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Chronic creatine supplementation alters depression-like behavior in rodents in a sex-dependent manner

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Abstract

Impairments in bioenergetic function, cellular resiliency, and structural plasticity are associated with the pathogenesis of mood disorders. Preliminary evidence suggests that creatine, an ergogenic compound known to promote cell survival and influence the production and utilization of energy in the brain, can improve mood in treatment resistant patients. The present study examined the effects of chronic creatine supplementation using the forced swim test (FST), an animal model selectively sensitive to antidepressants with clinical efficacy in humans. Thirty male (experiment 1) and thirty-six female (experiment 2) Sprague-Dawley rats were maintained on either chow alone or chow blended with either 2% w/w creatine monohydrate or 4% w/w creatine monohydrate for 5 weeks prior to the FST. Open field exploration and wire suspension tests were used to rule out general psychostimulant effects. Male rats maintained on 4% creatine displayed increased immobility in the FST as compared to controls with no differences by diet in the open field test, while female rats maintained on 4% creatine displayed decreased immobility in the FST and less anxiety in the open field test compared with controls. Open field and wire suspension tests confirmed that creatine supplementation did not produce differences in physical ability or motor function. The present findings suggest creatine supplementation alters depression-like behavior in the forced swim test in a sex-dependent manner in rodents, with female rats displaying an antidepressant-like response. While the mechanisms of action are unclear, sex-differences in creatine metabolism and the hormonal milieu are likely involved.

Keywords

creatine; creatine kinase; phosphocreatine; major depressive disorder; mood; forced swim test; antidepressant; treatment response; metabolic dysfunction; ATP

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DISCLOSURE/CONFLICT OF INTEREST

PF Renshaw is a consultant for Novartis, GSK, Roche, and Kyowa Hakko. He receives or has received research support from GSK, Roche, and Eli Lilly. PF Renshaw is an inventor on a patent application filed by McLean Hospital (Belmont, MA, USA) in 2001 that describes this use of creatine as a treatment for depression. No patent has issued, and this application has not been licensed. PJ Allen, KE D'Anci, and RB Kanarek have no financial or competing interests to declare.

INTRODUCTION

Changes in brain energy metabolism, neuronal plasticity, and cellular resiliency are associated with the pathogenesis of depressive disorders (Gabbay *et al*, 2007; Iosifescu *et al*, 2008; Volz *et al*, 1998; for review, see Fuchs *et al*, 2004; Kalia, 2005; Manji *et al*, 2001; McEwen, 2005). Additionally, evidence indicates that brain bioenergetics and cell survival pathways are necessary therapeutic targets for long-term clinical relief and symptom remission (Burroughs and French, 2007; Duman *et al*, 1999; Manji *et al*, 2000; Moretti *et al*, 2003). Taken together, a novel therapeutic strategy is to evaluate the effects on mood of agents that alter energy parameters and promote neuronal survival. In support of this supposition, two open-labeled studies found that creatine, a naturally occurring ergogenic compound, can significantly improve depressive mood (Amital *et al*, 2006; Roitman *et al*, 2007).

Creatine (*N*-aminoiminomethyl-*N*-methylglycine) is acquired from high protein foods or produced endogenously from the biosynthesis of arginine, glycine, or methionine in the liver, kidney, pancreas, and possibly the brain (Andres *et al*, 2008). The creatine-phosphocreatine (PCr) system serves as a spatial and temporal energy buffer in tissues with significant and fluctuating energy requirements, including the brain (Brosnan and Brosnan, 2007; McLeish and Kenyon, 2005; Walliman *et al*, 1992). Pools of cellular energy (ATP) are generated from the reaction $\text{PCr} + \text{ADP} \rightarrow \text{Creatine} + \text{ATP}$ and, conversely, energy is stored in the form of phosphocreatine: $\text{Creatine} + \text{ATP} \rightarrow \text{PCr} + \text{ADP}$ (Niklasson and Agren, 1984; Wyss and Kaddurah-Daouk, 2000). Creatine kinase (CK) isoenzymes, the catalysts of these reactions, are located in tissue where energy is greatly needed, making this enzyme system a critical regulator of energy homeostasis (Kuzhikandathil and Molloy, 1994; Walliman *et al*, 1992; Wilken *et al*, 1996).

Creatine may modify depressive behavior due to its pivotal role in buffering metabolic processes to prevent energy exhaustion and neuronal death. Creatine supplementation has been investigated in neurodegenerative and neurological diseases linked with mitochondrial dysfunction, such as Parkinson's Disease, Huntington's Disease, and Amyotrophic Lateral Sclerosis (see Andres *et al*, 2008 for review), and may be of particular relevance for psychiatric disorders in light of evidence that mitochondrial brain abnormalities occur in depressed subjects (Rezin *et al*, 2008; Shao *et al*, 2008). Findings from rodent research indicate that creatine exerts neuroprotective effects by buffering ATP levels against neurotoxic assaults from 3-NP, MPTP, malonate, and harmful levels of glutamate (Brustovetsky *et al*, 2001; Matthews *et al*, 1998; 1999; Roy *et al*, 2002). Moreover, creatine promotes neuronal growth and survival. For instance, *in vitro* culture studies demonstrate that supplementation enhances dopaminergic cell survival (Andres *et al*, 2005) and promotes differentiation of GABA-ergic neuronal precursors, potentially increasing cell soma size and number of primary neurites per neuron (Ducray *et al*, 2007).

Converging evidence indicates that alterations in high energy phosphate metabolism, particularly creatine and phosphocreatine levels, are associated with depression (Agren and Niklasson, 1988; Czéh *et al*, 2001; Iosifescu *et al*, 2008; Moore *et al*, 1997; Niklasson and Agren, 1984; Rezin *et al*, 2008; Volz *et al*, 1998). Phosphorus-magnetic resonance

spectroscopy (P-MRS) has shown increased phosphocreatine and decreased ATP in the frontal lobe and basal ganglia of depressed subjects *in vivo* (Kato *et al*, 1992; 1994; Moore *et al*, 1997; Volz *et al*, 1998). Additionally, creatine kinase activity and levels of brain creatine are inversely related to the severity of a depressive episode (Dager *et al*, 2004; Segal *et al*, 2007).

