 is initially ligated by extracellular bacterial lipopolysaccharide (LPS) and engages the intracellular adaptor MAL and MyD88. Subsequently a fraction of cell surface LPS bound TLR4/MD2 will undergo CD14 and clathrin dependent internalization and translocation to an early endosomal compartment (Zanoni et al., 2011). In this endosomal compartment TLR4, engages the signaling adaptors TRAM and TRIF. Much of the regulation of TLR4 internalization and transport remains to be discovered. A flow cytometric method for quantifying loss of TLR4 surface expression, as a metric for endosomal translocation was first described in Bone Marrow Derived Macrophages (BMDMs) by Jonathan Kagan and colleagues (Kagan et al., 2008). Here we describe a modified version of this protocol to enable quantitation of TLR4 internalization in thioglycollate-elicited peritoneal macrophages on which we have published (Rajaiah et al., 2015; Perkins et al., 2018).

### Materials and Reagents

#### A. Consumables

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Competing interests

We declare there are no competing interests related to this work.

1. Pipette tips
2. 10 ml syringes (BD, catalog number: 309604)
3. 18 G x 1 needles (BD, catalog number: 305195)
4. 50 ml conical tubes (Denville, catalog number: C1062-P)
5. FACS tubes (round), sterile, with caps (Corning, Falcon, catalog number: 352054)
6. FACS tubes (round), non-sterile, without caps
7. Cell strainers, 70  $\mu$ m, (Fisherbrand, catalog number: 22363548)
8. 0.22  $\mu$ m filters for filtering culture media and FACS buffer (Sarstedt, catalog number: 83.1823.001)

**B. Reagents**

1. Sterile Saline (Baxter, catalog number: 0338-0048-03)
2. Optional: ACK lysis buffer (Quality Biological, Inc., catalog number: 118-156-721)
3. Ultra pure LPS, purified from *E. coli* K235 by the hot phenol method as previously described (McIntire et al., 1967)
4. Unconjugated rat IgG2a,  $\lambda$  anti-mouse CD16/32 (FC receptor) antibody, clone 93 (Biolegend, catalog number: 101302)
5. PE-conjugated anti-mouse CD284 (TLR4) antibody, clone SA15-21 (Biolegend, catalog number: 145404)
6. PE-conjugated Isotype control antibody, Rat IgG2a,  $\kappa$ , clone RTK2758 (Biolegend, catalog number: 400507)
7. Optional: other antibodies for cellular identification (e.g. Alexafluor-conjugated anti mouse F480; Biolegend catalog number: 123119)
8. Optional: paraformaldehyde solution (PFA), 4% in PBS (Affymetrix, catalog number: 19943)
9. Sterile Phosphate Buffered Saline (PBS) (Corning, catalog number: 21-040-CV)
10. Culture Media (see Recipes)
  - a. RPMI 1640 (Corning, catalog number: 15-040-CV)
  - b. Fetal Bovine Serum (FBS) (Atlanta Biologicals, catalog number: S11550)
  - c. Penicillin/Streptomycin (Gibco, catalog number: 15140-122)
  - d. L-glutamine (Gibco, catalog number: 25030-081)
11. FACS buffer (see Recipes)

- a. Sterile Phosphate Buffered Saline (PBS) (Corning, catalog number: 21-040-CV)
- b. EDTA, 0.5 M (Quality Biological, Inc., catalog number: 351-027-101)
- c. Fetal Bovine Serum (FBS) (Atlanta Biologicals, catalog number: S11550)

## Equipment

1. Ice buckets
2. Pipettes
3. Light microscope with hemocytometer slide
4. 37 °C incubator with CO<sub>2</sub> control designated for cell culture work (no bacterial/viral work)
5. Biosafety Cabinet designated for cell culture work (no bacterial/viral work)  
Sterile Culture Hood
6. Vortex Genie
7. Refrigerated Swing Bucket Centrifuge with inserts for 50mL conical tubes and flow cytometry tubes
8. Flow Cytometer (e.g. Becton Dickinson LSR II)
9. Computer for analysis

