

# **HHS Public Access**

Author manuscript *Immunol Rev.* Author manuscript; available in PMC 2023 October 01.

Published in final edited form as:

Immunol Rev. 2022 October; 311(1): 39–49. doi:10.1111/imr.13121.

# Targetability of the neurovascular unit in inflammatory diseases of the central nervous system

Brandon C. Smith<sup>1,2</sup>, Rachel A. Tinkey<sup>1,3</sup>, Benjamin C. Shaw<sup>1</sup>, Jessica L. Williams<sup>1,4,\*</sup>

<sup>1</sup>Department of Neurosciences, Lerner Research Institute, Cleveland Clinic, Cleveland, OH USA

<sup>2</sup>Department of Biological, Geological, and Environmental Sciences, Cleveland State University, Cleveland, OH USA

<sup>3</sup>School of Biomedical Sciences, Kent State University, Kent, OH USA

<sup>4</sup>Brain Health Research Institute, Kent State University, Kent, OH USA

# Summary

The blood-brain barrier (BBB) is a selectively permeable barrier separating the periphery from the central nervous system (CNS). The BBB restricts the flow of most material into and out of the CNS, including many drugs that could be used as potent therapies. BBB permeability is modulated by several cells that are collectively called the neurovascular unit (NVU). The NVU consists of specialized CNS endothelial cells (ECs), pericytes, astrocytes, microglia, and neurons. CNS ECs maintain a complex "seal" via tight junctions, forming the BBB; breakdown of these tight junctions leads to BBB disruption. Pericytes control the vascular flow within capillaries and help maintain the basal lamina. Astrocytes control much of the flow of material that has moved beyond the CNS EC layer and can form a secondary barrier under inflammatory conditions. Microglia survey the border of the NVU for noxious material. Neuronal activity also plays a role in the maintenance of the BBB. Since astrocytes, pericytes, microglia, and neurons are all able to modulate the permeability of the BBB, understating the complex contributions of each member of the NVU will potentially uncover novel and effective methods for delivery of neurotherapies to the CNS.

# Keywords

blood-brain barrier; endothelial cell; tight junction; glia; pericyte; neuroinflammatory disease

# 1. Introduction

The blood-brain barrier (BBB) is a complex, highly regulated system with multiple cell types influencing its maintenance and formation. The BBB forms a privileged environment

All authors contributed to the writing, editing, and review of the manuscript.

Conflict of Interest

<sup>\*</sup>Corresponding author: Dr. Jessica L. Williams, Lerner Research Institute, Cleveland Clinic, 9500 Euclid Avenue/NC30, Cleveland, OH 44195. Tel: +1 216-445-3831, Fax: +1 216-444-7927; williaj39@ccf.org. Author Contributions

The authors declare no competing financial interests.

for the central nervous system (CNS), restricting entry of a wide breadth of potential hazards, including pathogens, immune cells, antibodies, and pharmaceuticals. This tight regulation protects the brain from external insults, but simultaneously prevents access of many therapeutics meant to treat neuroinflammatory and neurodegenerative disorders. As a result, multiple clinical trials have failed despite promise in preclinical studies, underscoring the need for a more complete understanding of the BBB and its modulatory mechanisms. In this review, we discuss the composition of the neurovascular unit (NVU), known mechanisms of BBB modulation, and potential therapeutic targets for neuroinflammatory disorders.

The existence of a selectively permeable BBB was first postulated at the turn of the 20<sup>th</sup> century when it was noted that water-soluble dyes injected into the periphery did not permeate the CNS<sup>1,2</sup>, and dyes injected into the CNS parenchyma did not exit to the periphery<sup>2,3</sup>. Further complexity of the BBB was demonstrated by seminal evidence indicating immune privilege, wherein immune cells are excluded and unable to surveille the CNS, over 100 years ago by Shirai<sup>4</sup> and later confirmed by Murphy and Sturm<sup>5</sup>. However, even now, the permeability, complexity, and dynamics of the BBB are incompletely understood<sup>6,7</sup>.

The BBB acts as a robust biological gateway, responsible for the flow of nearly all molecules and cells passing into and out of the CNS<sup>6,8,9</sup>. This selective permeability is critical in sparing the CNS from many circulating toxins and pathogens and allowing for availability of necessary nutrients to the CNS<sup>2,10,11</sup>. Instead of passing through fenestrations in endothelial cells (ECs) as it does in the peripheral vasculature, material is transported and scrutinized by one or more cells closely associated with the BBB<sup>11,12</sup>. However, it is this very system, designed to protect the CNS, that also hinders therapeutic development to treat CNS disorders including multiple sclerosis (MS), Alzheimer's disease (AD), Parkinson's disease, traumatic brain injuries, brain cancers, and others<sup>10,11,13</sup>. Several researchers have even employed artificial intelligence to determine the likelihood of small molecules to penetrate the BBB<sup>13,14</sup>.

This review will examine some of the NVU cell-specific effects on BBB permeability as well as the intercellular communication that is critical for proper function of the NVU. While other complex CNS barriers, including the blood-retinal barrier, the blood-cerebrospinal fluid barrier, and the arachnoid layer<sup>15</sup>, are also important to consider in therapeutic availability to the CNS, we will only focus on the BBB for the purposes of this review.

# 2. Structure and Organization of the NVU

The NVU is the fundamental multicellular unit supporting the BBB and consists of ECs, pericytes, astrocytes, microglia, and neurons (Fig. 1)<sup>6,7</sup>. The capillary lumen is the primary area of contact between the NVU and the periphery, and interfacing factors vary from cytokines and hormones to immune cells that have the potential to extravasate<sup>16,17</sup>. Forming the vessel lumen are specialized CNS ECs, which are semipermeable under homeostatic conditions due to the presence of tight junctions<sup>6,16,18–20</sup>. Perivascular mural cells, or

pericytes, have recently been shown to be more complex and varied than initially assumed. Pericytes are functionally heterogeneous<sup>21,22</sup> and further understanding of their diverse roles in the CNS is an active area of research. Pericytes were classically described to help govern vasoconstriction and dilation of capillaries<sup>23,24</sup>, which is highly regulated in the NVU, to limit BBB permeability. Recent studies are beginning to uncover pericyte coordination with other members of the NVU, particularly ECs and astrocytes<sup>21,24</sup>. Astrocytes are vital to the NVU and while the other members of the NVU have critical roles, it is largely the astrocytes that set the NVU apart from other capillary layers in the body<sup>25,26</sup>. Understanding the specific mechanisms by which astrocytes regulate tight junctions in CNS ECs could prove incredibly beneficial to optimizing BBB permeability for drug delivery. Astrocytes are relatively large and abundant CNS trophic cells<sup>27</sup> and their endfeet form an additional barrier around CNS capillaries<sup>6,25</sup>. The perivascular, or Virchow-Robin, space refers to the area between ECs and astrocytes and is a key checkpoint prior to entering the CNS parenchyma<sup>6,28</sup>. Microglia are a recent addition to our understanding of the NVU. Although astrocytes cover the majority of the perivascular space, there are some gaps in coverage. Recently, Kisler et al. used two-photon in vivo imaging to observe microglial processes covering many of these gaps<sup>29</sup>. How involved microglia are in the structure of the NVU is still being investigated. By area, neurons have a relatively minor role in the structure of the NVU, but can modulate BBB permeability via neuronal activity  $^{30,31}$ . Due to the complex nature of the NVU, it has proven difficult to study. In vitro models have been useful; however, the heterogeneity between species, individuals, and even CNS regions have added to this investigational barrier<sup>32,33</sup>. Here, we will discuss the components of the NVU and how each cell type may impact BBB permeability.

# 3. Endothelial Cells

ECs are found throughout the body and make up the walls of arteries, veins, and capillaries. They form a tube-like structure to allow passage of various blood cells and proteins<sup>34</sup>. ECs secrete and maintain the basal lamina - the extracellular matrix on which they reside<sup>35,36</sup>. The CNS ECs differ in several ways from peripheral ECs. Most notably, they lack the fenestrations found in peripheral vessels and instead create a continuous cellular barrier with significantly reduced permeability along the capillary lumen<sup>37</sup>. Instead, CNS ECs form tight junctions which prevent hydrophobic molecules from penetrating the endothelial layer unless transported through the cell. These tight junctions are responsible for much of the impermeability of the BBB. Tight junctions are composed primarily of claudin-5, occludin, and other junctional adhesion molecules<sup>38</sup> and bind to the actin cytoskeleton via ZO-1. ZO-2. or ZO- $3^{39}$ . Tight junctions are highly complex and can vary depending on the proteins coupled<sup>40,41</sup>. Transport of nutrients, waste, and signaling molecules between the CNS and the periphery is necessary under physiological conditions and is typically achieved by a myriad of EC transporters  $^{6,42}$ . In fact, these transporters make up 10–15% of the total protein in CNS ECs and allow specific molecules, peptides, and even cells to cross into the perivascular space, bypassing tight junctions<sup>33,43</sup>.

