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HIV-1 Tat-induced Astrocytic Extracellular Vesicle miR-7 Impairs Synaptic Architecture

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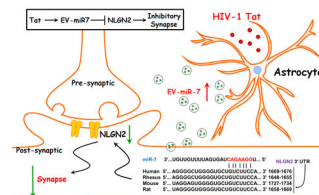
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Abstract

Although combination antiretroviral therapy (cART) has improved the health of millions of those living with HIV-1 (Human Immunodeficiency Virus, Type 1), the penetration into the central nervous system (CNS) of many such therapies is limited, thereby resulting in residual neurocognitive impairment commonly referred to as NeuroHIV. Additionally, while cART has successfully suppressed peripheral viremia, cytotoxicity associated with the presence of viral Transactivator of transcription (Tat) protein in tissues such as the brain, remains a significant concern. Our previous study has demonstrated that both HIV-1 Tat as well as opiates such as morphine, can directly induce synaptic alterations via independent pathways. Herein, we demonstrate that exposure of astrocytes to HIV-1 protein Tat mediates the induction and release of extracellular vesicle (EV) microRNA-7 (miR-7) that is taken up by neurons, leading in turn, to downregulation of neuronal neuroligin 2 (NLGN2) and ultimately to synaptic alterations. More importantly, we report that these impairments could be reversed by pretreatment of neurons with a neurotrophic factor platelet-derived growth factor-CC (PDGF-CC).

Graphical Abstract



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Conflict of Interest

The authors declare no competing financial interests in relation to the work described.

Keywords

HIV-1 Tat; Extracellular Vesicles; miRNA; Neuroligin; PDGF; synapse

INTRODUCTION

The clinical outlook for patients with HIV-1 (Human Immunodeficiency Virus, Type 1) has improved dramatically as an increasingly effective combination antiretroviral therapy (cART) options have become readily available (Maschke et al., 2000; Sacktor et al., 2002; Gray et al., 2003; McArthur et al., 2003). Paradoxically, however, increased survival rates resulting from therapy usage have resulted in an undesirable increase in the prevalence of minor neurocognitive impairment collectively referred to as NeuroHIV. Complications of the central nervous system (CNS) represent a substantial health burden in HIV-1 infected individuals because of the privileged immune status of the brain coupled with the limited accessibility of some ART regimens to penetrate the CNS (Ellis et al., 2007). Despite ART-mediated near complete suppression of virus replication, NeuroHIV still exists in almost 40% of infected patients (Simioni et al., 2010). Viral protein transactivator of transcription (Tat) that was first identified as a neurotoxin by Nath *et al.* (Nath et al., 1996) has been shown to exert diverse cytotoxic effects including, but not limited to, eliciting of neuroexcitatory responses (Haughey et al., 1999; Nath et al., 2000; Buscemi et al., 2007; Hui et al., 2012b; Hui et al., 2012a) as well as induction of dendritic/synaptic degeneration (Kim et al., 2008; Fitting et al., 2010; Mocchetti et al., 2012). Even in virally suppressed individuals that are on cART, mounting evidence implicates the presence of Tat in the lymph nodes and CNS compartments (Cowley et al., 2011). Viral Tat through its ability to exert excitotoxicity and inflammation is a contributor of CNS dysfunction (Gabuzda et al., 1986) by exerting toxicity in most cell types of the CNS including, but not limited to, the glia and the pyramidal hippocampal and cortical neurons (Aksenov et al., 2006). Studies have also demonstrated that exogenous Tat injections in the rodent brain cause a multitude of neuropathological (Bansal et al., 2000) and behavioral abnormalities mimicking aspects of NeuroHIV (Rappaport et al., 1999; Bruce-Keller et al., 2003).

Extracellular vesicles (EVs) play an essential role in mediating communication among diverse cell types and tissues, including the CNS (Cocucci et al., 2009; They et al., 2009; Gupta and Pulliam, 2014). Release kinetics of EVs, including their number, size distribution as well as their cargo, has been reported to be altered during various pathologies (Piccin et al., 2007; Shi et al., 2014) including cancer (Rabinowits et al., 2009; Taylor and Gercel-Taylor, 2011), neurological diseases (Gupta and Pulliam, 2014) and viral infections (Bhattarai et al., 2013; Narayanan et al., 2013). Elegant studies have also demonstrated that HIV-infected cells release EVs containing HIV proteins (Gag and Nef) and RNA (the trans-activation response element [TAR]) (Booth et al., 2006; Campbell et al., 2008; Shelton et al., 2012; Narayanan et al., 2013; Rahimian and He, 2016b; Sami Saribas et al., 2017) and that these viral components can be transferred to neighboring/distant cells leading to functional impairment of the recipient cells. Moreover, despite cART treatment, EVs from HIV-1-infected primary cells and patient biofluids such as plasma and cerebrospinal fluid continue

to express viral products (DeMarino et al., 2018), thereby underpinning the role of EVs in neurocognitive and immunological dysfunction in the era of cART.

Additionally, host microRNAs (miRNAs) and proteins that are released in EVs from donor cells during HIV-1 infection have also been well investigated (Hu et al., 2012; Konadu et al., 2015; Roth et al., 2015; Tang et al., 2016). For example, studies have shown upregulated expression of miR-21 in EVs isolated from the brains of macaques with simian immunodeficiency virus (SIV) encephalitis as well as from the brains of individuals with traumatic brain injury (TBI), which, in turn, leads to associated neurotoxicity (Yelamanchili et al., 2015) (Harrison et al., 2016). Findings from our group have also demonstrated the upregulated expression of miR-29b in HIV-1 Tat & morphine-stimulated astrocyte-derived EVs (ADEVs), resulting in neuronal apoptosis (Hu et al., 2012).

Members of the neuroligins (NLGNs) family of cell adhesion proteins are key regulators of synaptic structure, function, and remodeling (Sudhof, 2008; Li et al., 2017; Stogsdill et al., 2017) in the central neurons. Mutations in the NLGN family members have been implicated in autism spectrum disorders, thus suggesting their critical role as components of cellular machinery underlying neural circuit formation and function (Zhang et al., 2015; Martella et al., 2017).

In the current study, we observed upregulated expression of miR-7 in the astrocytes of SIV+/HIV+ brains. Furthermore, we also demonstrated that similar to the *in vivo* findings, astrocytes in culture that were exposed to HIV-1 Tat upregulated the induction and release of miR-7 in astrocyte-derived extracellular vesicles (ADEVs), and also that these exosomes were effectively taken up by rat primary hippocampal neurons, resulting in synaptic injury. Using bioinformatics and cellular analyses EV-miR-7 was found to target neuronal NLGN2. Interestingly, treatment of neuronal cells with the neurotropic factor platelet-derived growth factor-CC (PDGF-CC) was found to restore EV-miR-7-mediated synaptic injury. These findings thus suggest that PDGF-CC treatment could be considered as a neuroprotective strategy against Tat-EV-mediated alterations of synaptic densities.

RESULTS

Upregulated expression of miR-7 in the brains of SIV/HIV-infected subjects

As an initial approach, miRNA array profiling was performed on the RNAs isolated from the basal ganglia (BG) of both uninfected and SIV-infected rhesus macaques (RMs) using the LC Biosciences platform. We observed dysregulation of several key miRNAs in the archival brain tissues of infected rhesus macaques (Bokhari et al., 2011), compared with the uninfected macaques, with a significant increase in the expression of miRNAs controlling synaptic plasticity, such as miR-7 (Figure 1a). MiR-7 has been shown to play an important role in mediating neuronal homeostasis (Choi et al., 2015; Kleaveland et al., 2018). Based on the fact that there is disruption of neuronal plasticity in the brains of SIV-infected rhesus macaques and HIV-infected humans, we thus sought to assess the expression of miR-7 in the isolated from brain regions (BG and frontal cortices-FC) of SIV+/HIV+ subjects by qPCR. As shown in Figures 1b–1c, there was upregulated expression of miR-7 in the BG and FC of SIV+/HIV+ subjects compared with the uninfected controls. Upregulation of miR-7 in the

BG of SIV-infected RMs was also validated by *in situ* hybridization (ISH), wherein we found its expression primarily in the GFAP positive astrocytes (Figure 1d), compared with the uninfected controls. Additionally, as expected, there was also increased expression of GFAP in astrocytes in the brains of SIV-infected macaques compared with those from uninfected controls (Witwer et al., 2009; Lee et al., 2014).