## Software

1. FCS Express 6.0 Flow (De Novo Software; Glendale, CA USA)
2. GraphPad Prism 7 (Graph Pad Software; La Jolla, CA USA)

## Procedure

- A. Day 1
  1. Collect macrophages from previously thioglycollate injected animals by peritoneal lavage using  $2 \times 10$  ml of sterile saline, using the 10 ml syringe with an 18 G needle attached. We usually pool peritoneal exudates from 2–5 littermates of the same gender to ensure sufficient yield for all controls and time points. A typical protocol can be found here: <https://www.jove.com/video/1488/isolation-of-mouse-peritoneal-cavity-cells>
  2. Pellet cells by centrifugation at  $93\text{--}218 \times g$  (1,000 rpm on a Thermo Scientific TX-750 4 rotor) for 10 minutes, and resuspend in culture media.
  3. Count cells using a hemocytometer (or cell counter) and add  $1 \times 10^6\text{--}1.5 \times 10^6$  Thioglycollate elicited peritoneal macrophages (TEPMs)

to each sterile FACS tube in 1.5 ml culture media (see Recipes below). A typical experiment includes three time points in unstimulated and three timepoints in stimulated macrophages. Be sure to include 1 tube for unstained control, 1 tube per time point for isotype control, single-color controls if cells are to be co-stained with other cell markers, and 1–2 tubes per time point for unstimulated controls. Lightly recap FACS tubes to allow gas exchange in the incubator.

4. Allow TEPM to rest overnight in 37 °C incubator with 5% CO<sub>2</sub>.

**B.** Day 2

1. The following morning, add 100 ng/ml ultrapure *E. coli* K235 LPS to each tube to be used to measure TLR4 internalization (excepting unstimulated controls). Lightly recap FACS tubes and return to 37 °C incubator during stimulation.
2. At appropriate time points, transfer each tube of control or LPS stimulated TEPMs to an ice bath and add two volumes ice-cold culture media or FACS buffer (see Recipes below) to arrest TLR4 internalization.
3. Pellet cells in a pre-cooled swing bucket centrifuge at  $112\text{--}264 \times g$  (1,100 rpm on TX-750 4 rotor) for 5–10 min.

Note: For this and all subsequent washes: decant media, resuspend cells by gently running the flow tubes over an empty tube rack, and immediately return to ice bucket. If more time points will be collected, resuspend these cells in ice cold culture media and keep on ice until the last samples are removed from the incubator.

4. Wash cells twice more with ice-cold FACS buffer. Resuspend in 100 µl final volume of ice-cold FACS buffer.
5. Add 2 µg αFC CD16/32–(approx. 4 µl of Biolegend #101302) Ab and incubate 15 min on ice. This prevents FC mediated nonspecific antibody binding.
6. Add 0.5 µg PE labeled anti-TLR4 Ab or Isotype control (optional: co-stain with other antibodies to aid in identification of cellular subsets) and incubate 30 min on ice.
7. Wash cells 3x with ice-cold FACS buffer
8. Wash 1x with ice-cold PBS.
9. Either proceed to Flow Cytometry run immediately or fix cells using 1% PFA (final concentration) in 100 µl PBS. Resuspend cells in 0.5–1.0 ml PBS and pass samples through a 70 µm cell strainer into a clean flow tube (not required to be sterile) to remove clumps just prior to running on flow cytometer.