Preliminary findings suggest that creatine supplementation improves mood in humans (Amital *et al*, 2006; Roitman *et al*, 2007). Men and women with treatment resistant depression (Roitman *et al*, 2007) or PTSD with and without co-morbid depression (Amital *et al*, 2006) supplemented with creatine monohydrate for four weeks reported elevated mood on the Hamilton Depression Rating Scale (Hamilton, 1960). This evidence is encouraging, but these studies are methodologically limited and confounded by sex, co-morbidity, concurrent drug treatment, age, and severity of depression. Gender comparisons have not been reported in these studies due to small sample sizes, but the function and expression of CK is sexually dimorphic (Gledhill *et al*, 1988; Meltzer, 1971; Ramirez *et al*, 2002; Wong *et al*, 1983). Moreover, altered purine levels in depressed women, but not men, have been associated with treatment response (Renshaw *et al*, 2001), suggesting that creatine may be more beneficial for treating depressed females.

To resolve these issues, the objective of the present investigation was to determine the effects of long-term creatine supplementation on male (experiment 1) and female (experiment 2) treatment naive rats using the forced swim test (FST), a rodent behavioral assay that is selectively sensitive to antidepressants which have clinical efficacy in humans (Detke *et al*, 1995; Porsolt, 1978; Porsolt *et al*, 1977; 1978). Insight into the mechanisms associated with chronic creatine supplementation can be gained from the modified protocol of the FST, which distinguishes serotonergic and noradrenergic activity from selective increases swimming or climbing behavior, respectively (Detke *et al*, 1995).

MATERIALS AND METHODS

Animals and Housing

In experiment 1, thirty male Sprague Dawley rats (175–200 g), approximately 6 weeks old, were obtained from Charles River Laboratories (Raleigh, NC). Rats were housed individually in hanging stainless-steel cages in a climate controlled vivarium ($22 \pm 1^\circ\text{C}$) on a 12:12 h reverse light-dark cycle (lights on at 1900h). Animals were given 7 days to acclimate to handling and housing conditions prior to the beginning of the study. All experimental manipulations were conducted under red lights during the dark portion of the cycle, except as noted.

In experiment 2, thirty-six female Sprague Dawley rats (155–175 g), approximately 6 weeks old, were obtained from Charles River Laboratory (Raleigh, N.C.). Housing conditions, diet, and behavioral procedures were maintained exactly as in experiment 1. Vaginal smears were not obtained to ensure that females were not exposed to additional stressors not experienced by males.

All testing occurred between 1000 and 1800 hours, and animals were tested in random order. All procedures were approved by the Institutional Animal Care and Use Committee at Tufts University.

Diets

Animals were fed either chow alone or chow blended with 2% w/w creatine monohydrate or 4% w/w creatine monohydrate for 5 weeks prior to behavioral testing (n = 10 males per group; n = 12 females per group). The creatine-supplemented mixtures were made in-house using creatine monohydrate 98% (Sigma Chemical; 0 kcal/g) and ground Purina chow #5001 (3.4 kcal/g). The diets were mixed thoroughly each morning before the animals' food cups were refilled. Food was presented in Wahmann LC306 (Timonium, MD) stainless-steel food cups, which were covered with lids and clipped to the floor of the cage to protect against spillage. Glass water bottles with drip-proof stoppers were fitted at the front of each cage. All rats were weight-matched at the beginning of the experiment. Body weight, and food and water intakes were measured daily at the same time of day. Food intake was recorded to the nearest 0.1 g and reported in calories (to the nearest 0.1 kcal). Water intake and body weights were recorded to the nearest 1.0 g. Creatine intake was calculated by multiplying the amount of food consumed by the percentage of creatine added to the diet. The amount of creatine consumed was reported in grams per kilogram (g/kg).

Behavioral Testing

All behavioral measures were performed between 5 and 6 weeks after initial creatine supplementation. All procedures were conducted by the same experimenters under red lights, with the exception that the open field test was conducted under indirect dim white light (60 watts). All behavioral tests were set to start at the same time of day for all procedures (1000 h), and all animals were tested in random order. All behavioral measures were video-recorded for later scoring by two observers blind to dietary conditions (inter-rater reliability correlation > .90 for all tests).

Forced Swim Test (FST)

The FST was conducted over two days and consisted of a pre-test and test trial, referred to as trial 1 and trial 2. During the first trial, each rat was placed in a clear Plexiglas cylinder (25 cm in diameter by 65 cm in height) filled to 48 cm with 25 degree C° water for 15 minutes. Twenty-four hours after initial testing, each rat was placed in the same apparatus for 5 minutes. Each rat was tested in a clean container with fresh water, dried thoroughly upon trial completion, and placed back in its home cage. The container was drained and cleaned after each use.

The time-sampling technique developed by Detke *et al* (1995) was employed to score the first five minutes of behavior during trial 1 and the five minutes of behavior during trial 2. At the end of each 5-second period of time, two raters naive to diet conditions scored the predominant behavior of the rat (for a total of 60 intervals per trial). Four behaviors were rated: (1) immobility – defined as making only those movements necessary to stay afloat, (2) swimming – judged as active motions that propel the animal around the cylinder or motions that were more active than what is required to only stay afloat, (3) climbing – defined as

vigorous movement of the forepaws that is directed against the wall, (4) diving – defined as when the animal submerges its whole body and directs its head in a downward motion. Because diving occurred rarely in experiment 1 and experiment 2, and because diving has been previously shown to be an unreliable measure of antidepressant-like activity (Detke *et al*, 1995), diving counts are not reported. Latency to the onset of immobility was operationally defined as at least two consecutive five-second intervals (at least 10 seconds) of immobility. Latency to immobility is indicative of the individuals' sensitivity to an uncontrollable stressor (Carlezon *et al*, 2003; Leussis and Andersen, 2008). Specifically, decreased latency to immobility indicates the animal is more prone to depression-like behavior and the negative effects of forced swim stress, while increased latency indicates the opposite.

Body Temperature

Brain-type creatine kinase activity regulates body temperature rhythm and thermoregulation in rodents (Strejiger *et al*, 2009). Depletion of dietary creatine alters brown adipose tissue thermogenic activity (Yamashita *et al*, 1995), but the effects of excess dietary creatine on body temperature are unknown. To ensure that creatine supplementation does not alter behavioral response in the forced swim test, a digital rectal thermometer (Yellow Springs Instruments, Model 49TA) was used to record internal body temperatures before and after each swim trial.

Wire Suspension

Animals were allowed to rest after the FST for two days prior to psychomotor testing. The wire suspension test was used to rule out any potential confounds arising from differences in physical ability or motor function. The wire suspension task measured muscle strength and prehensile reflex, meaning the ability of the animal to grasp a taut horizontal wire (2 mm. in diameter, 62 cm. above the table top) with its forepaws and to remain suspended. Latency to drop from the wire was measured in seconds, with a maximum latency of 60 sec.