Tat-stimulated astrocyte-derived EVs (Tat-ADEVs) contain miR-7 that can be taken up by neurons

HIV-1 protein Tat has been previously reported to contribute to the pathogenesis of HIV-associated neurocognitive disorders (HAND) (Johnson et al., 2013; Steiner et al., 2015). Action of Tat involves its ability to cause both excitotoxicity & neuroinflammation in various CNS cells including the astrocytes (Aksenov et al., 2006), ultimately resulting in CNS dysfunction (Gabuzda et al., 1986). We thus sought to examine whether exposure of human primary astrocytes to HIV-1 Tat (as a surrogate of HIV-1 infection) could lead to upregulation of miR-7. As shown in Figure 2a, HIV Tat (100ng/ml) significantly upregulated the expression of miR-7 in human primary astrocytes. Of note, the upregulation (~3 fold) of miR-7 by Tat protein *in vitro* is much lower than that observed in brain tissues (~6 fold). This could be due to the addictive effect of Tat and Tat-ADEVs *in vivo* &/or due to cooperative changes in various CNS cells *in vivo*. Next, we sought to examine whether the Tat-stimulated astrocytes also upregulated the expression of miR-7 in EVs. To this end, we first isolated EVs from the conditioned media of human astrocyte A172 cells as well as rat and human primary astrocytes using the differential ultracentrifugation procedure previously reported by us (Yang et al., 2018a) (Hu et al., 2012; Hu et al., 2013), followed by characterization of ADEVs, using standard methodologies including transmission electron microscopy (TEM), western blotting for exosomal markers and nanosight analyses. As shown in Figure 2b, TEM images showed spherical-shaped EVs ranging in size from 40–100 nm in diameter. Additionally, immunoblotting of the ADEV lysates also revealed the presence of exosomal markers CD63 and Tsg101 (Figure 2c). ADEVs were also validated by ZetaView revealing a reasonable yield of EVs of the expected size [~100nm; Figure 2d]. In addition, we also sought to assess the expression of miR-7 in both control and HIV-1 Tat ADEVs by qPCR. Intriguingly, elevated miR-7 expression was also detected in ADEVs purified from conditioned media of astrocytes exposed to Tat protein (Figure 2e). The next step was to examine whether the miR-7 released in the EVs from Tat-stimulated astrocytes could actually be taken up by primary rat hippocampal neurons. For this EVs isolated from the conditioned media of control astrocytes were first loaded with Texas Red RNA followed by exposure of **primary rat hippocampal neurons (PRHNs)** to labeled EVs. As shown in Figure 2f, Texas Red labeling was detectable in neurons exposed to labeled ADEVs, thereby suggesting the uptake of ADEVs by neurons. In keeping with this data, PRHNs exposed to Tat-ADEVs isolated from rat primary astrocytes also demonstrated increased expression of cellular miR-7 compared with the neurons exposed to control-ADEVs or neurons not exposed to EVs (Figure 2g).

NLGN2 is a functional target of miR-7

Based on the Targetscan analysis NLGN2 is a predicted conserved target of miR-7 (Figure 3a). The next step, therefore, was to determine whether the transferred miR-7, once inside

the neurons, could directly bind to the 3'-untranslated region (3'UTR) of the target mRNA and inhibit its translation. For this HEK293 cells were transfected with an NLGN2-3'UTR luciferase reporter construct, wherein luciferase expression was regulated by the NLGN2 3'UTR, a miR-7 potential binding element (Figure 3a). We next sought to determine whether overexpressing miR-7 could downregulate luciferase activity in cells transfected with the NLGN2-3'UTR-luciferase constructs. For this, we first determined the transfection efficiency of HEK293 cells. As shown in Figures 3b and 3c, transfection of miR-7 resulted in >1000- fold increase in miR-7 without affecting cell viability. Co-transfection of HEK293 cells with both miR-7 and NLGN2-3'UTR-luciferase constructs resulted in a significant decrease in luciferase activity, thereby suggesting preferential binding of miR-7 with the 3'UTR of NLGN2. As a negative control, cells were also co-transfected with a construct containing mutations in the miR-7-binding region of the NLGN2 3'UTR, and as expected, in cells transfected with the NLGN2 3'UTR mutant, miR-7 failed to downregulate luciferase activity (Figure 3d). To validate the specificity of miR-7, we next exposed PRHNs to ADEVs isolated from rat primary astrocytes loaded with either control miRNA or miR-7 oligos, followed by assessing the expression of NLGN2 by western blotting. In PRHNs exposed to miR-7 loaded ADEVs, NLGN2 expression was downregulated compared with cells exposed to control ADEVs (Figure 3e). Intriguingly, neurons exposed to HIV Tat-ADEVs (A172 astrocytes) also demonstrated significant inhibition of NLGN2 expression compared with neurons exposed to control-ADEVs (Figure 3f).

miR-7 in Tat-ADEVs causes synaptic injury

Having demonstrated the transfer of EV-miR-7 from the astrocytes to neurons, the next step was to understand the functionality of miR-7 in neurons. PRHNs (20d *in vitro* (DIV20)) were exposed to EVs isolated from conditioned media of Tat-stimulated rat primary astrocytes for 24h. PRHNs were then subjected to synaptic density assessment by confocal microscopy imaging for both inhibitory (GAD65) and excitatory (vGlut1) synapses by immunostaining, followed by quantification. Intriguingly, neurons exposed to Tat-ADEVs demonstrated significant synaptic injury (increased loss of both inhibitory and excitatory synapses), compared with neurons exposed to control-ADEVs (Figures. 4a and 4b), with no change in cell viability (Figure 4c). Decreased expression of PSD95 in PRHNs exposed to Tat-ADEVs was further validated by western blotting (Figure 4d). Next, to determine the role of miR-7, PRHNs were exposed to ADEVs loaded with either control miR or miR-7 oligos followed by monitoring the expression of inhibitory synapse marker by immunostaining. As shown in Figures 4e and 4f, neurons exposed to miR-7 loaded ADEVs demonstrated a significant reduction of inhibitory synapses compared to neurons exposed to control-ADEVs. To further confirm that NLGN2 was the critical component in the Tat-ADEV-mediated synaptic loss, PRHNs were transfected with either NLGN2 overexpression construct or a control vector, followed by exposure to either control or Tat ADEVs for 24h. As shown in Figures 4g and 4h, Tat-ADEVs failed to decrease excitatory synapses in NLGN2 overexpressing neurons.

PDGF-CC restores the ADEV-miR-7-mediated synaptic injury

While the neuroprotective role of PDGF-CC in reversing neuronal apoptosis has been well documented by others (Tang et al., 2010) and us (Peng et al., 2012), whether this factor could also restore synaptic impairment has not been explored before. We thus rationalized

that exposure of neurons to PDGF-CC could lead to protection against ADEV-miR-7-mediated impairment of synaptic alterations. To verify this, PRHNs were pre-treated with PDGF-CC (20ng/ml) (Han et al., 2007; Donovan et al., 2013) for 24h followed by exposure of treated cells to ADEVs loaded with either control or miR-7 oligos. Twenty four hours later neurons were fixed and immunostained for synaptic markers followed by imaging by confocal microscopy to assess synaptic densities. As shown in Figures 5a and 5b, in the presence of PDGF-CC there was restoration of EV-miR-7-mediated decrease of inhibitory synapses. To examine the functional alterations induced by Tat-ADEVs followed by rescue with PDGF-CC, we also recorded frequencies of miniature excitatory postsynaptic currents (mEPSCs) as well as miniature inhibitory postsynaptic currents (mIPSCs) in PRHNs (DIV 19–21) exposed to either control-ADEVs or Tat-ADEVs or PDGF-CC or Tat-ADEVs plus PDGF-CC. As shown in Figures 5c–5f there was a decrease in the frequency of both mEPSCs and mIPSCs in PRHNs treated with Tat-ADEVs. In addition, and consistent with the data shown in Figures 5a and 5b, pre-treatment of PRHNs with PDGF-CC rescued the expression of both PSD95 and NLGN2 in the presence of either Tat-ADEVs (Figures 5g and 5h) or ADEV-miR-7 (Figures 5i and 5j).

Alteration of inhibitory and excitatory synapses and the levels of NLGN2 (inhibitory synaptic protein) in the brains of SIV/HIV-infected subjects

We further validated our findings in the archival brain tissues of both SIV-infected nonhuman primates as well as HIV-1-infected patients. Expression of inhibitory and excitatory synapses in the FC of macaque brains was examined by immunostaining the brain sections using antibodies specific for inhibitory postsynaptic marker Gephyrin and excitatory postsynaptic marker PSD95. As shown in Figures 6a and 6b, the numbers of both inhibitory and excitatory synaptic puncta were decreased in the FC of SIV-infected rhesus macaques compared with uninfected controls. Interestingly, there was also a decreased expression of NLGN2 in the FC of SIV-infected rhesus macaques compared with the uninfected controls (Figures 6c and 6d). In concordance with the macaque data, the numbers of both inhibitory and excitatory synaptic puncta were also found to be decreased in the FC of HIV-1-infected subjects compared with the uninfected controls (Figures 6e and 6f). Expression of NLGN2 was also significantly decreased in the FC of HIV-1-infected subjects compared with the uninfected controls (Figures 6g and 6h).

