



Published in final edited form as:

FASEB J. 2022 April ; 36(4): e22224. doi:10.1096/fj.202101686R.

## Liver Regeneration and Ethanol Detoxification: A New Link in YAP Regulation of ALDH1A1 During Alcohol-Related Hepatocyte Damage

Junmei Zhou<sup>1,&</sup>, Chunbao Sun<sup>2,&</sup>, Yang Lu<sup>3</sup>, Jinhui Wang<sup>4</sup>, Natacha Jn-Simon<sup>2</sup>, Chen Zhou<sup>5</sup>, Andrew Bryant<sup>6</sup>, Qi Cao<sup>7</sup>, Chenglong Li<sup>5</sup>, Bryon Petersen<sup>8</sup>, Liya Pi<sup>2,#</sup>

<sup>1</sup>Guangxi Key Laboratory of Molecular Medicine in Liver Injury and Repair, the Affiliated Hospital of Guilin Medical University, Guilin, Guangxi, China

<sup>2</sup>Department of Pathology, Tulane University, New Orleans, LA, USA

<sup>3</sup>Department of Systems Biology, Beckman Research Institute of the City of Hope, Duarte, CA USA

<sup>4</sup>Integrative Genomics Core, Beckman Research Institute of the City of Hope, Duarte, CA USA

<sup>5</sup>Department of Medical Chemistry, University of Florida, Gainesville, FL USA

<sup>6</sup>Department of Medicine, University of Florida, Gainesville, FL USA

<sup>7</sup>Department of Diagnostic Radiology and Nuclear Medicine, University of Maryland School of Medicine, Baltimore, MD USA

<sup>8</sup>Department of Pediatrics, University of Florida, Gainesville, FL USA

### Abstract

Yes-associated protein (YAP), a central effector in the Hippo pathway, is involved in the regulation of organ size, stem cell self-renewal, and tissue regeneration. In this study, we observed YAP activation in patients with alcoholic steatosis, hepatitis, and cirrhosis. Accumulation of this protein in the nucleus was also observed in murine livers that were damaged after chronic-plus-single binge or moderate ethanol ingestion combined with carbon tetrachloride intoxication (ethanol/CCl<sub>4</sub>). To understand the role of this transcriptional coactivator in alcohol-related liver injury, we knocked out the *Yap1* gene in hepatocytes of floxed homozygotes through adeno-associated virus (AAV8)-mediated deletion utilizing Cre recombinase. *Yap1* hepatocyte-specific knockouts (KO) exhibited hemorrhage, massive hepatic necrosis, enhanced oxidative stress, elevated hypoxia, and extensive infiltration of CD11b<sup>+</sup> inflammatory cells into hepatic microenvironments rich for connective tissue growth factor (Ctgf) during ethanol/CCl<sub>4</sub>-induced

**#Correspondence author:** Liya Pi, PhD, Department of Pathology, Tulane University School of Medicine, 1430 Tulane Ave, New Orleans, LA, USA, lpi@tulane.edu, Phone: (504)-988-2869.

Author Contributions

L Pi, B Petersen, Q Cao, A Bryant, and C Li designed research. J Zhou, C Sun, L Yang, N Jn-Simon, C Zhou, J Wang and L Pi performed experiments and analyzed data. L Pi and J Zhou wrote the article, and all authors read a draft of this article, suggested improvements, and approved the final version of this article.

<sup>&</sup>co-author with equal contribution

Conflict of Interests

The authors declare that there is no conflict of interest regarding publication of this manuscript.

liver damage. Analysis of whole-genome transcriptomics indicated upregulation of genes involved in hypoxia and extracellular matrix (ECM) remodeling, whereas genes related to hepatocyte proliferation, progenitor cell activation, and ethanol detoxification were downregulated in the damaged livers of *Yap1* KO. *Acetaldehyde dehydrogenase (Aldh)1a1*, a gene that encodes a detoxification enzyme for aldehyde substrates, was identified as a potential YAP target because this gene could be transcriptionally activated by a hyperactive YAP mutant. Ectopic expression of the human *ALDH1A1* gene caused increase in hepatocyte proliferation and decrease in hepatic necrosis, oxidative stress, ECM remodeling, and inflammation during ethanol/ $\text{CCl}_4$ -induced liver damage. Taken together, these observations indicated that YAP was crucial for liver repair during alcohol-associated injury. Its regulation of *ALDH1A1* represents a new link in liver regeneration and detoxification.

## Keywords

Yes-associated protein; ethanol; liver regeneration; carbon tetrachloride ( $\text{CCl}_4$ ); *Aldh1a1*

---

## INTRODUCTION

Drinking too much alcohol can have many adverse effects on the body, particularly to the liver that is the main site for ethanol metabolism. Alcohol dehydrogenase (ADH), catalase, and cytochrome P450 (CYP)2E1 are responsible for the first step of ethanol oxidation leading to acetaldehyde production in the liver. This toxic intermediate is eventually converted into acetic acid by cytosolic and mitochondrial aldehyde dehydrogenases (ALDH). Alcohol abuse can activate the microsomal ethanol oxidizing system to induce oxidative stress and lipid peroxidation leading to release of byproducts that form adducts, cause protein dysfunction, and promote DNA mutagenesis(1). Reactive oxygen species (ROS) attack hepatic cells and their signaling mediators leading to loss of hepatocyte function, inflammation, and abnormal liver repair. Consequently, a broad spectrum of liver pathologies starting from steatosis, hepatitis, fibrosis, to cirrhosis may occur in alcohol-associated liver disease (ALD). Ethanol impairs liver regeneration, accumulating with chronic inflammation and liver scarring. Understanding the molecular mechanisms underlying ethanol-induced hepatotoxicity and associated impairment of liver regeneration is essential for therapeutic intervention in ALD.

ALDH family1 member A1 (*ALDH1A1*) belongs to a group of nicotinamide-adenine dinucleotide phosphate-positive [ $\text{NAD(P)}^+$ ]-dependent enzymes that convert toxic aldehyde substrates to their corresponding carboxylic acids(2). It is a cytosolic enzyme and plays an important role in acetaldehyde removal(3). It has been considered as a metabolic marker for stem cells and provides drug protection and chemo-radiation resistance to cancer stem cells(4, 5). In the liver, upregulation of this enzyme is associated with a 57-month recurrence-free survival in patients with HBV-related hepatocellular carcinoma (HCC)(6). *Aldh1a1* has been shown to detoxify lipid peroxidation products such as 4-hydroxynonenal (HNE) and malondialdehyde (MDA), whereas hepatic inflammation downregulates this enzyme in rodents(7, 8). In addition, polymorphic variants of *ALDH1A1* resulting in low

enzyme activity have been reported to be associated with alcohol sensitivity(9). So far, the involvement of this enzyme during alcohol-induced liver injury is not fully understood.

Yes-associated protein (YAP) and its paralog - the WW domain-containing transcription regulator protein 1 (WWTR1), also termed TAZ (transcriptional co-activator with PDZ-binding motif), are central effectors in the Hippo pathway critical for regulation of liver development, repair, cell fate determination and tumorigenesis. Their activity is inhibited through phosphorylation by Hippo kinases including MST1/2 and LATS1/2(10). Phosphorylated YAP is inactive and undergoes proteasome degradation in the cytoplasm. YAP becomes accumulated upon liver injury. Active YAP is in an unphosphorylated form and can enter the nucleus where it binds to transcriptional factors, including members of the TEAD family, and activates target genes including connective tissue growth factor (Ctgf)(11). Upon transcriptional activation, the YAP targets can influence the cell cycle and enhance cell survival thereby promoting cell proliferation. High YAP activity has also been shown to reprogram hepatocytes into a progenitor-like state and control the fate of injured mouse hepatocytes *in vivo*(12, 13). Activation of YAP can attenuate hepatic damage and fibrosis in liver ischemia-reperfusion injury(14). Moreover, YAP can integrate metabolic and nutrient-sensing pathways leading to coordination of nutrient availability with the genetic program that sustains cell proliferation(15, 16). Emerging evidence has shown YAP dysregulation in alcoholic hepatitis(17). Hence, we examined expression patterns of this transcriptional regulator in human livers from patients with alcoholism, and in murine livers that were damaged after chronic-plus-single binge or moderate ethanol ingestion combined with acute intoxication by carbon tetrachloride (ethanol/CCl<sub>4</sub>). Hepatocyte-specific KO were utilized to determine the function of *Yap1* gene in liver regeneration following ethanol/CCl<sub>4</sub>. *Aldh1a1* was identified as one of the downregulated genes in the KO livers by RNA Sequencing analysis. YAP regulation of this detoxification gene and its function in alcohol-related liver injury were investigated.

## MATERIALS AND METHODS

### Animal Experiments

Binge ethanol drinking in combination with chronic ethanol exposure was performed to induce hepatocyte damage using the Lieber-DeCarli liquid diet (BioServ) according to Bertola et al(18). In brief, female C57Bl6 mice (8–10 weeks old) were fed *ad libitum* with the Lieber-DeCarli ethanol diet (ethanol-fed group) or an isocaloric maltose diet (pair-fed group) (Bio-Serv, Flemington, NJ) for 10 days. On day 11, an ethanol binge (single dose at 5 g/kg of body weight) was delivered through oral gavage. The animals were sacrificed at 0 or 9 hours after the binge. Parallel experiments were performed in controls that were fed isocaloric maltose and received gavage of equal volume of water (pair-fed/water).

Hepatocyte damage and liver regeneration were induced through combined treatment with ethanol and CCl<sub>4</sub> according to our previous report(19). Animals were allowed free access to diets with increasing concentrations of ethanol to 2% vol/vol (11% of Kcal) for 4 days. A single acute dose of CCl<sub>4</sub> (1 µl/g body weight) prediluted 1:3 in olive oil was given through intraperitoneal injection.

Hepatocyte-specific *Yap1* KO was generated through tail-vein injection into *Yap1<sup>flox/flox</sup>* mice (8-week-old age) with AAV8 virus expressing codon-improved Cre recombinase (iCre) under the control of the human thyroxine-binding globulin (TBG) promoter (AAV8-TBG-iCre). The virus was purchased from Vector Biolabs (Malvern, PA) and was given at  $5 \times 10^{12}$  plaque forming unit (pfu) per mouse. The same amount of AAV8-green fluorescence protein (GFP) was injected into animals as control. The *Yap1<sup>flox/flox</sup>* mice were in C57BL6 background and were derived from FLPase-mediated removal of the *lacZ* and neomycin reporters in *Yap1<sup>tm1a(KOMP)Mbp</sup>* animals that were purchased from the Knockout Mouse Project (KOMP) Repository. Three weeks after the viral administration, the AAV8-iCre infected *Yap1<sup>flox/flox</sup>* mice (hepatocyte-specific *Yap1* KO) and AAV8-GFP-infected controls were exposed to ethanol (1–2% vol/vol) followed by single acute dose of CCl<sub>4</sub> according to our previous report(19). The livers were harvested at 0, 7, 24, or 48 hours post the chemical induced liver injury.

ALDH1A1 was fused with FLAG tagged epitope in its C terminus (ALDH1A1:FLAG) and was expressed under the control of the chicken  $\beta$ -actin (CBA) promoter in AAV8 virus. AAV8- ALDH1A1:FLAG and AAV8-GFP ( $1 \times 10^{12}$  pfu/mouse) were transduced into 8-week-old animals. Three weeks after the viral administration, the infected mice were exposed to ethanol followed by acute CCl<sub>4</sub> injection as mentioned above. The livers were harvested at 48 hours post the chemical induced liver injury.

All animal protocols were approved by the Animal Care and Usage Committee at Tulane University and were conducted in compliance with their guidelines.