# 3.1. ECs and Immunity

Classically, it was thought that immune cells could not access the CNS parenchyma<sup>18,44</sup>. While mostly impermeable during homeostatic conditions, CD4<sup>+</sup> and CD8<sup>+</sup> T cells are allowed passage for immune surveillance of the CNS, facilitated by a highly regulated, multistep process through a tricellular junction<sup>45</sup>. This process involves endothelial ligands vascular cell adhesion molecule (VCAM)-1 and intercellular adhesion molecule (ICAM)-1 which recognize lymphocyte function-associated antigen 1 (LFA-1) and very late antigen-4 (VLA-4) on T cells. These interactions arrest T cells on the endothelium and allow migration against the flow of blood to these tricellular junctions and ultimately extravasation<sup>45,46</sup>. These tricellular junctions contain the proteins tricellulin and angulin-1 which direct and aid T cell diapedesis<sup>45</sup>. While CNS ECs express VCAM-1 and ICAM-1 under normal conditions, the expression is sparse, limiting the number of leukocytes that can traverse tricellular junctions<sup>45,47</sup>.

In response to infection, autoimmunity, or injury, the permeability of the BBB is significantly enhanced and can lead to severe inflammation<sup>48</sup>. This is partially due to an increase in the expression of adhesion molecules on the EC surface, primarily ICAM-1 and VCAM-1, which can arrest a greater number of immune cells for extravasation<sup>49</sup>. Inflammation can lead to break down of the tight junctions, allowing leukocytes to invade the perivascular space<sup>48</sup>. Many secreted inflammatory factors, cell damage signals, and pathogen components can alter tight junction integrity including CCL2 and transforming growth factor (TGF)- $\beta$ , which is known to alter expression of claudin-5, occludin, and ZO-1<sup>50-52</sup>. Likewise, tumor necrosis factor (TNF)-a, lipopolysaccharide (LPS), and mitochondrial damage can induce BBB permeability via actin filament rearrangement<sup>53–56</sup>. Prolonged dysfunction of the BBB, as in chronic inflammatory CNS diseases like MS or AD, can lead to permanent CNS tissue damage and neuroaxonal loss<sup>57–59</sup>. However, although overt and/or chronic inflammation can lead to loss of BBB integrity and extensive damage to the CNS, low to moderate amounts of inflammation can partially restore the BBB and limit peripheral immune cell infiltration into the CNS parenchyma, mitigating injury or infection<sup>57,60–62</sup>. Taking advantage of this immunological state and exploiting molecular signals used in the formation of tricellular junctions may be a novel avenue for therapeutic development.

#### 3.2. Targeting ECs

It is possible to permeate the BBB to treat neurological disorders, but the potential bystander effects of leaky BBB during diseases like glioblastoma, MS, AD, and others imposes significant risk. The BBB is disrupted in many neurodegenerative and psychological disorders. Additional access of immune cells, toxins, or other inflammatory mediators could enhance inflammation-mediated damage to the CNS. Additionally, BBB dysregulation disrupts homeostatic transport across the barrier, which provides trophic factors and controls osmotic regulation during physiological conditions<sup>63</sup>. Bypassing the ECs of the BBB is a difficult challenge; however, there are several other members of the NVU that introduce potentially novel avenues for drug targeting.

## 4. Pericytes

Proportionally, there is a higher density of pericytes in the CNS relative to peripheral organs<sup>24</sup>, suggesting they have a critical role in the CNS. As vascular mural cells, pericytes play a large role in the dilation of capillaries in the NVU. Of note, pericytes are found in the capillary bed and not surrounding arteries or veins, which house similar, but distinct, vascular mural cells, vascular smooth muscle cells<sup>64</sup>. In addition to vasodilation and vasoconstriction, pericytes also help form and maintain the basal lamina<sup>65,66</sup>. Control of vascular flow by pericytes can indirectly impact the BBB as changes in blood flow can allow for more or less cellular contact in the capillary lumen. Increases in cellular interaction can stretch and stress CNS ECs<sup>67–69</sup>, causing pericyte dysfunction, which is associated with aberrant immune cell trafficking<sup>70</sup>. Further, in mouse models deficient in functional pericytes, there is a loss of vascular control, dysfunctional tight junction regulation, and aberrant angiogenic sprouting, highlighting the importance of pericytes in maintaining a healthy BBB<sup>70–74</sup>.

New roles for pericytes continue to be uncovered, including their ability to maintain and produce elements of the basal lamina<sup>75–78</sup> and intricate regulation of tight junctions. Contrary to ECs, TGF- $\beta$  and angiopoietin-1 (Ang-1) signaling in pericytes enhances the expression of occludin on CNS ECs, reducing the permeability of the BBB. TGF- $\beta$  from pericytes also activates Smad4 signaling to upregulate bone morphogenic proteins (BMPs). BMPs can then ensure tight adherence of pericytes to ECs via N-cadherins, which is upregulated by vascular endothelial growth factor (VEGF), reinforcing the communication and physical interaction between pericytes and CNS ECs<sup>79–81</sup>. While basal levels of VEGF signaling can reduce BBB permeability, excessive VEGF signaling to pericytes can lead to downregulation of claudin-5 on ECs and dysregulate BBB tight junctions (Fig. 2)<sup>82–85</sup>.

#### 4.1. Heterogeneity of Pericytes

Similar to ECs, pericytes respond robustly to various signaling molecules. Inflammatory and non-inflammatory pericytes have been described and subdivided into Type-1 pericytes (PC1) and Type-2 pericytes (PC2). PC1s are non-inflammatory and tend to be the resting state of pericytes without injury or infection. PC2s tend to increase in frequency with and are highly responsive to inflammation<sup>74,86</sup>. It is likely, as is the case with astrocytes and microglia, that there is a spectrum of activation states, but as the study of pericytes is still in its infancy, this has not yet been fully elucidated. In young healthy patients, nearly all pericytes exhibit a non-inflammatory morphology<sup>72,86,87</sup>. However, aging results in an increase in the population of inflammatory pericytes, which is consistent with enhanced inflammation and BBB permeability associated with age<sup>86</sup>. Both pericyte subtypes contribute to the basal lamina, but PC2s tend to produce a more irregular basal lamina, impacting both ECs and astrocytes<sup>21,86</sup>. Additionally, PC2s produce less laminin-111 and laminin-211, which leads to cell hypertrophy and BBB disruption<sup>88,89</sup>. Exposing pericytes to inflammatory cytokines, LPS, or reactive oxygen species induces immunoreactivity in pericytes, altering their morphology, as well as inducing their separation from the basal lamina<sup>88-90</sup>. The dynamic morphology of pericytes suggests they have a critical role in the integrity and function of the BBB, which is altered in pathological states and in aging.

#### 4.2. Targeting Pericytes

Examining the targetability of pericytes and its potential influence on the BBB is difficult, but may prove promising. Affecting pericytes may be a subtler approach to drug penetrance rather than reducing the integrity of the EC layer itself. A deeper understanding of how pericytes communicate with CNS ECs and astrocytes could reveal potential nuanced approaches to bypass the BBB. While pericytes may be a promising target to leverage BBB permeability, much about pericyte mechanisms of BBB control remains unknown.

#### 5. Astrocytes

Astrocytes are large stellate cells with extensive processes that extend throughout the  $CNS^{27,91}$ . However, unlike CNS ECs and pericytes, astrocytes are unique in that only the ends of their processes are considered part of the NVU (Fig. 1). These endfeet cover roughly 90–95% of the area surrounding the BBB and have properties that are unique from the rest of the astrocyte<sup>92–94</sup>. Endfeet contain aquaporin-4 and the potassium channel K<sub>ir</sub>4.1 to modulate water and ion balance and, lacking tight junctions, astrocytic endfeet permit immune cell extravasation<sup>93,95,96</sup>. Although endfeet lack the barrier that tight junctions provide, ablation of astrocytes leads to rapid and extensive BBB deterioration<sup>25,94</sup>. And interestingly, early studies transplanting astrocytes outside of the CNS demonstrated that transplanted astrocytes develop a BBB-like morphology surrounding the peripheral vasculature<sup>97,98</sup>. In addition to the physical barrier that astrocytes provide, during homeostasis astrocytes are responsible for the transport of material from the perivascular space into the parenchyma<sup>6,25</sup>. This function makes them excellent and critical supportive cells in the maintenance of the BBB.

Astrocytes form gap junctions between endfeet forming a much "looser" network of connections compared to that of CNS ECs<sup>99</sup>. Gap junctions are formed by a hexamer of proteins called connexins. The material that is transported by a given gap junction is largely dependent on the specific combination of connexins that make up the junction<sup>100,101</sup>. Notable gap junctions in astrocytes are formed by connexin 30 and connexin 43, which help mediate glucose and lactate transport to distal neurons<sup>102</sup>. Astrocytes use these gap junctions to communicate with other cells of the NVU including neurons, microglia, and other astrocytes using ion gradients, electrical signals, and signaling molecules<sup>103</sup>.