Open Field Exploration

The open field test was designed to measure anxiety as well as behavioral responses such as locomotor activity and exploratory behaviors. One week after the forced swim test, rats were placed in a square apparatus (50 cm length × 50 cm width) divided into 5 cm² in a grid-like fashion. The center of the field was illuminated by a 60 watt bulb, which was the only direct source of light in the testing room. Each animal was placed in a randomized starting corner within the field and its behavior was recorded for five minutes. Scored behaviors included the number of rears, stretch attends, grooming, and the number of times the animal returned to its starting corner. Also recorded were latency to leave the home corner, time spent in all corners, time spent in the borders, and time spent in the center of the field.

Statistical Analysis

Data were analyzed using SPSS v.15.0 for Windows XP. Body weight, calories, creatine, and water intake were analyzed using repeated measures (mixed model) ANOVA, with diet group as the between subjects factor and day as the within subjects factor. Additionally,

weekly averages of daily body weight, calorie and creatine consumption, and water intake were analyzed using one-way ANOVA.

Forced swim behavior and body temperature were analyzed using separate repeated measures (mixed model) ANOVA, with diet as the between subjects factor and trial as the within subjects variable. Separate one-way ANOVAs were also performed for each trial of forced swim behavior when significant effects were detected (i.e. latency to immobility, total immobility, swimming, climbing). Open field and wire suspension performance were analyzed using one-way ANOVA. Planned *post hoc* comparisons were made using least-significant differences (LSD) tests. Significance was defined as a *p*-value of < 0.05.

RESULTS

Experiment 1. Effect of creatine on forced swim test responsivity and locomotor performance in males

Diets and Body Weight—Average creatine intake (g/kg) significantly decreased over six weeks of supplementation (+2% creatine: $F(5,45) = 42.78, p < .0001$; +4% creatine: $F(5,45) = 23.71, p < .0001$). During the first week of supplementation, males fed +2% creatine consumed an average (\pm SEM) of 1.90 ± 0.02 g/kg daily, and males fed +4% creatine consumed an average of 4.17 ± 0.10 g/kg daily. The mean decline in creatine consumption over the course of six weeks for the +2% and +4% diets were 0.70 ± 0.2 g/kg and 1.53 ± 0.05 g/kg, respectively.

There were no significant diet by day interactions for body weight, calories, or water intake over six weeks of creatine supplementation (Figure 1a–c, respectively). Analyses of average daily water consumption by week showed a significant main effect of diet during week 1 ($F(2, 27) = 8.332, p = .002$) and during week 2 ($F(2, 27) = 4.5999, p = .019$). *Post hoc* tests indicated that rats given + 4% creatine drank significantly more water than rats in all other groups (Figure 1c), but this effect was no longer significant by the third week. There were no significant differences for the remainder of the study or during behavioral testing days.

Forced Swim Test

Trial: Repeated measures ANOVAs were conducted on all three diet groups together to determine the changes across time and between diet groups. Overall, there was a significant increase in total immobility over the two testing sessions ($F(1,27) = 33.30, p < .001$). Analysis of each diet group separately using paired t-tests indicated that the all groups significantly increased total counts of immobility (0%: $t(9) = -2.27, p = .049$; 2%: $t(9) = -3.57, p = .006$; 4%: $t(9) = -4.81, p = .001$). There was also a significant decrease in latency to the onset of immobility over the two testing sessions ($F(1,27) = 17.16, p < .001$). Analysis of each diet group separately indicated that the +2% diet ($t(9) = 3.27, p = .01$) and the +4% diet (4%: $t(9) = 2.47, p = .035$) significantly decreased latency to immobility. The +0% group decreased latency to immobility, but this group did not reach statistical significance. Climbing counts significantly decreased over the two testing sessions ($F(1,27) = 34.02, p < .001$). Analysis of each diet group separately indicated that the all groups significantly

reduced climbing behavior (+0%: $t(9) = 4.16, p = .002$; +2%: $t(9) = 3.49, p = .007$; +4%: $t(9) = 2.90, p = .018$. There were no significant changes in swimming behavior over time.

Diet: Latency to the onset of immobility differed significantly as a function of diet collapsed across trials ($F(2, 27) = 11.90, p < .001$). *Post hoc* analysis indicated that rats supplemented with +4% creatine more rapidly displayed immobility in the FST than rats given +0% creatine ($p < .001$) or +2% creatine ($p < .001$; Figure 2a). Mean counts of total immobility differed significantly as a function of diet collapsed across trials ($F(2, 27) = 4.392, p < .05$). *Post hoc* analyses indicated that rats supplemented with +4% creatine spent significantly more time immobile than rats fed +0% creatine ($p = .011$) or +2% creatine ($p = .027$; Figure 2b–c). There was no significant diet by trial interaction for latency to onset of immobility or mean counts of immobility.

Mean swimming counts differed significantly as a function of diet collapsed across trials ($F(2,27) = 9.841, p < .001$), but there was not a significant diet by trial interaction. *Post hoc* testing indicated that the +4% creatine group swam significantly less than the +0% creatine controls ($p < .001$) and the +2% creatine group ($p < .001$; Figure 2b–c). Mean climbing counts did not differ as a function of diet or diet over two trials (Figure 2b–c).

When data were analyzed separately for each FST trial, latency to immobility differed significantly as a function of diet during both trial 1 ($F(2,27) = 7.77, p = .002$) and trial 2 ($F(2,27) = 8.579, p = .001$). *Post hoc* analyses indicated that rats supplemented with +4% creatine more rapidly displayed immobility during trial 1 and trial 2 than rats given +0% creatine (trial 1: $p = .003$; trial 2: $p < .0001$) or +2% creatine (trial 1: $p = .001$, trial 2: $p = .037$). Immobility counts differed significantly during trial 2 only ($F(2,27) = 4.35, p = .023$). *Post hoc* analyses indicated that males fed +4% creatine spent more time immobile than males fed +0% creatine ($p = .009$) or +2% creatine ($p = .035$). Swimming counts differed significantly during trial 1 ($F(2,27) = 5.44, p = .01$) and trial 2 ($F(2,27) = 8.65, p = .001$). *Post hoc* analyses indicated that animals given +4% creatine swam significantly less compared to animals given +0% creatine during trial 1 and trial 2 (T1: $p = .011$, T2: $p < .0001$) and swam significantly less than males fed +2% creatine during T1 ($p = .006$). Climbing counts did not differ during trial 1 or trial 2.

