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Synthetic Lethality in Ovarian Cancer

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Abstract

Ovarian cancers include several distinct malignancies which differ with respect to clinicopathological features and prognosis. High-grade serous cancer is the most common histological subtype and accounts for most ovarian cancer deaths. HGSOC is treated with surgery and platinum-based chemotherapy, but most patients relapse and succumb to chemoresistant disease. The genetic concept of synthetic lethality, in which the synergy of mutations in multiple genes results in cell death, provides a framework to design novel therapeutic approaches to overcome chemoresistance in ovarian cancer. Recent progress in understanding the genomic architecture and hereditary drivers of ovarian cancer has shown potential for synthetic lethality strategies designed around homologous DNA repair. Clinical trials have validated high response rates for poly (ADP-ribose) polymerase (PARP) inhibitors in patients with *BRCA1* or *BRCA2* mutations. Here we discuss the biological rationale behind targeting BRCA-PARP synthetic lethality based on genetic context in ovarian cancer and how this approach is being assessed in the clinic. Applying the concept of synthetic lethality to target non-BRCA-mutant cancers is an ongoing challenge, and we discuss novel approaches to target ovarian cancer using synthetic lethality in combination with and beyond PARP inhibitors. This review will also describe obstacles for synthetic lethality in ovarian cancer and new opportunities to develop potent targeted drugs for ovarian cancer patients.

Keywords

Ovarian cancer; Synthetic lethality; PARP inhibitors

Introduction

In 2021, approximately 21,410 American women will receive a new diagnosis of ovarian cancer and about 13,770 women will die from the disease, according to estimates from the American Cancer Society (1). Although, ovarian cancers account for only 2.5% of all

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malignancies among females, most women present at initial diagnosis with advanced stage (corresponding to stages III and IV) where 5-year survival rates range 26–42%

Epithelial ovarian carcinoma (EOC) is a heterogeneous disease classified into several major morphological subtypes: serous carcinomas (SC), endometrioid carcinomas (EC), mucinous carcinomas (MC), and clear-cell carcinomas (CCC), with distinct epidemiology, etiology, morphology, and prognosis (2). Diverse genetic and epigenetic alterations distinguish tumorigenesis and disease progression among the various subtypes of EOC. High-grade serous ovarian cancers (HGSC) are the most common and aggressive major EOC subtype, accounting for 70–80% of all cases (3). According to recent studies, the majority of HGSCs arise from abnormal fallopian tube epithelial (FTE) cells that undergo early genetic and epigenetic changes which induce migration to and colonization of the ovary, usually eventually manifesting as an ovarian tumor (4). Despite tumor heterogeneity across subtypes and within single tumors, insights into the molecular biology of EOC has produced new drug targeting strategies in recent years, especially for the most common serous subtype.

Standard treatment for newly diagnosed advanced ovarian cancer incorporates cytoreductive surgery and platinum-based chemotherapy, usually combined with a taxane, and with or without concurrent and maintenance bevacizumab, a clinically approved humanized monoclonal antibody against vascular endothelial growth factor (VEGF) (5). The assessment and integration of predictive biomarkers to achieve optimal debulking with no residual disease (R0 resection) continues to need extensive research, also indicating that it is essential to consider tumor biology when determining choice and sequence of surgery/systemic therapy. Patients with HGSOC respond very well to initial chemotherapy, with several randomized clinical trials evaluating dose, dose density, choice of platinum and/or a taxane, schedule, route of administration, and sequence of chemotherapy agents (6). Bevacizumab has been used as an adjunct to standard chemotherapy for first-line treatment of advanced ovarian cancer and has also been utilized in second-line regimens for platinum-sensitive and platinum-resistant disease (7). While most patients respond to this treatment paradigm, advanced disease cases typically relapse after a chemotherapy-free interval ranging from 10 to 26 months, whereupon development of chemoresistant tumors is a concern. While clinical trials of molecular targeted agents have mostly focused on treatment of recurrent disease, recent trials have increasingly looked at a role for incorporating precision oncology into upfront therapy as well. (8–11). This approach uses small-molecule inhibitors to target ‘oncogene addictions’ or oncogenic pathways integral to the survival of tumor cells to prevent the outgrowth of resistant clones to cytotoxic chemotherapy.

Over the past decade, PARP inhibitors (PARPi) have led the way in new targeted approaches in the treatment of ovarian cancer. Poly(ADP ribose) polymerase (PARP) 1 and 2 enzymes play critical roles in the repair of single-strand DNA breaks (SSB) and maintenance of genomic integrity predominantly through the base excision repair (BER) pathway (12). PARPi block this DNA repair pathway by taking advantage of concomitant defects in homologous recombination repair (HRR), a pathway responsible for the high-fidelity repair of double-strand breaks (DSB) in DNA. By binding to and trapping PARP1 and PARP2 on DNA at the sites of SSBs, PARPi produce persistent SSB and collapsed replication forks, which then turn into unresolvable DSB and cause cell death. Initially, PARP

inhibitors were presumed to function exclusively by inhibiting the catalytic activity of PARP1/PARP2. Later, the activity of PARP inhibitors in locking on to damaged DNA and “PARP-trapping” thereby providing an additional mechanism to catalytic inhibition that resulted in cytotoxicity was described. Select inhibitors were subsequently demonstrated to trap PARP1 and PARP2 on sites of DNA damage by means of a toxic allosteric effect (13,14). Trapped and stabilized PARP-DNA complexes prevent DNA repair and allow degeneration of stalled replication forks into double-stranded DNA breaks, thereby killing cancer cells more effectively than catalytic inhibition. However, not all clinical-stage PARP inhibitors have equivalent PARP-trapping activity despite displaying identical capacities to inhibit PARP catalytic activity. The ability to trap PARP closely correlates with each drug’s ability to kill cancer cells. Clinical PARP inhibitors can be ranked by their ability to trap PARP which parallels their cytotoxic potency (from the most to the least potent): talazoparib >> niraparib > olaparib = rucaparib >> veliparib (13,14).

Breast related cancer antigen (BRCA) proteins BRCA1 and BRCA2 are essential to the homologous recombination (HR) pathway and inheritance of one defective copy of either of these genes predisposes individuals to breast and ovarian cancers (15). In HR-deficient cells, PARP inhibition leads to termination of two functional DNA repair pathways (HRR and BER) resulting in a reliance on the error-prone non-homologous end joining (NHEJ) for DNA repair, accumulation of genetic damage, and ultimately cell death. Approximately 15–30% of women with ovarian cancer carry a germline or somatic BRCA mutation, and PARPi have shown benefit among these populations in both the first and second-line maintenance settings and treatment of recurrence (16). With our improving understanding of the HR pathway and its role in gynecologic malignancies, the potential applications of PARPi continue to expand. Currently, three FDA-approved PARPi are used in women with ovarian cancer: olaparib, rucaparib, and niraparib. Multiple completed and ongoing studies employing other PARPi such as veliparib and talazoparib have also shown encouraging results in clinical trials ([NCT01472783](#), [NCT02470585](#), [NCT01540565](#), [NCT01286987](#)). The clinical success of PARP inhibitors shows the potential of “synthetic lethality,” i.e. when concurrent perturbation of two genes produces a lethal outcome while perturbation of either single gene results in viable outcomes (17). Identification and characterization of robust synthetic lethal gene pairs is fundamental to exploiting synthetic lethality in cancer treatment. This review summarizes the relevance and ramifications of synthetic lethality as an engine for ovarian cancer drug target discovery and discusses potential applications for exploiting this concept for personalized medicine.