#### 5.1. Tight Junction Modulation

Astrocytes release a variety of trophic factors that help maintain a functional NVU<sup>104–106</sup>. Many of the factors released under physiologic conditions increase the amount and order of tight junction proteins between BBB ECs<sup>107,108</sup>. For instance, astrocytes secrete sonic hedgehog (Shh), which increases tight junctions in CNS ECs by inducing Patch1 signaling<sup>108</sup>. Other astrocytic factors that can enhance tight junctions in ECs include Wnt signaling, TGF $\beta$ , and apolipoprotein E (ApoE)<sup>105,107,109–111</sup>. ApoE and TGF $\beta$  may work indirectly through pericytes to impact EC tight junctions<sup>112,113</sup>. Additionally, astrocytes produce Ang-1, which signals to EC Tie2 to further upregulate tight junction proteins and reduce adhesion molecules, reducing the likelihood of leukocyte entry<sup>114,115</sup>. Further, astrocytes are incredibly responsive to inflammatory stimuli<sup>116,117</sup>, and during

inflammation, it has been shown that BBB ECs down-regulate claudin-5, while astrocytes upregulate tight junction proteins claudin-1, claudin-4, and JAM-A<sup>118,119</sup>. This suggests that astrocyte endfeet form a secondary barrier to prevent excessive immune cell entry past the perivascular space, regulating access to the CNS parenchyma<sup>118</sup>. This accumulation of peripheral immune cells in the perivascular space is often referred to as perivascular cuffing and is commonly seen in MS lesions<sup>118,120</sup>.

However, although astrocytes provide an impressive physical barrier, activated astrocytes can also secrete trophic factors, chemokines, and cytokines that dysregulate EC tight junctions and recruit peripheral immune cells (Fig. 2)<sup>25,39,120,121</sup>. These factors can include VEGF, nitric oxide, MMPs, endothelins, and glutamate; all of which can downregulate endothelial tight junction proteins<sup>39,122,123</sup>. Importantly, astrocytes also promote a return to homeostasis following an inflammatory event, secreting several beneficial trophic factors including Shh, astrocyte-derived Ang-1, glial-derived neurotrophic factor (GDNF), insulin-like growth factor-1, ApoE, and retinoic acid. These factors not only prevent EC apoptosis, but also stimulate tight junction formation and a return to a homeostatic state<sup>117,122</sup>. Taken together, astrocytes orchestrate a complex modulation of the BBB and may represent an amenable cell type for drug manipulation.

#### 5.2. Astrocytes as an Immunologic Barrier

Since astrocytes are highly responsive to immune stimuli, they have the ability to upregulate a wide array of chemokines and adhesion molecules that can serve as an immunologic barrier to prevent excessive influx of inflammatory cells during a CNS insult (Fig. 2)<sup>121,124</sup>. Interestingly, in the event immune cells breach the BBB, enter the perivascular space, and cross astrocyte endfeet into the parenchyma, astrocytes can express immune checkpoint molecules, or inhibitory receptors, which can induce exhaustion and death of leukocytes<sup>125,126</sup>, providing the CNS multiple layers of immunologic protection. As an example, astrocytes are known to upregulate the immune checkpoint protein programmed death ligand 1 (PD-L1) in response to inflammatory cytokines, primarily interferons<sup>127</sup>. PD-L1 typically signals to cells via programmed death 1 (PD-1) to reduce activation and induce apoptosis<sup>128,129</sup>. While many CNS cells, including microglia and even neurons, express PD-1, it is highly enriched on immune cells, making them particularly susceptible to exhaustion<sup>127–130</sup>. In summary, astrocytes act not only as a physical barrier, but also a trophic and immunologic barrier to limit and control the activation state of immune cells that enter the CNS during neuroinflammation.

#### 5.3. Targeting Astrocytes

Astrocytes have great potential as a therapeutic target for BBB maintenance. Their ability to weaken EC tight junctions while maintaining a barrier could prove beneficial in the development of drugs to bypass traditional CNS barriers. There are many examples of astrocytes guiding various cell types and molecules across the BBB. This principle —if better understood—could be leveraged to advance CNS drug permeability. Similar to pericytes, astrocytes have great influence on the maintenance of EC tight junctions. Elucidating the molecular control that astrocytes have over BBB permeability will inevitably lead to more avenues of drug delivery.

# 6. Microglia

Microglia are the resident immune cells of the CNS. As such, their functions are multifaceted. While they share many similarities with peripheral macrophages, they have many distinct characteristics, including their origins. Macrophages are generated in the bone marrow and circulate in the blood, while microglia migrate from the embryonic yolk sac<sup>131</sup>. In general, microglia are responsible for initial responses to injury and infection, clearance of waste, synaptic pruning and maintenance, as well as providing several trophic factors to other cells of the NVU<sup>131</sup>. Although it has been long appreciated that microglia are vital to the health of the CNS, surveying along the BBB and responding quickly to breaches, they are a relatively recent addition to the NVU and provide coverage of BBB areas not wrapped by astrocytes<sup>29,132</sup>. In addition, microglia are known to signal to other members of the NVU and communicate with peripheral immune cells<sup>6</sup>, attracting and/or activating them within the CNS parenchyma during injury and infection as sentinels of the CNS<sup>133–136</sup>. Activated microglia and peripheral immune cells can secrete a number of factors that modulate BBB integrity (Fig. 2). This is discussed in a number of reviews, but as an example, TNFa and TGF- $\beta$  can be secreted from reactive microglia under certain conditions which can either increase or decreases BBB integrity, respectively<sup>124,137,138</sup>. While ablation or depletion of microglia did not result in overt BBB breakdown<sup>139</sup>, recently, microglia were described to intricately associate with CNS capillaries and contribute to blood flow regulation and vasodilation<sup>140–142</sup>. Additionally, microglia are incredibly motile and migrate quickly in response to injury, BBB leakage, and/or inflammation<sup>143</sup>. Finally, similar to astrocytes, microglia can upregulate PD-L1 in response to neuroinflammation<sup>144,145</sup>, providing multiple mechanisms of potential control of the BBB by microglia, although there is still much to learn about the intricacies of microglia and their impact on BBB permeability.

Given their emerging role in the NVU, the mechanisms underlying microglial regulation of the BBB remain incompletely described. Targeting microglia may modify CNS capillary permeability or other NVU cells to temporarily allow access to the CNS, but there is still much to learn about this exciting new potential target.

# 7. Neurons

Neurons are the functional unit of the CNS and most other cells in the CNS support them, either directly or indirectly<sup>27</sup>. Despite their indispensable role in the CNS, as a member of the NVU, they do not directly provide a physical barrier, but instead release a number of factors that modulate other NVU cells<sup>27,146</sup> using primarily their axons and dendrites<sup>6</sup>. Interestingly, like astrocytes, neurons can induce a BBB-like barrier in neighboring ECs<sup>97,98</sup>, suggesting neurons influence the formation and maintenance of the BBB *in vivo*<sup>147,148</sup>. Astrocytes and neurons are highly communicative<sup>149,150</sup>, providing a potential route of BBB control. Additionally, neural activity can influence the integrity of the BBB<sup>151–153</sup>, although it is unclear if neural activity directly impacts ECs or if neuronal signals are propagated through astrocytes<sup>154</sup>.

Neuroinflammation can alter the activity of neurons, which in turn can lead to alterations in the BBB<sup>151,155</sup>. As mentioned above, neurons express PD-1, to which astrocytes and

microglia can signal via PD-L1 expression<sup>128-130</sup>. Neuronal PD-1 signaling does not induce apoptosis, but instead hyperpolarizes the neuronal membrane to inhibit action potential frequency<sup>130</sup>. This in turn could lead to a reduction in activity-dependent transport across the BBB<sup>156</sup>. Neurons also produce Wnt ligands which reduce BBB permeability by increasing tight junction proteins in ECs<sup>157</sup>. While neurons partially modulate the BBB via activity, they largely rely on communication with other NVU members (Fig. 2). While more information is necessary to determine how neurons might control the BBB, neurons represent an unlikely target for BBB permeability modulation given their moderate impact on the BBB and other essential functions.