Body Temperature

Body temperature declined significantly after exposure to each trial of the forced swim test (Trial 1: $F(1, 27) = 176.11, p < .0001$; Trial 2: $F(1,27) = 760.40, p < .0001$), but this hypothermic effect did not vary as a function of diet ($p > .05$). The average change in body temperature (\pm SEM) after exposure to trial 1 (15 minutes) for males fed +0% creatine, +2% creatine, or +4% creatine was 3.89 ± 0.67 C°, 5.66 ± 0.34 C°, and 4.70 ± 0.77 C° respectively. The average change in body temperature after exposure to trial 2 (5 minutes) for males fed +0% creatine, +2% creatine, or +4% creatine was 3.84 ± 0.25 C°, 3.61 ± 0.25 C°, and $3.47 \pm .19$ C°, respectively.

Wire Suspension

Total wire suspension time did not differ significantly as a function of diet (Figure 3a).

Open Field Exploration

Open field activity did not vary as a function of diet (all $p > .05$; Figure 3b–c). There were no significant differences in time to leave the home corner, time spent in all corners, time spent in the borders, or time spent in the center. There were no significant differences in behavioral counts, specifically the number of rears, stretch attends, episodes of grooming, or returns to home corner.

Experiment 2. Effect of creatine on forced swim test responsivity and locomotor performance in females

Unequal n—After the forced swim test, one female animal in the 2% creatine condition died due to causes unrelated to the test procedure. Behavioral data reflect this unequal n .

Diets and Body Weights—Average creatine intake (g/kg) significantly decreased over six weeks of supplementation (+2%: $F(5,50) = 33.09, p < .0001$; +4%: $F(5,50) = 49.60, p < .0001$). During the first week of supplementation, females fed +2% creatine consumed an average (\pm SEM) of 1.89 ± 0.03 g/kg daily, and females fed +4% creatine consumed an average of 3.77 ± 0.08 g/kg daily. The mean decline in creatine consumption over the course of six weeks for the +2% and +4% diets were 0.35 ± 0.04 g/kg and 0.70 ± 0.07 g/kg, respectively.

There were no significant diet by day interactions for body weight, calories, or water intake over 6 weeks of creatine supplementation (Figure 4a–c, respectively).

Forced Swim Test

Trial: Repeated measures ANOVAs were conducted on all three diet groups together to determine the changes across time and between diet groups. Overall, there was a significant increase in total immobility over the two testing sessions ($F(1,33) = 44.62, p < .001$). Analysis of each diet group separately using paired t-tests indicated that the all groups significantly increased total counts of immobility (0%: $t(11) = -4.41, p = .008$; 2%: $t(11) = -3.44, p = .005$; 4%: $t(11) = -3.83, p = .003$). There was also a significant decrease in latency to the onset of immobility over the two testing sessions ($F(1,33) = 50.82, p < .001$). Analysis of each diet group separately indicated that the all groups significantly decreased latency to immobility (0%: $t(11) = 3.23, p = .008$; 2%: $t(11) = 4.30, p = .001$; 4%: $t(11) = 5.06, p < .001$). There was no overall difference in swimming counts across two trials. Analysis of each diet group separately indicated that females fed the +0% diet significantly decreased swimming activity over time ($t(11) = 2.84, p = .016$), but there were no changes in swimming behavior by the +2% and +4% groups. There was a significant decrease in climbing behavior over the two testing sessions ($F(1,33) = 29.33, p < .0001$). Analysis of each diet separately indicated that all groups significantly reduced climbing activity in the second trial (0%: $t(11) = 2.65, p = .023$; 2%: $t(11) = 3.28, p = .007$; 4%: $t(11) = 3.41, p = .006$).

Diet: Latency to immobility differed significantly as a function of dietary condition collapsed across trials ($F(2, 33) = 5.956, p = .006$). *Post hoc* analyses indicated that the +0% creatine controls more rapidly displayed immobility in the FST than rats in the +4% creatine

condition ($p < .001$; Figure 5a). Mean counts of total immobility differed significantly as a function of dietary condition collapsed across trials ($F(2, 33) = 3.328, p = .048$). *Post hoc* analyses indicated that rats supplemented with +4% creatine spent significantly less time immobile than the +0% creatine controls ($p = .021$), but did not differ significantly from rats given +2% creatine ($p = .654$; Figure 5b–c). There was no significant diet by trial interaction for latency to onset of immobility or mean counts of immobility.

Mean swimming counts did not differ as a function of diet across trials, but there was a significant diet by trial interaction ($F(2,33) = 4.28, p = .022$). *Post hoc* analysis indicates that the +0% creatine controls reduced total swimming activity over time while rats given +2% and +4% creatine showed similar counts during both trials (Figure 5b–c). Mean climbing counts did not differ as a function of diet or diet by trial interaction.

When data were analyzed separately for each FST trial by diet, latency to immobility differed significantly as a function of diet during both trial 1 ($F(2,33) = 6.07, p = .006$) and trial 2 ($F(2,33) = 3.28, p = .049$). *Post hoc* analyses indicated that females fed +4% creatine less rapidly displayed immobility compared with females fed +0% creatine (Trial 1: $p = .001$; Trial 2: $p = .019$). Swimming counts differed significantly during trial 2 only ($F(2,33) = 3.42, p = .045$). *Post hoc* analyses indicated that females fed +2% creatine swam significantly more than females fed +0% creatine ($p = .013$). Immobility counts and climbing counts did not differ significantly during trial 1 or trial 2.

Body Temperature

Body temperature declined significantly after exposure to each trial of the forced swim test (Trial 1: $F(1,33) = 607.10, p < .0001$; Trial 2: $F(1,33) = 708.17; p < .0001$), but this hypothermic effect did not vary as a function of diet ($p > .05$). The average change in body temperature (\pm SEM) after exposure to trial 1 (15 minutes) for females fed +0% creatine, +2% creatine, or +4% creatine was 7.47 ± 0.44 C°, 7.53 ± 0.80 C°, and 8.10 ± 0.26 C° respectively. The average change in body temperature after exposure to trial 2 (5 minutes) for females fed +0% creatine, +2% creatine, or +4% creatine was 3.18 ± 0.23 C°, 3.47 ± 0.23 C°, and 3.82 ± 0.23 C°, respectively.

Wire Suspension

Total wire suspension time did not differ significantly as a function of diet ($p > .05$; Figure 6a).

Open Field Test

Open field activity did differ as a function of diet (Figure 6b–c). There were significant differences in time spent in all corners ($F(2,32) = 3.357, p = .047$) and time spent in the borders ($F(2,32) = 3.448, p = .044$). *Post hoc* analyses indicates that rats given +4% creatine spent significantly less time in the corners than the +0% creatine group ($p = .016$), while the +4% creatine group spent significantly more time in the borders than both +0% creatine ($p = .022$) and +2% creatine ($p = .044$) groups. There were no significant differences in time to leave the home corner, latency to enter center, or time spent in the center. There were also

no significant differences in behavioral counts, specifically the number of rears, stretch attends, or returns to home corner.