## Data analysis

1. Flow cytometry analysis
  - a. For flow cytometry analysis, select single cells by forward scatter (FSC) profile (Height and Area of peaks). Set “stop gate” to 20,000 single events for each sample. Leave “record gate” to record all events.
  - b. For analysis, gate events on FSC-A and SSC-A profile to exclude dead cells and cellular debris.
  - c. TLR level can be visualized in a histogram for PE intensity.
  - d. Data (all events) are exported as FCS3.0 files.
2. Data analysis for the measurement of TLR4 endocytosis.
  - a. Open data files in FCS Express 6.0 Flow to fine tune “single cell” and “live macrophage” gates.
  - b. Record the median fluorescence intensity (MFI) for each sample.
  - c. Subtract the MFI of the unstained cells (autofluorescence) from the MFI of each sample.
  - d. Average the MFI of the “0 min time point” TLR4-stained samples (do this for each genotype, if multiple strains of mice were used)—and normalize the MFI data to the average (Time 0 min = 100%).
  - e. The internalization rate is calculated as the % of TLR4 lost from the surface over time.
3. Expected results: In wild type mice, the level of TLR4 surface expression in unstimulated cells should remain constant over the course of the experiment, and be well above the level of the isotype control-stained cells. LPS-stimulated cells should lose surface expression at a steady rate (at least over the first 90 min).

## Notes

1. We strongly recommend testing different lots of thioglycollate before starting these experiments, as endotoxin contamination is common and will pre-activate macrophages.
2. The yield of thioglycollate-elicited macrophages can vary based not only on the lot of thioglycollate, but also on the experience level of the lab member performing the peritoneal lavage, and the age and strain of the mice. The following table shows the range of peritoneal lavage cell yields from our laboratory for 6–8 week old mice of commonly used strains.

Strain	Novice	Experienced
C57BL/6J	0.8–2.1 × 10 <sup>7</sup>	1.4–9.2 × 10 <sup>7</sup>

Strain	Novice	Experienced
		Median: $4.1 \times 10^7$
TLR4 <sup>-/-</sup>	N/A	0.7–3.2 $\times 10^7$ Median: $2.1 \times 10^7$
Ticam1 <sup>-/-</sup>	N/A	2.2–3.2 $\times 10^7$
Ifnb <sup>-/-</sup>	N/A	1.6–2.8 $\times 10^7$

3. If the cell pellets from the peritoneal lavage look bloody, we perform red blood cell lysis using ACK buffer, followed by two washes with sterile PBS before resuspending in culture media.
4. It is important to allow the macrophages to rest overnight after isolation in order to achieve consistent kinetics of internalization from experiment to experiment.
5. Inspect each tube before adding LPS. Any tubes that have acidified or become cloudy in the overnight incubation are probably contaminated and should be discarded.
6. The success of this experiment depends on keeping the cells cold to prevent further internalization of the TLR4 complex. At cold temperatures, the cell membranes change physical properties from fluid to gel (Thewalt and Bloom, 1992; Spink et al., 1996). Protein complexes required for internalization will still form inside the membrane and as soon as membranes liquidize, internalization will proceed. Therefore, please be sure to pre-cool centrifuges and have an ice bucket and ice-cold FACS buffer or media available for rapid-cooling of cells at the time they are removed from the incubator. Minimize the time that the lid of the centrifuge is open, avoid touching the bottom of the flow cytometry tubes, and return tubes to the ice bucket immediately when removing from centrifuge. Without touching the bottom, you can hold up the tubes against a light to visually inspect the tubes after their centrifugation: Pellets of cold cells have higher opacity. Experiments can also be performed in a cold-room.
7. This assay is compatible with surface staining of other cell markers. If intracellular staining is required, we recommend setting up a parallel set of samples for permeabilization.