Origins of Synthetic Lethality

In 1922, Calvin Bridges made an interesting observation that simultaneous mutation of a pair of genes in a fruit fly would result in death, but mutation of either gene alone did not significantly affect survival (18). Two decades later, a similar phenomenon was confirmed by Theodore Dobzhansky (19) and John Lucchesi, also in *Drosophila* (20). Together these findings contributed to the evolution of the term synthetic lethality, prompting genetic studies in yeast to understand the mechanism. In the context of cancer, a genetic alteration, such as a defect in a tumor suppressor gene, causes a second gene to be indispensable for tumor cell viability; therefore, inhibition of the second gene product should be lethal

to tumor cells without affecting healthy and non-malignant cells (Figure 1). A synthetic lethal combination that includes a cancer-specific mutation is occasionally referred to as 'non-oncogene addiction' since the mutant tumor cell requires functionality of the synthetic lethal partner gene for survival (21).

Genetic Context in Ovarian Cancer Therapy: Advent of PARP Inhibitors

Ovarian cancer presents several challenges to precision oncology approaches to therapy. As noted, patients usually present with late-stage disease, and there is great heterogeneity both within and among tumor types. The disease is characterized by recurrent copy number alterations, low frequency of somatic oncogenic mutations, and highly complex genomic profiles (4,22). In addition, rather than an orderly progression from localized to systemic disease, advanced ovarian cancer also propagates via a multitude of independent "shedding" events within the peritoneal cavity (23). This heterogeneity leads to a sporadic distribution of genetically unique subpopulations across and within the tumor (spatial heterogeneity) as well as temporal alterations in the molecular architecture of cancer cells (temporal heterogeneity). For these reasons, establishing common features of the ovarian cancer genomic landscape has been essential for designing widely applicable treatment strategies.

Genomic instability is one of the common themes among high grade serous ovarian cancers. Genomic instability in HGSOC drives the development of further variants and increases development of genetically diverse subclones within the tumor. Genomic instability is associated with treatment resistance and poor prognosis if subclones develop pro-survival characteristics (24). A mutator phenotype in cancer cells refers to mutations in genes that function to maintain genomic stability and result in accumulating unrepaired DNA damage (25). Higher levels of genomic instability are actually linked to increased response rates to platinum-based chemotherapy and PARPi and improved survival outcomes. Cells attempt to repair damaged DNA and single-strand breaks (SSB) through several mechanisms including BER, nucleotide excision repair, mismatch repair, NHEJ, and HR pathways (26), with NHEJ and HR mechanisms targeting and repairing DSBs. NHEJ causes binding proteins to attach to open ends of the DNA resulting in stabilization and induces rejoining, albeit without regard for the reading frame (27), making it error-prone. SSB repair requires the engagement of several proteins, including polyadenosine diphosphate [ADP] ribose polymerase-1 (PARP1) and -2 (PARP2) that catalyze the addition of poly (ADP)-ribose from nicotinamide adenine dinucleotide (NAD⁺) molecules to proteins and to PARP1/2 themselves. PARP1 is responsible for the majority of PARylation in malignant and non-malignant cells which is critical to several cellular functions including DNA repair and chromatin regulation (28). In 2011, comprehensive analysis performed by The Cancer Genome Atlas (TCGA) consortium revealed that the *BRCA1/2* genes play a role in up to 30% of HGSOCs, irrespective of germline status (29). Further analysis of DNA damage-repair mechanisms revealed at least one mutation in genes related to HR pathways, exhibiting a "mutator phenotype" in nearly half of the tested tumors suggesting these tumors might also benefit from PARPi as a potential therapy.

Current genomic stratification of ovarian tumors reveals that about 20% of HGSOCs possess a germline or somatic mutation in *BRCA1/2* with smaller contributions from mutations

or epigenetic silencing of other genes related to HR pathways (22). BRCA1 is known to play an important role in DSB repair via the HR pathway; BRCA1-deficient cells display impaired HR and possess an inability to repair defective chromosomes (30). In a similar manner, BRCA2 has also been implicated in HR through formation of complexes with RAD51 to provide genomic stability. Defective HR repair mechanisms, arising from deficiencies in key repair proteins such as BRCA1/2, RAD51, and CHK1, cause cells to be highly reliant on PARP activity and therefore highly sensitive to PARP inhibition (31). PARP inhibition compromises SSB repair and BER, and, in cells lacking intact HR mechanisms (e.g., *BRCA1* and *BRCA2* mutants), these amplify into DSBs and persistent DNA lesions, resulting in cell death. BRCA1 or BRCA2-deficient cells are, respectively, 57- and 133-fold more sensitive to PARP inhibition than wild-type cells (32). Thus, HR-deficient cells can be targeted with PARPi to further impair DNA repair mechanisms, resulting in synthetic lethality (Figure 1). PARPi were originally proposed to function as chemosensitizers in combination with other agents; however, the level of BRCA selectivity attained with PARPi alone in pre-clinical studies (BRCA-mutant tumor cells were as much as 1,000 times more sensitive to PARPi than BRCA-wild type cells) provided the impetus for PARPi testing as single agents in clinical trials (32).

PARP Inhibitors as maintenance therapy in Recurrent Ovarian Cancer

Homologous recombination deficiency (HRD) is a pivotal determinant of platinum sensitivity in HGSOs and sensitivity to platinum correlates with sensitivity to olaparib (33). In 2014, the European Medical Agency approved olaparib as maintenance therapy for patients with platinum-sensitive recurrent serous ovarian cancer based on a series of trials that started with study [NCT00753545](#), also called Study 19 (34). Of the 265 patients who underwent randomization, 136 were assigned to the olaparib group. Progression-free survival was significantly improved with olaparib treatment versus the placebo (8.4 months vs. 4.8 months from randomization upon completion of chemotherapy). It was also observed that regardless of subgroup, patients in the olaparib group had a lower risk of progression. The olaparib treatment cohort also reported the following adverse effects at grade 1 or 2: nausea, fatigue, vomiting, and anemia. This was followed by three large randomized phase 3 trials that confirmed the progression-free survival (PFS) benefit shown in Study 19 with the use of maintenance PARPi following response to platinum therapy in the recurrent setting. These studies resulted in FDA and EMA approvals for olaparib, niraparib, and rucaparib.

Olaparib

FDA approval of olaparib was primarily based on a non-randomized clinical trial in 298 patients with deleterious or suspected deleterious germline BRCA mutation-associated cancer and recurrent cancers, that included 193 patients with ovarian cancer (35). Patients were treated with olaparib capsules at a dose of 400 mg orally twice daily until disease progression or intolerance to therapy. Median PFS was 7 months and the proportion of patients who were progression-free at 6 months was 54.6%. After a median follow-up of 6.5 years, there was also a clinically significant improvement in overall survival (OS) in the study population (HR 0.73, 95% CI 0.55–0.95; nominal $p=0.021$) and in patients with a BRCA mutations (HR 0.62, 95% CI 0.42–0.93; $p=0.021$). Further, responses to olaparib

were seen in 30% of patients who had platinum-resistant disease, suggesting that there is not always cross resistance between platinum-resistant tumors and PARP inhibition.