DISCUSSION

Creatine supplementation alters depression-like behavior in the forced swim test in a sex-dependent manner in rodents. Male rats given +4% creatine more rapidly displayed immobility and spent significantly more time immobile than rats in the +0% or +2% creatine conditions, suggesting that males fed +4% creatine are more vulnerable to the negative effects of forced swim stress and are prone to depression-like behavior. In comparison, female rats supplemented with +4% creatine displayed immobility less rapidly and spent significantly less time immobile compared to the +0% creatine condition, suggesting that +4% creatine attenuates the negative effects of forced swim stress and exerts antidepressant-like effects in female rats. Of note is the behavioral specificity of these findings. Muscle strength, locomotion, and body temperature did not differ as a function of diet in both males and females. Additionally, females supplemented with +4% creatine displayed modest reductions in anxiety compared to rats given +0% and +2% creatine without altering locomotor activity.

A complementary pattern of findings from swimming and climbing counts indicate that dietary creatine is associated with serotonergic activity in males and females in opposite directions. Detke and colleagues (1995) have shown that serotonergic antidepressants selectively increase swimming behavior, while noradrenergic antidepressants selectively increase climbing behavior. Moreover, antagonists of the serotonergic system prevent both the reduction of immobility and increase in swimming behavior usually observed in rats administered specific serotonin reuptake inhibitors (SSRIs; Page et al., 1999; Wang et al., 2008). In the present studies, males fed +4% creatine showed significantly reduced swimming activity with no changes in climbing activity compared with controls. Females fed +2% and +4% creatine maintained higher swimming activity over two trials compared to females fed +0% creatine with no differences in climbing activity. Ongoing studies are examining the relationship between creatine and serotonergic agents, specifically to evaluate the usefulness of creatine as an adjuvant to SSRI drug therapy.

Importantly, male and female animals supplemented with +4% creatine displayed differential responsivity to forced swim stress during the initial trial of the procedure. These findings contrast with the original purpose of trial 1 of the FST as described by Porsolt and colleagues (1977), which was to induce “behavioral despair,” a phenomenon characterized by reduced latency for rats to become immobile and increased total immobility during the second trial. Antidepressant agents are typically administered after the pre-test trial and before the test trial, resulting in an anti-immobility effect at certain doses. Accordingly, the first trial of the FST procedure is not often quantified or reported because the experimental manipulation takes place following the induction of a state change in the animal. The present experimental design was different in that the rats were administered creatine daily for five weeks prior to the initial FST, the average duration of treatment before clinical effects are observed in humans. The finding that chronic creatine supplementation altered behavior in the first trial is consistent with previous research that has found that chronic antidepressant

treatment can induce significant changes in forced swimming behavior during the first trial (Armario *et al*, 1988; Lahmame *et al*, 1996; Overstreet *et al*, 2004).

It is important to note that there were no statistical comparisons between males and females because each sex was studied at different times. Moreover, to be consistent with previous research, male and female diets were enhanced with equivalent percentages of creatine (+2% and +4% w/w), but this translated into significantly different dosages of creatine per kilogram of body weight given that males eat larger quantities of chow and gain substantially more weight over a six week period of time. The fact that males and females received different absolute doses that declined over the course of the study could have had an effect on behavior. It is critical to know if rodent behavior varies at different doses or if there is a threshold dose after which the effects begin to occur.

The present observation that male rats fed +4% creatine are more prone to depression-like behavior is consistent with evidence that suggests that creatine supplementation alters brain chemistry in a manner that may be conducive to depressive behavior. Unmedicated depressed male and female patients have increased absolute concentrations of total creatine (creatine and phosphocreatine) in the left and right frontal lobe (Frey *et al*, 2003). Further, Lyoo and colleagues (2003) reported that oral creatine supplementation in humans significantly increased creatine and PCr reserves while it decreased beta gamma-nucleotide triphosphate (β -NTP) levels, which reflect lowered ATP levels. Reduced total ATP and lowered creatine kinase activity are characteristic of depression (Moore *et al*, 1997, Volz *et al*, 1998). High levels of creatine supplementation may cause the brain to make compensatory changes to accommodate increases in the high-energy phosphate pool. Specifically, it is hypothesized that a shift in the creatine kinase equilibrium reaction down-regulates the synthesis of ATP in response to increased PCr to maintain energy homeostasis (Iosfescu *et al*, 2008; Lyoo *et al*, 2003). The present finding has important clinical implications. Specifically, high baseline levels of PCr and low levels of ATP may be unique to depressed patients who respond more favorably to antidepressant treatment (Iosfescu *et al*, 2008). Supplementation with creatine can induce equivalent changes in brain energetic parameters in healthy adults (Lyoo *et al*, 2003), suggesting the possibility that creatine supplementation can modify brain high-energy phosphate metabolism to improve treatment response in patients with depression. Moreover, the effectiveness of antidepressant drugs may be related to their actions upon phosphorylating enzymes like creatine kinase (Moretti *et al*, 2003). Altogether, although creatine may be pro-depressant in male rats as a dietary supplement alone, it may sensitize the brain to antidepressant treatment, thereby increasing therapeutic efficacy.

In contrast to the results for males, the highest dose of creatine (+4%) produced antidepressant-like effects in female rats. This result is consistent with findings that performance in animal models of depression and responses to antidepressant treatment are sex-dependent (Brotto *et al*, 2000; Drossopoulou *et al*, 2004; Lifschytz *et al*, 2006). However, sex comparisons have not been made between male and female depressed patients in human clinical trials evaluating the antidepressant effect of creatine supplementation due to the small sample sizes (Amital *et al*, 2006; Roitman *et al*, 2007). As such, the mechanisms underlying the sex-specific effects of creatine supplementation in rats have yet

to be elucidated but may involve sex differences in general metabolic processes, the hormonal milieu, or the stress response. The differential response to creatine supplementation may be explained by sexually dimorphic patterns of CK expression, levels of CK activity or total creatine concentrations (Gledhill *et al*, 1988; Meltzer, 1971; Ramirez *et al*, 2002; Wong *et al*, 1983). The brain-type CK (BB-CK) is a favored marker of estrogenic action (Reiss and Kaye, 1981). In females, estrogen rapidly up-regulates the synthesis of BB-CK in regions densely populated by estrogen receptors, such as the preoptic area, anterior hypothalamus, and median eminence (Malnick *et al*, 1983). Ramirez and colleagues (2002) observed that only female rats displayed cyclical increases and decreases in cerebral CK activity, which may reflect cyclical changes in the estrous cycle. Further, it was reported that female rats express a larger number and variance of cytosolic CK in the cerebrum and cerebellum, which increases the efficiency of ATP replenishment, via the creatine-PCr system, to meet sudden energy needs. One potential implication is that estrogen or its receptors modulate the therapeutic effect of creatine observed in female rats via the sexually dimorphic expression and function of CK.