### Human ALD Livers and Histological Analysis

Liver tissues from normal livers and ALD patients were purchased from Sekisui XenoTech (Kansas city, KS). Frozen tissues and fixed sections were used to examine expression patterns of YAP protein. Histological analyses for human or animal livers were performed with standard protocols using the following primary antibodies. Rabbit anti-YAP (Cell Signaling, Danvers, MA), rabbit anti-4-hydroxynonenal (HNE) (Millipore, Burlington, MA), rabbit anti-Ctgf (Abcam, Waltham, MA), rabbit anti-hepatocyte nuclear factor (Hnf4 $\alpha$ ) (Santa Cruz Technologies, Paso Robles, CA), rabbit anti-Ki67 (Abcam), rabbit anti-CD11b (Abcam), mouse anti- $\alpha$  smooth muscle actin (SMA) (Sigma, St. Louis, MO), rabbit anti-von Willebrand factor (vWF) (Dako, Santa Clara, CA), mouse anti-pimonidazole adduct (Hypoxyprobe, Burlington, MA), rat anti-CD163 (Thermo Fisher Scientific, Waltham, MA), and rabbit anti-collagen IV (Thermo Fisher Scientific). Detection was carried out according to the manufacturer's instructions using the ABC-Elite kit with ImmPACT DAB substrate (Vector Laboratories, Burlingame, CA). In addition, IHC for either Ctgf and CD11b in liver sections was detected using a VECTASTAIN ABC-AP kit and Vector Alkaline Phosphatase Red substrate (Vector Laboratories). Alexa Fluor 488 or 594 conjugated donkey anti-rabbit secondary antibody (Invitrogen, Carlsbad, CA) were used for the immunofluorescence staining. Necrosis was revealed by H&E staining. Collagen deposition was measured by Sirius Red staining according to previous publication(20). For estimation of YAP-stained positive areas, hepatic necrosis, Hnf4 $\alpha$ <sup>+</sup> hepatocytes, and Ki67<sup>+</sup> proliferating hepatocytes, images were captured with CellSens software using an Olympus BX 51 upright fluorescence

microscope outfitted with an Olympus DP80 camera, Plan Fluorite objectives and an LED transmitted light source (Olympus, Waltham, MA). DAB stained areas were quantified from 10 random fields of images (200x magnification) using Image J software (<http://rsb.info.nih.gov/ij/>) and IHC profiler according to published methods(21).

### RNA Isolation, Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR), and RNA-Sequencing (RNA-Seq) Analysis

Total RNA isolation, RT-PCR and Quantitative RT-PCR (Q-RT-PCR) analyses, and RNA-Seq were performed in the same conditions according to our previous publication(19). The raw sequencing data and the processed data have been deposited in the Gene Expression Omnibus (GEO) database under the accession number GSE123309 for *Yap1* hepatocyte-specific knockouts that received the ethanol/ $\text{CCl}_4$ -induced liver injury. The related raw and processed data for the chronic ethanol feeding-plus-single binge model are also in GEO database (under review). Primers used in RT-PCR analysis were reported(20), except for the following genes. *Dehydrogenase/Reductase 3 (Dhrs3)* primers were 5' GCGCTGGTAGTGTTCCTC 3' and 5' GGTGTTGACATGCTGGGACTT 3'; *Aldh1a1* were 5' AACACAGGTTGGCAAGTTAATCA 3' and 5' TGCGACACAACATTGGCCTT 3'; *Yap1* were 5' TTCAATGCCGTCATGAACC 3' and 5' ATCCTGAGTCATGGCTTGCT 3'; *iCre* were 5' GTGCAAGCTGAACAACAGGA 3' and 5' CCAGCATCCACATTCTCCTT 3'; *Adh1* were 5' CCATCGAGGACATAGAAGTCGC 3' and 5' TGGTTTCACACAAGTCACCCC 3'; *Hypoxia-inducible factor 1a (Hif1a)* were 5' GTCCCAGCTACGAAGTTACAGC 3' and 5' CAGTGCAGGATACACAAGGTTT 3'; *Vascular endothelial growth factor (Vegf)-a* were 5' GCACATAGAGAGAATGAGCTTCC 3' and 5' CTCCGCTCTGAACAAGGCT 3'; *Integrin subunit alpha M (Itgam)* were 5' ATGGACGCTGATGGCAATACC 3' and 5' TCCCCATTACGTCTCCCA 3'; *Epithelial cell adhesion molecule (Epcam)* were 5' CTGGCGTCTAAATGCTTGGC 3' and 5' CCTTGTCGGTTCTTCGGACTC 3'; *Axin-related protein 2 (Axin2)* were 5' ATGAGTAGCGCCGTGTTAGTG 3' and 5' GGGCATAGGTTTGGTGGACT 3'; *Leucine rich repeat containing G protein-coupled receptor 5 (Lgr5)* were 5' GGACCAGATGCGATACCGC 3' and 5' CAGAGGCGATGTAGGAGACTG 3'; *T-Box 3 (Tbx3)* were 5' AGATCCGGTTATCCCTGGGAC 3' and 5' CAGCAGCCCCACTAACTG 3'; *Cytokeratine 19 (Krt19)* were 5' GTTCAGTACGCATTGGGTCAG 3' and 5' GAGGACGAGGTCACGAAGC 3'; *GFP* were 5' CAAATGGGCGGTAGGC 3' and 5' AGCGTGGATGGCGTCT 3'.

### Hepatotoxicity Measurement

Hepatic MDA in liver tissues were extracted and quantified based on the fluorescence intensity with excitation at 532 nm and emission at 553 nm according to manufacturer's instruction in a Lipid Peroxidation Assay Kit (Sigma).

ROS generation was determined in mouse livers by using 2',7' -dichlorofluorescein diacetate (DCFHDA) that reacts with ROS to produce 2', 7' -dichlorofluorescein (DCF), a highly fluorescent compound. In brief, mouse livers were perfused to remove blood. The fluorescent product DCF was measured with excitation at 484 nm and emission at 530 nm after homogenization of the liver extracts.

For measurement of hepatic Aldh activity, cytosolic fractions were isolated by centrifugation at 108,000xg from homogenized mouse livers. The Aldh activity was measured using fluorometric detection kit (Sigma) with excitation wavelength at 535 nm and emission wavelength at 587 nm.

For acetaldehyde determination, the liver was perfused and homogenized in PBS. After centrifugation, the liver supernatants were mixed with AldeLight™ blue reaction mixture in Amplite™ Fluorimetric Aldehyde Quantitation kit (AAT Bioquest, Inc., Sunnyvale, CA) and fluorescence was measured at 365 nm for excitation wavelength and 435 nm for emission wavelength.

For assessment of liver hypoxia, animals that received ethanol/CCl<sub>4</sub> cotreatment were intraperitoneally injected with pimonidazole (120 mg/Kg) 1 hour before euthanasia. The harvested livers were paraffin embedded. Pimonidazole-adducts were stained in IHC using a kit from Hypoxyprobe.

Alanine aminotransferase (ALT) in sera was measured using a kit from Sigma according to manufacturer's instruction.

### Western Blotting

Total proteins were extracted from mouse livers or cultured cells in RIPA buffer containing proteinase inhibitors (Sigma). Nuclear fractions were isolated using NE-PER™ Nuclear and Cytoplasmic Extraction Reagents (ThermoFisher Scientific, Waltham, MA). Nuclear fractions (10 µg) or total protein lysates (50 µg) were boiled in 1x Laemmli buffer containing 5% β-mercaptoethanol, separated on 4–12% Bis-Tris protein gels (Novex, Carlsbad, CA), and electro-transferred onto polyvinylidene difluoride membrane for immunoblotting. Primary antibodies used were mouse anti-Aldh1a1 (Santa Cruz Technologies), rabbit anti-Yap and anti-phosphorylated Yap at serine 127 residue (p-YAP) (Cell Signaling), rabbit anti-Adh1 (Santa Cruz Technologies), mouse anti-FLAG antibody (sigma), mouse anti-Cyclin D1 (Santa Cruz Technologies), rabbit anti-Hif1α (ThermoFisher Scientific), rabbit anti-fibronectin (Fn1) (Abcam), rabbit anti-Vegfa, rabbit anti-CD11b (Abcam), mouse anti-Aldh1a1 (Cell Signaling), and Dhfr3 (Proteintech, Rosemont, IL), mouse anti-Actin (Abcam), and rabbit anti-GAPDH (Abcam). Detection was carried out using horseradish peroxidase-conjugated secondary antibodies (Santa Cruz biotechnologies) and the ECL Plus kit (Amersham Biosciences, Piscataway, NJ).

### Electrophoretic Mobility Shift Assay and Luciferase Reporter Assay

Electrophoretic Mobility Shift Assay (EMSA) was carried out using Pierce™ Biotin 3' End DNA labeling kit (ThermoFisher Scientific) to generate probes. A wild type *Aldh1a1* probe was generated using primers 5' AGCAGGAAAAGGGAATGGAAAAAAA 3' (sense) and 5' TTTTTTTTCCATTCCCTTTTCCTGCT 3' (antisense). 5' AGCAGGAAAAGactagGAAAAAAA 3' (sense) and 5' TTTTTTTTCctagCTTTTCCTGCT 3' (antisense) were for a mutant probe. A mobility shift reaction mixture was set up using LightShift™ Chemiluminescent EMSA kit (ThermoFisher Scientific). It contained crude nuclear extracts overexpressing murine Yap protein that was tagged with Myc epitope at its C terminal region (Yap:Myc) (2 mg), poly(dIdC)

(1mg), 0.1 mg of sonicated denatured salmon sperm DNA, biotin-labeled wild type or mutant probe (4 pmol). A cold wild type probe (20 fmol) was added in some experiments to determine binding specificities of tested probes. Complexes in reaction mixtures (20  $\mu$ l) were separated in polyacrylamide gel and electro-transferred onto a nylon membrane for immunoblotting. The biotin end-labeled DNA probe was detected using streptavidin conjugated to horseradish peroxidase and chemiluminescent substrate according to previous publication(19).

The reporter plasmid carrying Gaussia luciferase (Gluc) under the control of the mouse *Aldh1a1* promoter (*Aldh1a1p-Gluc*) (NM\_027406) was from Genecopoeia (Rockville, MD). This plasmid also expressed secreted alkaline phosphatase (SEAP) that enabled transfection normalization. The *Aldh1a1p-Gluc* plasmid was co-transfected in DMEM medium carrying 10% FBS together with constructs that express a human YAP2 protein or the YAP2<sup>S127A</sup> mutant in which the phosphorylation site at serine 127 residue was mutated to alanine (Addgene, Watertown, MA). GLuc activity was measured in proliferating cells at 24 hours post transfection using Secrete-Pair™ Dual Luminescence Assay Kit (Genecopoeia).

### In vivo bioluminescence imaging

The mouse *Aldh1a1* promoter was cloned to the 5' region of a firefly luciferase gene at KpnI and BamHI restriction sites in AAV vector. The resulting clone was transfected together with pACG2c8 and phelper plasmids into AAV293 cells (Cell Biolabs, San Diego, CA). AAV8 virus expressing *Aldh1a1* promoter-driven luciferase (AAV8-*Aldh1a1p-luc*) was generated and purified. AAV8-*Aldh1a1p-luc* ( $1 \times 10^{12}$  pfu/mouse) was administrated into mouse livers through tail vein injection. Three weeks later, animals were subjected to chronic-ethanol-plus single binge according to Bertola et al(18). Thirty-six hours before the binge, the plasmid carrying FLAG:YAP2<sup>S127A</sup> (10 ug/mouse) was delivered into mice through hydrodynamic tail vein injection. AAV8-*Aldh1a1p-luc* activities were measured at 0 or 9 hours after the binge by IVIS *in vivo* imaging system (Perkin Elmer, Waltham, MA) after intraperitoneal injection of D-luciferin (150 mg/kg body weight) (ThermoFisher Scientific).

### Statistical Analysis

GraphPad Prism 6.0 (GraphPad Software, San Diego CA) was used for statistical analysis. Statistical significance ( $P < 0.05$ ) was evaluated using the unpaired *t*-test and one-way analysis of variance (ANOVA).

## RESULTS

### YAP activation during ALD progression or in experimental models of ethanol-associated liver injury

Alcohol abuse can cause a broad spectrum of liver pathology starting from fat accumulation, alcoholic steatohepatitis (ASH), to alcoholic cirrhosis (AC). We examined expression pattern of YAP protein in total liver homogenates from six ALD patients who suffered from alcoholic steatosis (AS), ASH, and AC. Elevated levels of total YAP protein were found

in all tested ALD livers in comparison to two normal controls (Figure 1A). Given the fact that YAP is inactivated through phosphorylation, we also determined the levels of inactive YAP protein using a specific antibody that recognized the phosphorylation site at serine 127. The inactive phosphorylated YAP was fluctuated in the ALD livers compared to normal controls, but percentage of the active nonphosphorylated form were significantly increased in total liver homogenates of patients with AS, ASH, and AC after quantitative analysis using densitometry (Figure 1B). Further IHC in ASH livers showed appearance of YAP in parenchyma of ASH livers (Figure 1C), although there was constitutive expression of this protein in normal portal tracts (Supplemental Figure 1). These results demonstrated YAP activation during ALD progression.