DISCUSSION

In the current study, we found that exposure of astrocytes to HIV-1 Tat-induced the expression and release of miR-7 in the ADEVs, which, upon being taken up by the neurons, resulted in the loss of both inhibitory and excitatory synapse via downregulation of neuronal NLGN2. We validated our findings in the brains of SIV-infected nonhuman primates as well as HIV-1-infected patients.

ADEVs are critical regulators of neuronal and microglial functions as well as peripheral immune responses (Hu et al., 2012; Dickens et al., 2017; Chaudhuri et al., 2018; Yang et al., 2018b). For example, Dickens *et al.* have demonstrated that ADEVs from IL-1 β stimulated astrocytes can rapidly cross the blood-brain barrier (BBB) and enter the peripheral

circulation and the liver, leading, in turn, to the regulation of proinflammatory cytokine expression via suppression of PPAR α , ultimately promoting the transmigration of leukocytes into the CNS (Dickens et al., 2017). Furthermore, ADEVs released from IL-1 β and TNF α stimulated astrocytes are found to be enriched with miRNAs, including, but not limited to, miR-125a-5p and miR-16-5p, which upon uptake by the neurons can downregulate neuronal NTKR3, ultimately resulting in reduction of dendritic growth, dendritic complexity, reduced spike rates and burst activity (Chaudhuri et al., 2018). We have previously reported that HIV-1 Tat mediates the induction and release of miR-9 in ADEVs, which upon uptake by the microglia, result in enhanced migration of these latter cells (Yang et al., 2018a). Herein, we expand our findings and demonstrate that exposure of astrocytes to HIV Tat resulted in increased expression and release of several miRNAs in the EV cargo, specifically the brain-enriched miR-7, which upon uptake by the hippocampal neurons, resulted in the loss of inhibitory synapses.

Synapses play a critical role in establishing and maintaining neural circuits thereby permitting targeted information transfer throughout the brain. A large portfolio of synaptic adhesion/organizing proteins (SAPs) exists in the mammalian brain that are involved in synapse development and maintenance. SAPs play critical roles in cell adhesion and in organizing protein interaction networks. Members of the NLGNs family of SAPs that are present on the postsynaptic membrane are vital regulators of synaptic structure, function, and remodeling (Sudhof, 2008; Li et al., 2017; Stogsdill et al., 2017) in the central neurons. SAPs maintain a tightrope balance of excitatory/inhibitory synapses in the brain (Durieux et al., 2016). Dysfunction of SAPs such as NLGNs is associated with several CNS diseases, including autism and epilepsy (Lionel et al., 2013; Nakanishi et al., 2017). While impairments in excitatory and inhibitory synapse balance are a hallmark of various neurodevelopmental and neuropsychiatric disorders and have been implicated in CNS complications associated with HIV infection and opiate abuse (Kim et al., 2008; Fitting et al., 2013; Fitting et al., 2014; Hahn et al., 2015; Gangwar et al., 2017; Xiao et al., 2017), the role of SAPs in the context of HIV-1 infection remains elusive. Interesting emerging studies have implicated astrocytes, one of the most abundant cell types in the CNS, as regulators of NLGNs (Rudenko, 2017). Previous findings by others (Rahimian and He, 2016a; Dickens et al., 2017) and us (Hu et al., 2012; Hu et al., 2018; Yang et al., 2018b) have demonstrated that EVs released from astrocytes are conduits for delivering cargo to donor cells, culminating into a broad spectrum of functional changes in the recipient cells. In the current study, we found that the expression of NLGN2 was significantly reduced in the archival brain tissues of HIV/SIV-infected subjects compared with uninfected controls. Furthermore, we demonstrated that HIV-1 Tat-mediated release of astrocytic EV-miR-7 downregulated neuronal NLGN2, which could be a functional link between the NLGN2 and cognitive dysfunction associated with HIV-1.

In the CNS, neuronal homeostasis is a delicate balance between neurotrophic vs. neurotoxic factors. Various factors such as BDNF, FGF, and GDNF have been implicated in neuronal protection against neurotoxins (Alzheimer and Werner, 2002; Almeida et al., 2005; Deierborg et al., 2008; Colafrancesco and Villoslada, 2011). Elegant studies have suggested downregulation of BDNF following HIV-1 infection (Bachis and Mocchetti, 2005; Mocchetti et al., 2007; Nosheny et al., 2007; Avdoshina et al., 2011; Bachis et al., 2012).

Findings from our lab have identified yet another factor, PDGF that plays a crucial role in reversing neuronal toxicity mediated by HIV-1 Tat and gp120 (Yao et al., 2009). We previously demonstrated that treatment of morphine-exposed neurons with PDGF-BB ameliorated morphine-mediated synaptic alterations in hippocampal neurons through inhibition of ER stress and autophagy pathways, which leads to the preservation of excitatory and inhibitory synapses (Cai et al., 2016). Our current data suggest that pre-treatment of Tat-ADEV-exposed neurons with PDGFCC ameliorated Tat-ADEV-mediated synaptic loss in the hippocampal neurons. We also demonstrated that pre-treatment of PRHNs with PDGF-CC rescued downregulated expression of both PSD95 and NLGN2 in Tat-ADEVs-stimulated PRHNs, thereby underpinning the role of PDGF-CC as a neuroprotective agent for HIV-1 Tat and opiate-mediated synaptodendritic injury.

Taken together, our results demonstrate that exposure of PRHNs to Tat-ADEVs resulted in decreased expression of neuronal NLGN2 which, in turn, resulted in the loss of both excitatory and inhibitory synaptic densities in the hippocampal neurons. Moreover, we also demonstrate a neuroprotective role of PDGF-CC in rescuing Tat-ADEV-mediated synaptic impairment. It can thus be envisioned that therapeutic strategies aimed at activating PDGF-CC signaling could be beneficial in reversing HIV-1-mediated cognitive decline.

MATERIALS AND METHODS

Animals

All animal procedures were performed in strict accordance with the protocols approved by the Institutional Animal Care and Use Committee at the University of Nebraska Medical Center and the National Institutes of Health. Sprague-Dawley rats and C57BL/6N mice were purchased from Charles River Laboratories (Wilmington, MA, USA) and housed under conditions of constant temperature and humidity on a 12 h light, 12 h dark cycle, with lights on at 0700 h. Food and water were available *ad libitum*. Pregnant rats were purchased from Charles River Laboratories.

Reagents and Brain Tissues

Endotoxin-free Recombinant HIV-1 Tat-101 (1032–10) was purchased from ImmunoDX, LLC (Woburn, MA). Recombinant human PDGF-CC Protein was purchased from R&D Systems (Minneapolis, MN, USA). miR-7 (UGGAAGACUAGUGAUUUUGUUGU) and negative control (ACCAUAUUGCGCGUAUAGUCGC) oligonucleotides were purchased from Integrated DNA Technologies Inc. (Coralville, IA).

HIV–, HIV+ patients' brain tissues were obtained from the UNMC PEN Brain Banking for Study of Neurologic Diseases (IRB: 056–00-FB). Archival macaque brain tissues were collected as described in our previous study (Bokhari et al., 2011).

Primary neuron cultures

All experimental procedures were approved by the University of Nebraska Medical Center Institutional Animal Care and Use Committee. Primary rat hippocampal neurons were obtained from E18 rat hippocampi as previously described (Arikath et al., 2009; Beaudoin

et al., 2012; Cai et al., 2016). In brief, after dissociation of the dissected brain hippocampi in HBSS (14170112; Invitrogen, Carlsbad, CA, USA) supplemented with 0.25% trypsin (LS003707; Worthington Biochemical Corporation; Lakewood, NJ, USA), primary neurons were plated at a density of 7.5×10^4 on poly-L-lysine-coated 18-mm glass coverslips. Cultures were maintained in Neurobasal medium (21103049; Invitrogen, Carlsbad, CA, USA) with B27 supplement (17504044; Invitrogen), Glutamax (3505061; Invitrogen, Carlsbad, CA, USA), and penicillin-streptomycin (15070063; Invitrogen, Carlsbad, CA, USA) at 37°C with 5% CO₂. Cytosine β-D-arabinofuranoside (1 μM; C1768; Sigma-Aldrich, St. Louis, MO, USA) was added on DIV 2, and all media were replaced on DIV 3. Half of the media was replaced twice every week. Transfection was performed on DIV 12 using Lipofectamine 2000 (11668027; Invitrogen, Carlsbad, CA, USA) as indicated by the manufacturer.

Cell culture and cell lines

The human astrocytic cell line A172 (no. CRL-1620; American Type Culture Collection (ATCC)) was cultured as described previously (Bethel-Brown et al., 2012) and maintained in Dulbecco's modified Eagle's medium (DMEM) with high glucose containing 10% heated-inactivated fetal bovine serum, 2 mM glutamine, penicillin (100 units/ml), streptomycin (100 μg/ml), essential amino acids and vitamins. In this study, A172 cells were used within 30 passages.