Estrogen is a plausible consideration, as there is a number of clinical and experimental evidence that estrogenic metabolites mediate depressive behavior (Carranza-Lira and Valentino-Figueroa, 1999; Cohen *et al*, 2003; Estrada-Camarena *et al*, 2003; Halbreich and Kahn, 2001; Rodriguez-Landa *et al*, 2007; Walf *et al*, 2004). An important question is whether creatine can exert antidepressant effects in the absence of estrogen. It is hypothesized that estrogen plays a protective role in the brain by enhancing energy metabolism and utilization through direct stimulation of protein kinase-mediated phosphorylation in the mitochondria of neurons (for review, see Chen *et al*, 2005; Klinge, 2008; Vasudevan and Pfaff, 2008). If the antidepressant-like effects of creatine depend on the presence of estrogen, the estrogen receptor ER β isoform may be involved because it is localized within mitochondria of neurons (Yang *et al*, 2004), and it is known to have a distinct role in mediating depressive behavior (Walf and Frye, 2006; Walf *et al*, 2008; 2004; Rocha *et al*, 2005). Taken together, it may be possible that the sex-specific differences by diet observed in the FST are a consequence of estrogenic influences on creatine kinase activity, but this question cannot be answered with the present findings. This is an important avenue for future research as depression occurs twice as frequently in women than in men, a finding that is consistent across history and cultures (Bebbington *et al*, 2003; Kessler, 2003). Moreover, depressed women are more severely impaired, experience longer episodes, and relapse more frequently than depressed men (Kornstein *et al*, 2000; Nolen-Hoeksema, 1987). Ongoing research is planned to evaluate the relationship between sex hormones and alterations in depression-like behavior in response to creatine supplementation.

Future research should extend the present findings to clarify the effects of both exogenous and endogenous creatine on the central nervous system. An important consideration in the interpretation of the present results is the choice of animal model. The key rationale of the present research was based on human clinical trials, which found that creatine had mood-elevating effects in depressed patients resistant to antidepressant therapies (Roitman *et al*, 2007) and patients with comorbid PTSD and depression (Amital *et al*, 2006). The FST was chosen to evaluate animal behavior because it demonstrates superior construct and

predictive validities among models that detect antidepressant-like effects (Dalvi and Lucki, 1999). As such, it would be the best model to answer the present research question of whether creatine can exert antidepressant-like effects in rodents. However, the “mood state” of the animal during the time of treatment and testing may alter the behavioral outcomes. A concern in animal literature is the use of antidepressant models that subject otherwise healthy animals to acute stress. Stress has been studied extensively as a precipitating factor in the onset of depression (Caspi *et al*, 2003; Duman and Monteggia, 2006; Pittenger and Duman, 2008, Sapolsky, 2000), and the effects of a given treatment may vary greatly depending on whether the subject has a “healthy” versus “depressed” brain, the latter of which is metabolically impaired. The possibility remains that metabolic impairments resulting from chronic stress could alter the effects of creatine on behavioral responsiveness in the FST. Accordingly, use of an animal model of depression, such as the unpredictable chronic mild stress paradigm (Willner *et al*, 1987; Willner, 1997), would further clarify the effects of creatine on depression-like behavior in rats.

Dosages and time course are two more notable considerations for behavioral studies. Ample research has established that dietary creatine, in daily doses ranging from +0.25% w/w to +7% w/w, is safe and well-tolerated in rodents (Horn *et al*, 1998). However, the majority of animal research has failed to include more than one dose of creatine, with the majority using +2% w/w or less (Pe a-Altamira *et al*, 2005; Rooney *et al*, 2002; Roy *et al*, 2002; Sullivan *et al*, 2000). There is need to evaluate the dose-response curve for creatine using a more reliable method of administration, such as the use of gavage (as with Ozkan *et al*, 2005) or intracerebroventricular injection (as used by Koga *et al*, 2005). Additionally, rats in the +4% creatine group consumed approximately 2 to 4 g/kg daily, an amount that is at least five to ten times the highest dose usually taken by humans. For instance, clinical trials in humans with a number of neurological conditions, such as GAMT deficiency or mitochondrial myopathy, treated patients with 0.13 g/kg to 0.8 g/kg daily (for review, see Andres *et al*, 2008).

Additionally, housing and handling procedures may have influenced behavior in the present studies. Animals were handled extensively prior to behavioral testing and housed singly. The majority of forced swim test studies house animals in groups, allowing for social interaction. The present study housed rats in isolation, which has been shown to increase immobility in the forced swim test (Grippio *et al*, 2008; Sáenz *et al*, 2006), alter responsiveness to the effects of antidepressant drugs (Wongwitdecha *et al*, 2006), and reduce hippocampal neuroplasticity factors linked to depression, such as brain derived neurotrophic factor, neuropeptide Y, and serotonin receptors, in a sex-specific manner (Bjørnebekk *et al*, 2008). Additionally, evidence indicates that isolation affects these changes in a sex-specific manner. For instance, in rats, group-housed females exhibit similar levels of immobility as isolated females and more immobility compared to group-housed males; conversely, males left in isolation show significantly more depression-like behavior compared to group-housed males, indicating male rats are more vulnerable to social isolation stress (Leussis and Andersen, 2008). Further, socially isolated male mice show enhanced 5-HT_{1A} receptor binding compared to females in areas throughout the brain, including the hippocampus, septum, cortical amygdala, and in the periaqueductal grey,

suggesting isolation housing could differentially alter antidepressant response in a sex-dependent manner (Schiller *et al.*, 2006).

There is reason for concern if long-term creatine supplementation can induce depressive behavior by means of altering brain chemistry, as creatine monohydrate is one of the most popular dietary supplements on the market, with a 400 million dollar industry to support its use (Dalbo *et al.*, 2008). Still, the majority of literature examining the effects of creatine supplementation on cognitive function has been positive for both human and animals. In humans, daily supplementation between 5–8g/day (up to 6 weeks) reduces mental fatigue (Watanabe *et al.*, 2002), enhances recognition memory (Ferrier *et al.*, 2000) and working memory (Rae *et al.*, 2003), and attenuates performance decline from significant sleep deprivation (McMorris *et al.*, 2006). In animals, creatine supplementation lessens stress-responses (e.g. vocalizations, spontaneous activity, corticosterone concentration) of chicks subjected to social-separation stress (Koga *et al.*, 2005). It remains to be seen whether creatine is making male animals more adaptive to forced swim test stress, as becoming immobile more rapidly may be an advantageous coping strategy in terms of energy conservation.