The NIAAA mouse model of chronic-plus-single-binge ethanol feeding mimics hepatocyte damage, inflammation and steatosis in patients with acute-on-chronic alcoholic liver injury(18). We utilized this model and fed C57BL6 mice 5% ethanol-containing liquid diet followed by ethanol binge. Then RNA-Seq was performed to identify genes or pathways that were differentially expressed at 9 hours after binge in comparison to the livers without binge in ethanol-fed mice. Functional enrichment analysis detected upregulation of gene signatures for triglyceride catabolic process, smooth muscle cell proliferation, inflammatory response, and Hippo signaling pathway, whereas genes in oxidation-reduction process and retinol metabolism were downregulated in the ethanol/binge-treated livers (Table 1 and Supplemental Table 1). To monitor active Yap content, we isolated nuclear fractions from total liver homogenates of mice that were exposed to ethanol-fed/binge or pair-fed/water. As shown in Figure 1D, accumulation of nuclear Yap was observed at 4 and 9 hours post binge in ethanol-fed livers but not in controls. IHC demonstrated Yap accumulation in pericentral hepatocytes where ethanol was presumably metabolized in these ethanol-fed/binge livers (Figure 1E).

CCl<sub>4</sub> and ethanol are metabolized in pericentral hepatocytes and induce hypoxia, oxidative stress, and hepatic damage. Moderate ethanol feeding in combination with CCl<sub>4</sub> has been modeled in mice to investigate the effect of ethanol on hepatic apoptosis, liver regeneration, and fibrosis(22, 23). We have demonstrated that the ethanol/CCl<sub>4</sub> cotreatment can induce Yap in damaged livers(19). To further determine active Yap content, we extracted nuclear fractions from total liver homogenates and detected elevation of nuclear Yap protein within 2 days post CCl<sub>4</sub> intoxication in ethanol-fed livers (Figure 1D). Furthermore, IHC labeled plenty of nuclear Yap protein in pericentral hepatocytes within the first day post the chemical induced liver injury (Figure 1E). These observations indicated accumulation of active Yap in pericentral hepatocytes during ethanol-associated liver injury.

### **Hepatocyte-specific deletion of Yap1 reduces hepatocyte proliferation and cellular reprogramming to progenitor phenotypes during ethanol/CCl<sub>4</sub>-induced liver injury**

Hepatocytes are the main parenchymal cells to mediate liver regeneration. They harbor a heterogeneous capacity to proliferate. Subpopulations of pericentral hepatocytes express some early liver progenitor markers, such as *Tbx3* and *Axin2*, and show self-renewal abilities during homeostasis(24). On the other hand, Hnf4 $\alpha$ <sup>+</sup> periportal hepatocytes can reconstitute the livers after chronic injury, such as CCl<sub>4</sub> intoxication(25). Hnf4 $\alpha$  was



deregulated in damaged hepatocytes after ethanol/CCl<sub>4</sub>-induced damage(19). To explore *Yap* function in regulation of hepatocyte proliferation and progenitor activation, we generated hepatocyte specific KO or controls through transduction of equal amounts of AAV8-TBG-iCre or AAV8-GFP viruses into *Yap1*<sup>fl<sup>ox</sup>/fl<sup>ox</sup></sup> homozygotes prior to moderate ethanol feeding and acute CCl<sub>4</sub> intoxication. Expression of *iCre* and *GFP* transgenes, and loss of *Yap1* at mRNA and protein levels were detected in KO by RT-PCR analysis, immunoblotting, and IHC (Figure 2A–C). Q-RT-PCR and immunoblotting analyses demonstrated significant reduction of *Ccnd1* mRNA and Cyclin protein in the hepatocyte specific KO (Figure 2B and C). IHC revealed significantly enhanced loss of Hnf4α<sup>+</sup> periportal populations and exacerbated necrosis in liver parenchyma of the damaged hepatocyte-specific KO. There was concomitant reduction of hepatocyte proliferation as indicated by the decreased number of Ki67<sup>+</sup> proliferating hepatocytes in the KO compared with controls (Figure 2C). RNA-Seq and functional enrichment analysis demonstrated downregulation of progenitor-related markers including *Epcam*, *Lgr5*, *Axin*, *Tbx3*, and the biliary marker *cytokeratin 19 (Krt19)* in the damaged *Yap1* KO. Q-RT-PCR analysis verified 33% downregulation of *Epcam*, 87% decrease of *Lgr5*, 58% reduction of *Axin2*, 59% loss of *Tbx3*, and 82% decrease of *Krt19* transcript in damaged *Yap1* KO (Figure 2D). Collectively, these results indicated that *Yap* loss affected hepatocyte proliferation and cellular programming that regulated progenitor-related genes during liver regeneration following ethanol/CCl<sub>4</sub> cotreatment.

### Hepatocyte-specific deletion of *Yap1* enhances hemorrhages, hypoxia, aberrant extracellular matrix (ECM) remodeling, and recruitment of CD11b<sup>+</sup> inflammatory cells during ethanol/CCl<sub>4</sub>-induced liver injury

Hemorrhages can result from a breach in the hepatic parenchyma. Consistent with their impaired regeneration, the *Yap1* KO developed morphologically distinguishable hemorrhage at 48 hours post CCl<sub>4</sub> intoxication after moderate ethanol pre-exposure (Figure 3A). This hemorrhage was associated with abnormal ECM remodeling as detected by dual staining for the endothelial marker vWF and the activated hepatic stellate cell (HSC) marker αSMA. Spontaneous lesions positive for vWF and αSMA staining were found around pericentral zones only in the damaged *Yap1* KO livers at 7 hours and became more evident than controls at the 48-hour time point (Figure 3A). Since ethanol metabolism consumes oxygen and causes hypoxia leading to the stabilization of the oxygen-sensitive transcription factor Hif1α(26), we performed IHC for the hypoxia marker pimonidazole and found greater stained areas in ethanol-fed *Yap1* KO livers than controls at 7 and 48 hours post CCl<sub>4</sub> (Figure 3A). RNA-Seq and functional enrichment analysis detected upregulation of gene signatures for hypoxia, angiogenesis, smooth muscle cell proliferation, and cell migration (Table 1). Q-RT-PCR analysis showed that *Yap1* KO livers had about 2.31-fold increase of *Hif1α*, 2.91-fold upregulation of *Vegf-a*, 4.83-fold increase of *Ctgf*, 2.15-fold elevation of *Fn1*, 2.41-fold increase of *Col4a1*, 2.13-fold upregulation of *Col4a2*, and 2.43-fold induction of *Itgam* that encodes integrin αM subunit for CD11b (Figure 3B). Accordingly, Hif1α, Vegf-a, Ctgf, Fn1, and Itgam proteins were elevated, and these elevations were confirmed after quantifications of band intensities based on Western analyses (Figure 3C). Further Sirius Red staining revealed larger areas of collagen deposition near perivenous areas in the ethanol-fed *Yap1* KO livers than CT controls at 48 hours post CCl<sub>4</sub> (Supplemental Figure 2). Enhanced ECM remodeling in these damaged *Yap1* KO livers was also verified after the

immunofluorescent staining for collagen IV in Supplemental Figure 3. We then monitored inflammatory cell infiltration based on IHC for CD11b since this cell surface protein is an adhesion receptor for Ctgf(27). CD11b<sup>+</sup> inflammatory cells were tightly associated with damaged pericentral hepatocytes, whereas the majority of CD11b<sup>+</sup> inflammatory cells were in Ctgf-enriched parenchyma of *Yap1* KO livers where integrity of pericentral veins were disrupted (Figure 3D). M2-type macrophages have been shown to control tissue repair by increasing the expression of growth factors such as Vegf-a and transforming growth factor (Tgf)- $\beta$ (28, 29). Therefore, we examined the extent of M2 macrophage activation based on the immunofluorescent staining for cluster of differentiation 163 (CD163) protein. CD163 is a known marker for activated M2 macrophages and modulates inflammatory responses during physio-pathological conditions in many organs including the livers(30–32). More CD163<sup>+</sup> activated M2 macrophages were found in the ethanol-fed *Yap1* KO livers than CT controls at 48 hours post CCl<sub>4</sub> (Supplemental Figure 4). These observations indicated that *Yap1* deficiency was associated with enhancement of hemorrhages, hypoxia, activation of stromal cells (vascular endothelial cells, HSC, and inflammatory cells), and aberrant ECM remodeling.

### Hepatocyte-specific deletion of Yap1 enhances susceptibility to hepatotoxicity induced by ethanol/CCl<sub>4</sub> cotreatment

Transient accumulation of Yap was found in peri-central hepatocytes at 7 hours post the acute intoxication and in periportal hepatocytes at 48 hours post ethanol/CCl<sub>4</sub>, whereas the hepatocyte-derived *Yap* was lost in KO that received the same treatment during these time points (Figure 4A). IHC revealed 18% increase of areas that were positive for Yap staining in the damaged CT livers at 48 hours compared with those at 7 hours post ethanol/CCl<sub>4</sub>. KO livers lost this increase and stained negative for Yap in hepatocytes at both time points. The reactive aldehyde HNE is a major bioactive byproduct of lipid peroxidation and oxidative stress generated during metabolism of chemicals such as ethanol and CCl<sub>4</sub>. Increased intensity and areas positive for HNE staining were observed as indicated by in damaged livers of the ethanol-fed *Yap1* KO at 7 to 48 hours post CCl<sub>4</sub> (Figure 4A). Areas that were positive for HNE staining in the ethanol-fed KO livers had 10 to 26% increases in comparison to CT at 7 to 48 hours post CCl<sub>4</sub>. This enhancement of lipid peroxidation was verified based on quantification of hepatic MDA concentration ( $0.36 \pm 0.02$  in the ethanol pre-exposed *Yap1* KO versus  $0.24 \pm 0.04$  nmol/mg protein in controls) at 48 hours post the chemical induced injury (Figure 4B). Increased ROS production was observed ( $12.01 \pm 1.50$  pmol/min/mg protein in the KO livers versus  $5.81 \pm 0.51$  pmol/min/mg protein in controls) (Figure 4C), indicating stronger oxidative stress in the *Yap1* KO. Elevation of hepatic acetaldehyde was also found ( $89 \pm 21$  nmol/mg in the *Yap1* KO livers versus  $48 \pm 14$  nmol/mg in the control livers) (Figure 4D), indicating a slower removal of this toxic intermediate in absence of *Yap* during ethanol/CCl<sub>4</sub> combined treatment. *Adh1*, *Aldh1a1* and *Dhrs3* encode cytosolic enzymes in ethanol and retinal metabolisms. There was a 1.81-fold decrease of cytosolic Aldh activity in the damaged KO livers (Figure 4E). Additional qRT-PCR and immunoblotting analyses demonstrated that the *Yap1* KO livers had downregulation of *Adh1*, *Aldh1a1*, and *Dhrs3* at mRNA and protein levels (Figure 4F and G). Further functional enrichment analysis detected 3.615-fold decrease of the oxidation-reduction pathway (p=6.78E-12) and 7.64-fold reduction of retinal metabolism

( $p=2.47E-08$ ) in the ethanol-fed *Yap1* KO livers at 48 hours post CCl<sub>4</sub> intoxication (Table 1). Related genes in the corresponding pathways identified in the functional enrichment analysis were listed in Supplemental Table 1. Collectively, these results indicated that hepatocyte-specific deletion of the *Yap1* gene enhanced susceptibility to hepatotoxicity during ethanol/CCl<sub>4</sub>-induced injury, likely due to alteration of detoxification pathways for these chemicals.