## Recipes

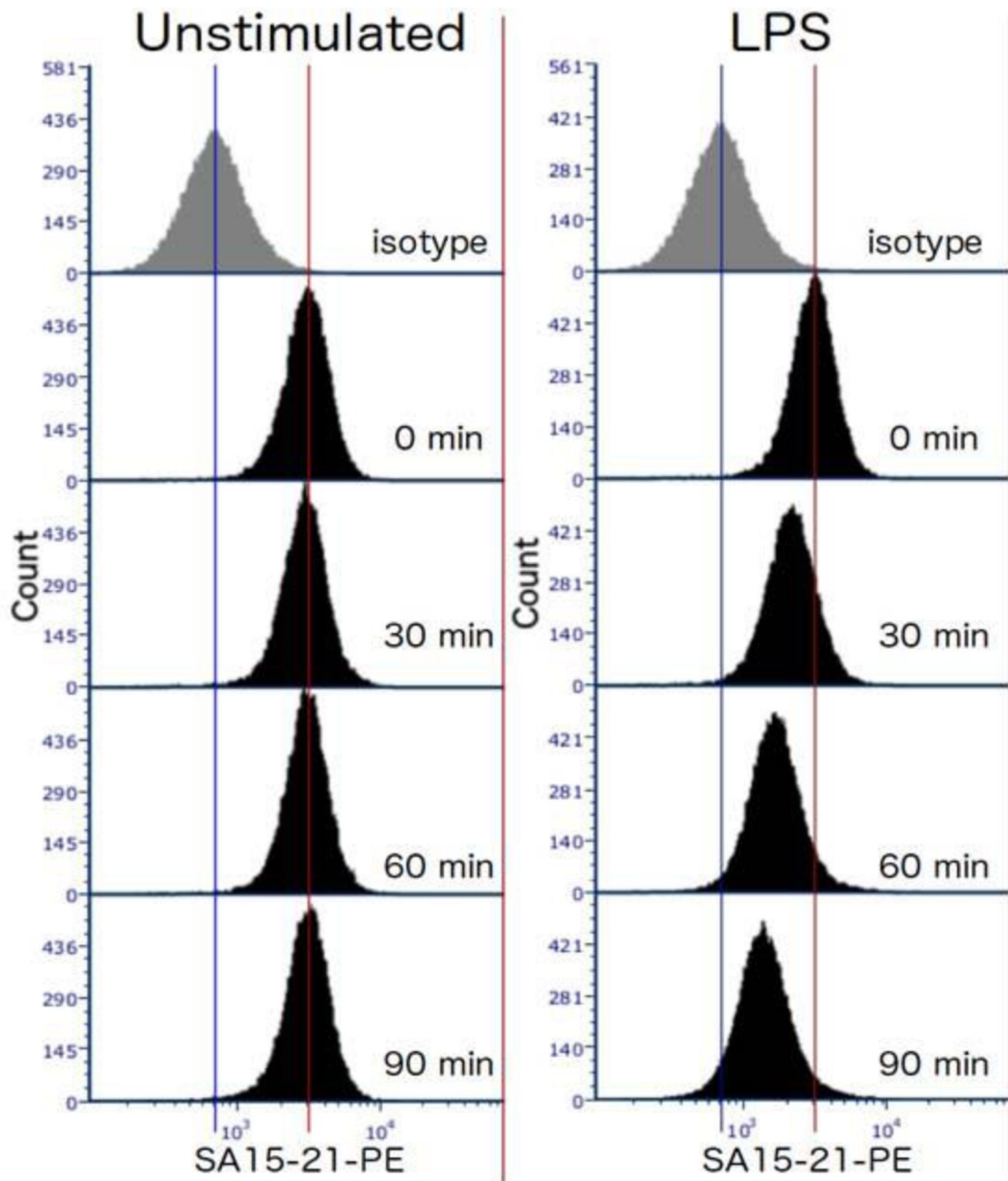
1. Culture Media
  - a. RPMI 1640 + 2% FBS + 2 mM L-glutamine + 1% Pen/Strep
  - b. Filter sterilize through 0.22  $\mu\text{m}$  filter
  - c. Store at 4 °C for up to three weeks
2. FACS buffer
  - a. Ice-cold PBS + 0.5% FBS + 2 mM EDTA
  - b. Store at 4 °C for up to three weeks

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**Figure 1.**

Representative data showing TLR4 internalization in response to 100ng/ml E coli LPS in TEPMs at the indicated time points. TEPMs were stained with anti-mouse TLR4 antibody and analyzed by flow cytometry.