The present study offers evidence that creatine supplementation has a beneficial effect on depression-like behavior in females and may sensitize the brain to antidepressant treatment. Altered bioenergetics may be an important avenue for enhancing therapeutic efficacy in depression. Further studies are needed to determine the nature of endogenous creatine in the brain and the mechanisms by which it may influence affective processes.

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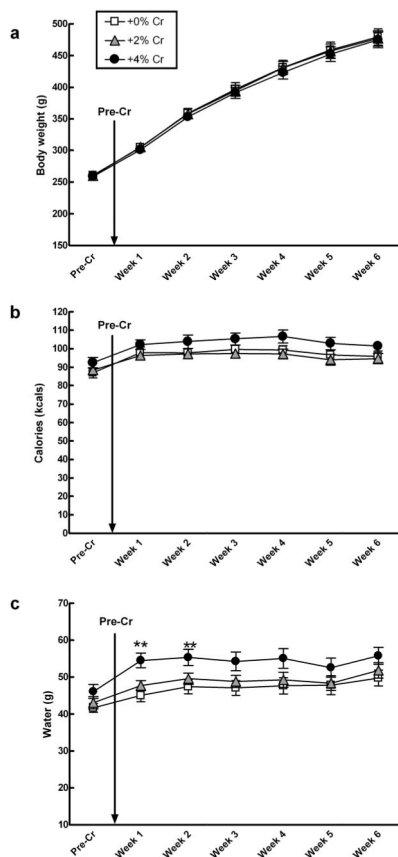


Figure 1. Mean (± SEM) body weight, caloric intake, and water consumption of males did not vary as a function of creatine supplementation over six weeks (a–c). Separate one-way ANOVAs for weekly averages of daily water consumption indicate that rats given +4% creatine drank more water during week 1 (** p < .001 vs. +0%, ** p < .008 vs. +2%) and week 2 (** p < .007 vs. +0%, *p < .041 vs. +2%), but this effect was no longer significant by the third week. There were no significant differences for the remainder of the study or during behavioral testing days.

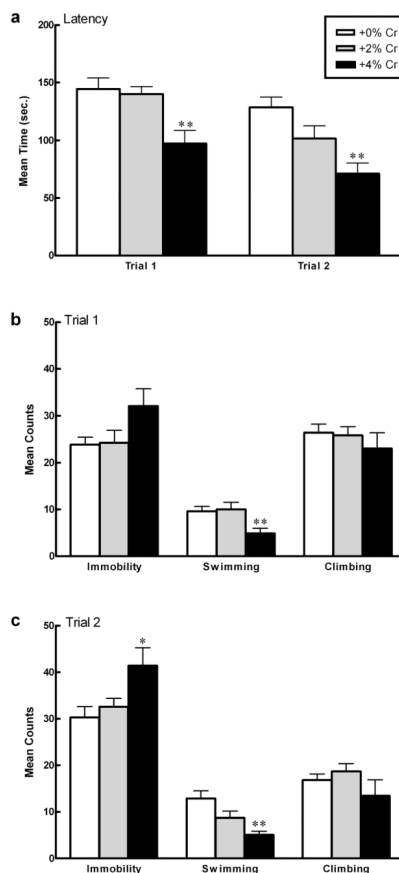


Figure 2. Mean (\pm SEM) latency to immobility (a), immobility counts (b–c), swim counts (b–c), and climbing counts (b–c) of males as a function of creatine supplementation. Note that significance values displayed on graphs are derived from separate one-way ANOVAs for each forced swim test trial compared to +0% controls. Repeated measures analyses indicate that: (a, top panel) +4% creatine group more rapidly displayed immobility across trials (** $p < .001$, +4% vs. +0%; ** $p < .001$, +4% vs. +2%). (b–c) Complementary pattern of results as measured by total immobility (* $p = .011$; +4% vs +0%; * $p = .027$, +4% vs +2% creatine), swimming (** $p < .001$, +4% vs. +0%; ** $p < .001$, +4% vs. +2%), and climbing ($p > .05$, n.s.). Data are presented as latencies to become immobile in seconds (a), or total counts of immobility, swimming, or climbing (b–c).

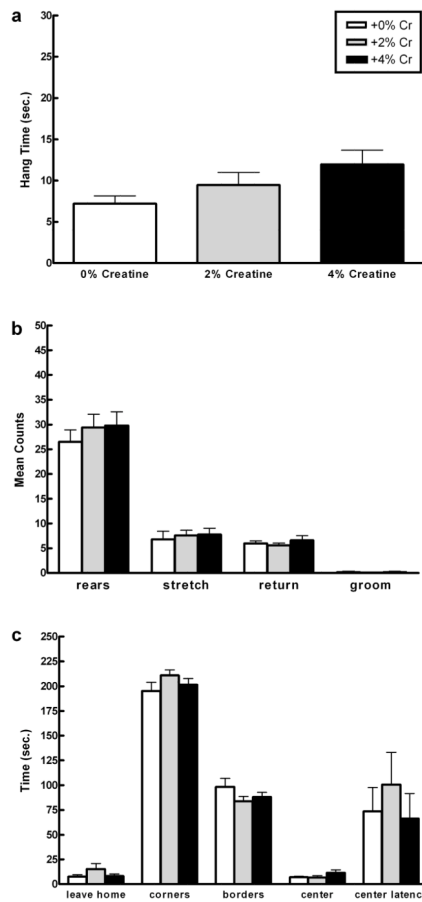


Figure 3. Mean (\pm SEM) wire suspension (a) and open field (b–c) performance of males as a function of creatine supplementation. Locomotion, anxiety-like behavior, and muscle strength did not vary by diet. Data for time in center and latency to enter center are represented by a subset of the total number of rats (+0%: $n = 9$; +2%: $n = 8$; +4% $n = 9$).