### The *Aldh1a1* promoter contains a cis-element for YAP and can be induced by a hyperactive mutant of this transcriptional coactivator

The human *ALDH1A1* promoter contains a cis-acting element for TEAD binding in a TAZ-dependent manner(33). Since YAP and TAZ are paralogous proteins with overlapping functions, we examined the mouse *Aldh1a1* promoter (*Aldh1a1p*) and identified a putative YAP/TEAD binding site “GGAATG” located at –317 to –322 upstream of the transcription initiation site (Figure 5A). To test if Yap is able to be associated with this region, we performed EMSA assays using a biotinylated probe containing the consensus sequences and incubated it with nuclear extracts of HEK293 cells that overexpressed the murine Yap:Myc protein. This biotinylated *Aldh1a1p* probe bound to Yap:Myc protein and formed complexes, which were detected as a shift band in gel shift assays. The complexes could be disrupted with addition of excessive cold wild-type probe (Figure 5B). Moreover, the interaction of *Aldh1a1p* with Yap:Myc was specific because a biotinylated mutant probe in which the putative consensus sequences were replaced to a SpeI site “actagt” was unable to form complexes (Figure 5A and B).

The regulation of *Aldh1a1* promoter was further examined using YAP and mutant isoforms. Phosphorylation site at serine residue 127 has been known to control YAP cytoplasm retention. Abolishment of this phosphorylation site by alanine substitution gives rise to a hyperactive YAP<sup>S127A</sup> that escapes cytoplasmic sequestration and constitutively turns on signaling in nucleus(34, 35). YAP comprises two major splicing isoforms, YAP1 containing a single WW domain and YAP2 with two WW domains(36, 37). YAP2 was chosen for analysis in this study because it shares a similar protein structure with its mouse counterpart, which contains two WW domains(38). We tested effects of YAP2 protein or the YAP2<sup>S127A</sup> mutant on *Aldh1a1* promoter activity. The two proteins were tagged with FLAG epitope at their N terminal regions (FLAG:YAP2 and FLAG:YAP2<sup>S127A</sup>). SEAP was co-expressed as internal control for normalization of transfection efficiency in the luciferase systems. Dual reporter analysis showed that the *Aldh1a1* promoter activity as indicated by ratio between Gluc and SEAP could be induced by the FLAG:YAP2<sup>S127A</sup> mutant in the rat hepatic progenitor cell line WB-F344 and the human hepatocellular carcinoma cell line Huh7 (Figure 5C). Paradoxically, slight reduction of *Aldh1a1* promoter mediated by FLAG:YAP2 was also observed (Figure 5C). Considering similar overexpression of FLAG:YAP2 and FLAG:YAP2<sup>S127A</sup> proteins in the two types of cells (Figure 5D), these different patterns indicated that FLAG:YAP2 mediated negative regulation of *Aldh1a1* promoter through unknown mechanisms, whereas the hyperactive FLAG:YAP2<sup>S127A</sup> protein transcriptionally activated this detoxification gene. To further verify the regulation of *Aldh1a1* promoter by FLAG:YAP2<sup>S127A</sup> protein *in vivo*, we generated AAV8-*Aldh1a1p*-luc virus and transduced it into mouse livers. A strong luciferase activity driven by *Aldh1a1* promoter was specifically found in transduced livers (Figure 5E). We then performed

bioluminescence imaging and measured change of *Aldh1a1p* activity at 0 and 9 hours after binge in ethanol-fed mice that expressed FLAG:YAP2<sup>S127A</sup>. Quantitative analysis showed an increase of AAV8-*Aldh1a1p*-luc activity in all three tested mice according to elevated bioluminescent signals at 9 hours after binge (Figure 5F). These data supported our observations that *Aldh1a1* promoter could be transcriptionally activated by YAP2<sup>S127A</sup>.

### **Ectopic ALDH1A1 promotes hepatocyte proliferation, protects the livers from hepatotoxicity, reduces ECM remodeling, and attenuates recruitment of CD11b<sup>+</sup> inflammatory cells during ethanol/CCl<sub>4</sub>-induced liver damage**

Previous reports show potential roles of ALDH1A1 to prevent oxidative stress-induced pathology due to its remarkably low Km for HNE and MDA(8). To study *in vivo* function of this enzyme, we ectopically expressed AAV8-ALDH1A1:FLAG in ethanol-fed livers that received one acute dose of CCl<sub>4</sub>. Elevation of *ALDH1A1* mRNA and protein were confirmed at 48 hours after the chemical-induced liver damage compared to AAV-GFP infected controls (Figure 6A and B). ALT level in sera was also significantly reduced ( $480.0 \pm 86.86$  in the AAV8-ALDH1A1:FLAG infected mice versus  $668.3 \pm 96.43$  Unit/L in AAV8-GFP infected controls) (Figure 6C). ROS production in the AAV8-ALDH1A1:FLAG-infected livers had about 22% decrease (Figure 6D). Lower levels of hepatic MDA ( $0.075 \pm 0.002$  nmol/mg protein in AAV8-ALDH1A1:FLAG infected livers versus  $0.240 \pm 0.030$  nmol/mg protein in AAV8-GFP infected controls) were also observed (Figure 6E), indicating ectopic ALDH1A1 ameliorated lipid peroxidation. Further IHC confirmed that ectopic ALDH1A1 was associated with decreased necrosis in H&E staining, reduced lipid peroxidation in HNE staining, and attenuated numbers of proliferating hepatocytes in Ki67 staining (Figure 6F). Consistent with the increased hepatocyte proliferation in AAV8-ALDH1A1:FLAG-transduced livers, we detected increased levels of *Ccnd1* mRNA and Cyclin D1 protein (Figure 6A and B). Significant reduction of *Ctgf*, *Fn1*, and *Itgam* at mRNA and protein levels were observed after ectopic expression of ALDH1A1:FLAG protein (Figure 6A and B). Further immunofluorescent staining for  $\alpha$ SMA, vWF, and CD11b demonstrated reduction in ECM remodeling and infiltration of inflammatory cells in the ethanol/CCl<sub>4</sub>-damaged livers (Figure 6F). Additional Sirius Red staining verified less collagen deposition near perivenous areas in the AAV8-ALDH1A1:FLAG-transduced ethanol-fed livers than controls at 48 hours post CCl<sub>4</sub> (Supplemental Figure 5). Areas that were immunoreactive for collagen IV were also smaller in the livers that expressed ectopic ALDH1A1 than controls (Supplemental Figure 6). Collectively, these data indicated that ectopic ALDH1A1 promoted liver regeneration, protected the transduced livers from hepatocyte death and oxidative stress, attenuated ECM remodeling, and reduced inflammation during ethanol/CCl<sub>4</sub>-induced liver injury.

## **DISCUSSION**

YAP is an important regulator in liver regeneration following chemical and surgical-induced damage. Consistent with our recent findings(19), this study demonstrated Yap activation upon hepatocyte damage caused by insults such as ethanol and CCl<sub>4</sub>. Impaired regenerative responses were observed in hepatocyte-specific *Yap1* KO as summarized in Figure 7. Su et al have also characterized hepatocyte-specific KO in which the murine *Yap* gene was

deleted using albumin-promoter-driven Cre during liver damage after a single injection of CCl<sub>4</sub>(39). They have found that *Yap* null hepatocytes proliferate significantly less than their WT counterparts throughout the repair phase at days 1–3 after the injury. In bile duct ligation-induced injury, the liver-specific *Yap* deletion suppresses hepatocyte proliferation, enhances hepatocyte necrosis, and compromises bile duct proliferation(40). Active human YAP is also accumulated in duct cells and parallels fibrosis progression in non-alcoholic fatty liver disease(41). Similarly, deletion of *Yap/Taz* genes in murine livers has been known to reduce liver regeneration after partial hepatectomy owing to reduced ability of cell cycle entry and hepatocyte proliferation(42, 43). Moreover, YAP can regulate hepatocyte fate determination during injury(44). Additionally, YAP has been considered as a stress sensor to maintain homeostasis in response to hepatic insults such as ethanol by eliminating injured cells(13). In presence of second signal such as tissue damage or inflammation, YAP switches this homeostatic function from injured cell elimination to growth promotion leading to clonal expansion and hepatocyte reprogramming into progenitor phenotypes for liver regeneration(41, 45). Notably, YAP protein levels and phosphorylation seemed to correlate with alcoholic liver disease severity when we characterized its expression patterns in AS, ASH, and AC patients in this study. YAP's roles from cell elimination to growth stimulation likely occur during the ALD progression and these diverse functions explain dynamic distributions of this protein during liver injury caused by CCl<sub>4</sub> and ethanol in this study. We found *Yap* induction in pericentral hepatocytes at 7 hours post CCl<sub>4</sub> and at 9 hours post chronic-ethanol-plus-single binge, whereas this molecule was re-distributed in periportal zones at 48 hours post CCl<sub>4</sub> in ethanol-fed wild type livers. Considering that pericentral zones are damaged areas where CCl<sub>4</sub> and ethanol are metabolized, it is conceivable that the pericentral-derived *Yap* functions to selectively eliminate injured hepatocytes and the periportal-derived *Yap* works to promote hepatocyte proliferation for liver regeneration in response to these chemical-induced injuries.

Another important finding of this study is about potential roles of hepatocyte-derived *Yap* against hypoxia, stromal cell activation and inflammatory cell infiltration. Abnormal ECM remodeling has been reported in mice deficient for *Yap* after CCl<sub>4</sub> intoxication(39). Consistent with the previous report, this study demonstrated enhancement of necrosis and severe hypoxia as well as abnormal ECM remodeling and inflammatory cell infiltration in hepatocyte-specific *Yap1* KO after ethanol/CCl<sub>4</sub> treatment. Hepatotoxins such as ethanol and CCl<sub>4</sub> cause hypoxia in the hepatic pericentral area due to competitive oxygen consumption by their metabolisms. HIF1 $\alpha$  is normally degraded but becomes stabilized in response to low oxygen tension and pro-inflammatory signals. Active HIF1 $\alpha$  is translocated into the nucleus and induces expression of pro-angiogenic genes including *Vegf-a*, *Ctgf*, and *fibronectin* for ECM remodeling. Upregulation of *Hif1 $\alpha$*  and its downstream targets in angiogenesis might result from increased hepatotoxicity due to downregulation of detoxification enzymes (*Aldh1*, *Aldh1a1* and *Dhrs3*) in *Yap* null hepatocytes as shown in Figure 7. In addition, emerging evidence has shown a link of *Yap* regulation with oxidative stress. Weak oxidative stress defense can result from *Yap* decrease due to acetaminophen-induced glutathione depletion(46). *Yap* acts as a nuclear co-factor and induces antioxidant genes leading to enhanced cardiomyocyte survival and reduced oxidative stress after ischemia/reperfusion(47). *Yap* overexpression protects

cardiomyocytes against H<sub>2</sub>O<sub>2</sub>-induced cell death, whereas *Yap1* deficiency exacerbates injury in response to chronic myocardial infarction(48). Therefore, it is easy to postulate that transcriptional regulation of antioxidant and detoxification enzymes by Yap provides protective mechanisms against hypoxia and cellular toxicity caused by lipid peroxidation/ROS-triggered signaling during tissue injury.

Ethanol abuse causes alcoholic liver disease through oxidative stress, acetaldehyde toxicity, and altered retinoid metabolism. This study demonstrated that forced expression of ALDH1A1 reduced hepatotoxicity by attenuating production of ROS, lipid peroxidation products (HNE, MDA et c), and acetaldehyde. ALDH1A1 can not only detoxify acetaldehyde in ethanol oxidation but also convert retinaldehyde into retinoic acid (RA) (49). RA is a potent inducer of hepatocyte proliferation(50, 51). This RA-mediated action may contribute to the enhanced proliferation of hepatocytes observed in AAV8-ALDH1A1:FLAG-transduced livers after ethanol/CCl<sub>4</sub> treatment. Furthermore, *Dhrs3* is also involved in RA metabolism. This enzyme converts retinaldehyde into retinol and inhibits RA synthesis(52, 53). Interestingly, the promoter of *Dhrs3* gene contains regulatory elements for YAP/TEAD binding(53). Thus, YAP may regulate both *Aldh1a1* and *Dhrs3* promoters. Loss of this regulation may explain why *Aldh1a1* and *Dhrs3* as well as retinol metabolism (mmu00830) were downregulated in *Yap1* KO livers during ethanol/CCl<sub>4</sub>-induced liver injury.