Rat primary astrocytes were prepared from whole brains of postnatal (1- to 3-day-old) rats and plated on poly-D-lysine pre-coated cell culture flasks containing DMEM (10% fetal bovine serum, 100 U/ml penicillin and 100 μg/ml streptomycin). The cells were grown in a humidified atmosphere of 5% CO₂/95% air at 37 °C. In this study, rat primary astrocytes were used from passage 2 to 5. Immunocytochemical analyses demonstrated that the cultures comprised of >95% GFAP-positive astrocytes.

Human primary astrocytes obtained from ScienCell Research Laboratories were cultured in astrocyte medium containing astrocyte growth supplement, 2% heat-inactivated FBS and 10 U/ml penicillin-streptomycin and incubated at 37°C in a 5% CO₂-humidified incubator. Human primary astrocytes were used under 10 passages according to the manufacturer's instructions. The purity of primary astrocytes was assessed by staining with GFAP. Only cells with a purity of greater than 95% were used in our studies.

EV Isolation

EVs were prepared from the supernatant of primary astrocytes and by differential centrifugations as previously described (Hu et al., 2012; Hu et al., 2013; Hu et al., 2018). Briefly, primary astrocytes were treated with HIV Tat protein for 24 hours. Then, conditioned media were harvested, centrifuged at 1,000 g for 10 min to eliminate cells, and again spun at 10,000 g for 30 min, followed by filtration through 0.22 μm filter to remove cell debris. EVs were pelleted by ultracentrifugation (Beckman Ti70 rotor, Brea, CA, USA) at 100,000g for 70 min. EVs were assessed for their protein content using BCA Protein Assay Kit (Pierce, Rockford, IL, USA). TSG101 and CD63 were detected by western blot as

exosome markers. EVs were further quantified using the ZetaView nanoparticle tracking analysis system (Particle Metrix, Mebane NC).

Cell and EV transfection

Plasmid transfections were performed using Lipofectamine 2000 (Life Technologies, catalog 11668027, Grand Island, NY, USA) according to the manufacturer's instructions. Briefly, cells were transfected with a plasmid (500ng) mixed with 2 μ l of Lipofectamine 2000 diluted in 100 μ l of opti-MEM (Life Technologies, catalog #31985062, Grand Island, NY, USA). The resulting plasmid-lipid complexes were added to the cells, incubated for 6 h, and the medium changed into fresh DMEM. Next, the medium was changed to 10% FBS-containing medium for 20 h incubation. The transfected cells were then ready for use in experiments. pNICE-NL2A was a gift from Peter Scheiffele (Addgene plasmid # 15259, Watertown, Massachusetts, USA) (Chih et al., 2006). The pCAGGs-IRES-eGFP plasmid has been previously described and validated (Arikkath et al., 2008). EVs were transfected with miRNA using Exo-Fect™ Exosome Transfection Reagent (SBI, System Biosciences, Palo Alto, CA, USA) according to the manufacturer's instructions.

MTT assay

Cell viability was measured using the MTT method as reported previously (Hu et al., 2012). Briefly, cells were seeded in a 96-well plate at a density of 5,000 cells per well for 24 h followed by transfection with either control or miR-7 oligoes for additional 24 h and incubated with 20 μ l MTT tetrazolium salt dissolved in Hank's balanced salt solution at a final concentration of 5 mg/ml added to each well and incubated in a CO₂ incubator for 1–4 h. Finally, the medium was aspirated from each well and 200 μ l of dimethyl sulfoxide was added to dissolve the formazan crystals, and the absorbance of each well was obtained using a plate counter at test and reference wavelengths of 570 and 630 nm, respectively.

Immunocytochemistry

For immunocytochemistry, primary rat hippocampal neurons were plated on coverslips treated with the respective agents for 24–48 h, followed by fixation with 4% PFA and 4% sucrose in PBS for 20 min at room temperature and permeabilization with 0.1% Triton X-100 (BP151–1; Thermo Fisher Scientific, Grand Island, NY, USA). Neurons were incubated in 5% normal goat serum in PBS buffer for 1 h at room temperature followed by addition of respective antibodies: PSD95 (1:500; catalog no. ab18258, lot no. RRID: AB_444362; Abcam, Cambridge, MA, USA), gephyrin (1:1,000; catalog number 147 011, lot number RRID: AB_88771; Synaptic Systems, Goettingen, Germany), vGlut1 (1:4,000; catalog no. AB5905, lot no. RRID: AB_23017501; EMD Millipore, Burlington, MA, USA), and GAD65 (1:100; catalog no. GAD-6, lot no. RRID: AB_528264; Developmental Studies Hybridoma Bank, Iowa City, IA, USA) overnight at 4°C. This was followed by the addition of the secondary Alexa Fluor 555 goat anti-rabbit (1:500; catalog no. A27039, lot no. RRID: AB_2536100; Thermo Fisher Scientific, Grand Island, NY, USA), Alexa Fluor 555 goat anti-mouse (1:500; catalog no. A-21424, lot no. RRID: AB_2535845; Thermo Fisher Scientific, Grand Island, NY, USA) or Alexa Fluor 488 goat anti-guinea pig (1:500; catalog no. A-11073, lot no. RRID: AB_2534117; Thermo Fisher Scientific, Grand Island, NY, USA), and Alexa Fluor 555 goat anti-rabbit (1:500; catalog no. A27039, lot no. RRID:

AB_2536100; Thermo Fisher Scientific, Grand Island, NY, USA), respectively. Cells were mounted with Prolong Gold antifade reagent with DAPI (P36935; Thermo Fisher Scientific, Grand Island, NY, USA).

Immunohistochemistry

Animals under anesthesia were perfused with chilled 4% PFA. Sections encompassing the entire hippocampus were sectioned at 12 μ m on a cryostat and incubated with a blocking buffer containing 5% normal goat serum and 0.2% Triton X-100 (Thermo Fisher Scientific, Grand Island, NY, USA) for 1 h at room temperature followed by addition of respective antibody pairs: vGlut1 (1:4,000; catalog no. AB5905, lot no. RRID: AB_2301751; EMD Millipore, Burlington, MA, USA) and GAD65 (1:100; catalog no. GAD-6, lot no. RRID: AB_528264; Developmental Studies Hybridoma Bank, Iowa City, IA, USA) and PSD95 (1:500; catalog no. ab18258, lot no. RRID: AB_444362; Abcam, Cambridge, MA, USA) and gephyrin (1:1,000; catalog no. 147 011, lot no. RRID: AB_88771; Synaptic Systems, Goettingen, Germany) and incubated overnight at 4°C. Next day, the sections were washed followed by incubation with Alexa Fluor 488 goat anti-guinea pig (1:500; catalog no. A-11073, lot no. RRID: AB_2534117; Thermo Fisher Scientific, Grand Island, NY, USA) and Alexa Fluor 555 goat anti-rabbit (1:500; catalog no. A27039, lot no. RRID: AB_2536100; Thermo Fisher Scientific, Grand Island, NY, USA) or Alexa Fluor 488 goat anti-rabbit (1:500; catalog no. A11008, lot no. RRID: AB_10563748; Thermo Fisher Scientific, Grand Island, NY, USA) and Alexa Fluor 555 goat anti-mouse (1:500; catalog no. A-21424, lot no. RRID: AB_2535845; Thermo Fisher Scientific, Grand Island, NY, USA), respectively, in 0.5% BSA and 0.2% Triton X-100 at room temperature for 2 h, followed by mounting with Prolong Gold antifade reagent (P36935; Thermo Fisher Scientific, Grand Island, NY, USA).

In situ hybridization and immunostaining

Macaque brain sections were deparaffinized and subject to antigen retrieval with PBS. Sections were pre-hybridized in hybridization buffer (50% formamide, 10 mM Tris-HCl, pH 8.0, 200 μ g/mL yeast tRNA, 1X Denhardt's solution, 600 mM NaCl, 0.25% SDS, 1 mM EDTA, 100 μ g/ml salmon sperm DNA) for 1h at 37°C in a humidified chamber. LNA modified miR-7, labeled at both the 5' and 3' ends with digoxigenin (Exiqon, Hilden, Germany), was diluted to final concentration of 2 pM in hybridization buffer, heated to 65°C for 5 min, and separately hybridized to the sections at 37°C overnight. The slides were then washed three times in 2XSSC and twice in 0.2XSSC at 42°C. They were then blocked with 1% BSA, 3% normal goat serum in 1XPBS for 1h at room temperature and incubated with anti-digoxigenin conjugated with horseradish peroxidase (1:200, Roche Diagnostics GmbH, Mannheim, Germany) and anti-GFAP(1:500, Sigma, St. Louis, MO, USA) antibodies overnight at 4°C. The slides were washed twice with 1XTBS and incubated with Alexa Fluor 488 goat anti-rabbit IgG (1:400, Invitrogen, Carlsbad, CA, USA) antibody for 1h at room temperature. This was followed by 3 times 1X TBS washes and signal amplification using TSA Cy5 kit (PerkinElmer, Waltham, MA, USA) according to the manufacturer's protocol. The slides were mounted in Prolong Gold anti-fade reagent with DAPI (Invitrogen, Carlsbad, CA, USA). The specificity of the miR-7 signal in FISH experiments was

confirmed by comparing with a scrambled control. Unlike the miR-7, the scramble probe showed no signal in astrocytes.

