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Integrin expression and function in the response of primary culture hepatic stellate cells to connective tissue growth factor (CCN2)

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Abstract

Production of connective tissue growth factor (CCN2, also known as CTGF) is a hallmark of hepatic fibrosis. This study examined early primary cultures of hepatic stellate cells (HSC) for (i) CCN2 regulation of its cognate receptor integrin subunits; and (ii) interactions between CCN2 and integrin $\alpha 5\beta 1$, heparan sulfate proteoglycans (HSPG) or fibronectin in supporting cell adhesion. HSC were isolated from healthy male Balb/c mice. mRNA levels of CCN2 or $\alpha 5$, $\beta 1$, αv or $\beta 3$ integrin subunits were measured in Day 1–7 primary culture HSC, and Day 3 or Day 7 cells treated with recombinant CCN2 or CCN2 small interfering RNA. Interactions between CCN2 and integrin $\alpha 5\beta 1$, HSPG or fibronectin were investigated using an *in vitro* cell adhesion assay. Coincident with autonomous activation over the first 7 days, primary culture HSC increasingly expressed mRNA for CCN2 or integrin subunits. Addition of exogenous CCN2 or knockdown of endogenous CCN2 differentially regulated integrin gene expression in Day 3 *versus* Day 7 cells. Either full length CCN2 (“CCN2₁₋₄”) or residues 247–349 containing module 4 alone (“CCN2₄”) supported Day 3 cell adhesion in an integrin $\alpha 5\beta 1$ - and HSPG-dependent fashion. Adhesion of Day 3 cells to fibronectin was promoted in an integrin $\alpha 5\beta 1$ -dependent manner by CCN2₁₋₄ or CCN2₄, while fibronectin promoted HSPG-dependent HSC adhesion to CCN2₁₋₄ or CCN2₄. These findings suggest CCN2 regulates integrin expression in primary culture HSC and supports HSC adhesion *via* its binding of cell surface integrin $\alpha 5\beta 1$, a novel CCN2 receptor in primary culture HSC which interacts co-operatively with HSPG or fibronectin.

Keywords

connective tissue growth factor; CCN2; hepatic stellate cells; cell adhesion; integrin; fibronectin

INTRODUCTION

Liver fibrosis or cirrhosis is a common result of chronic liver disease such as that caused by excessive alcohol consumption or hepatitis virus infection. A main feature of liver fibrosis is overt deposition of scar tissue due to an increased expression/synthesis or impaired degradation of extracellular matrix (ECM) components such as collagen types I and III or

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fibronectin (FN), the latter of which is among first such proteins to be deposited in the space of Disse and around the central veins in experimental cirrhosis [1,2]. Hepatic stellate cells (HSC) are a principal source of hepatic ECM, especially during fibrosing liver injury when they become transcriptionally activated and highly responsive to growth factors and chemokines that are released by injured hepatocytes or infiltrating macrophages [3,4]. In their activated state, HSC are highly proliferative, chemotactic and pro-fibrogenic, the latter of which is due to the action transforming growth factor beta 1 (TGF- β 1) which stimulates collagen production and is highly over-expressed in many fibrotic tissues [1–4]. Recently, connective tissue growth factor (CCN2, also known as CTGF), a mosaic matricellular protein containing four evolutionarily conserved structural modules, has emerged as an important player in the fibrotic response because it acts downstream of TGF- β and interacts co-operatively with TGF- β to enhance fibrotic pathways [5,6], although the underlying mechanisms are complex and likely highly dependent on the cell-type and environmental context [5,7]. Nonetheless, CCN2 expression is increased in activated HSC *in vitro* as well as in fibrotic liver tissue and down-regulation of CCN2 gene expression by small interfering RNA (siRNA) or short hairpin RNA reverses, at least partially, HSC activation and the amount of collagen deposition in rodent models of hepatic fibrosis [5,8–13].

The exact mechanisms by which CCN2 drives HSC activation and liver fibrosis are unclear even though it is well-documented that CCN2 plays important roles in cell proliferation, cell survival and apoptosis, cell adhesion and migration [7,14–17]. We and others have previously reported that integrins α v β 3 and α 5 β 1 are CCN2 receptors on, respectively, rat HSC or rat pancreatic stellate cells [15,17–19] and that the principal binding sites for these receptors reside in the C-terminal 103 residues of CCN2, a region that is also the location of one or more heparin-binding domains which function to regulate CCN2 activity via their engagement of cell-surface heparan sulfate proteoglycans (HSPG) [15]. To date, there are no published data regarding the regulation or role of CCN2 receptors in mouse HSC (mHSC) as a function of activation despite the increasing use of mouse models of CCN2 action in fibrosis. Therefore, in this study primary mHSC were used to determine CCN2 regulation of its cognate receptor integrin subunits and to analyze interactions between CCN2 and integrin α 5 β 1, HSPG, or FN in supporting HSC adhesion.