Aldehydes may arise from metabolisms of amino acids, alcohols, lipids, vitamins, cytotoxic drugs, and environmental agents in the liver. These toxic intermediates easily engage in numerous secondary reactions that attack cells, resulting in protein or DNA adduct formation as well as lipid peroxidation, with varying degrees of toxicity at the site of generation or when transported to distant cells. The liver serves as the primary metabolism site and produces enzymes such as *Aldh1a1* for hepatic protection after injury. High levels of *Aldh* isoforms are expressed in mouse livers, whereas *Aldh1a1* deficiency in Hepa-1c1c7 cells enhances susceptibility to aldehyde-induced cell death, accumulation of acrolein-protein adducts, caspase 3 activation and generating oxidative stress(7). Moreover, regulation of *ALDH1A1* by YAP and TAZ in some cancer cells has been reported(33, 54). Taking into considerations that ALDH activity is often altered in cirrhotic livers of AC patients(55), understanding the regulation of ALDH1A1 by YAP has therapeutic implications against impaired liver regeneration and ethanol-induced hepatotoxicity during development of ALD.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## ACKNOWLEDGMENT

We want to thank Ms. Marda Jorgensen and Haven Thomas for technical assistance about immunohistochemistry. We also thank Drs. Chen Ling and Gai Ran in AAV production.

### Financial Support

This study is supported by National Institutes of Health NIAAA KO1AA024174 and RO1AA028035 grants as well as Children Miracle Research Foundation grant awarded to LP. This study is also partially supported by the National Natural Science Foundation of China (82060124), the Natural Science Foundation of Guangxi (2019JJA140718), and the Guangxi Distinguished Experts Special Fund (2019B12).

### Data Availability Statement

All data supporting the findings of this study are available within the paper and within its supplementary materials published online.

### Abbreviations

<b>Yap</b>	Yes-associated protein
<b>p-YAP</b>	phosphorylated Yap at serine 127 residue
<b>CCl<sub>4</sub></b>	carbon tetrachloride
<b>iCre</b>	codon-improved Cre recombinase
<b>ctgf</b>	connective tissue growth factor
<b>ADH</b>	alcohol dehydrogenase
<b>ALDH</b>	aldehyde dehydrogenases
<b>ROS</b>	reactive oxygen species
<b>ALD</b>	alcohol-associated liver disease
<b>TBG</b>	thyroxine-binding globulin
<b>KOMP</b>	knockout Mouse Project
<b>GEO</b>	Gene Expression Omnibus
<b>HNE</b>	4-hydroxynonenal
<b>Hnf4<math>\alpha</math></b>	hepatocyte nuclear factor 4 $\alpha$
<b>vWF</b>	von Willebrand factor
<b>Hif1<math>\alpha</math></b>	hypoxia-inducible factor 1 $\alpha$
<b>Itgam</b>	Integrin subunit alpha M
<b>Vegf-a</b>	vascular endothelial growth factor-a
<b>ECM</b>	extracellular matrix
<b>Dhrs3</b>	dehydrogenase/Reductase 3
<b>AS</b>	alcoholic steatosis
<b>ASH</b>	alcoholic steatohepatitis

<b>AC</b>	alcoholic cirrhosis
<b>MDA</b>	malondialdehyde
<b>HSC</b>	hepatic stellate cell
<b><math>\alpha</math>SMA</b>	alpha smooth muscle actin
<b>Tbx3</b>	T-Box 3
<b>Axin2</b>	Axin-related protein 2
<b>Epcam</b>	epithelial cell adhesion molecule
<b>Lgr5</b>	Leucine rich repeat containing G protein-coupled receptor 5
<b>Krt19</b>	cytokeratin 19
<b>WWTR1</b>	the WW domain-containing transcription regulator protein 1
<b>TAZ</b>	transcriptional co-activator with PDZ-binding motif
<b>CYP2E1</b>	cytochrome P450
<b>GFP</b>	Green fluorescence protein
<b>KO</b>	Knockout
<b>CBA</b>	chicken $\beta$ -actin
<b>pfu</b>	plaque forming unit

## REFERENCES

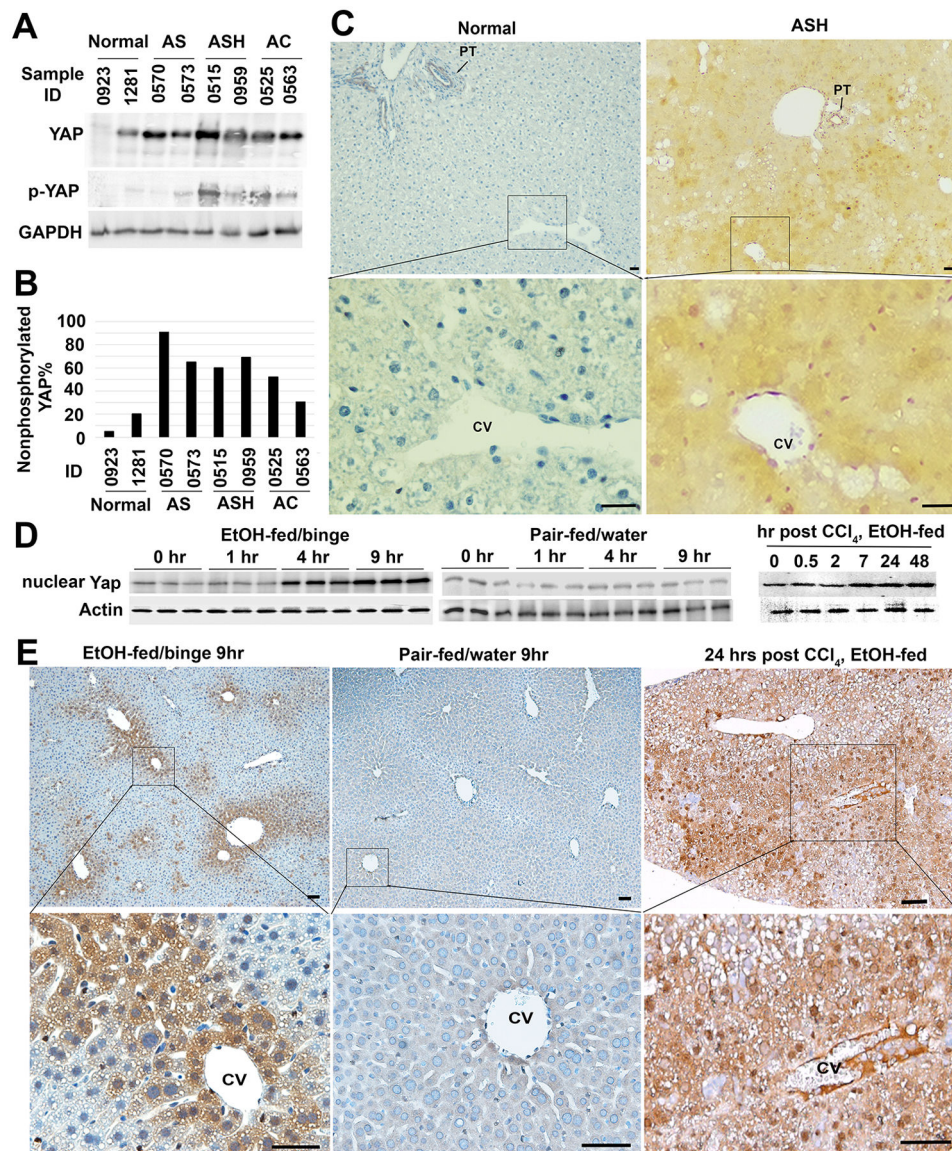
1. Lieber CS, and DeCarli LM (1968) Ethanol oxidation by hepatic microsomes: adaptive increase after ethanol feeding. *Science* 162, 917–918 [PubMed: 4386718]
2. Chen Y, Thompson DC, Koppaka V, Jester JV, and Vasiliou V (2013) Ocular aldehyde dehydrogenases: protection against ultraviolet damage and maintenance of transparency for vision. *Prog Retin Eye Res* 33, 28–39 [PubMed: 23098688]
3. Marchitti SA, Deitrich RA, and Vasiliou V (2007) Neurotoxicity and metabolism of the catecholamine-derived 3,4-dihydroxyphenylacetaldehyde and 3,4-dihydroxyphenylglycolaldehyde: the role of aldehyde dehydrogenase. *Pharmacol Rev* 59, 125–150 [PubMed: 17379813]
4. Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, Brown M, Jacquemier J, Viens P, Kleer CG, Liu S, Schott A, Hayes D, Birnbaum D, Wicha MS, and Dontu G (2007) ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. *Cell Stem Cell* 1, 555–567 [PubMed: 18371393]
5. Steg AD, Bevis KS, Katre AA, Ziebarth A, Dobbin ZC, Alvarez RD, Zhang K, Conner M, and Landen CN (2012) Stem cell pathways contribute to clinical chemoresistance in ovarian cancer. *Clin Cancer Res* 18, 869–881 [PubMed: 22142828]
6. Suzuki E, Chiba T, Zen Y, Miyagi S, Tada M, Kanai F, Imazeki F, Miyazaki M, Iwama A, and Yokosuka O (2012) Aldehyde dehydrogenase 1 is associated with recurrence-free survival but not stem cell-like properties in hepatocellular carcinoma. *Hepatology* 55, 1100–1111 [PubMed: 22583771]
7. Makia NL, Bojang P, Falkner KC, Conklin DJ, and Prough RA (2011) Murine hepatic aldehyde dehydrogenase 1a1 is a major contributor to oxidation of aldehydes formed by lipid peroxidation. *Chem Biol Interact* 191, 278–287 [PubMed: 21256123]



8. Rebolledo-Rios R, Venton G, Sánchez-Redondo S, Iglesias IFC, Fournet G, González E, Romero Fernández W, Borroto Escuela DO, Di Stefano B, Penarroche-Díaz R, Martin G, Ceylan I, Costello R, and Perez-Alea M (2020) Dual disruption of aldehyde dehydrogenases 1 and 3 promotes functional changes in the glutathione redox system and enhances chemosensitivity in nonsmall cell lung cancer. *Oncogene* 39, 2756–2771 [PubMed: 32015486]
9. Vasiliou V, and Pappa A (2000) Polymorphisms of human aldehyde dehydrogenases. Consequences for drug metabolism and disease. *Pharmacology* 61, 192–198 [PubMed: 10971205]
10. Meng Z, Moroishi T, and Guan KL (2016) Mechanisms of Hippo pathway regulation. *Genes Dev* 30, 1–17 [PubMed: 26728553]
11. Zhao B, Ye X, Yu J, Li L, Li W, Li S, Yu J, Lin JD, Wang CY, Chinnaiyan AM, Lai ZC, and Guan KL (2008) TEAD mediates YAP-dependent gene induction and growth control. *Genes Dev* 22, 1962–1971 [PubMed: 18579750]
12. Yimlamai D, Christodoulou C, Galli GG, Yanger K, Pepe-Mooney B, Gurung B, Shrestha K, Cahan P, Stanger BZ, and Camargo FD (2014) Hippo pathway activity influences liver cell fate. *Cell* 157, 1324–1338 [PubMed: 24906150]
13. Miyamura N, Hata S, Itoh T, Tanaka M, Nishio M, Itoh M, Ogawa Y, Terai S, Sakaida I, Suzuki A, Miyajima A, and Nishina H (2017) YAP determines the cell fate of injured mouse hepatocytes in vivo. *Nat Commun* 8, 16017 [PubMed: 28681838]
14. Liu Y, Lu T, Zhang C, Xu J, Xue Z, Busuttill RW, Xu N, Xia Q, Kupiec-Weglinski JW, and Ji H (2019) Activation of YAP attenuates hepatic damage and fibrosis in liver ischemia-reperfusion injury. *J Hepatol* 71, 719–730 [PubMed: 31201834]
15. Mo JS, Meng Z, Kim YC, Park HW, Hansen CG, Kim S, Lim DS, and Guan KL (2015) Cellular energy stress induces AMPK-mediated regulation of YAP and the Hippo pathway. *Nature cell biology* 17, 500–510 [PubMed: 25751140]
16. Enzo E, Santinon G, Pocaterra A, Aragona M, Bresolin S, Forcato M, Grifoni D, Pession A, Zanconato F, Guzzo G, Biciato S, and Dupont S (2015) Aerobic glycolysis tunes YAP/TAZ transcriptional activity. *The EMBO journal* 34, 1349–1370 [PubMed: 25796446]
17. Bou Saleh M, Louvet A, Ntandja-Wandji LC, Boleslawski E, Gnemmi V, Lassailly G, Truant S, Maggiotto F, Ningarhari M, Artru F, Anglo E, Sancho-Bru P, Corlu A, Argemi J, Dubois-Chevalier J, Dharancy S, Eeckhoutte J, Bataller R, Mathurin P, and Dubuquoy L (2021) Loss of hepatocyte identity following aberrant YAP activation: A key mechanism in alcoholic hepatitis. *J Hepatol* 75, 912–923 [PubMed: 34129887]
18. Bertola A, Mathews S, Ki SH, Wang H, and Gao B (2013) Mouse model of chronic and binge ethanol feeding (the NIAAA model). *Nat Protoc* 8, 627–637 [PubMed: 23449255]
19. Zhou J, Sun X, Yang L, Wang L, Ran G, Wang J, Cao Q, Wu L, Bryant A, Ling C, and Pi L (2020) Hepatocyte nuclear factor 4alpha negatively regulates connective tissue growth factor during liver regeneration. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 34, 4970–4983 [PubMed: 32057145]
20. Pi L, Robinson PM, Jorgensen M, Oh SH, Brown AR, Weinreb PH, Trinh TL, Yianni P, Liu C, Leask A, Violette SM, Scott EW, Schultz GS, and Petersen BE (2015) Connective tissue growth factor and integrin alphavbeta6: a new pair of regulators critical for ductular reaction and biliary fibrosis in mice. *Hepatology* 61, 678–691 [PubMed: 25203810]
21. Varghese F, Bukhari AB, Malhotra R, and De A (2014) IHC Profiler: an open source plugin for the quantitative evaluation and automated scoring of immunohistochemistry images of human tissue samples. *PloS one* 9, e96801 [PubMed: 24802416]
22. Chiang DJ, Roychowdhury S, Bush K, McMullen MR, Pisano S, Niese K, Olman MA, Pritchard MT, and Nagy LE (2013) Adenosine 2A receptor antagonist prevented and reversed liver fibrosis in a mouse model of ethanol-exacerbated liver fibrosis. *PLoS One* 8, e69114 [PubMed: 23874883]
23. Deshpande KT, Liu S, McCracken JM, Jiang L, Gaw TE, Kaydo LN, Richard ZC, O’Neil MF, and Pritchard MT (2016) Moderate (2%, v/v) Ethanol Feeding Alters Hepatic Wound Healing after Acute Carbon Tetrachloride Exposure in Mice. *Biomolecules* 6, 5 [PubMed: 26751492]
24. Wang B, Zhao L, Fish M, Logan CY, and Nusse R (2015) Self-renewing diploid Axin2(+) cells fuel homeostatic renewal of the liver. *Nature* 524, 180–185 [PubMed: 26245375]