# 8. Future Directions

Each cellular component of the NVU has both unique and overlapping roles in maintaining BBB permeability. ECs are the initial barrier to the periphery, reinforced with tight junctions, specialized to transport material between the periphery and the CNS. Pericytes, and to some extent, microglia, regulate vascular flow and, while they may also have a structural role, pericytes provide critical trophic support to ECs. Astrocytes, and potentially neurons, are critical to the formation of the BBB, aiding in tight junction formation. Microglia and neurons also potentially modulate BBB permeability through complex signaling networks, although this is an emerging field. The intricate dynamics of BBB formation, function, and maintenance has created a literal and figurative barrier when it comes to therapy development for the treatment of chronic neurologic diseases such as glioblastoma, MS, and AD. While it is possible to increase the permeability of the BBB, this typically results in detrimental off target effects. The BBB is disrupted in many neurological diseases, thus additional interventional disruption would likely prove deleterious as unwanted CNS "intruders" including immune cells and large molecules are able to enter the CNS without proper scrutiny, leading to a cascade of osmotic, trophic, and inflammatory dysregulation. Thus, optimizing targeted and temporary entry of therapeutics into the CNS with minimal BBB dysregulation is the holy grail of nextgeneration neurotherapeutics. Promising modalities include encapsulating drugs of interest in a form that allows them to be transported through the BBB and astrocyte endfeet or creating a transient passage that is quickly repaired. Both of these methods are active areas of research using nanoparticles and ultrasonic disruption of the BBB, respectively<sup>12,158,159</sup>. Ultimately, a deeper understanding of how each of the NVU components modulate CNS drug accessibility may shed new light on actionable therapeutic modalities.

#### Acknowledgements

We apologize to authors whose work could not be referenced in this review due to scope and space limitations. Graphical illustrations were made using BioRender (https://biorender.com/). This work was supported by grants from the CSU CD-CAVS training grant NIH T32 HL150389 (awarded to Brandon C. Smith), NIH K00 NS120365 (awarded to Benjamin C. Shaw), NIH R01 NS119178 (awarded to Jessica L. Williams).

# References

1. Biedl A & Kraus R Über eine bisher unbekannte toxische Wirkung der Gallensäuren auf das Zentralnervensystem. Zentralblatt inn Med 19, 1185–1200 (1898).

- 2. Goldmann EE Vitalfärbung am Zentralnervensystem: Beitrag zur Physio-Pathologie des Plexus chorioideus und der Hirnhäute. (Königl. Akademie der Wissenschaften, 1913).
- Dreser H Ueber den experimentellen Nachweis der Vertiefung und Verlangsamung der Athemzüge nach therapeutischen Heroingaben. Archiv für die gesamte Physiologie des Menschen und der Tiere 80, 86–95, doi:10.1007/BF01661930 (1900).
- 4. Shirai Y On the transplantation of the rat sarcoma in adult heterogenous animals. Jap Med World 1, 14–15 (1921).
- Murphy JB & Sturm E Conditions determining the transplantability of tissues in the brain. J. Exp. Med 38, 183–197, doi:doi:10.1084/jem.38.2.183 (1923). [PubMed: 19868782]
- Segarra M, Aburto MR & Acker-Palmer A Blood–Brain Barrier Dynamics to Maintain Brain Homeostasis. Trends in Neurosciences 44, 393–405, doi:10.1016/j.tins.2020.12.002 (2021). [PubMed: 33423792]
- Salmina AB et al. Blood–Brain Barrier Breakdown in Stress and Neurodegeneration: Biochemical Mechanisms and New Models for Translational Research. Biochemistry (Moscow) 86, 746–760, doi:10.1134/S0006297921060122 (2021). [PubMed: 34225598]
- Kolá ová H, Ambr zová B, Švihálková Šindlerová L, Klinke A & Kubala L Modulation of Endothelial Glycocalyx Structure under Inflammatory Conditions. Mediators of Inflammation 2014, 694312, doi:10.1155/2014/694312 (2014).
- Kutuzov N, Flyvbjerg H & Lauritzen M Contributions of the glycocalyx, endothelium, and extravascular compartment to the blood-brain barrier. Proceedings of the National Academy of Sciences 115, E9429–E9438, doi:doi:10.1073/pnas.1802155115 (2018).
- Schinkel AH, Wagenaar E, Mol CA & van Deemter L P-glycoprotein in the blood-brain barrier of mice influences the brain penetration and pharmacological activity of many drugs. The Journal of Clinical Investigation 97, 2517–2524, doi:10.1172/JCI118699 (1996). [PubMed: 8647944]
- 11. Li J et al. Development of Novel Therapeutics Targeting the Blood–Brain Barrier: From Barrier to Carrier. Advanced Science 8, 2101090, doi:10.1002/advs.202101090 (2021).
- 12. Anthony DP et al. Targeting receptor-ligand chemistry for drug delivery across blood-brain barrier in brain diseases. Life Sci 274, 119326, doi:doi:10.1016/j.lfs.2021.119326 (2021).
- Nicolazzo JA, Charman SA & Charman WN Methods to assess drug permeability across the blood-brain barrier. Journal of Pharmacy and Pharmacology 58, 281–293, doi:10.1211/ jpp.58.3.0001 (2006). [PubMed: 16536894]
- Liu L et al. Prediction of the Blood–Brain Barrier (BBB) Permeability of Chemicals Based on Machine-Learning and Ensemble Methods. Chemical Research in Toxicology 34, 1456–1467, doi:10.1021/acs.chemrestox.0c00343 (2021). [PubMed: 34047182]
- Strazielle N & Ghersi-Egea JF Physiology of Blood–Brain Interfaces in Relation to Brain Disposition of Small Compounds and Macromolecules. Molecular Pharmaceutics 10, 1473–1491, doi:10.1021/mp300518e (2013). [PubMed: 23298398]
- Ando Y et al. Brain-Specific Ultrastructure of Capillary Endothelial Glycocalyx and Its Possible Contribution for Blood Brain Barrier. Sci. Rep 8, 17523, doi:doi:10.1038/s41598-018-35976-2 (2018). [PubMed: 30504908]
- 17. Zaragozá R Transport of Amino Acids Across the Blood-Brain Barrier. Front. Physiol 11, 973, doi:10.3389/fphys.2020.00973 (2020). [PubMed: 33071801]
- Pachter JS, de Vries HE & Fabry Z The Blood-Brain Barrier and Its Role in Immune Privilege in the Central Nervous System. J. Neuropathol. Exp. Neurol 62, 593–604, doi:doi:10.1093/jnen/ 62.6.593 (2003). [PubMed: 12834104]
- Wang W, Dentler WL & Borchardt RT VEGF increases BMEC monolayer permeability by affecting occludin expression and tight junction assembly. American Journal of Physiology-Heart and Circulatory Physiology 280, H434–H440, doi:10.1152/ajpheart.2001.280.1.H434 (2001). [PubMed: 11123261]
- 20. Liu W-Y, Wang Z-B, Zhang L-C, Wei X & Li L Tight Junction in Blood-Brain Barrier: An Overview of Structure, Regulation, and Regulator Substances. CNS Neuroscience & Therapeutics 18, 609–615, doi:10.1111/j.1755-5949.2012.00340.x (2012). [PubMed: 22686334]