Subsequent studies have expanded on the utility of olaparib in the recurrent setting. An adaptive study, Study 24 ([NCT00777582](#)), enrolling patients with advanced solid BRCA-mutated ovarian and breast tumors was carried out to determine the optimal dose of the olaparib tablet formulation for use in phase III trials and determined that the olaparib 300mg b.i.d. matched or surpassed the dose of 400mg b.i.d. capsules (36). The use of olaparib as maintenance therapy was demonstrated by the SOLO2 phase III trial ([NCT01874353](#)) study wherein a significant PFS improvement (19.1 months with maintenance olaparib vs. 5.5 months with placebo, HR = 0.30 [95% CI: 0.22–0.41]) with no detrimental effect on quality of life and low-grade, manageable toxicities was observed in patients with platinum-sensitive, relapsed ovarian cancer and a germline *BRCA1/2* mutation who had previously received at least 2 rounds of chemotherapy (37). Patients were randomly assigned olaparib (300 mg in two 150 mg tablets, twice daily) or placebo tablets using an interactive voice and web response system and stratification was based on response to prior platinum-based chemotherapy (complete vs partial) and length of platinum-free interval (>6–12 months vs >12 months). In SOLO3, a phase III trial of olaparib tablets versus non-platinum chemotherapy in patients with germline BRCA-mutated platinum-sensitive relapsed ovarian cancer who had received at least 2 prior lines of platinum-based chemotherapy, olaparib was associated with statistically significant and clinically relevant improvements in ORR and PFS (38). Olaparib has also been tested in combination with other agents to determine efficacy as therapy for recurrent ovarian cancers. These include platinum agents such as carboplatin (39–41) and anti-angiogenic agents such as cediranib (42). There are a multitude of ongoing clinical studies that aim to test usage of several FDA-approved drugs or agents in combination with olaparib for platinum-sensitive and resistant recurrent ovarian cancers.

Niraparib

Approval for niraparib maintenance therapy was based on the findings from the NOVA trial ([NCT01847274](#)) (43–45). Of 203 patients with a germline BRCA mutation (gBRCAm), 99 had a partial response and 104 had a complete response to their last platinum-based therapy; of 350 patients without a confirmed gBRCAm (non-gBRCAm), 173 had a partial response and 177 had a complete response. In this randomized, double-blind, phase 3 trial, patients received niraparib 300 mg or placebo once daily until progression of disease or death. While there was a more substantial benefit in the germline BRCA-mutated cohort (21.0 vs. 5.5 months, HR = 0.26; 95% CI: 0.17–0.41), patients treated with niraparib in the non-gBRCAm cohort had a PFS advantage of almost 6 months as well (9.3 vs. 3.9 months, HR = 0.45; 95% CI: 0.34–0.61). Patients achieved clinical benefit from maintenance treatment with niraparib regardless of response to the last platinum-based therapy and the incidence of any-grade and grade 3 or greater adverse events was manageable. Following the NOVA trial, the principal findings were replicated among a Chinese population in the NORA trial (46). In this study, 265 patients were randomized 2:1 to receive niraparib or placebo. Notably, most patients received a dose of 200 mg daily, which was found to be a better tolerated dose than 300 mg in the NOVA trial. Again, PFS was significantly longer (18.3 vs. 5.4 months, HR = 0.32 [95% CI: 0.23–0.45]).

Rucaparib

Finally, ARIEL3 (NCT01968213) demonstrated a PFS advantage among all patients who received rucaparib maintenance therapy vs. placebo (10.8 vs. 5.4 months, HR = 0.36; [95% CI: 0.30–0.45]), with a significantly pronounced effect in women with HRD (13.6 vs 5.4 months, HR = 0.32; [95% CI: 0.24–0.32]) (47). In this randomized, double-blind, placebo-controlled, phase 3 trial, 375 out of 564 patients were allocated to oral rucaparib 600 mg twice daily in 28-day cycles using a computer-generated sequence, stratified by HR repair gene mutation status, progression-free interval after the penultimate platinum-based regimen, and best response to the most recent platinum-based regimen. The most common treatment-based adverse effects were anemia and decreased hemoglobin concentration.

The potency of PARPi as maintenance therapy for recurrent ovarian cancer was consistently demonstrated in these studies. Moreover, the overall tolerability of these drugs was determined, with familiar rare but severe adverse events such as secondary malignancies occurring at a rate of < 1.5% which was equivalent across all trials. Currently, the National Comprehensive Cancer Network guidelines recommend the consideration of PARPi for all patients with platinum-sensitive recurrent ovarian cancer irrespective of BRCA status (48) and there are several studies underway testing combinations of synthetic lethality-based PARPi in varying stages of ovarian cancer (Table 1). Benefits of PARPi appear strongest in women with gBRCAm with platinum-sensitive disease. In addition, HRD causes characteristic genomic scar signatures, and the “HRD score” is the sum of these scar signature scores which correlates with sensitivity to PARPi (49–51). The HRD score is defined by determining mutations of BRCA1 and BRCA2, the methylation of their promoters, the level of expression of BRCA1 at the mRNA level, and loss of heterozygosity affecting the loci coding for BRCA1 and BRCA2. Studies are needed to validate HRD assays as predictive biomarkers for benefit to PARPi to determine and personalize the type of drug that will be the most beneficial to each patient and predict when during the treatment regimen to use them.

PARP Inhibitors in Frontline Therapy

Based on the success of PARPi maintenance therapy after treatment for recurrent HGSOc, use in the frontline was investigated (52). SOLO1 (NCT01844986) is an international, randomized, double-blind, phase 3 trial that demonstrated a substantial PFS benefit with the use of olaparib tablets (300 mg, twice daily) in patients with newly-diagnosed advanced HGSOc or endometrioid ovarian cancer with germline or somatic *BRCA 1/2* mutations who had a complete or partial clinical response after platinum-based chemotherapy (53). After a median follow-up of 41 months, the risk of disease progression or death was 70% lower with olaparib than with placebo (HR = 0.30; [95% CI: 0.23–0.41]), with the median PFS not yet reached (vs. 13.8 months for placebo). A sensitivity analysis performed to assess for attrition bias showed that the median PFS was approximately 36 months longer in the olaparib group compared to the placebo group. Despite the longer duration of treatment, the safety profile of olaparib in the SOLO1 trial was consistent with that observed in the SOLO2 trial in patients with relapsed disease. Based on the SOLO1 study, the first FDA approval for PARPi in a frontline setting was granted for olaparib in December 2018.

Similarly, PRIMA ([NCT02655016](#)) compared niraparib maintenance with placebo following chemotherapy for newly diagnosed ovarian cancer (54). Of the 733 patients who underwent randomization, 373 (50.9%) had HR-deficient tumors. The PFS among women with HRD compared to placebo was 21.9 months vs. 10.4 months (HR = 0.43; [95% CI: 0.31–0.59], with a PFS advantage of 13.8 months (niraparib) vs. 8.2 months (placebo) (HR = 0.62; [95% CI: 0.50–0.76] in the overall population. Target enrollment has been completed for the phase 3 ATHENA trial, which will evaluate rucaparib (Rubraca) as frontline maintenance therapy in patients with newly diagnosed advanced ovarian cancer. Data are expected in 2021.