Microscopy acquisition and spine architecture quantitation

Imaging was performed on an inverted LSM 700 microscope with a 40× objective and a digital zoom of 4× (Carl Zeiss, Thornwood, NY, USA). Contrast and brightness of overall images were consistent in each set of experiments. For primary neurons in vitro, the number of synapses and dendritic spines were counted manually in Zen 2010 SP1 (Carl Zeiss, Thornwood, NY, USA) using a digital zoom of 4× with Z-stack projection. These measurements were obtained on one dendrite per neuron within 80 μm from the cell body. Data were obtained from four independent cultures. The numbers of synapses and dendritic spines are indicated on the bar graph. For in vivo, the number of synapses in the striatum lucidum and radiatum of the hippocampus was measured using Bitplane Imaris 7.6.5 software (RRID: SCR_007370; Imaris, Concord, MA, USA).

Electrophysiology

Whole-cell recordings were performed on primary hippocampal neurons (DIV 19–21) seeded on coverslips with an Axopatch 200B (Molecular Devices, Sunnyvale, CA, USA). Signal was filtered at 2 kHz and digitized at 10 kHz using an Axon Digidata 1440A analog-to-digital board (Molecular Devices, San Jose, CA, USA). Recordings with a pipette access resistance less than 20 mOhm and that of less than 20% changes during the duration of recording were included. The external solution contained: 150 mM NaCl, 3 mM KCl, 10 mM HEPES, 6 mM mannitol, 0.02 mM EDTA, 1.5 mM MgCl₂, and 2.5 mM CaCl₂ (pH 7.4). Glass pipettes with a resistance of 3–5 mOhm were filled with an internal solution consisting of 110 mM cesium gluconate, 30 mM CsCl, 5 mM HEPES, 4 mM NaCl, 0.5 mM CaCl₂, 2 mM MgCl₂, 5 mM BAPTA, 2 mM Na₂ATP, and 0.3 mM Na₂GTP (pH 7.35). 5 mM QX-314 was added in pipette solution to block voltage-gated sodium channels from recorded cell. Miniature excitatory postsynaptic currents (mEPSCs) were recorded at –70 mV in the presence of 0.5 μM tetrodotoxin and 100 μM picrotoxin. Miniature inhibitory postsynaptic currents (mIPSCs) were recorded at 0 mV in the presence of 0.5 μM tetrodotoxin, 10 μM CNQX, and 100 μM DL-AP5. The mEPSC and mIPSC recordings were analyzed using MiniAnalysis software (SynapseSoft, Atlanta, GA, USA) with an amplitude threshold set at 5 pA. Frequency of the miniature currents was measured.

Western blotting

Treated cells or tissue were lysed using the Mammalian Cell Lysis kit (Sigma, St. Louis, MO, USA) and quantified using the BCA Protein Assay kit (Pierce, Rockford, IL, USA). Equal amounts of the corresponding proteins were electrophoresed in a sodium dodecyl sulfatepolyacrylamide gel (10–12%) under reducing conditions followed by transfer to PVDF membranes. The blots were blocked with 5% non-fat dry milk in phosphate buffered saline. Western blots were then probed with antibodies recognizing the CD63 antibody (1:1000; Abcam, Cambridge, MA, USA; ab216130), TSG101 (1:1000; Abcam, Cambridge, MA, USA; ab125011), and β-actin (1:4000; Sigma, St. Louis, MO, USA; A5316). The secondary antibodies were alkaline phosphatase conjugated to goat anti-mouse/rabbit IgG (1:5000). Signals were detected by chemiluminescence and imaged on the FLA-5100

(Fujifilm, Valhalla, NY, USA) digital image scanner; densitometry was performed utilizing Image J software (NIH) (Schneider et al., 2012).

RNA isolation and TaqMan™ MicroRNA Assays

RNA was purified from homogenized tissue, cells or EVs using the Quick-RNATM MiniPrep Kit (ZYMO Research, CA, USA) according to the manufacturers' instructions followed by RNA elution in 50 µL of RNase/DNA-free water. The column purified total RNA was used to perform the TaqMan™ MicroRNA Assays (Applied Biosystems, CA, USA) as reported earlier (Periyasamy et al., 2018).

TaqMan® MicroRNA Reverse Transcription Kit (PN 4366596) was utilized according to the provided protocol. The reaction system (15 µL) was set as follows: 10 mM dNTPs (with dTTP) 1.5 µL, MultiScribe™ Reverse Transcriptase 50 U/µL 1.0 µL, 10X Reverse Transcription Buffer 1.5 µL, RNase Inhibitor, 20 U/µL 0.2 µL, nuclease-free water to 7 µL, total RNA 5 µL (~10 ng), and 3 µL of RT (5X) primer. The tube containing the reaction mixture was centrifuged and loaded onto the thermal cycler: 16 °C for 30 min, 42 °C for 30 min, and 85 °C for 5 min. The reverse transcription product was then diluted 1:10 for the following PCR reaction: TaqMan® PCR primer (20X) 1 µL, RT reaction product 1.5 µL, TaqMan® 2X Universal PCR Master Mix, No AmpErase UNGa 10 µL (PN 4324018), and distilled water up to 20 µL. The PCR conditions were as follows: hold 95 °C for 10 min, then 40 cycles for 95 °C for 15 s and 60 °C for 1 min. All reactions were run in triplicate. The expression levels of miR-7 were calculated by normalizing to U6 snRNA as reported previously (Hu et al., 2012; Yao et al., 2014; Hu et al., 2017).

MiRNA Arrays

Total RNA was isolated from brain specimens as described above, followed by miRNA array analysis on the LC Biosciences platform (Rhesus monkey miRNA Array, Version No.: miRRhesusMonkey_21).

Electron microscopy (EM)

EV pellets were prepared for negative staining and immunogold double-labeling employing a slightly modified procedure. Using wide-bore tips, 3 µL of EV pellet was gently placed on 200-mesh formvar-coated copper grids, allowed to adsorb for 4–5 min, and processed for standard uranyl acetate staining. In the last step, the grid was washed with three changes of PBS and allowed to semi-dry at room temperature before observation in TEM (Hitachi H7500 TEM, Tokyo, Japan).

Luciferase activity assays

A 37 bp NLGN2 3'UTR segment (sense 5' - TCGAGGCGCCGCTAGGGGTGGGGTGCTGTCTTCCAT -3' and antisense 3' - CCGCCGGCGATCCCCACCCACGACAGAAGGTAGATC-5') containing the putative miR-7 target site was cloned into the *XhoI* and *XbaI* sites of the pmirGLO vector. For pmirGLO-NLGN2 3'UTR-miR-7-target-mutant segment (sense 5' - TCGAGGCGCCGCTAGGGGTGGGGTGCTCAGAAGGAT-3' and antisense 3' - CTAGATCCTTCTGACACCCACCCCTAGCGGCCGCC-5'), the miR-7 target site

(GTCTTCC) within the NLGN2–3'UTR was changed to (CAGAAGG). Followed the manufacturer's protocol (Promega), HEK293 cells were seeded in 24-well plates. MiR-7 or scramble miRNA-control were co-transfected with pmirGLO-NLGN2–3'UTR-miR-7-target or pmirGLO-NLGN2–3'UTR-miR-7-target-mutant luciferase reporter vector by using lipofectamine 3000 (Invitrogen). Two days post transfection, luciferase activity was assessed using the Dual-Luciferase Reporter Assay (Promega). Firefly luciferase activity was normalized by renilla luciferase activity and expressed as a percentage of the control (Triplicated independent experiment, performed in 3 wells each time).

Statistical analysis

All data are presented as mean \pm SD and were analyzed using the Student's *t*-test or analysis of variance (ANOVA) model with post hoc test to compare means between each group. All data were graphed, and statistical analyses were performed using Prism 6 (GraphPad Software, La Jolla, CA, USA). Results were considered statistically significant if probability levels were <0.05 .