- 21. Bohannon DG, Long D & Kim W-K Understanding the Heterogeneity of Human Pericyte Subsets in Blood–Brain Barrier Homeostasis and Neurological Diseases. Cells 10, 890, doi:doi:10.3390/ cells10040890 (2021). [PubMed: 33919664]
- Rustenhoven J et al. TGF-beta1 regulates human brain pericyte inflammatory processes involved in neurovasculature function. Journal of neuroinflammation 13, 37, doi:10.1186/s12974-016-0503-0 (2016). [PubMed: 26867675]
- 23. Zimmermann KW Der feinere Bau der Blutcapillaren. Zeitschrift für Anatomie und Entwicklungsgeschichte 68, 29–109, doi:10.1007/BF02593544 (1923).
- 24. Hamilton N, Attwell D & Hall C Pericyte-mediated regulation of capillary diameter: a component of neurovascular coupling in health and disease. Front. Neuroenergetics 2, doi:doi:10.3389/ fnene.2010.00005 (2010).
- 25. Heithoff BP et al. Astrocytes are necessary for blood–brain barrier maintenance in the adult mouse brain. Glia 69, 436–472, doi:10.1002/glia.23908 (2021). [PubMed: 32955153]
- 26. Rubin LL et al. A cell culture model of the blood-brain barrier. The Journal of cell biology 115, 1725–1735, doi:10.1083/jcb.115.6.1725 (1991). [PubMed: 1661734]
- 27. Bear M, Connors B & Paradiso MA Neuroscience: Exploring the Brain, Enhanced Edition. (Jones & Bartlett Learning, 2020).
- Ueno M et al. Blood-brain barrier permeability in the periventricular areas of the normal mouse brain. Acta Neuropathologica 99, 385–392, doi:10.1007/s004010051140 (2000). [PubMed: 10787037]
- 29. Kisler K, Nikolakopoulou AM & Zlokovic BV Microglia have a grip on brain microvasculature. Nature Communications 12, 5290, doi:10.1038/s41467-021-25595-3 (2021).
- Cauli B et al. Cortical GABA Interneurons in Neurovascular Coupling: Relays for Subcortical Vasoactive Pathways. The Journal of Neuroscience 24, 8940–8949, doi:10.1523/ jneurosci.3065-04.2004 (2004). [PubMed: 15483113]
- McConnell HL, Kersch CN, Woltjer RL & Neuwelt EA The Translational Significance of the Neurovascular Unit. J. Biol. Chem 292, 762–770, doi:doi:10.1074/jbc.R116.760215 (2017). [PubMed: 27920202]
- 32. Bagchi S et al. *In vitro* blood-brain barrier models for drug screening and permeation studies: an overview. Drug Des. Devel. Ther 13, 3591–3605, doi:doi:10.2147/DDDT.S218708 (2019).
- Wong A et al. The blood-brain barrier: an engineering perspective. Front. Neuroeng 6, 7, doi:doi:10.3389/fneng.2013.00007 (2013). [PubMed: 24009582]
- Jaffe EA Cell biology of endothelial cells. Human Pathology 18, 234–239, doi:10.1016/ S0046-8177(87)80005-9 (1987). [PubMed: 3546072]
- Kramer RH, Bensch KG, Davison PM & Karasek MA Basal lamina formation by cultured microvascular endothelial cells. Journal of Cell Biology 99, 692–698, doi:10.1083/jcb.99.2.692 (1984). [PubMed: 6746743]
- Wolburg H, Noell S, Mack A, Wolburg-Buchholz K & Fallier-Becker P Brain endothelial cells and the glio-vascular complex. Cell and Tissue Research 335, 75–96, doi:10.1007/s00441-008-0658-9 (2009). [PubMed: 18633647]
- Ruck T, Bittner S & Meuth SG Blood-brain barrier modeling: challenges and perspectives. Neural Regen Res 10, 889–891, doi:10.4103/1673-5374.158342 (2015). [PubMed: 26199600]
- Shayan G, Choi YS, Shusta EV, Shuler ML & Lee KH Murine in vitro model of the blood–brain barrier for evaluating drug transport. European Journal of Pharmaceutical Sciences 42, 148–155, doi:10.1016/j.ejps.2010.11.005 (2011). [PubMed: 21078386]
- Greene C, Hanley N & Campbell M Claudin-5: gatekeeper of neurological function. Fluids and Barriers of the CNS 16, 3, doi:10.1186/s12987-019-0123-z (2019). [PubMed: 30691500]
- 40. Branca JJV et al. Effects of Cadmium on ZO-1 Tight Junction Integrity of the Blood Brain Barrier. International Journal of Molecular Sciences 20, 6010 (2019).
- 41. Lochhead JJ, Yang J, Ronaldson PT & Davis TP Structure, Function, and Regulation of the Blood-Brain Barrier Tight Junction in Central Nervous System Disorders. Front. Physiol 11, 914, doi:doi:10.3389/fphys.2020.00914 (2020). [PubMed: 32848858]
- 42. Nitta T et al. Size-selective loosening of the blood-brain barrier in claudin-5-deficient mice. J. Cell Biol 161, 653–660, doi:doi:10.1083/jcb.200302070 (2003). [PubMed: 12743111]

- 43. Enerson BE & Drewes LR The Rat Blood—Brain Barrier Transcriptome. Journal of Cerebral Blood Flow & Metabolism 26, 959–973, doi:10.1038/sj.jcbfm.9600249 (2006). [PubMed: 16306934]
- 44. Fabry Z et al. Differential activation of Th1 and Th2 CD4+ cells by murine brain microvessel endothelial cells and smooth muscle/pericytes. The Journal of Immunology 151, 38–47 (1993). [PubMed: 8100844]
- 45. Castro Dias M et al. Brain endothelial tricellular junctions as novel sites for T cell diapedesis across the blood–brain barrier. Journal of Cell Science 134, jcs253880, doi:10.1242/jcs.253880 (2021).
- 46. Greenwood J, Wang Y & Calder VL Lymphocyte adhesion and transendothelial migration in the central nervous system: the role of LFA-1, ICAM-1, VLA-4 and VCAM-1. off. Immunology 86, 408–415 (1995). [PubMed: 8550078]
- Cinamon G, Shinder V & Alon R Shear forces promote lymphocyte migration across vascular endothelium bearing apical chemokines. Nature Immunology 2, 515–522, doi:10.1038/88710 (2001). [PubMed: 11376338]
- 48. Ronaldson PT, Demarco KM, Sanchez-Covarrubias L, Solinsky CM & Davis TP Transforming growth factor-beta signaling alters substrate permeability and tight junction protein expression at the blood-brain barrier during inflammatory pain. Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism 29, 1084– 1098, doi:10.1038/jcbfm.2009.32 (2009).
- Majerova P et al. Trafficking of immune cells across the blood-brain barrier is modulated by neurofibrillary pathology in tauopathies. PLoS One 14, e0217216, doi:doi:10.1371/ journal.pone.0217216 (2019).
- Arima M et al. Claudin-5 Redistribution Induced by Inflammation Leads to Anti-VEGF–Resistant Diabetic Macular Edema. Diabetes 69, 981–999, doi:10.2337/db19-1121 (2020). [PubMed: 32139595]
- Wang J et al. Blood-brain barrier disruption and inflammation reaction in mice after chronic exposure to Microcystin-LR. Science of The Total Environment 689, 662–678, doi:10.1016/ j.scitotenv.2019.06.387 (2019). [PubMed: 31279213]
- 52. Greene C, Hanley N & Campbell M Claudin-5: gatekeeper of neurological function. Fluids and Barriers of the CNS 16, 1–15 (2019). [PubMed: 30616618]
- Xaio H, Banks WA, Niehoff ML & Morley JE Effect of LPS on the permeability of the blood– brain barrier to insulin. Brain Research 896, 36–42, doi:10.1016/S0006-8993(00)03247-9 (2001). [PubMed: 11277970]
- 54. Hansson E Actin Filament Reorganization in Astrocyte Networks is a Key Functional Step in Neuroinflammation Resulting in Persistent Pain: Novel Findings on Network Restoration. Neurochem. Res 40, 372–379, doi:doi:10.1007/s11064-014-1363-6 (2015). [PubMed: 24952067]
- Descamps L, Coisne C, Dehouck B, Cecchelli R & Torpier G Protective effect of glial cells against lipopolysaccharide-mediated blood-brain barrier injury. Glia 42, 46–58, doi:10.1002/glia.10205 (2003). [PubMed: 12594736]
- 56. Hoshi Y, Uchida Y, Tachikawa M, Ohtsuki S & Terasaki T Actin filament-associated protein 1 (AFAP-1) is a key mediator in inflammatory signaling-induced rapid attenuation of intrinsic P-gp function in human brain capillary endothelial cells. Journal of Neurochemistry 141, 247–262, doi:10.1111/jnc.13960 (2017). [PubMed: 28112407]
- Skike CEV et al. Inhibition of mTOR protects the blood-brain barrier in models of Alzheimer's disease and vascular cognitive impairment. American Journal of Physiology-Heart and Circulatory Physiology 314, H693–H703, doi:10.1152/ajpheart.00570.2017 (2018). [PubMed: 29351469]
- Kurz C, Walker L, Rauchmann B-S & Perneczky R Dysfunction of the blood–brain barrier in Alzheimer's disease: Evidence from human studies. Neuropathology and Applied Neurobiology 48, e12782, doi:10.1111/nan.12782 (2022). [PubMed: 34823269]
- 59. Nishihara H et al. Intrinsic blood–brain barrier dysfunction contributes to multiple sclerosis pathogenesis. Brain, awac019, doi:10.1093/brain/awac019 (2022).
- 60. Pan W et al. Cytokine signaling modulates blood-brain barrier function. Curr Pharm Des 17, 3729–3740, doi:10.2174/138161211798220918 (2011). [PubMed: 21834767]