25. Font-Burgada J, Shalapour S, Ramaswamy S, Hsueh B, Rossell D, Umemura A, Taniguchi K, Nakagawa H, Valasek MA, Ye L, Kopp JL, Sander M, Carter H, Deisseroth K, Verma IM, and Karin M (2015) Hybrid Periportal Hepatocytes Regenerate the Injured Liver without Giving Rise to Cancer. *Cell* 162, 766–779 [PubMed: 26276631]
26. Roychowdhury S, Chiang DJ, McMullen MR, and Nagy LE (2014) Moderate, chronic ethanol feeding exacerbates carbon-tetrachloride-induced hepatic fibrosis via hepatocyte-specific hypoxia inducible factor 1alpha. *Pharmacol Res Perspect* 2, e00061 [PubMed: 25089199]
27. Schober JM, Chen N, Grzeszkiewicz TM, Jovanovic I, Emeson EE, Ugarova TP, Ye RD, Lau LF, and Lam SC (2002) Identification of integrin alpha(M)beta(2) as an adhesion receptor on peripheral blood monocytes for Cyr61 (CCN1) and connective tissue growth factor (CCN2): immediate-early gene products expressed in atherosclerotic lesions. *Blood* 99, 4457–4465 [PubMed: 12036876]
28. Lai YS, Wahyuningtyas R, Aui SP, and Chang KT (2019) Autocrine VEGF signalling on M2 macrophages regulates PD-L1 expression for immunomodulation of T cells. *J Cell Mol Med* 23, 1257–1267 [PubMed: 30456891]
29. Sica A, Invernizzi P, and Mantovani A (2014) Macrophage plasticity and polarization in liver homeostasis and pathology. *Hepatology* 59, 2034–2042 [PubMed: 24115204]
30. Nielsen MC, Hvidbjerg Gantzel R, Claria J, Trebicka J, Moller HJ, and Gronbaek H (2020) Macrophage Activation Markers, CD163 and CD206, in Acute-on-Chronic Liver Failure. *Cells* 9
31. Polfliet MM, Fabriek BO, Daniels WP, Dijkstra CD, and van den Berg TK (2006) The rat macrophage scavenger receptor CD163: expression, regulation and role in inflammatory mediator production. *Immunobiology* 211, 419–425 [PubMed: 16920481]
32. Svendsen P, Graversen JH, Etzerodt A, Hager H, Roge R, Gronbaek H, Christensen EI, Moller HJ, Vilstrup H, and Moestrup SK (2017) Antibody-Directed Glucocorticoid Targeting to CD163 in M2-type Macrophages Attenuates Fructose-Induced Liver Inflammatory Changes. *Mol Ther Methods Clin Dev* 4, 50–61 [PubMed: 28344991]
33. Zhao AY, Dai YJ, Lian JF, Huang Y, Lin JG, Dai YB, and Xu TW (2018) YAP regulates ALDH1A1 expression and stem cell property of bladder cancer cells. *Onco Targets Ther* 11, 6657–6663 [PubMed: 30349299]
34. Zhao B, Wei X, Li W, Udan RS, Yang Q, Kim J, Xie J, Ikenoue T, Yu J, Li L, Zheng P, Ye K, Chinnaiyan A, Halder G, Lai ZC, and Guan KL (2007) Inactivation of YAP oncoprotein by the Hippo pathway is involved in cell contact inhibition and tissue growth control. *Genes Dev* 21, 2747–2761 [PubMed: 17974916]
35. Imajo M, Miyatake K, Imura A, Miyamoto A, and Nishida E (2012) A molecular mechanism that links Hippo signalling to the inhibition of Wnt/ $\beta$ -catenin signalling. *Embo j* 31, 1109–1122 [PubMed: 22234184]
36. Webb C, Upadhyay A, Giuntini F, Eggleston I, Furutani-Seiki M, Ishima R, and Bagby S (2011) Structural features and ligand binding properties of tandem WW domains from YAP and TAZ, nuclear effectors of the Hippo pathway. *Biochemistry* 50, 3300–3309 [PubMed: 21417403]
37. Hoshino M, Qi ML, Yoshimura N, Miyashita T, Tagawa K, Wada Y, Enokido Y, Marubuchi S, Harjes P, Arai N, Oyanagi K, Blandino G, Sudol M, Rich T, Kanazawa I, Wanker EE, Saitoe M, and Okazawa H (2006) Transcriptional repression induces a slowly progressive atypical neuronal death associated with changes of YAP isoforms and p73. *J Cell Biol* 172, 589–604 [PubMed: 16461361]
38. Sudol M, Bork P, Einbond A, Kastury K, Druck T, Negrini M, Huebner K, and Lehman D (1995) Characterization of the mammalian YAP (Yes-associated protein) gene and its role in defining a novel protein module, the WW domain. *J Biol Chem* 270, 14733–14741 [PubMed: 7782338]
39. Su T, Bondar T, Zhou X, Zhang C, He H, and Medzhitov R (2015) Two-signal requirement for growth-promoting function of Yap in hepatocytes. *Elife* 4
40. Bai H, Zhang N, Xu Y, Chen Q, Khan M, Potter JJ, Nayar SK, Cornish T, Alpini G, Bronk S, Pan D, and Anders RA (2012) Yes-associated protein regulates the hepatic response after bile duct ligation. *Hepatology* 56, 1097–1107 [PubMed: 22886419]

41. Machado MV, Michelotti GA, Pereira TA, Xie G, Premont R, Cortez-Pinto H, and Diehl AM (2015) Accumulation of duct cells with activated YAP parallels fibrosis progression in non-alcoholic fatty liver disease. *J Hepatol* 63, 962–970 [PubMed: 26070409]
42. Moya IM, and Halder G (2019) Hippo-YAP/TAZ signalling in organ regeneration and regenerative medicine. *Nat Rev Mol Cell Biol* 20, 211–226 [PubMed: 30546055]
43. Lu L, Finegold MJ, and Johnson RL (2018) Hippo pathway coactivators Yap and Taz are required to coordinate mammalian liver regeneration. *Exp Mol Med* 50, e423 [PubMed: 29303509]
44. Yimlamai D, Christodoulou C, Galli GG, Yanger K, Pepe-Mooney B, Gurung B, Shrestha K, Cahan P, Stanger BZ, and Camargo FD (2014) Hippo pathway activity influences liver cell fate. *Cell* 157, 1324–1338 [PubMed: 24906150]
45. Swiderska-Syn M, Xie G, Michelotti GA, Jewell ML, Premont RT, Syn WK, and Diehl AM (2016) Hedgehog regulates yes-associated protein 1 in regenerating mouse liver. *Hepatology* 64, 232–244 [PubMed: 26970079]
46. Wu H, Xiao Y, Zhang S, Ji S, Wei L, Fan F, Geng J, Tian J, Sun X, Qin F, Jin C, Lin J, Yin ZY, Zhang T, Luo L, Li Y, Song S, Lin SC, Deng X, Camargo F, Avruch J, Chen L, and Zhou D (2013) The Ets transcription factor GABP is a component of the hippo pathway essential for growth and antioxidant defense. *Cell Rep* 3, 1663–1677 [PubMed: 23684612]
47. Shao D, Zhai P, Del Re DP, Sciarretta S, Yabuta N, Nojima H, Lim DS, Pan D, and Sadoshima J (2014) A functional interaction between Hippo-YAP signalling and FoxO1 mediates the oxidative stress response. *Nat Commun* 5, 3315 [PubMed: 24525530]
48. Del Re DP, Yang Y, Nakano N, Cho J, Zhai P, Yamamoto T, Zhang N, Yabuta N, Nojima H, Pan D, and Sadoshima J (2013) Yes-associated protein isoform 1 (Yap1) promotes cardiomyocyte survival and growth to protect against myocardial ischemic injury. *J Biol Chem* 288, 3977–3988 [PubMed: 23275380]
49. Fan X, Molotkov A, Manabe S, Donmoyer CM, Deltour L, Foglio MH, Cuenca AE, Blaner WS, Lipton SA, and Duester G (2003) Targeted disruption of Aldh1a1 (Raldh1) provides evidence for a complex mechanism of retinoic acid synthesis in the developing retina. *Mol Cell Biol* 23, 4637–4648 [PubMed: 12808103]
50. Shabtai Y, and Fainsod A (2018) Competition between ethanol clearance and retinoic acid biosynthesis in the induction of fetal alcohol syndrome. *Biochemistry and cell biology = Biochimie et biologie cellulaire* 96, 148–160 [PubMed: 28982012]
51. Liu HX, Ly I, Hu Y, and Wan YJ (2014) Retinoic acid regulates cell cycle genes and accelerates normal mouse liver regeneration. *Biochemical pharmacology* 91, 256–265 [PubMed: 25087568]
52. Adams MK, Belyaeva OV, Wu L, and Kedishvili NY (2014) The retinaldehyde reductase activity of DHRS3 is reciprocally activated by retinol dehydrogenase 10 to control retinoid homeostasis. *The Journal of biological chemistry* 289, 14868–14880 [PubMed: 24733397]
53. Xiao Y, Hill MC, Zhang M, Martin TJ, Morikawa Y, Wang S, Moise AR, Wythe JD, and Martin JF (2018) Hippo Signaling Plays an Essential Role in Cell State Transitions during Cardiac Fibroblast Development. *Developmental cell* 45, 153–169 e156 [PubMed: 29689192]
54. Yu J, Alharbi A, Shan H, Hao Y, Snetsinger B, Rauh M, and Yang X (2017) TAZ induces lung cancer stem cell properties and tumorigenesis by up-regulating ALDH1A1. *Oncotarget* 8
55. Vidal F, Toda R, Gutierrez C, Broch M, Fernandez-Muixi F, Lorenzo A, and Richart C (1998) Influence of chronic alcohol abuse and liver disease on hepatic aldehyde dehydrogenase activity. *Alcohol* 15, 3–8 [PubMed: 9426831]