The frontline setting has also seen addition of another PARPi, veliparib. Veliparib was evaluated in the VELIA phase 3 trial ([NCT02470585](#)) assessing its efficacy when added to first-line chemotherapy followed by veliparib maintenance therapy (55). In this international, phase 3, placebo-controlled trial, patients received chemotherapy plus placebo followed by placebo maintenance (control), chemotherapy plus veliparib followed by placebo maintenance (veliparib combination only), or chemotherapy plus veliparib followed by veliparib maintenance (veliparib throughout). A PFS advantage of 34.7 months in the veliparib throughout group vs. 22.0 months in the control group (HR = 0.44; [95% CI: 0.28–0.68] was observed in the BRCAm cohort, with a similar PFS advantage seen in the HRD cohort as well. The role of veliparib maintenance therapy alone without its effects during induction therapy has not been established. Based on the aforementioned studies, it is likely that the indication for frontline maintenance therapy with a PARPi will be expanded to women with HRD and potentially all patients with newly diagnosed advanced ovarian cancer (Figure 2).

PARPi combinations

There has been interest in combining PARPi maintenance with antiangiogenic maintenance therapy. PAOLA-1 ([NCT02477644](#)) evaluated concomitant olaparib and bevacizumab first-line maintenance therapy following platinum-based chemotherapy with concurrent bevacizumab in a randomized, double-blind, international phase 3 trial (9). Patients received first-line platinum chemotherapy plus bevacizumab and were randomized to receive maintenance placebo or olaparib plus bevacizumab maintenance, regardless of BRCA mutation status. Preliminary results demonstrated a median PFS of 22.1 months in the olaparib/bevacizumab cohort vs. 16.6 months in the placebo/bevacizumab cohort ($p < 0.0001$). However, the benefits of PFS were observed only in patients with BRCA mutations and HRD. Given that no trial cohort assessed PARPi therapy without bevacizumab, and only those with HRD and BRCAm benefited from the addition of olaparib, it is difficult to determine if and how much additional benefit is achieved by adding bevacizumab.

Trials are also underway looking at combining PARPi with immunotherapy (Table 1). This is based on three key observations: (1) BRCA1/2-mutated tumors have higher neoantigen loads, more tumor-infiltrating lymphocytes, and overexpress immune checkpoint modulators, PD-1 and PDL1 when compared to HR-proficient tumors (56,57); (2) PARPi elicits a STING-dependent antitumor immune response independent of its effects on DNA repair (58); and (3) PARPi causes tumor cells to increase expression of the immune checkpoint molecule PDL1 (59). DNA repair defects that have resulted (directly or

indirectly) from genomic instability are transcribed and translated to encode novel peptide sequences that are unique to the cancer cell. During normal protein degradation, these novel peptides may be bound by major histocompatibility complex (MHC) proteins that present them on the cell surface as ‘neoantigens’ and this large proportion of mutant neoantigens in mismatch repair–deficient cancers makes them sensitive to immune checkpoint blockade, regardless of the cancer tissue of origin. The MEDIOLA trial (NCT02734004) examined a combination of olaparib and durvalumab in germline BRCA1/2-mutated platinum-sensitive recurrent EOC and found an overall response rate (ORR) of 71.9% (60). The TAPACIO trial (NCT02657889) is looking at niraparib plus pembrolizumab in patients with platinum resistant ovarian cancer irrespective of germline BRCA1/2 status (61). The investigators have reported an ORR of 25% among all platinum-resistant patients and ORR of 45% among patients with somatic BRCA mutations. Two large ongoing clinical trials are also looking at possible PARPi/immunotherapy in the upfront setting. The FIRST trial (NCT03602859) has recently concluded accrual looking at niraparib/dostarlimab followed up niraparib/dostarlimab maintenance, while the ATHENA trial (NCT03522246) is examining rucaparib/nivolumab in the maintenance setting.

Synthetic Lethality beyond PARP inhibitors

Several mechanisms of PARPi resistance have been described and they may be HR-dependent or independent. HR-dependent mechanisms include restoration of BRCA function via secondary or reversion mutations (62) or restoration of HR through other pathways that are independent of BRCA (63). HR-independent mechanisms entail increased replication fork stabilization (64), upregulation of pathways that favor cell survival (65), upregulation of drug efflux pumps (66), and alterations in PARP activity (67). This calls for identifying novel synthetic lethal combinations to overcome PARP resistance mechanisms or sensitize cancer cells to PARP inhibitors, thereby improving clinical effectiveness and patient outcome.

The potential of synthetic lethality to produce genotype-specific cell inhibition has promoted further interest in identifying novel synthetic lethal combinations for ovarian cancer therapy (Table 1). One such approach focuses on loss of function mutations in tumor suppressor genes. For example, the tumor suppressor TP53 (p53) is mutated in most cancers, including over 96% of HGSOCs (68). Several lethal approaches to targeting p53-mutant tumor cells have been proposed, including targeting the p38 mitogen-activated protein kinase MK2 (69), components of the PI3-kinase signaling pathway (70), the DNA-damage checkpoint kinase CHK1 (71,72), and a key determinant of replication fork stability, ATR (ataxia telangiectasia and RAD3-related) (73,74). Wang and Simon identified potentially druggable genes synthetically lethal for p53 using three microarray datasets for gene expression profiles of the NCI-60 cancer cell lines, one next-generation sequencing (RNA-Seq) dataset from the Cancer Genome Atlas (TCGA) project, and one gene expression data from the Cancer Cell Line Encyclopedia (CCLE) project. They demonstrated that pre-screening of potential synthetic lethal genes using gene expression profiles is a promising approach for improving the efficiency of synthetic lethal RNAi screening (75). The role and function of p53 as a key driver of tumorigenesis has led to new proposed innovative synthetic lethality strategies against p53-defective cancers (76). Several drugs targeting mutant p53

have entered clinical trials. One strategy is targeting the chaperone protein Hsp90, which is required for proper folding of the mutant p53 protein. For example, GANNET53 was a trial of the Hsp90-inhibitor Ganetespiib given in combination with paclitaxel. Unfortunately, the Phase II trial failed to show an improvement from the combination therapy over paclitaxel alone, but similar strategies are still in development (77).

Another new strategy for synthetic lethality-based therapies is targeting protein coding genes that are essential for chromatin regulation and reorganization. One promising target includes the SWI/SNF chromatin remodeling complex (78). Helming *et al* demonstrated that partial loss of ARID1 (AT-Rich Interaction Domain) function via mutation of *ARID1A* alleles or, less frequently, *ARID1B* alleles can drive cancer growth but at the same time creates a specific vulnerability compared to non-mutant cells, suggesting ARID1B as a potential target for cancers that contain inactivating ARID1A mutations (79). Dasatinib, an oral dual BCR/ABL and Src family tyrosine kinase inhibitor approved for use in patients with chronic myelogenous leukemia (CML), was found to be consistently lethal in ovarian cancer cells after ARID1A knockdown in ovarian clear cell carcinoma (80). These results implicate ARID1B as a potential therapeutic target for ovarian cancers using synergistic lethality approaches. For example, BET (bromodomain and extra terminal domain) inhibitors cause a reduction in the expression of multiple SWI/SNF members including ARID1B, which produces synthetic lethality with ARID1A loss (81). This strategy is now being tested in a clinical trial of PLX2853, a BET inhibitor, for ARID1A-deficient ovarian cancers (NCT04493619).