- Zamudio F et al. TDP-43 mediated blood-brain barrier permeability and leukocyte infiltration promote neurodegeneration in a low-grade systemic inflammation mouse model. Journal of Neuroinflammation 17, 283, doi:10.1186/s12974-020-01952-9 (2020). [PubMed: 32979923]
- 62. Saand AR, Yu F, Chen J & Chou SHY Systemic inflammation in hemorrhagic strokes -A novel neurological sign and therapeutic target? J. Cereb. Blood Flow Metab 39, 959–988, doi:doi:10.1177/0271678X19841443 (2019). [PubMed: 30961425]
- Villaseñor R, Lampe J, Schwaninger M & Collin L Intracellular transport and regulation of transcytosis across the blood–brain barrier. Cellular and Molecular Life Sciences 76, 1081–1092, doi:10.1007/s00018-018-2982-x (2019). [PubMed: 30523362]
- 64. Grubb S, Lauritzen M & Aalkjær C Brain capillary pericytes and neurovascular coupling. Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology 254, 110893, doi:10.1016/j.cbpa.2020.110893 (2021).
- 65. Kim S et al. Human bone marrow-derived mesenchymal stem cells play a role as a vascular pericyte in the reconstruction of human BBB on the angiogenesis microfluidic chip. Biomaterials 279, 121210, doi:10.1016/j.biomaterials.2021.121210 (2021).
- 66. Xu L, Nirwane A & Yao Y Basement membrane and blood-brain barrier. Stroke Vasc Neurol 4, 78–82, doi:10.1136/svn-2018-000198 (2018). [PubMed: 31338215]
- 67. Peppiatt CM, Howarth C, Mobbs P & Attwell D Bidirectional control of CNS capillary diameter by pericytes. Nature 443, 700–704, doi:10.1038/nature05193 (2006). [PubMed: 17036005]
- Sá-Pereira I, Brites D & Brito MA Neurovascular Unit: a Focus on Pericytes. Molecular Neurobiology 45, 327–347, doi:10.1007/s12035-012-8244-2 (2012). [PubMed: 22371274]
- Ferro MP, Heilshorn SC & Owens RM Materials for blood brain barrier modeling in vitro. Materials Science and Engineering: R: Reports 140, 100522, doi:10.1016/j.mser.2019.100522 (2020).
- Török O et al. Pericytes regulate vascular immune homeostasis in the CNS. Proceedings of the National Academy of Sciences 118, e2016587118, doi:10.1073/pnas.2016587118 (2021).
- 71. Mäe MA et al. Single-Cell Analysis of Blood-Brain Barrier Response to Pericyte Loss. Circulation Research 128, e46–e62, doi:doi:10.1161/CIRCRESAHA.120.317473 (2021). [PubMed: 33375813]
- 72. Dubrac A et al. NCK-dependent pericyte migration promotes pathological neovascularization in ischemic retinopathy. Nature Communications 9, 3463, doi:10.1038/s41467-018-05926-7 (2018).
- Warmke N et al. Pericyte Insulin Receptors Modulate Retinal Vascular Remodeling and Endothelial Angiopoietin Signaling. Endocrinology 162, bqab182, doi:10.1210/endocr/bqab182 (2021).
- Vazquez-Liebanas E et al. Adult-induced genetic ablation distinguishes PDGFB roles in bloodbrain barrier maintenance and development. Journal of Cerebral Blood Flow & Metabolism 42, 264–279, doi:10.1177/0271678x211056395 (2022). [PubMed: 34689641]
- Rudziak P, Ellis CG & Kowalewska PM Role and Molecular Mechanisms of Pericytes in Regulation of Leukocyte Diapedesis in Inflamed Tissues. Mediators of Inflammation 2019, 4123605, doi:10.1155/2019/4123605 (2019).
- 76. Nag TC, Gorla S, Kumari C & Roy TS Aging of the human choriocapillaris: Evidence that early pericyte damage can trigger endothelial changes. Experimental Eye Research 212, 108771, doi:10.1016/j.exer.2021.108771 (2021).
- 77. Kisler K et al. Acute Ablation of Cortical Pericytes Leads to Rapid Neurovascular Uncoupling. Front. Cell. Neurosci 14, doi:doi:10.3389/fncel.2020.00027 (2020).
- Nikolakopoulou AM et al. Pericyte loss leads to circulatory failure and pleiotrophin depletion causing neuron loss. Nat. Neurosci 22, 1089–1098, doi:doi:10.1038/s41593-019-0434-z (2019). [PubMed: 31235908]
- Gerhardt H, Wolburg H & Redies C N-cadherin mediates pericytic-endothelial interaction during brain angiogenesis in the chicken. Developmental Dynamics 218, 472– 479, doi:10.1002/1097-0177(200007)218:3<472::AID-DVDY1008>3.0.CO;2-# (2000). [PubMed: 10878612]
- 80. Schofield CL, Rodrigo-Navarro A, Dalby MJ, Van Agtmael T & Salmeron-Sanchez M Biochemical- and Biophysical-Induced Barriergenesis in the Blood–Brain Barrier: A Review of

Barriergenic Factors for Use in In Vitro Models. Advanced NanoBiomed Research 1, 2000068, doi:10.1002/anbr.202000068 (2021).

- Tillet E et al. N-cadherin deficiency impairs pericyte recruitment, and not endothelial differentiation or sprouting, in embryonic stem cell-derived angiogenesis. Exp. Cell Res 310, 392–400, doi:doi:10.1016/j.yexcr.2005.08.021 (2005). [PubMed: 16202998]
- 82. Kuo Y-C, Lee C-L & Rajesh R Regulation of human brain vascular pericytes and human astrocytes in a blood–brain barrier model using human brain microvascular endothelial cells: Expression of TGF-β1, VEGF, MMP-9 and P-gp. Journal of the Taiwan Institute of Chemical Engineers 86, 9–17, doi:10.1016/j.jtice.2018.03.003 (2018).
- Zheng Z, Chopp M & Chen J Multifaceted roles of pericytes in central nervous system homeostasis and disease. Journal of Cerebral Blood Flow & Metabolism 40, 1381–1401, doi:10.1177/0271678X20911331 (2020). [PubMed: 32208803]
- 84. Laredo F, Plebanski J & Tedeschi A Pericytes: Problems and Promises for CNS Repair. Front. Cell. Neurosci 13, 546, doi:doi:10.3389/fncel.2019.00546 (2019). [PubMed: 31866833]
- Geranmayeh MH, Rahbarghazi R & Farhoudi M Targeting pericytes for neurovascular regeneration. Cell Communication and Signaling 17, 26, doi:10.1186/s12964-019-0340-8 (2019). [PubMed: 30894190]
- 86. Bohannon DG et al. A subtype of cerebrovascular pericytes is associated with blood-brain barrier disruption that develops during normal aging and simian immunodeficiency virus infection. Neurobiol Aging 96, 128–136, doi:10.1016/j.neurobiolaging.2020.08.006 (2020). [PubMed: 33002766]
- Escartin C et al. Reactive astrocyte nomenclature, definitions, and future directions. Nature Neuroscience 24, 312–325, doi:10.1038/s41593-020-00783-4 (2021). [PubMed: 33589835]
- Gautam J, Cao Y & Yao Y Pericytic Laminin Maintains Blood-Brain Barrier Integrity in an Age-Dependent Manner. Translational Stroke Research 11, 228–242, doi:10.1007/s12975-019-00709-8 (2020). [PubMed: 31292838]
- Yao Y, Chen Z-L, Norris EH & Strickland S Astrocytic laminin regulates pericyte differentiation and maintains blood brain barrier integrity. Nature communications 5, 3413–3413, doi:10.1038/ ncomms4413 (2014).
- Yamanaka G et al. The Neuroinflammatory Role of Pericytes in Epilepsy. Biomedicines 9, 759 (2021). [PubMed: 34209145]
- Miller SJ Astrocyte Heterogeneity in the Adult Central Nervous System. Front. Cell. Neurosci 12, 401, doi:10.3389/fncel.2018.00401 (2018). [PubMed: 30524236]
- Mathiisen TM, Lehre KP, Danbolt NC & Ottersen OP The perivascular astroglial sheath provides a complete covering of the brain microvessels: an electron microscopic 3D reconstruction. Glia 58, 1094–1103, doi:10.1002/glia.20990 (2010). [PubMed: 20468051]
- Wolburg-Buchholz K et al. Loss of astrocyte polarity marks blood-brain barrier impairment during experimental autoimmune encephalomyelitis. Acta Neuropathol 118, 219–233, doi:doi:10.1007/ s00401-009-0558-4 (2009). [PubMed: 19533155]
- 94. Kubotera H et al. Astrocytic endfeet re-cover blood vessels after removal by laser ablation. Sci. Rep 9, 1263, doi:10.1038/s41598-018-37419-4 (2019). [PubMed: 30718555]
- 95. Higashi K et al. An inwardly rectifying K+ channel, Kir4.1, expressed in astrocytes surrounds synapses and blood vessels in brain. American Journal of Physiology Cell Physiology 281, C922– C931, doi:doi:10.1152/ajpcell.2001.281.3.C922 (2001). [PubMed: 11502569]
- 96. Warth A, Mittelbronn M & Wolburg H Redistribution of the water channel protein aquaporin-4 and the K+ channel protein Kir4.1 differs in low- and high-grade human brain tumors. Acta Neuropathologica 109, 418–426, doi:10.1007/s00401-005-0984-x (2005). [PubMed: 15723236]
- 97. Kaisar MA et al. New experimental models of the blood-brain barrier for CNS drug discovery. Expert Opinion on Drug Discovery 12, 89–103, doi:doi:10.1080/17460441.2017.1253676 (2017). [PubMed: 27782770]
- Ponio JB-D et al. Instruction of Circulating Endothelial Progenitors In Vitro towards Specialized Blood-Brain Barrier and Arterial Phenotypes. PLOS ONE 9, e84179, doi:10.1371/ journal.pone.0084179 (2014).