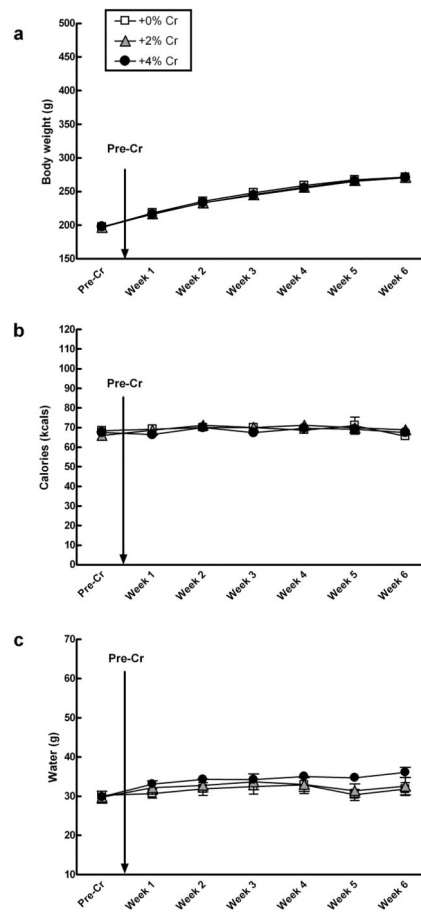


Figure 4.

Mean (\pm SEM) body weight, caloric intake, and water consumption of females did not vary as a function of creatine supplementation over six weeks (a–c). Separate one-way ANOVAs indicate that there were no differences in body weight (a), caloric intake (b), or water consumption (c) per week.

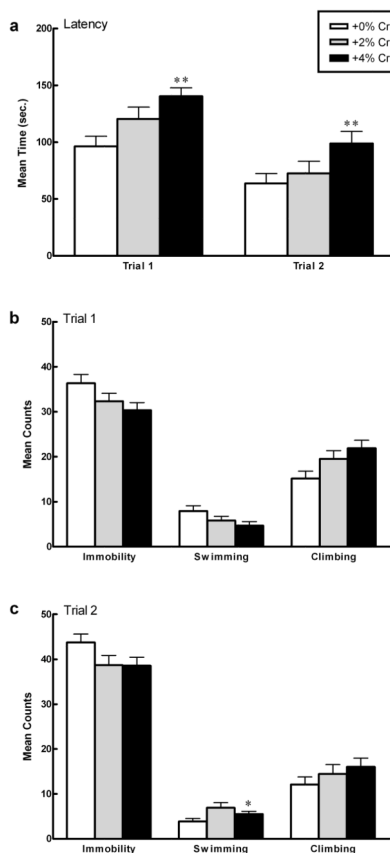


Figure 5.

Mean (\pm SEM) latency to immobility (a), immobility counts (b–c), swimming counts (b–c), and climbing counts (b–c) of females as a function of creatine supplementation. Note that significance values displayed on graphs are derived from separate one-way ANOVAs for each forced swim test trial compared to +0% controls. Repeated measures analyses indicate that: (a, top panel) +4% creatine group displayed immobility less rapidly across trials compared with controls (** $p < .001$, +4% vs. +0%; $p > .05$, +4% vs. +2%). (b–c) Complementary pattern of results as measured by total immobility (* $p = .021$, +4% vs. +0%; $p > .05$, +4% vs. +2%), swimming (** $p < .001$, +4% vs. +0%; $p > .05$, +4% vs. +2%), and climbing (n.s., all groups $p > .05$). Data are presented as latencies to become immobile in seconds (a), or total counts of immobility, swimming, or climbing (b–c).

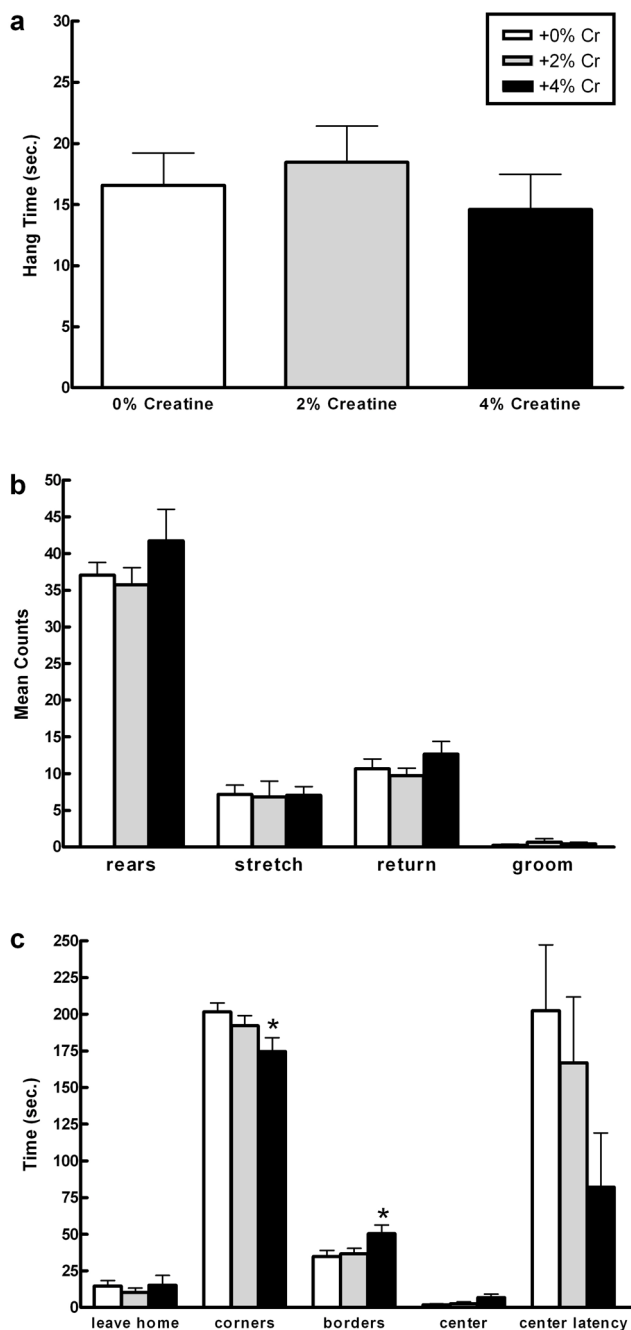


Figure 6.

Mean (\pm SEM) wire suspension (a) and open field (b–c) performance of females as a function of creatine supplementation. One-way ANOVAs indicate that (c, bottom panel) anxiety-like behavior varied by diet. Females fed +4% creatine spent significantly less time in the corners compared with controls ($*p = 0.016$, +4% vs. +0%) and significantly more time in the borders compared with animals fed +0% or +2% creatine ($*p = .022$, +4% vs. +0%; $*p = .044$, +4% vs. +2%), suggesting +4% creatine had a modest anxiolytic-like effect in females. Locomotion and muscle strength did not vary by diet (a–c). Data for time in

center and latency to enter center are represented by a subset of the total number of rats (+0%: $n = 5$; +2%: $n = 6$; +4% $n = 5$).

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