Other pathways involved in DNA damage response (DDR) and DNA repair targeting are producing new opportunities for synthetic lethality interventions. The lessons learned from PARPi inhibitors have helped inform DDR inhibitor development. For example, WEE1 protein kinase has been described as an inhibitor of CDK 1 and 2 resulting in temporary cell cycle arrest and DNA damage repair. In ovarian and endometrial cancers WEE1 inhibitor induce mitotic lethality, rendering p53-deficient cells sensitive to radiation and DNA damaging agents including PARPi and chemotherapeutic agents (82). The WEE1 inhibitor adavosertib demonstrated preclinical synergy with gemcitabine in early phase clinical trials (83,84). Adavosertib efficacy was demonstrated as a monotherapy in a phase 1 clinical trial with 25 refractory solid tumor patients where two patients with BRCA mutations had partial response (85) and also as a combinatorial modality with chemotherapies which resulted in 53% disease stabilization and 10% partial response (86). A phase 2 trial with 24 p53-mutated ovarian cancer patients receiving adavosertib plus carboplatin reported that the overall response rate was 43% including one patient (5%) with a prolonged complete response (87). More recently, in a double-blind, randomized, placebo-controlled, phase 2 trial, women with measurable recurrent platinum-resistant or platinum-refractory high-grade serous ovarian cancer, the combination of adavosertib and gemcitabine resulted in an improvement in progression-free survival and overall survival (88). This observed clinical efficacy of a WEE1 inhibitor combined with existing treatments supports ongoing assessment of DNA damage response drugs in HGSOC.

Certainly, additional opportunities for synthetic lethality have yet to be explored. Heinzel *et al* identified a set of drug combinations currently not tested in late stage ovarian cancer

clinical trials which might produce synthetic lethal interactions and therefore could be worthy of further studies in ovarian cancer (89). This data was mined by mapping a set of synthetic lethal interactions from publicly available data on yeast screens to their respective human orthologs and was complemented by a set of predicted synthetic lethal interactions based on a set of protein meta-data such as molecular pathways. Further, the group established that twelve of the tested drug combinations addressed a synthetic lethal interaction with the anti-VEGF inhibitor bevacizumab in combination with paclitaxel, targeting the synthetic lethal pair between VEGFA and BCL2. They also identified a set of 84 drug combinations with PARPi currently not tested in phase III or IV trials in the context of ovarian cancer which address 102 synthetic lethal interactions.

Conclusions and Future Directions

About 20 years ago, Hartwell *et al.* first proposed that synthetic lethality-based anticancer drug targets can be identified in model organisms, such as yeast (90). Despite the promising nature of synthetic lethality-based therapeutics, the technology faces three major shortcomings that deter translation into the clinic. The first considerable impediment has been the identification of robust, clinically relevant synthetic lethal interactions due to false positive and false negative interactions predicted by high-throughput screening technologies. This calls for follow-up studies to cross-validate potential targets using other experimental strategies such as combinatorial drug testing. Second, these genetic interactions induce lethality, rendering it a challenge to recover and identify mutants. Third, the regulation of signaling pathways and the intracellular environment are heterogeneous due to the genetic and metabolic differences between different cell types making most synthetic lethal interactions condition-dependent and rare. Large numbers of mutant gene-pair combinations are required to understand crosstalk between these pathways and establish potent synthetic lethal interactions. Owing to these reasons, most large-scale synthetic lethal genetic interaction screens have been carried out in budding yeast or fission yeast, as these screens can survey a much larger interaction space and can identify potential cancer-relevant synthetic lethal interactions for direct testing in human cell lines, thereby reducing the number of genetic interaction pairs that need to be tested in human cells. These screens can be constructed by mining genetic networks from existing model organisms for synthetic lethal interactions together with cancer-relevant genes. Furthermore, breakthroughs in RNA interference (RNAi) and CRISPR technology and advances in genome sequencing to rapidly identify genetic and epigenetic changes that differentiate tumor cells from non-tumor cells in a patient have rendered large-scale unbiased synthetic lethality screening directly in human cells feasible.

In conclusion, synthetic lethality provides a means to selectively target tumor cells and spare the patient's non-malignant cells, resulting in favorable therapeutic indices. Additionally, synthetic lethal interactions can facilitate the indirect targeting of non-druggable cancer mutations via identification of a druggable synthetic lethal gene partner, thereby broadening the repertoire of anticancer therapeutic targets. Strategies to exploit synthetic lethality promise to be an area of active research in ovarian cancer for years to come.

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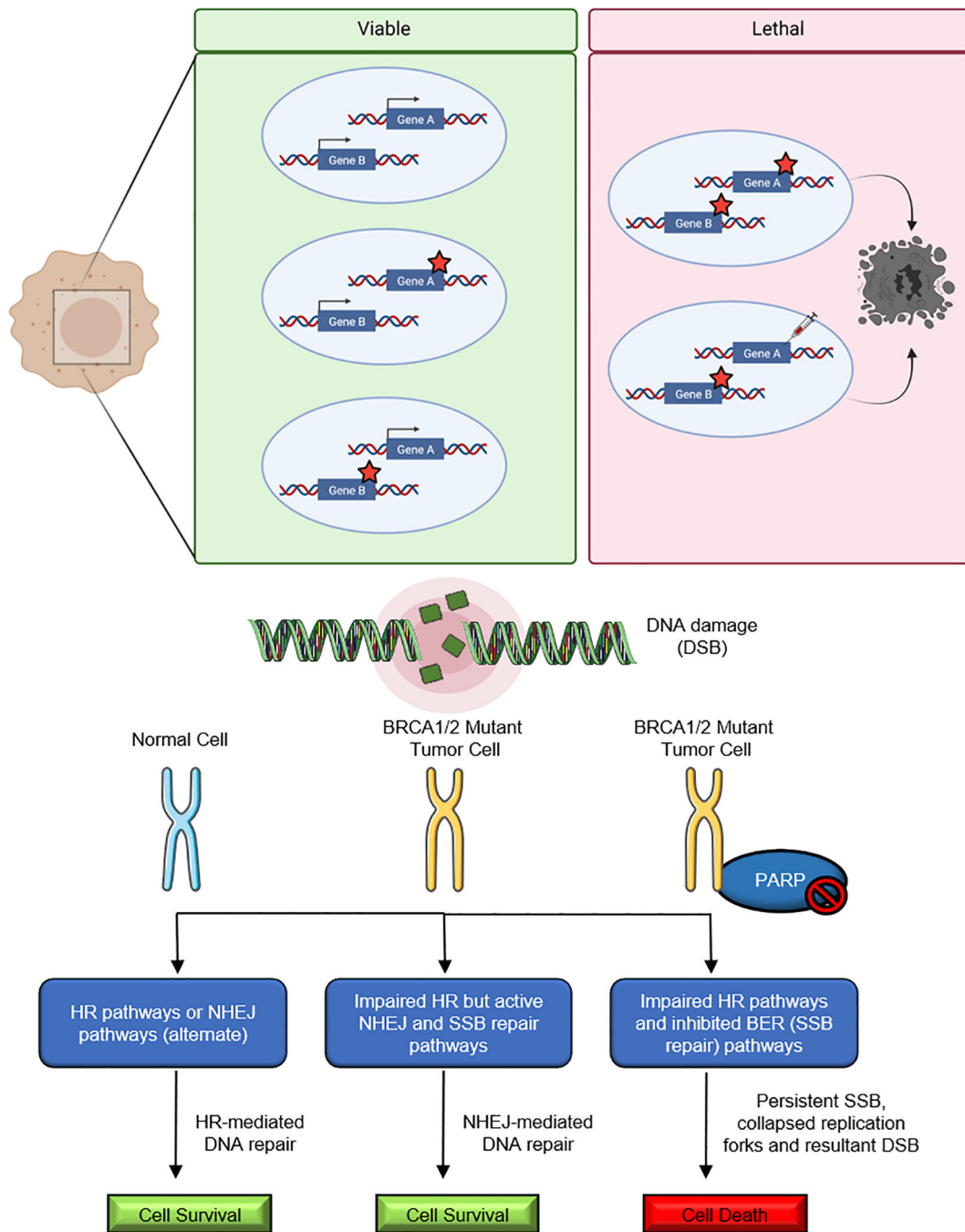