**Figure 1. YAP activation during progression of alcohol-related liver disease and in experimental models of alcohol-associated liver injury**

(A) Total YAP and its phosphorylated form at serine 127 residue in six patients with alcoholic liver disease were examined with specific rabbit antibodies in Western analysis. (B) Densitometrical analysis was performed to calculate percentage of the active nonphosphorylated YAP based on band intensity of phosphorylated (inactive) and total forms of this protein in (A). AS: alcoholic steatosis; ASH: alcoholic steatohepatitis; and AC: alcoholic cirrhosis. (C) IHC detected YAP accumulation in ASH livers. Scale bar: 50  $\mu$ M. (D) Immunoblotting and densitometry analyses showed Yap accumulation in nuclear fraction of 5% ethanol-fed animals that received ethanol binge (EtOH-fed/binge), but not in pair-fed livers that received water gavage (pair-fed/water). Accumulation of nuclear Yap was also found in livers that were exposed to moderate ethanol and one acute dose of CCl<sub>4</sub> intoxication. Quantification was performed based on three independent experiments from 5 animals per group. \* $P$ <0.05. (E) IHC detected abundant Yap protein in pericentral

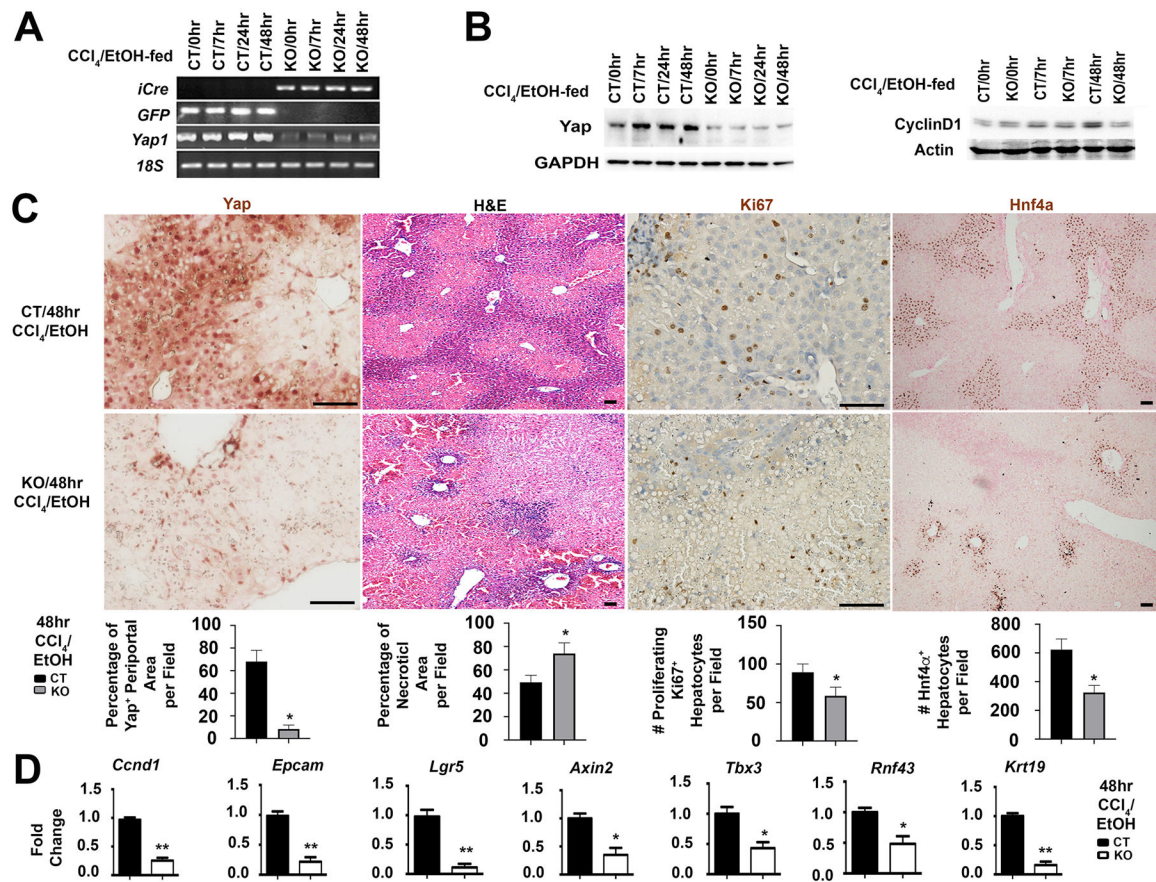
hepatocytes of ethanol-fed livers at 9 hours after binge or 24 hours post CCl<sub>4</sub> intoxication.  
Scale bar: 100 μM.

Author Manuscript

Author Manuscript

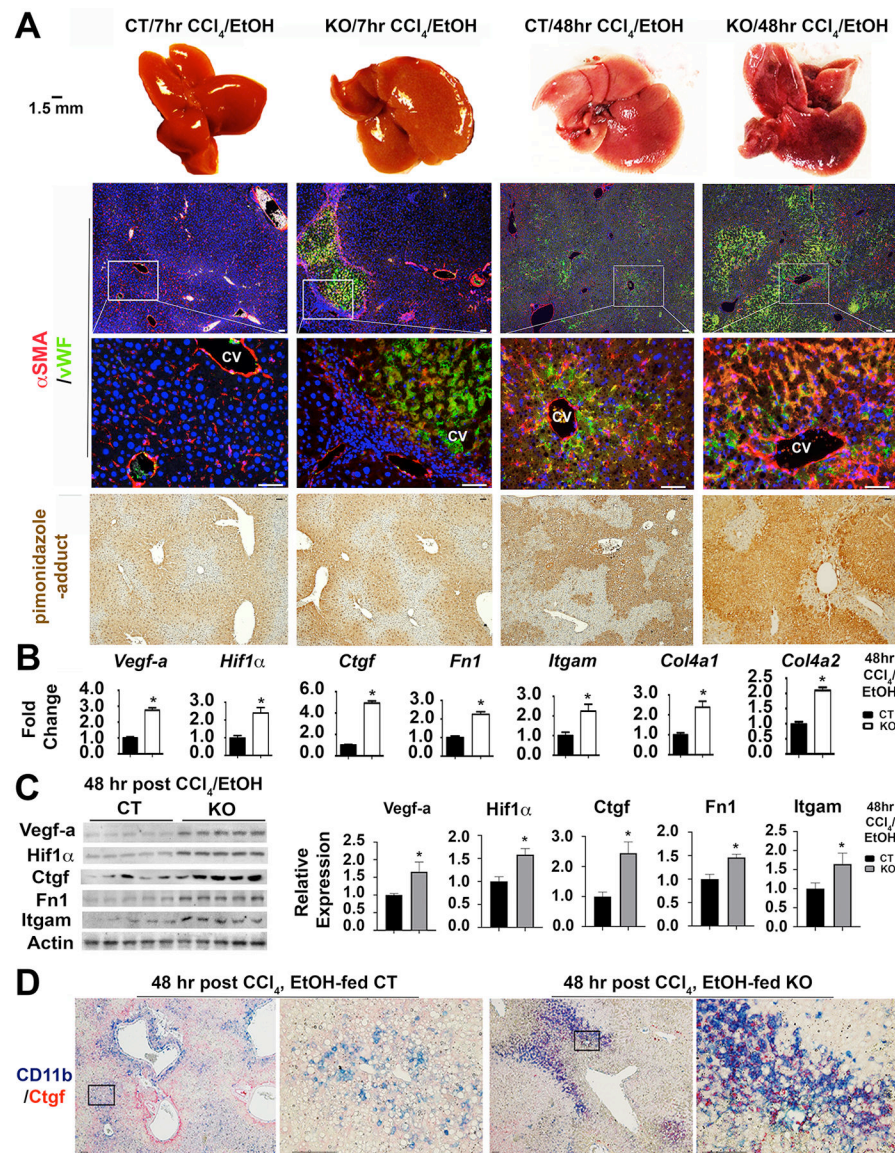
Author Manuscript

Author Manuscript



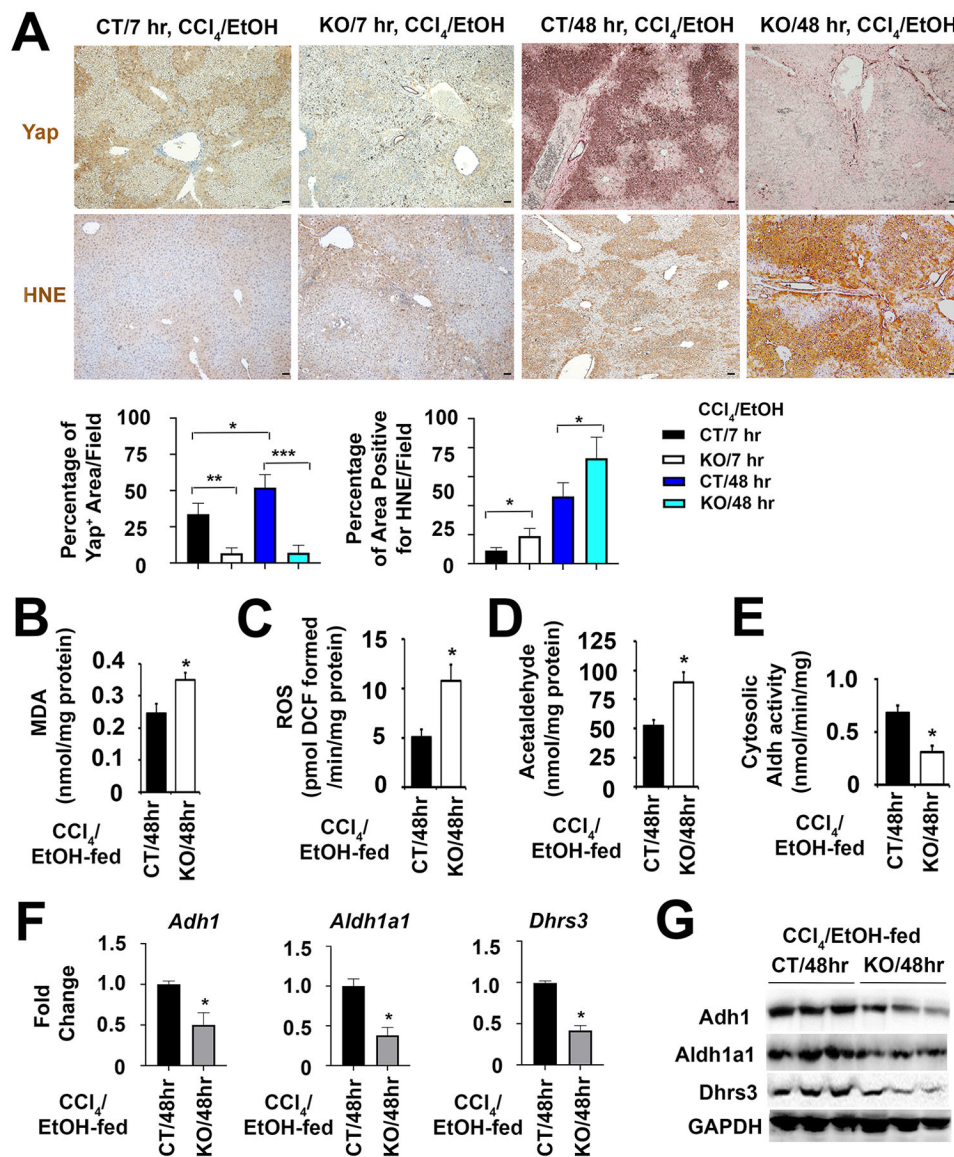
**Figure 2. *Yap1* deletion inhibits hepatocyte proliferation and downregulates progenitor related genes after ethanol/CCl<sub>4</sub> treatment.**

Control (CT) animals or hepatocyte-specific knockouts (KO) were generated by tail vein injection of AAV8-GFP or AAV8-iCre on floxed homozygotes followed by ethanol-feeding and a single dose of CCl<sub>4</sub> intoxication (n=5/per group per time point). *Yap1* deletion in hepatocytes was confirmed in RT-PCR analysis (A), Western blotting (B), and IHC (C). *Yap* loss in periportal hepatocytes of KO livers at 48 hours after ethanol/CCl<sub>4</sub>-induced injury was associated with massive hepatic necrosis shown in H&E staining, reduced proliferating hepatocytes in Ki67 staining, and decreased number of Hnf4a<sup>+</sup> periportal cells. Scale bar: 100 μm. Values were means ± SEM based on quantification of images from more than 10 fields per mouse (n=5 mice per group). (D) Q-RT-PCR analysis detected downregulation of *Ccnd1* and progenitor related transcripts in the ethanol-fed *Yap1* KO livers at 48 hours post CCl<sub>4</sub> intoxication. Values represent means ± SD in relation to controls from three independent experiments. \**P*<0.05, \*\**P*<0.01.