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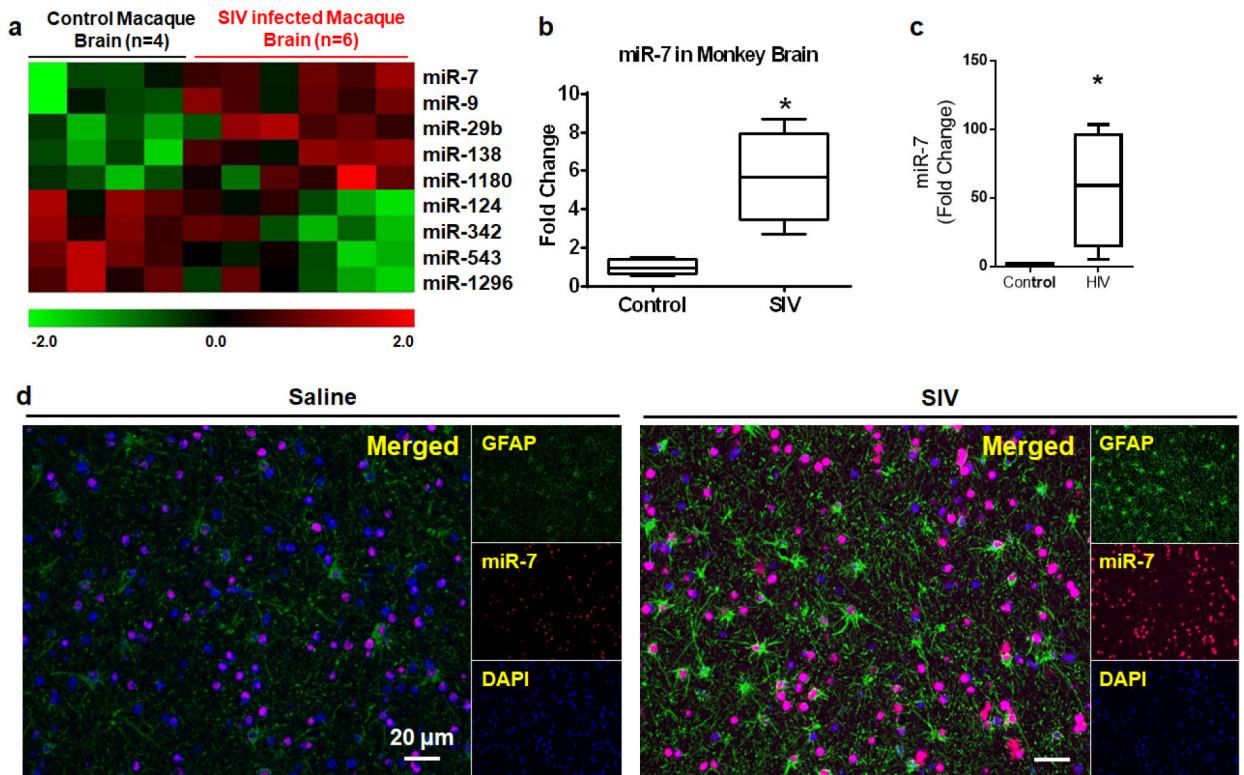


Figure 1. Upregulation of miR-7 in the SIV+/HIV+ brains.

(a) Unbiased heatmap correlating the expression of various miRNAs in the BG of SIV+ macaques, with colors indicating relative expression levels. Real-time PCR analysis of miR-7 expression in the brains of uninfected and SIV+ (b) or HIV+ (c) subjects. (d) *In situ* hybridization demonstrating increased expression of miR-7 in the astrocytes of SIV-infected macaques. * $p < 0.05$ vs control.

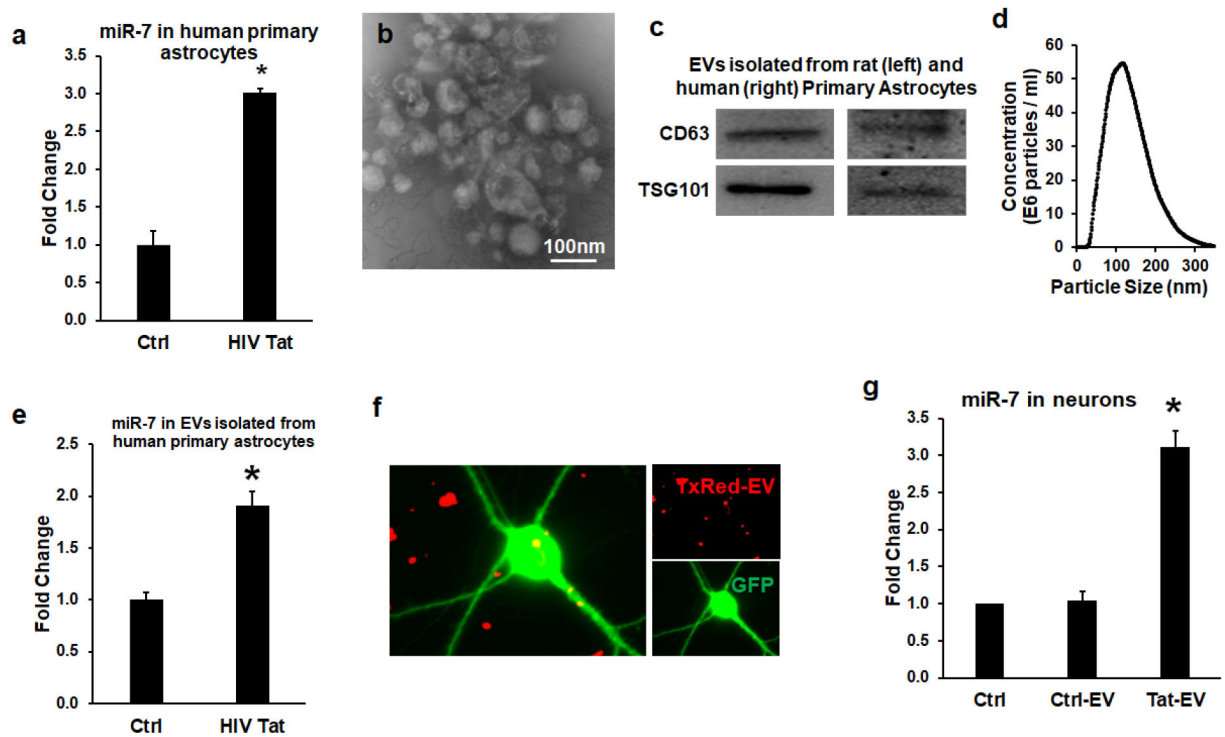


Figure 2. miR-7 was upregulated in the EVs released from HIV-1 Tat-stimulated astrocytes.

(a) Real-time PCR analysis of miR-7 expression in human primary astrocytes stimulated with HIV-1 Tat for 24 h. (b) Western blot characterization of astrocyte EVs. Protein isolated from astrocyte EVs was separated on SDS-PAGE and electroblotted onto nitrocellulose membrane. Blots were probed with exosome marker antibody against TSG101 and CD63. (c) Electron micrograph of EVs isolated from A172 cells. Scale bar=100nm. (d) Size and particle distribution plots of isolated EVs from cell culture by Nanosight Tracking Analysis (NTA). (e) Real-time PCR analysis of miR-7 expression in EVs isolated from HIV-1 Tat stimulated human primary astrocytes for 24 h. (f) Confocal images of rat primary neurons cultured with TxRed labeled ADEVs. Scale bar = 5 μm. (g) Expression levels of miR-7 in untreated (control), EVs from control astrocytes (control-EV), and Tat-stimulated astrocytes (Tat-EV) treated rat primary neurons were measured by qPCR. Bars represent mean ± SD from 3 independent experiments. *p < 0.05 vs control..