Figure 1: The concept of synthetic lethality and exploiting the synthetic lethal relationship between PARP1 and BRCA1/2 for the targeted treatment of HR-deficient human tumors.

(a) The loss, inhibition or the overexpression (either of which is denoted by a red star) of either of the protein products of gene A or B alone does not affect cell viability. Mutation, pharmacological inhibition, or overexpression of the protein product of gene B in cells with previous mutations in gene A results in synthetic lethality-based cell death. (b) HR can serve as a backup DNA repair pathway to resolve DSBs resulting from replication fork collapse. In normal cells, base modifications are repaired using base excision repair prior to S-phase entry. The newly synthesized sister chromatid can serve as a template for HR-mediated

repair in BRCA1/2-proficient cells. In BRCA1/2-deficient cancer cells HR-mediated repair of PARP inhibitor-induced DSBs is not available. In this condition, tumors could still rely on the alternative error prone NHEJ pathway to repair DSBs and survive. Use of PARP inhibitors will cause unrepaired SSBs, progressive genomic instability, and synthetic lethality-based cell death in BRCA-mutated cancers.

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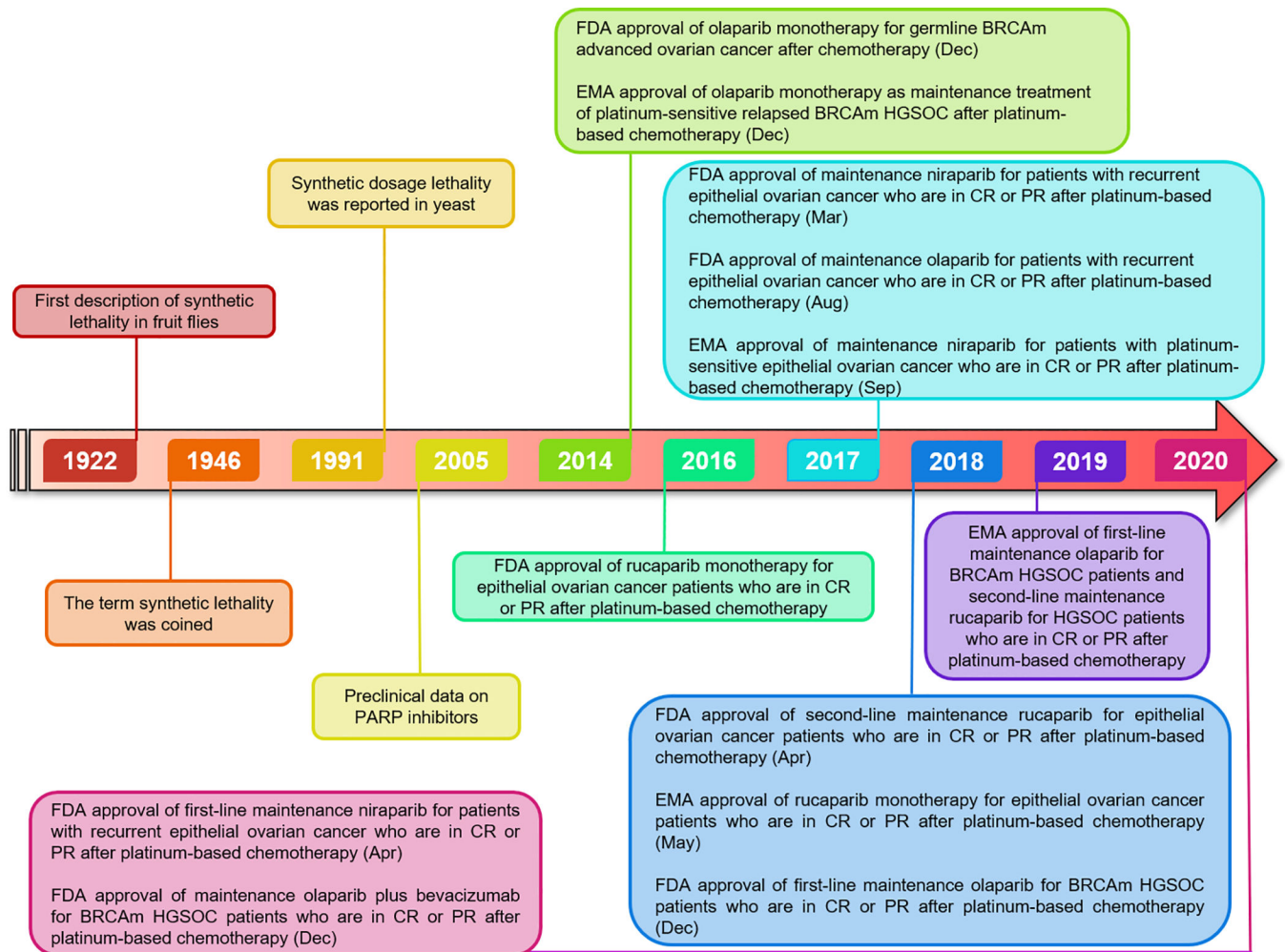


Figure 2: Timeline of landmark synthetic lethality advances in ovarian cancer.

Several expanded concepts of synthetic lethality have been constantly proposed and studied. After Ashworth and Helleday demonstrated synthetic lethality of PARP inhibitors in BRCA1/2-mutated (BRCAm) tumors, several PARP inhibitors have been clinically tested for first-line, second-line, and maintenance therapy in ovarian cancer. FDA - U.S. Food and Drug Administration, EMA - European Medicines Agency, CR - complete response, PR - partial response

Table 1:

Ongoing clinical trials related to synthetic lethal interactions involving PARPi, combination of PARPi with immunotherapies, and targeting other synthetic lethal interactions in OC beyond PARPi.