**Figure 3. *Yap1* deficiency in hepatocytes enhances hypoxia, vascular remodeling, and infiltration of CD11b<sup>+</sup> inflammatory cells into Ctgf enriched microenvironments after ethanol/CCl<sub>4</sub> treatment.**

(A) Macroscopic visualization shows hemorrhage in ethanol-fed *Yap1* KO livers at 48 hours post CCl<sub>4</sub> intoxication (first row). Vascular endothelial cells and activated hepatic stellate cells were stained with vWF and αSMA (second and third rows). Hypoxia was stained based on pimonidazole-adducts (fourth row). Scale bar: 100 μm. Q-RT-PCR analysis (B) and Western blotting (C) detected upregulation of gene signatures for hypoxia and vascular remodeling. Values in (B) represent means ± SD from five animals. \**P*<0.05. Relative levels of Vegf-a, Hif1α, Ctgf, Fn1, and Itgam proteins in KO in comparison to CT groups (n=5 per group) were quantified based on densitometrical analyses of band intensities in Western analyses and were expressed as means ± SD. \**P*<0.05. (D) Dual staining showed extensive infiltration of CD11b<sup>+</sup> macrophages into Ctgf enriched microenvironments in the damaged *Yap1* KO livers. Scale bar: 100 μm.

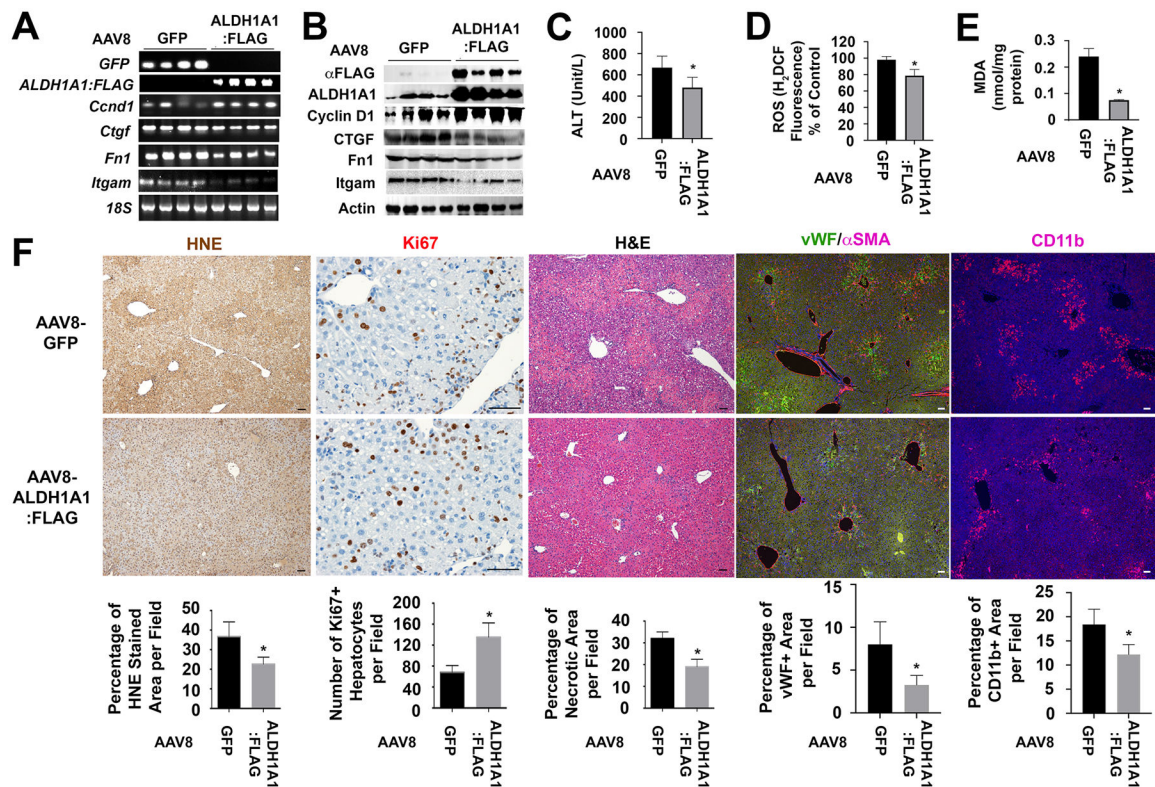


**Figure 4.** *Yap1* loss in hepatocytes is associated with enhanced hepatotoxicity and downregulation of detoxification enzymes including *Aldh1a1* after ethanol/ $\text{CCl}_4$  treatment.

(A) The ethanol-fed *Yap1* null livers lost both pericentral and periportal induction of Yap protein post  $\text{CCl}_4$  intoxication, and exhibited elevated levels of lipid peroxidation as indicated by stronger staining of HNE-adducts than controls. Scale bar: 100  $\mu\text{m}$ . Areas immunoreactive to antibodies for Yap or HNE were quantified and the percentages of positive areas were calculated as means  $\pm$  SEM based on quantification of images from more than 10 fields (100x magnification) from 5 mice per group. \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ . (B-E) Enhanced production of hepatic MDA (B), ROS (C), and acetaldehyde (D) were associated with decreased cytosolic Aldh activity (E) in the damaged *Yap1* KO livers. (F and G) Downregulation of *Adh1*, *Aldh1a1*, and *Dhars3* were found at mRNA in Q-RT-PCR analysis (F) and protein levels in immunoblotting (G). Values in (B-F) represent means  $\pm$  SD (n=5 mice per group). \* $P < 0.05$ .

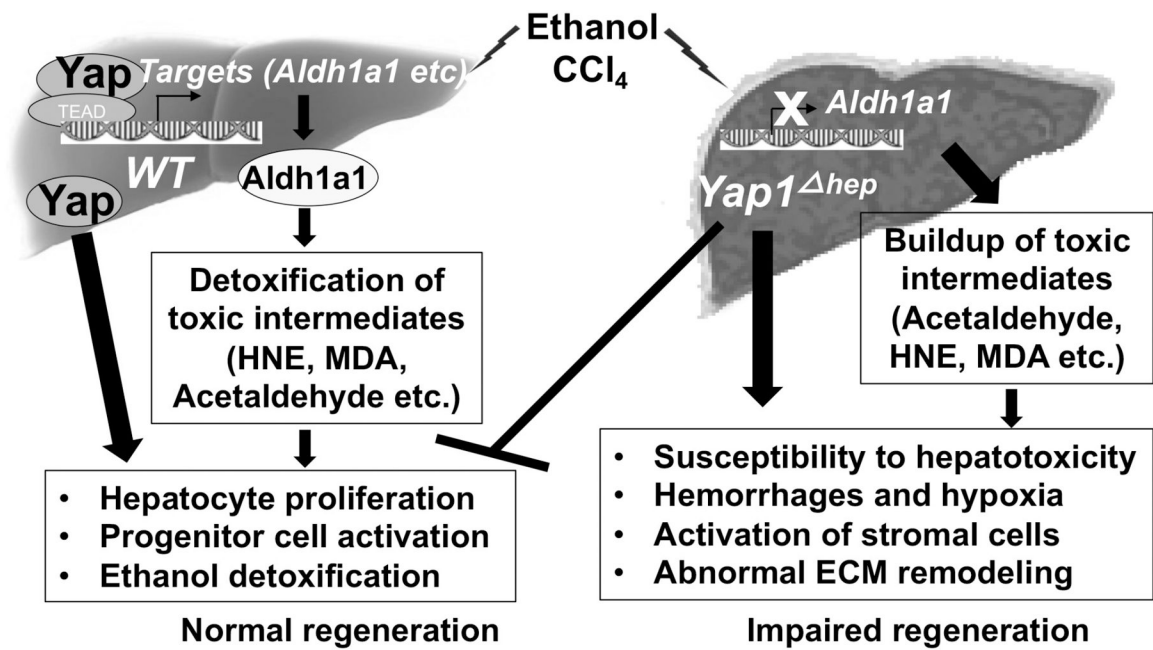






**Figure 6. Ectopic expression of ALDH1A1 reduces oxidative stress, decreases hypoxia, attenuates vascular remodeling, ameliorates hepatic inflammation, and enhances hepatocyte proliferation during ethanol/CCl<sub>4</sub>-induced liver damage.**

(A and B) RT-PCR analysis and Western blotting detected AAV8-ALDH1A1:FLAG or AAV8-GFP in transduced ethanol-fed livers at 48 hours post CCl<sub>4</sub> intoxication. Ectopic expression of ALDH1A1 was associated with upregulation of *Cyclin D1* and downregulation of *Ctgf*, *Fn1*, and *Itgam* at mRNA and protein levels. (C-E) Reduction of serum ALT (C), hepatic MDA (D), and ROS (E) were observed in the EtOH/CCl<sub>4</sub>-damaged AAV8-ALDH1A1:FLAG-transduced livers. Values in (C-E) represent means  $\pm$  SD (n=5 mice per group). \**P*<0.05. (F) IHC showed that ectopic expression of ALDH1A1 enhanced number of proliferating hepatocytes (Ki67 staining) and caused reduction in lipid peroxidation (HNE staining), hepatic necrosis (H&E staining), vascular remodeling ( $\alpha$ SMA/vWF staining), and inflammation (CD11b staining) during ethanol/CCl<sub>4</sub>-induced liver injury. Values were means  $\pm$  SEM based on quantification of images from more than 10 fields (200x magnification) from 5 mice per group. \**P*<0.05.



**Figure 7. A diagram illustrating different regenerative responses in hepatocyte *Yap1* specific KO and wild type mice during liver injury.**

*Yap* protein is activated upon hepatocyte damage caused by insults such as ethanol and carbon tetrachloride. Active *Yap* has been known to form complexes with TEAD transcriptional factors in nucleus and mediate transcriptional reprogramming leading to hepatocyte proliferation and progenitor cell activation. Moreover, the active *Yap* can transcriptionally activate target genes, including *Aldh1a1* that encodes an enzyme for removal of toxic aldehydes during hepatocyte damage. However, *Yap1* hepatocyte specific KO loses normal regeneration ability. Without *Yap*, target genes such as *Aldh1a1* can't be induced for effective removal of toxic aldehydes. The impaired regeneration eventually causes susceptibility to hepatotoxicity, hemorrhages, hypoxia, activation of stromal cells and abnormal ECM remodeling.

**Table 1.**

Functional enrichment analyses identify genes and pathways that are upregulated or downregulated during indicated liver injury.

Injury Type	Term	Fold Enrichment	P Value	Change
chronic-ethanol-binge-induced liver injury in wild type mice	GO:0055114~oxidation-reduction process	-3.49554532	4.72E-13	Down
	Mmu00830:Retinol metabolism	-5.194388162	4.33E-05	Down
	GO:0019433~triglyceride catabolic process	11.02560976	0.00531723	Up
	GO:0048661~positive regulation of smooth muscle cell proliferation	5.088742964	0.00638916	Up
	GO:0050729~positive regulation of inflammatory response	5.25029036	0.005597808	Up
	mmu04390:Hippo signaling pathway	2.302091518	0.057852084	Up
ethanol/CCl <sub>4</sub> cotreatment in the hepatocyte-specific <i>Yap1</i> KO	GO:0055114~oxidation-reduction process	-3.61466496	6.78E-12	Down
	Mmu00830:Retinol metabolism	-7.63762278	2.47E-08	Down
	GO:0045766~positive regulation of angiogenesis	8.160437763	1.08E-09	Up
	GO:0048661~positive regulation of smooth muscle cell proliferation	7.714163823	5.61E-06	Up
	GO:0016477~cell migration	3.5541697	0.001	Up