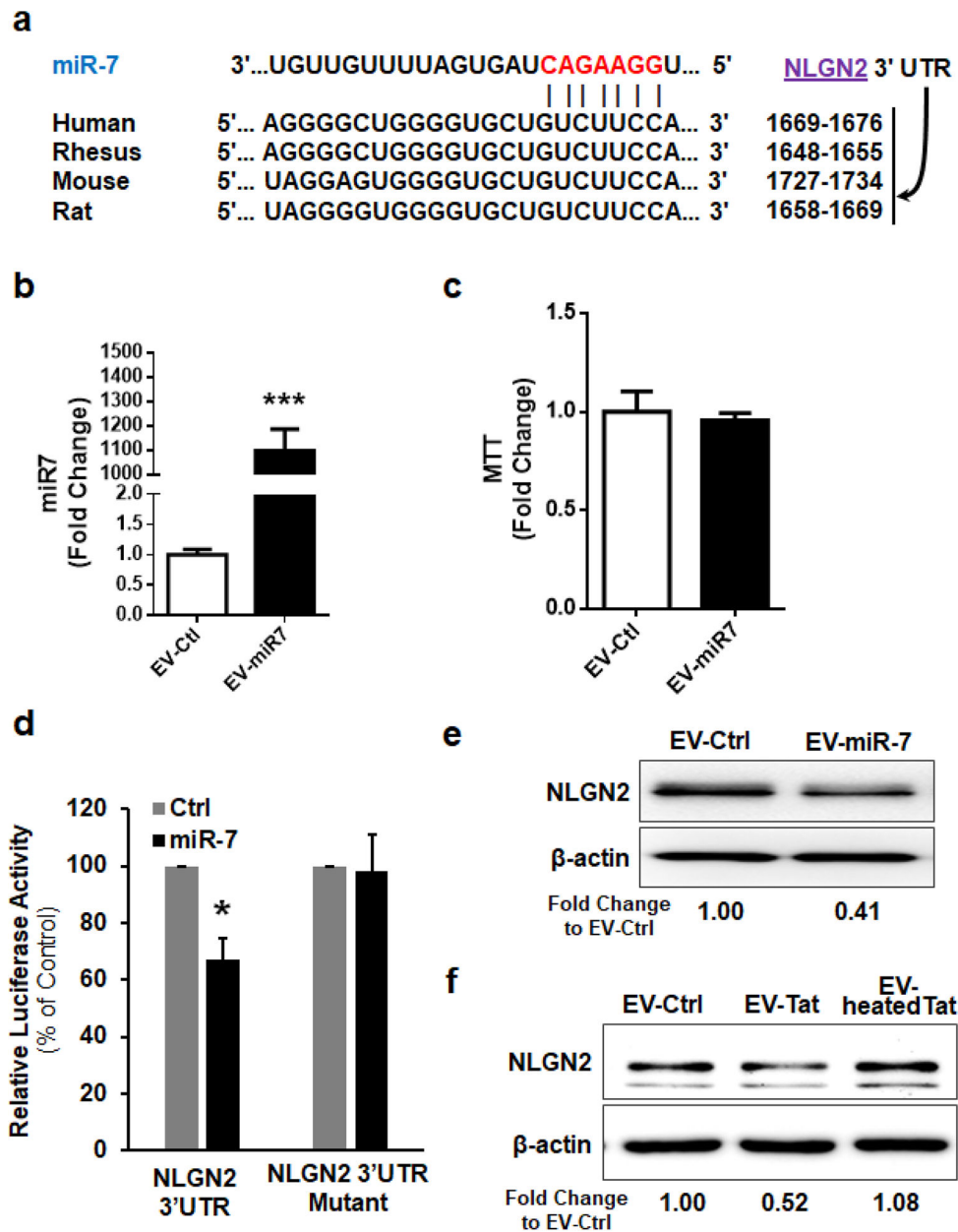


Figure 3. EV-miR-7 targets neuronal NLGN2.

(a) Putative miR-7 binding site in NLGN2 3'UTR. (b) HEK293 cells were transfected with 200 nM mature miR-7 oligos using Lipofectamine 2000 for 24 h. Transfection efficiency was determined using qPCR and was normalized to U6 small nuclear RNA (U6). (c) Cell viability was measured using MTT assay. (d) Relative luciferase activity of WT/3'UTR mutant constructs of NLGN2 co-transfected with miR-control/miR-7. (d, e) Western blot analysis of NLGN2 protein levels in rat primary neurons treated with EVs (100 EVs per cell) isolated from astrocytes loaded with control or miR-7 oligo (2.5 pmol per well) (e) and EVs isolated from astrocytes under indicated conditions (f). *p < 0.05 vs control.

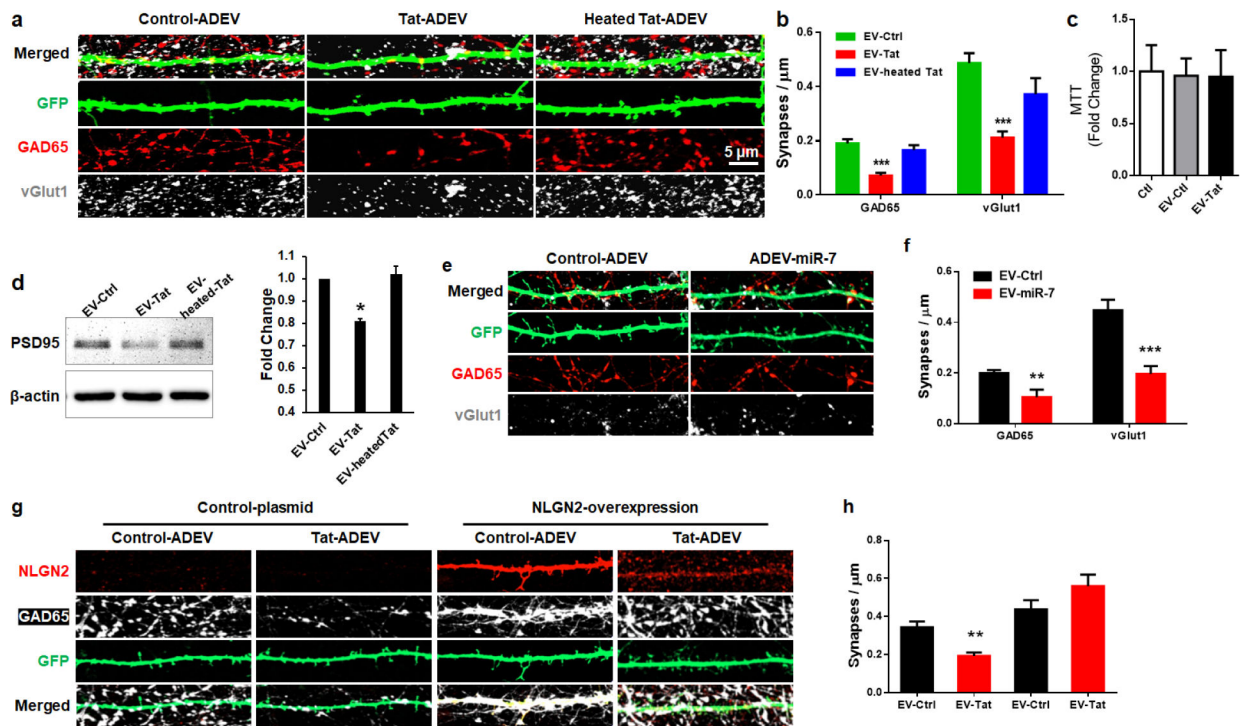


Figure 4. miR-7 in Tat-ADEVs causes synaptic injury.

(a) Representative confocal images of GFP-expressing PRHNs exposed to indicated ADEVs (24h), stained with GAD65 and vGlut1 & (b) quantification of excitatory & inhibitory synapses. (c) Cell viability was measured using MTT assay. (d) Representative western blot image and quantification of PSD95 in PRHNs exposed to indicated ADEVs. (e) Representative confocal images of GFP-expressing PRHNs exposed to ADEVs loaded with either control or miR-7 oligos (24h) & stained with GAD65 & vGlut1 & (f) quantification of excitatory and inhibitory synapses. (g) Representative confocal images of GFP and NLGN2-expressing PRHNs exposed to the indicated ADEVs (24h) & stained with NLGN2 & PSD95 & (h) quantification of puncta for each condition. One way ANOVA with post hoc test. Bars represent mean \pm SD from 3 independent experiments. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$ vs control; # $p < 0.05$ vs Tat-ADEV group.