Ongoing clinical trials related to synthetic lethal interactions involving PARPi for OC		
Trial	Phase	Design
<i>Olaparib</i>		
NCT02282020 SOLO3	III	Randomized olaparib tablets po bid vs physician choice single agent non-platinum-based chemo for gBRCAm platinum-sensitive relapsed HGSOE/EOC following 2 platinum-based chemo with progression 6 months after last platinum
NCT02446600	III	Randomized platinum-based chemo (carboplatin + paclitaxel; carboplatin + gemcitabine; carboplatin + PLD) vs olaparib vs olaparib + cediranib for platinum-sensitive relapsed HGSOE/EOC or gBRCAm HGSOE with any number of platinum-based chemo and 1 non-platinum therapy with CR to last platinum
NCT03106987 OReO	III	Randomized olaparib vs placebo maintenance re-treatment for relapsed non-mucinous EOC, who have had disease progression following maintenance therapy with a PARPi and a CR/PR to subsequent platinum-based chemotherapy
NCT03278717 ICON 9	III	Randomized maintenance olaparib + cediranib vs olaparib alone for relapsed OC with disease progressed more than 6 months after first line chemotherapy or CR/PR to 4 cycles of platinum-based chemotherapy
NCT03402841 OPINION	IIIb	Non-randomized, non-comparative olaparib maintenance for platinum-sensitive relapsed non-gBRCAm HGSOE/EOC
NCT02502266	II/III	Randomized physician choice chemo (paclitaxel; PLD; topotecan) vs olaparib + cediranib vs olaparib vs cediranib for platinum-resistant or refractory relapsed, HGSOE/EOC non-gBRCAm or HGSOE gBRCAm with 3 prior regimens and 1 non-platinum
NCT02340611	II	Non-randomized, non-comparative cediranib+olaparib after disease progression on olaparib alone in OC
NCT03117933 OCTOVA	II	Randomized olaparib vs olaparib + cediranib vs weekly paclitaxel for BRCAm platinum-resistant OC
NCT03470805	II	Non-randomized, non-comparative olaparib maintenance after response to trabectedin-PLD in recurrent gBRCAm or sBRCAm HGSOE/EOC
NCT03161132 ROLANDO	II	Non-randomized, non-comparative olaparib + PLD for platinum resistant advanced OC
NCT03314740 BAROCCO	II	Randomized weekly paclitaxel vs cediranib-olaparib with continuous schedule vs cediranib-olaparib with intermittent schedule for platinum refractory or resistant recurrent HGSOE
NCT03462342 CAPRI	II	Non-randomized, non-comparative ATR inhibitor AZD6738 + olaparib for recurrent HGSOE (platinum-sensitive or -resistant)

NCT03579316	II	Randomized non-comparative adavosertib AZD1775 alone or with olaparib for recurrent OC during olaparib progression
NCT04065269 ATARI	II	ATR inhibitor in combination with olaparib in gynecological cancers with ARID1A loss
NCT02345265	II	Non-randomized, non-comparative olaparib + cediranib for recurrent OC
NCT04261465 NUVOLA	II	Neoadjuvant chemotherapy in unresectable OC with olaparib and weekly carboplatin plus paclitaxel
NCT02489006 NEO	II	Randomized, neoadjuvant olaparib for platinum sensitive recurrent HGSOE prior to surgery and chemotherapy
NCT02983799	II	Non-randomized, non-comparative olaparib for platinum-sensitive or partially platinum-sensitive, relapsed, HGSOE/EOC with at least 1 prior line of platinum-based chemotherapy, in gBRCAm, sBRCAm, or HRD subgroups
NCT02889900 CONCERTO	IIb	Non-randomized, non-comparative cediranib + olaparib for recurrent platinum resistant OC without gBRCAm
NCT01116648	I/II	To determine the safety and best dose of cediranib + olaparib for recurrent OC
NCT02855697 MOLTO	I	Non-randomized, non-comparative multi-maintenance olaparib for platinum sensitive relapsed gBRCAm HGSOE/EOC with 2 or more courses of maintenance olaparib
NCT01445418	I	Non-randomized, non-comparative olaparib + carboplatin for gBRCAm and sporadic OC
NCT03162627	I	Evaluation of the combination of selumetinib and olaparib in OC with Ras pathway alterations and tumors with PARP resistance
NCT01623349	I	To determine the safety of oral PI3kinase Inhibitor BKM120 or BYL719 + olaparib for recurrent HGSOE
NCT02898207	I	To determine the safety and best dose of olaparib + HSP90 inhibitor onalespib for recurrent OC
NCT01650376	Ib	To determine the MTD of olaparib + weekly carboplatin and paclitaxel in relapsed OC
NCT02208375	Ib	To determine the MTD of olaparib + oral mTORC1/2 inhibitor AZD2014 or AKT inhibitor AZD5363 for recurrent OC
NCT04586335	Ib	Study to evaluate the safety, tolerability, pharmacokinetics and clinical activity of CYH33, a PI3K inhibitor in combination with olaparib in advanced solid tumors
<i>Niraparib</i>		
NCT02655016 PRIMA	III	Randomized niraparib po qd vs placebo for first-line maintenance HGSOE/EOC stage III-IV with CR/PR to front-line platinum-based chemo
NCT03602859	III	Randomized comparison of platinum-based therapy with TSR-042 and niraparib vs standard of care platinum-based therapy as first-line of treatment in stage III or IV EOC
NCT04217798	II	Study to evaluate the efficacy and safety of niraparib combined with oral etoposide in platinum resistant/ refractory recurrent OC

NCT04284852 NEOPRIMA	II	Niraparib maintenance in patients with advanced OC at neoadjuvant setting	
NCT04507841	II	Niraparib for the neoadjuvant treatment of unresectable OC	
NCT02354586 QUADRA	II	Non-randomized, non-comparative niraparib po qd for platinum-resistant or heavily pretreated HGSOC following 3 or 4 prior regimens, a response 6ms to first-line platinum-based chemo	
NCT04376073 ANNIE	II	Niraparib in combination with anlotinib in platinum-resistant recurrent OC	
NCT04826198 REVOCAN	I/II	Study to assess the safety and efficacy of AsDNATM, a DNA repair inhibitor and niraparib in relapsed platinum-sensitive OC	
NCT04502602 iNNOVATE	I/b	Niraparib and neratinib in advanced solid tumors with an expansion cohort in platinum-resistant OC	
NCT03154281	I	Evaluation of the safety and tolerability of niraparib in combination with everolimus in advanced OC	
NCT03586661	I	Niraparib with the PI3K inhibitor copanlisib for recurrent OC	
NCT04267939	Ib	To determine the maximum tolerated and/or recommended phase 2 dose of the ATR inhibitor BAY 1895344 in combination with niraparib in patients with recurrent OC	
Rucaparib			
NCT02855944 ARIEL 4	III	Randomized rucaparib po bid vs chemotherapy (carboplatin/paclitaxel, carboplatin/gemcitabine, cisplatin/gemcitabine, paclitaxel, carboplatin, cisplatin) for relapsed or progressing BRCAm HGSOC 2 prior regimens	
NCT03522246 ATHENA	III	Randomized study evaluating rucaparib and nivolumab as maintenance treatment following response to front-line platinum-based chemotherapy in OC	
NCT03992131 SEASTAR	I/II	Study to assess the safety, tolerability, pharmacokinetics, and preliminary efficacy of oral rucaparib in combination with other anticancer agents in OC	
NCT03552471	I	Mirvetuximab soravtansine (IMGN853) and rucaparib for recurrent OC	
NCT03840200	Ib	Evaluating the safety and efficacy of ipatasertib in combination with rucaparib in advanced OC	
Ongoing clinical trials combining PARPi with immunotherapies for OC			
Combinatorial Drug	Trial	Phase	Design
<i>Olaparib</i>	NCT02477644 PAOLA-1	III	Randomized olaparib vs placebo for advanced IIIB - IV HGSOC/EOC with standard first-line platinum-taxane chemotherapy and bevacizumab concurrent and in maintenance, 3 cycles of bevacizumab in combination with the 3 last cycles of platinum-based chemotherapy
	NCT03740165	III	Randomized study of chemotherapy with or without pembrolizumab followed by maintenance with olaparib or placebo for the first-line treatment of BRCA non-mutated advanced EOC