- 99. Huang X et al. Astroglial Connexins in Neurodegenerative Diseases. Front. Mol. Neurosci 14, 657514, doi:doi:10.3389/fnmol.2021.657514 (2021).
- 100. Van Campenhout R et al. Mechanisms Underlying Connexin Hemichannel Activation in Disease. International Journal of Molecular Sciences 22, 3503 (2021). [PubMed: 33800706]
- 101. Peng B, Xu C, Wang S, Zhang Y & Li W The Role of Connexin Hemichannels in Inflammatory Diseases. Biology 11, 237 (2022). [PubMed: 35205103]
- 102. Rouach N, Koulakoff A, Abudara V, Willecke K & Giaume C Astroglial Metabolic Networks Sustain Hippocampal Synaptic Transmission. Science 322, 1551–1555, doi:doi:10.1126/ science.1164022 (2008). [PubMed: 19056987]
- 103. Xing L, Yang T, Cui S & Chen G Connexin Hemichannels in Astrocytes: Role in CNS Disorders. Front. Mol. Neurosci 12, 23, doi:doi:10.3389/fnmol.2019.00023 (2019). [PubMed: 30787868]
- 104. Spampinato SF, Merlo S, Sano Y, Kanda T & Sortino MA Astrocytes contribute to Aβ-induced blood-brain barrier damage through activation of endothelial MMP9. J. Neurochem 142, 464– 477, doi:doi:10.1111/jnc.14068 (2017). [PubMed: 28488764]
- 105. Song S et al. Activation of endothelial Wnt/β-catenin signaling by protective astrocytes repairs BBB damage in ischemic stroke. Prog. Neurobiol 199, 101963, doi:doi:10.1016/ j.pneurobio.2020.101963 (2021).
- 106. Han RT, Kim RD, Molofsky AV & Liddelow SA Astrocyte-immune cell interactions in physiology and pathology. Immunity 54, 211–224, doi:10.1016/j.immuni.2021.01.013 (2021). [PubMed: 33567261]
- 107. Fu J et al. Astrocyte-Derived TGFβ1 Facilitates Blood–Brain Barrier Function via Non-Canonical Hedgehog Signaling in Brain Microvascular Endothelial Cells. Brain Sciences 11, 77 (2021). [PubMed: 33430164]
- 108. Feng S et al. RhoA/ROCK-2 Pathway Inhibition and Tight Junction Protein Upregulation by Catalpol Suppresses Lipopolysaccaride-Induced Disruption of Blood-Brain Barrier Permeability. Molecules 23, 2371 (2018).
- 109. Liu C-C et al. Tau and apolipoprotein E modulate cerebrovascular tight junction integrity independent of cerebral amyloid angiopathy in Alzheimer's disease. Alzheimer's & Dementia 16, 1372–1383, doi:10.1002/alz.12104 (2020).
- 110. Jackson RJ et al. APOE4 derived from astrocytes leads to blood-brain barrier impairment. Brain, awab478, doi:10.1093/brain/awab478 (2021).
- 111. Guérit S et al. Astrocyte-derived Wnt growth factors are required for endothelial blood-brain barrier maintenance. Prog. Neurobiol 199, 101937, doi:doi:10.1016/j.pneurobio.2020.101937 (2021).
- 112. Montagne A et al. APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. Nature 581, 71–76, doi:10.1038/s41586-020-2247-3 (2020). [PubMed: 32376954]
- 113. Yamazaki Y et al. ApoE (Apolipoprotein E) in Brain Pericytes Regulates Endothelial Function in an Isoform-Dependent Manner by Modulating Basement Membrane Components. Arteriosclerosis, Thrombosis, and Vascular Biology 40, 128–144, doi:10.1161/ ATVBAHA.119.313169 (2020). [PubMed: 31665905]
- 114. Zhang ZG, Zhang L, Croll SD & Chopp M Angiopoietin-1 reduces cerebral blood vessel leakage and ischemic lesion volume after focal cerebral embolic ischemia in mice. Neuroscience 113, 683–687, doi:10.1016/S0306-4522(02)00175-6 (2002). [PubMed: 12150788]
- 115. Venkat P et al. Angiopoietin-1 Mimetic Peptide Promotes Neuroprotection after Stroke in Type
  1 Diabetic Rats. Cell Transplantation 27, 1744–1752, doi:10.1177/0963689718791568 (2018).
  [PubMed: 30124060]
- 116. Giovannoni F & Quintana FJ The Role of Astrocytes in CNS Inflammation. Trends Immunol 41, 805–819, doi:doi:10.1016/j.it.2020.07.007 (2020). [PubMed: 32800705]
- 117. Smith BC, Sinyuk M, Jenkins JE, Psenicka MW & Williams JL The impact of regional astrocyte interferon-γ signaling during chronic autoimmunity: a novel role for the immunoproteasome. Journal of Neuroinflammation 17, 184, doi:10.1186/s12974-020-01861-x (2020). [PubMed: 32532298]

- 118. Horng S et al. Astrocytic tight junctions control inflammatory CNS lesion pathogenesis. The Journal of clinical investigation 127, 3136–3151, doi:10.1172/JCI91301 (2017). [PubMed: 28737509]
- 119. Neuhaus W et al. The pivotal role of astrocytes in an in vitro stroke model of the bloodbrain barrier. Front. Cell. Neurosci 8, 352, doi:doi:10.3389/fncel.2014.00352 (2014). [PubMed: 25389390]
- 120. McCandless EE, Wang Q, Woerner BM, Harper JM & Klein RS CXCL12 Limits Inflammation by Localizing Mononuclear Infiltrates to the Perivascular Space during Experimental Autoimmune Encephalomyelitis. The Journal of Immunology 177, 8053–8064, doi:10.4049/ jimmunol.177.11.8053 (2006). [PubMed: 17114479]
- 121. Williams JL et al. Astrocyte-T cell crosstalk regulates region-specific neuroinflammation. Glia 68, 1361–1374, doi:10.1002/glia.23783 (2020). [PubMed: 31961459]
- 122. Michinaga S & Koyama Y Dual Roles of Astrocyte-Derived Factors in Regulation of Blood-Brain Barrier Function after Brain Damage. International Journal of Molecular Sciences 20, 571 (2019).
- 123. Kim GW et al. Neurodegeneration in striatum induced by the mitochondrial toxin 3nitropropionic acid: role of matrix metalloproteinase-9 in early blood-brain barrier disruption? Journal of Neuroscience 23, 8733–8742 (2003). [PubMed: 14507973]
- 124. Psenicka MW, Smith BC, Tinkey RA & Williams JL Connecting Neuroinflammation and Neurodegeneration in Multiple Sclerosis: Are Oligodendrocyte Precursor Cells a Nexus of Disease? Frontiers in Cellular Neuroscience 15, doi:10.3389/fncel.2021.654284 (2021).
- 125. Gao X et al. PD-L1-expressing astrocytes act as a gate-keeper for neuroinflammation in the central nervous system of mice with traumatic brain injury. bioRxiv, 2021.2011.2004.467368, doi:10.1101/2021.11.04.467368 (2021).
- 126. Gao X et al. PD-L1 signaling in reactive astrocytes counteracts neuroinflammation and ameliorates neuronal damage after traumatic brain injury. Journal of Neuroinflammation 19, 43, doi:10.1186/s12974-022-02398-x (2022). [PubMed: 35135580]
- 127. Kummer MP et al. Microglial PD-1 stimulation by astrocytic PD-L1 suppresses neuroinflammation and Alzheimer's disease pathology. The EMBO Journal 40, e108662, doi:doi:10.15252/embj.2021108662 (2021).
- 128. Keir ME, Butte MJ, Freeman GJ & Sharpe AH PD-1 and its ligands in tolerance and immunity. Annu. Rev. Immunol 26, 677–704, doi:doi:10.1146/annurev.immunol.26.021607.090331 (2008). [PubMed: 18173375]
- 129. Latchman Y et al. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. Nat. Immunol 2, 261–268, doi:10.1038/85330 (2001). [PubMed: 11224527]
- 130. Chen G et al. PD-L1 inhibits acute and chronic pain by suppressing nociceptive neuron activity via PD-1. Nature neuroscience 20, 917–926, doi:10.1038/nn.4571 (2017). [PubMed: 28530662]
- 131. Li Q & Barres BA Microglia and macrophages in brain homeostasis and disease. Nature Reviews Immunology 18, 225–242, doi:10.1038/nri.2017.125 (2018).
- Mondo E et al. A Developmental Analysis of Juxtavascular Microglia Dynamics and Interactions with the Vasculature. J. Neurosci 40, 6503–6521, doi:doi:10.1523/jneurosci.3006-19.2020 (2020). [PubMed: 32661024]
- 133. Shigemoto-Mogami Y, Hoshikawa K & Sato K Activated Microglia Disrupt the Blood-Brain Barrier and Induce Chemokines and Cytokines in a Rat in vitro Model. Front. Cell. Neurosci 12, 494, doi:doi:10.3389/fncel.2018.00494 (2018). [PubMed: 30618641]
- 134. da Fonseca ACC et al. The impact of microglial activation on blood-brain barrier in brain diseases. Front. Cell. Neurosci 8, doi:doi:10.3389/fncel.2014.00362 (2014).
- 135. Halder SK & Milner R Mild hypoxia triggers transient blood–brain barrier disruption: a fundamental protective role for microglia. Acta Neuropathologica Communications 8, 175, doi:10.1186/s40478-020-01051-z (2020). [PubMed: 33115539]
- 136. Ulvestad E et al. Reactive microglia in multiple sclerosis lesions have an increased expression of receptors for the Fc part of IgG. Journal of the Neurological Sciences 121, 125–131, doi:10.1016/0022-510X(94)90340-9 (1994). [PubMed: 8158203]