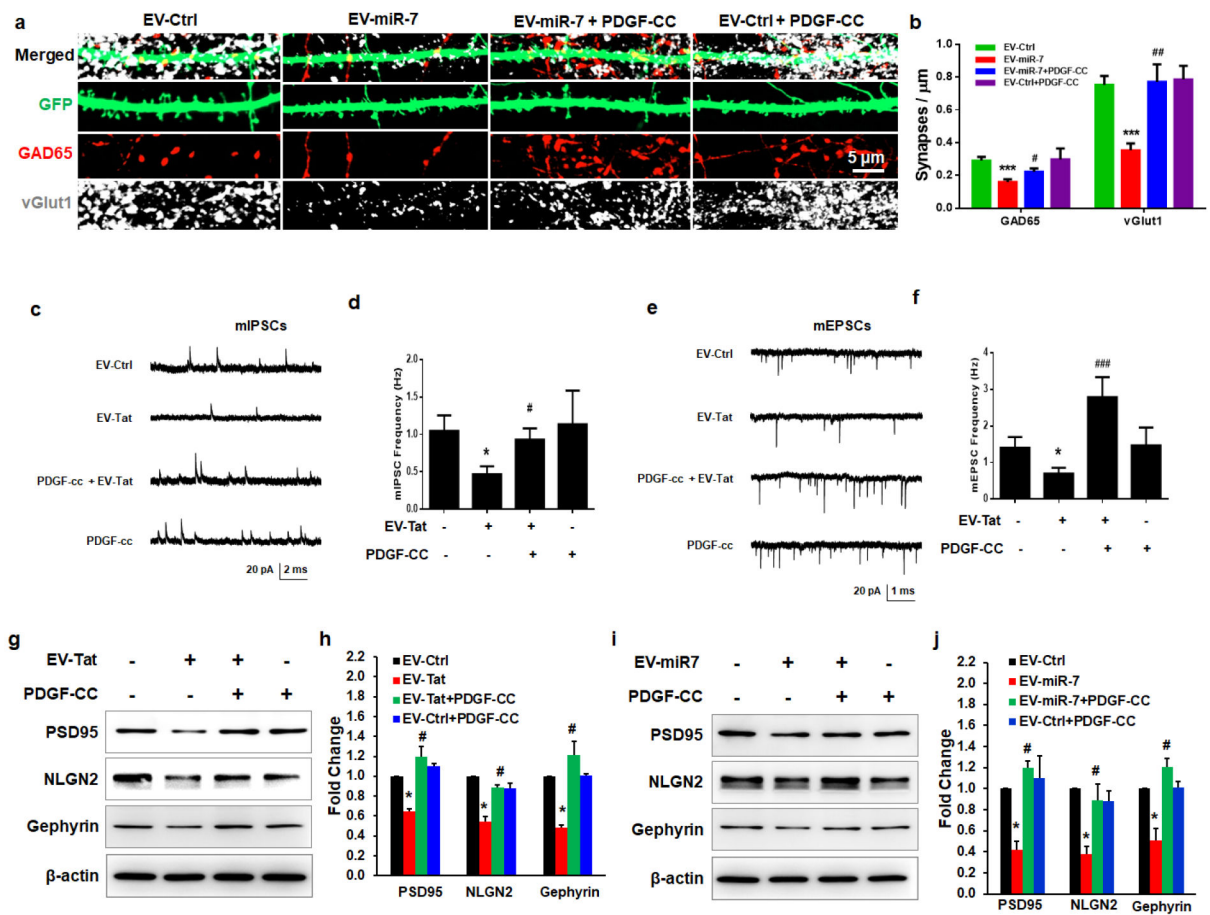


Figure 5. Protective effect of PDGF-CC in EV-miR-7-mediated synaptic alterations.

Representative confocal images of GFP-expressing primary rat hippocampal neurons exposed to EV-miR-7 followed by treatment with PDGF-CC (20 ng/ml) and stained with vGlut1 & GAD65 (a) and quantification of excitatory & inhibitory synapses (b).

Representative traces of whole-cell voltage-clamp recording showing mEPSC (c), mIPSC (e), mean mEPSC frequencies (d), and mean frequencies of mIPSCs (Hz; f) in primary rat neurons (DIV 19–21) treated with combinations of ADEVs and PDGF-CC as indicated. (g, i) Representative western blot (h, j) and quantification of PSD95, NLGN2, and Gephyrin in the cell lysates of neurons exposed to ADEVs followed by treatment with PDGF-CC (20 ng/ml) for an additional 24 h. One way ANOVA with post hoc test. Bars represent mean \pm SD from 3 independent experiments. * $p < 0.05$; ** $p < 0.01$ vs control; # $p < 0.05$, ## $p < 0.01$ versus treated group.

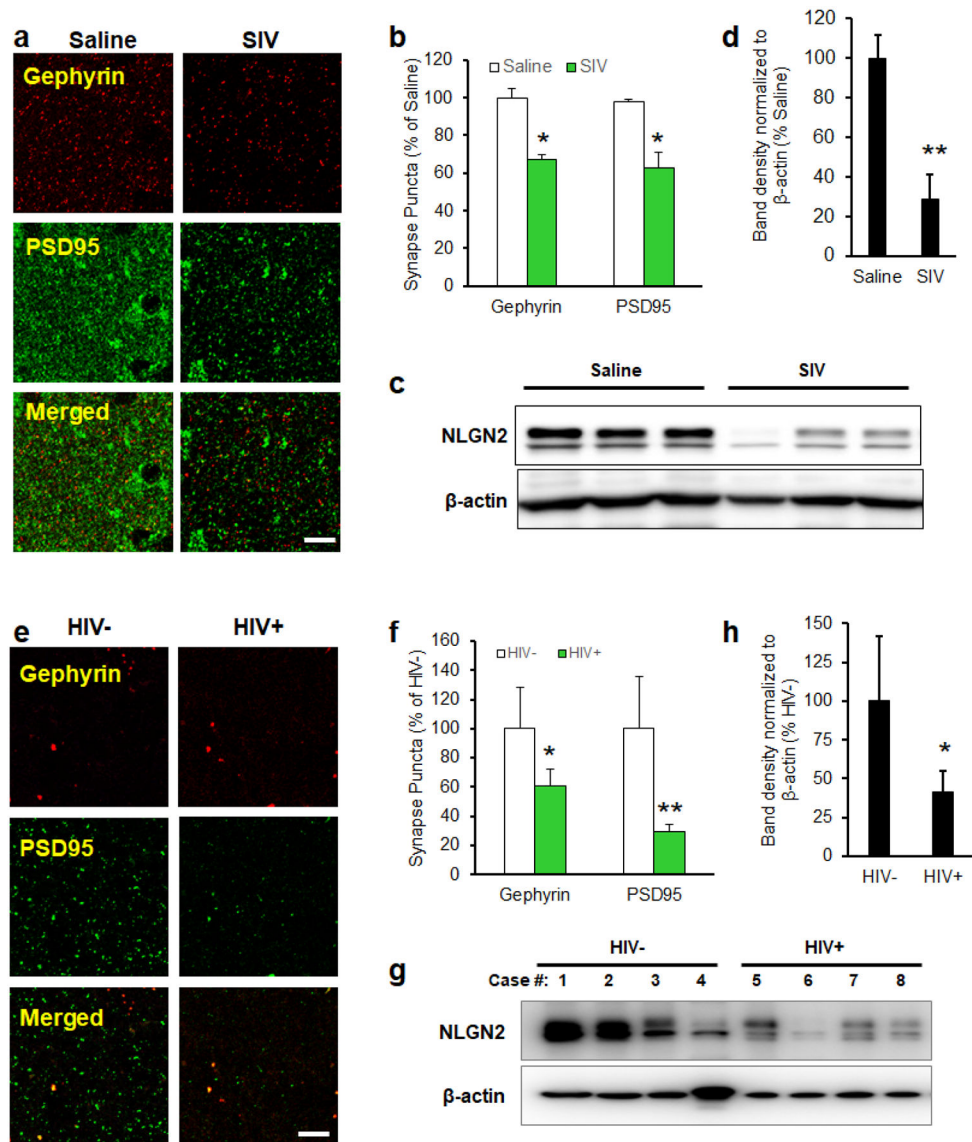


Figure 6. Alteration of inhibitory and excitatory synapses and the levels of NLGN2 in the brains of SIV/HIV-infected subjects.

(a, d) Representative confocal images of the frontal cortex regions of SIV/HIV-infected subjects were stained with Gephyrin (inhibitory postsynaptic marker) and PSD95 (excitatory postsynaptic marker). (b, e) Quantification of Gephyrin and PSD95 puncta. (c, d and g, h) Expression level of NLGN2 is significantly reduced in the brains of SIV/HIV-infected subjects. Bars represent mean \pm SD from 3 independent experiments. * $p < 0.05$; ** $p < 0.01$ vs. uninfected controls.

Table 1:

Clinical characteristics of the patients included in this study

Case Number	HIV Infection	Age	Gender	Race	Neuropathology	Other Pathologies
1(99-112)	Negative	47	Female	Caucasian	N/A	Synovial sarcoma and fibrinous pericarditis
2(99-113)	Negative	66	Male	Caucasian	N/A	Massive pulmonary consolidation secondary to diffuse alveolar damage (ARDS). Cryptogenic cirrhosis complicated with portal hn, upper GI bleed due to ulcers, hepatic encephalopathy, corpulmonate, chronic renal failure.
3(99-114)	Negative	58	Female	Caucasian	N/A	End-stage liver disease secondary to non-alcoholic steatohepatitis
4(99-132)	Negative	55	Male	Caucasian	N/A	Sudden Cardiac death in a setting of moderate-severe coronary artery disease
5(01-126)	HIV+	44	Male	Caucasian	Dementia	Brain: microglial activation, microglial nodules, and astrocytosis. Microglial nodules found in striatum, inferior temporoal lobe & midbrain. Widespread gliosis & microglial activation. No multinucleated giant cells. No other infectious processes, with exception of microglial nodules. Pt. Has 10 yrs hx of AIDS. No clinical dementia.
6(96-100)	HIV+	55	Male	Caucasian	N/A	Cortical brain thrombosis. Intracerebral hemorrh. And necrosis. AIDS
7(96-102)	HIV+	35	Male	Caucasian	N/A	Atypical leptomeningeal lymphocytosis, consistent with non-hodgkins lymphoma, small cell type. Cause: AIDS
8(96-103)	HIV+	54	Male	Caucasian	N/A	Cause of death is AIDS. No pathologic alterations found in brain