Author Manuscript	NCT03737643 DUO-O	III	Randomized multicentre study of durvalumab in combination with chemotherapy and bevacizumab, followed by maintenance durvalumab, bevacizumab and olaparib in newly diagnosed advanced OC
	NCT04034927	II	Randomized olaparib vs olaparib plus tremelimumab in platinum-sensitive recurrent OC
	NCT03699449 AMBITION	II	Randomized olaparib+cediranib, durvalumab + olaparib, durvalumab + chemotherapy, durvalumab + tremelimumab + chemotherapy; a biomarker-driven targeted therapy for HRD platinum-resistant recurrent OC
	NCT04361370	II	Olaparib maintenance with pembrolizumab & bevacizumab in BRCA non-mutated patients with platinum-sensitive recurrent OC
	NCT04739800	II	Randomized triplet therapy trial (a PD-L1 inhibitor (durvalumab) MEDI4736 in combination with olaparib and cediranib) compared to olaparib and cediranib or durvalumab and cediranib or standard of care chemotherapy platinum-resistant recurrent EOC after prior bevacizumab
	NCT02571725	I-II	Non-randomized, non-comparative olaparib and CTLA-4 blockade tremelimumab for BRCAm recurrent OC
	NCT02953457	I/II	Non-randomized, non-comparative olaparib together with durvalumab and tremelimumab for gBRCAm recurrent or refractory OC
	NCT02734004 MEDIOLA	I/II	Anti-PD-L1 antibody MEDI4736 in combination with olaparib in advanced OC
	NCT02484404	I/II	Anti-PD-L1 antibody MEDI4736 in combination with olaparib and/or cediranib for advanced or recurrent OC
	NCT02121990	I	Dose-escalation study of IP cisplatin, IV/IP paclitaxel, IV bevacizumab, and oral olaparib for newly diagnosed OC
<i>Niraparib</i>	NCT03598270 ANITA	III	Randomized trial of platinum-based chemotherapy with or without atezolizumab followed by niraparib maintenance with or without atezolizumab in recurrent OC
	NCT04679064 NiCHE-MITO33	III	Randomized niraparib-TSR-042 (Dostarlimab) vs physician's choice chemo in recurrent OC
	NCT04556071 AVANIRA 3	II	Study to evaluate the efficacy and safety of niraparib combined with bevacizumab in platinum-refractory/resistant recurrent OC
	NCT03326193	II	Non-randomized, non-comparative bevacizumab-niraparib for first-line maintenance HGSOC/EOC with CR/PR to front-line platinum-based chemo + bevacizumab 1 debulking surgery
	NCT03574779	II	Study to evaluate the safety and efficacy of novel treatment combinations (niraparib + TSR042 + bevacizumab) in recurrent OC

	NCT02354131 AVANOVA	I/II	Randomized niraparib vs bevacizumab-niraparib for platinum-sensitive relapsed HGSOC/EOC
	NCT02657889 TOPACIO KEYNOTE 162	I/II	Niraparib in combination with pembrolizumab (MK-3475) in recurrent OC
	NCT03695380	Ib	Study of cobimetinib and niraparib, with or without atezolizumab in advanced platinum-sensitive OC
	NCT04673448	Ib	Niraparib and neoadjuvant PD-1 inhibitor dostarlimab (TSR-042) in BRCAm OC
Rucaparib	NCT04227522	III	Rucaparib maintenance after bevacizumab maintenance following carboplatin-based first-line chemotherapy in OC
	NCT02873962	II	Safety lead-in of nivolumab in combination with bevacizumab or in combination with bevacizumab and rucaparib for relapsed EOC
	NCT03552471	I	To determine the safety and best dose of mirvetuximab soravtansine and rucaparib camsylate for recurrent OC
Ongoing clinical trials targeting synthetic lethal interactions in OC beyond PARPi			
ATR	NCT02487095	I/II	Trial of topotecan with ATRi VX-970 (M6620) in OC
	NCT04616534	I	Gemcitabine combined with the BAY 1895344 ATRi with expansion cohorts in advanced OC
	NCT02627443	I	Dose escalation and expansion cohort of carboplatin and gemcitabine with or without ATRi berzosertib M6620 (VX-970) in first or second recurrence platinum-sensitive EOC
Chk1/2	NCT02203513	II	Study of the Chk1/2 inhibitor (LY2606368) In <i>BRCA1/2m</i> HGSOC
	NCT02797964	I/II	Chk1 inhibitor (SRA737) administered orally in <i>BRCA1/2m</i> advanced OC
WEE1	NCT02101775	II	Randomized trial comparing gemcitabine monotherapy to gemcitabine in combination with WEE1 inhibitor (MK-1775) in recurrent platinum-resistant EOC
PI3K	NCT04711161	I/Ib	Evaluation of the safety, pharmacokinetics and efficacy of GRN-300, a salt-inducible kinase inhibitor, alone and in combination with paclitaxel, in recurrent OC
	NCT03719326	I/Ib	Dose-escalation, and dose-expansion study to evaluate the safety, tolerability, pharmacokinetics and clinical activity of etrumadenant (AB928) in combination with pegylated liposomal doxorubicin (PLD) with or without PI3K inhibitor IPI-549 in OC
Akt	NCT04374630 PROFECTA II	II	Study to assess the efficacy and safety of Akt inhibitor afuresertib plus paclitaxel vs paclitaxel in platinum-resistant OC
p53	NCT03113487	II	Modified vaccinia virus ankara vaccine expressing p53 and pembrolizumab in recurrent OC

	NCT02272790	II	Study of adavosertib plus chemotherapy in p53 mutated platinum-resistant OC
	NCT04489706	N/A	Trial to evaluate the efficacy, safety, and tolerability of arsenic trioxide in recurrent metastatic OC with P53 mutation
ARID1A	NCT04493619	I/II	Study of BET inhibitor PLX2853 monotherapy in ARID1A mutated advanced gynecological malignancies and study of PLX2853/carboplatin combination in platinum-resistant EOC

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