- 137. Chen A-Q et al. Microglia-derived TNF-α mediates endothelial necroptosis aggravating blood brain-barrier disruption after ischemic stroke. Cell Death & Disease 10, 487, doi:10.1038/ s41419-019-1716-9 (2019). [PubMed: 31221990]
- 138. Kuroda E et al. Mouse Bone Marrow-derived Microglia-like Cells Secrete Transforming Growth Factor-β1 and Promote Microglial Aβ Phagocytosis and Reduction of Brain Aβ. Neuroscience 438, 217–228, doi:10.1016/j.neuroscience.2020.05.004 (2020). [PubMed: 32522344]
- 139. Kerkhofs D et al. Pharmacological depletion of microglia and perivascular macrophages prevents Vascular Cognitive Impairment in Ang II-induced hypertension. Theranostics 10, 9512–9527, doi:10.7150/thno.44394 (2020). [PubMed: 32863942]
- 140. Bisht K et al. Capillary-associated microglia regulate vascular structure and function through PANX1-P2RY12 coupling in mice. Nature Communications 12, 5289, doi:10.1038/ s41467-021-25590-8 (2021).
- 141. Császár E et al. Microglia modulate blood flow, neurovascular coupling, and hypoperfusion via purinergic actions. The Journal of experimental medicine 219, doi:10.1084/jem.20211071 (2022).
- 142. Halder SK & Milner R A critical role for microglia in maintaining vascular integrity in the hypoxic spinal cord. Proceedings of the National Academy of Sciences of the United States of America 116, 26029–26037, doi:10.1073/pnas.1912178116 (2019). [PubMed: 31772011]
- 143. Davalos D et al. Fibrinogen-induced perivascular microglial clustering is required for the development of axonal damage in neuroinflammation. Nature Communications 3, 1227, doi:10.1038/ncomms2230 (2012).
- 144. Chen Q et al. Enhanced Expression of PD-L1 on Microglia After Surgical Brain Injury Exerts Self-Protection from Inflammation and Promotes Neurological Repair. Neurochemical Research 44, 2470–2481, doi:10.1007/s11064-019-02864-8 (2019). [PubMed: 31482256]
- 145. Wang L-L et al. The roles of the PD-1/PD-L1 pathway at immunologically privileged sites. American Journal of Reproductive Immunology 78, e12710, doi:10.1111/aji.12710 (2017).
- 146. Zhang Y et al. An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. The Journal of neuroscience : the official journal of the Society for Neuroscience 34, 11929–11947, doi:10.1523/jneurosci.1860-14.2014 (2014). [PubMed: 25186741]
- 147. Tontsch U & Bauer H-C Glial cells and neurons induce blood-brain barrier related enzymes in cultured cerebral endothelial cells. Brain Research 539, 247–253, doi:10.1016/0006-8993(91)91628-E (1991). [PubMed: 1675906]
- 148. Stanness KA et al. A new model of the blood–brain barrier: co-culture of neuronal, endothelial and glial cells under dynamic conditions. Neuroreport 10, 3725–3731, doi:doi:10.1097/00001756-199912160-00001 (1999). [PubMed: 10716199]
- 149. Barreto GE, Gonzalez J, Torres Y & Morales L Astrocytic-neuronal crosstalk: Implications for neuroprotection from brain injury. Neuroscience Research 71, 107–113, doi:10.1016/ j.neures.2011.06.004 (2011). [PubMed: 21693140]
- 150. Mulica P, Grünewald A & Pereira SL Astrocyte-Neuron Metabolic Crosstalk in Neurodegeneration: A Mitochondrial Perspective. Front. Endocrinol. (Lausanne) 12, 668517, doi:doi:10.3389/fendo.2021.668517 (2021).
- 151. Kaplan L, Chow BW & Gu C Neuronal regulation of the blood-brain barrier and neurovascular coupling. Nature Reviews Neuroscience 21, 416–432, doi:10.1038/s41583-020-0322-2 (2020). [PubMed: 32636528]
- 152. Yang G, Zhang Y, Ross ME & Iadecola C Attenuation of activity-induced increases in cerebellar blood flow in mice lacking neuronal nitric oxide synthase. American Journal of Physiology-Heart and Circulatory Physiology 285, H298–H304, doi:10.1152/ajpheart.00043.2003 (2003). [PubMed: 12623792]
- 153. Hogan-Cann AD, Lu P & Anderson CM Endothelial NMDA receptors mediate activity-dependent brain hemodynamic responses in mice. Proceedings of the National Academy of Sciences 116, 10229–10231, doi:10.1073/pnas.1902647116 (2019).
- 154. Lia A et al. Calcium Signals in Astrocyte Microdomains, a Decade of Great Advances. Front. Cell. Neurosci 15, 673433, doi:doi:10.3389/fncel.2021.673433 (2021).

- 155. Semmler A et al. Sepsis causes neuroinflammation and concomitant decrease of cerebral metabolism. Journal of Neuroinflammation 5, 38, doi:10.1186/1742-2094-5-38 (2008). [PubMed: 18793399]
- 156. Pulido RS et al. Neuronal Activity Regulates Blood-Brain Barrier Efflux Transport through Endothelial Circadian Genes. Neuron 108, 937–952.e937, doi:doi:10.1016/j.neuron.2020.09.002 (2020). [PubMed: 32979312]
- 157. Salinas PC & Zou Y Wnt signaling in neural circuit assembly. Annual review of neuroscience 31, 339–358 (2008).
- 158. Gasca-Salas C et al. Blood-brain barrier opening with focused ultrasound in Parkinson's disease dementia. Nature Communications 12, 779, doi:10.1038/s41467-021-21022-9 (2021).
- 159. Zhang W, Mehta A, Tong Z, Esser L & Voelcker NH Development of Polymeric Nanoparticles for Blood–Brain Barrier Transfer—Strategies and Challenges. Advanced Science 8, 2003937, doi:10.1002/advs.202003937 (2021).



#### Figure 1. The cellular components of the neurovascular unit.

The NVU is composed of a complex network of cells that are functionally diverse. ECs form the walls of blood vessels and capillaries and contribute to the formation and maintenance of the basal lamina and extracellular matrix. In addition, tight junctions formed between ECs and expression of adhesion molecules regulate BBB permeability. Pericytes reside in the capillary bed and, with regard to BBB integrity, are primarily responsible for modulation of vascular flow as well as structural changes in tight junctions and the extracellular matrix. Astrocyte endfeet cover 90–95% of the area surrounding the BBB and contribute to a variety of processes that include, but are not limited to, osmotic homeostasis, trophic factor concentration, molecular transport into and out of the perivascular space, and formation of gap junctions under neuroinflammatory conditions. Microglia have been found to cover the remaining space around BBB ECs, respond to injury and infection, and regulate tight junction formation between ECs. Neurons predominantly communicate with astrocyte endfeet and aid in regulation of tight junctions and molecular transport.



#### Figure 2: The NVU interaction network.

Cells within the NVU interact through intricate signaling mechanisms that allow for proper functioning of the BBB. ECs receive a variety of protective signals from other NVU cells that upregulate tight junction formation thus enhancing BBB integrity. These factors include, but are not limited to, TGF $\beta$ , Ang-1, APOE, Shh, Wnts, glial-derived neurotrophic factor, insulin-like growth factor (IGF)-1, and retinoic acid. Contrastingly, ECs may also receive signals that downregulate tight junction proteins, particularly during inflammatory events, such as TNF $\alpha$ , NO, MMPs, endothelins, and glutamate that lead to increased BBB permeability. Importantly, ECs also maintain the ability to signal to pericytes through VEGF and N-cadherin, which are necessary for BBB maintenance. Immune checkpoint proteins, such as the PD-1/PD-L1 complex, regulate the cellular activity of microglia and neurons to modulate BBB integrity as well as dampen the inflammatory activity of infiltrating peripheral immune cells, which increases BBB permeability. Finally, neuronal activity is a critical modulator of cellular and molecular transport across the BBB.