MATERIALS AND METHODS

Reagents

Heparinase I, bovine serum albumin (BSA) and anti-alpha-smooth muscle actin (α -SMA) were purchased from Sigma-Aldrich Co. (St. Louis, MO); Dulbecco's Minimum Essential Medium (DMEM), DMEM/F-12, fetal bovine serum, penicillin/streptomycin, L-glutamine, and trypsin/EDTA were from Mediatech, Inc. (Manassas, VA); OptiPrepTM, Trizol[®], DNA ladder, CyQuant NF Cell Proliferation Assay Kit, Alexa Fluor[®] 568 goat anti-chicken secondary antibody, SuperScriptTM II reverse transcriptase and Platinum[®] Taq DNA Polymerase were from Invitrogen (Carlsbad, CA); Collagenase type IV was from Worthington Biochemical Corporation (Lakewood, NJ); Protease Inhibitor Cocktail (EDTA-free), Pronase E and DNase I were from Roche Diagnostics GmbH (Mannheim, Germany); Silencer[®] Select siRNAs (scrambled, targeting GAPDH or CCN2), siPORTTM NeoFXTM Transfection Agent, Oligo(dT)₁₂₋₁₆ and SYBR[®] Green PCR Master Mix reagent were from Applied Biosystems (Foster City, CA); FN was from EMD Chemicals Inc. (Gibbstown, NJ); anti-antibodies were from Millipore Corporation (Billerica, MA); anti-mouse IgG Texas Red conjugated secondary antibody was from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA); recombinant full length human CCN2 containing all 4 structural modules (termed "CCN2₁₋₄") and an isoform of CCN2 comprising a naturally occurring C-terminal fragment of 103 residues (amino acids 247–349) and which contains module 4 alone (termed "CCN2₄") were produced as we have previously described [20,21]; recombinant human

TGF- β 1 was from eBioscience, Inc. (San Diego, CA). Anti-CCN2 IgY was produced by immunization of chickens with human CCN2 produced in this laboratory.

Isolation and primary culture of mHSC

HSC were isolated from healthy male Balb/c mice (Harlan Laboratories, Indianapolis, IN), 6–8 weeks of age, by a modification of previously described methodology [22]. This protocol was approved by the Institutional Animal Care and Use Committee of The Research Institute at Nationwide Children's Hospital (Columbus, OH). mHSC were harvested by gradient centrifugation using OptiPrep™, resuspended in DMEM/F12 containing 20% fetal bovine serum, 100 U/ml penicillin and 100 mg/ml streptomycin, and plated into plastic tissue culture dishes or tissue culture plates (Corning Incorporated, Corning, NY), and maintained at 37°C in a humidified atmosphere of 5% CO₂/95% air. The next day, cells were replaced with fresh DMEM/F12 containing 10% fetal bovine serum, and medium replacement was then repeated every other day if applicable. By analysis through autofluorescence of vitamin A and trypan blue exclusion, the purity and vitality of the isolated HSCs were more than 95% and 97%, respectively (data not shown).

RNA extraction and reverse transcription

Total cellular RNA was extracted using Trizol Reagent, and reverse transcribed to produce cDNA using Superscript II Reverse Transcriptase and Oligo(dT)₁₂₋₁₆ primers according to the manufactures' directions.

Polymerase chain reaction (PCR) and quantitative real-time PCR

The primer sequences were as follows: α -SMA: 5'-TTC GTT ACT ACT GCT GAG CGT GAG A-3' (sense) and 5'-AAA GAT GGC TGG AAG AGG GTC-3' (antisense) (200 bp); β -actin: 5'-TGT TAC CAA CTG GGA CGA CA-3' (sense) and 5'-CTT TTC ACG GTT GGC CTT AG-3' (antisense) (130 bp); CCN2: 5'-CCA GGA AGT AAG GGA CAC GA-3' (sense) and 5'-GGT TCT CAC TTT GGT GGG AT-3' (antisense) (148 bp); integrin α 5: 5'-CTT AGA ATC CGA GAC TGG GC-3' (sense) and 5'-TCA GGA CAA ATC CCA AAC AA-3' (antisense) (126 bp); integrin α v: 5'-TCC CAG GAC TGT ATT GCT GA-3' (sense) and 5'-CAG TGT GGG CGA AGT AAA TG-3' (antisense) (119 bp); integrin β 1: 5'-TCA CAA ATA CTG TAA GCT GTC C-3' (sense) and 5'-AGC ACT GTC AAA ATG AAA AGG C-3' (antisense) (9200 bp); integrin β 3: 5'-ACT GTG CCT GCC AGG CCT TT-3' (sense) and 5'-TCC TTG GGG CTG CAC TCT TC-3' (antisense) (169 bp). Primers were validated by PCR and products were confirmed by 1.5 % agarose gel electrophoresis using ethidium bromide staining. To measure gene expression, first strand cDNA products mixed with primers and SYBR® Green PCR Master Mix reagent were subjected to quantitative real-time PCR on an Applied Biosystems 7500 Real-Time PCR System, and the cycling parameters were: 50°C for 2 min, 95°C for 10 min, followed by 40 cycles of 95°C for 15 sec and 60°C for 1 min. Samples were run in triplicate, and expression values were normalized to housekeeping gene β -actin primers using the $\Delta\Delta$ Ct method.

Immunofluorescence

Cells grown on coverslips in 6-well or 12-well tissue culture plates were fixed with 4% paraformaldehyde for 15 min at 4°C. After blocking with 5% BSA in PBS, cells were incubated with primary antibody against α -SMA or CCN2 in blocking solution for 1 hr at 37°C, then secondary antibody conjugated with Texas red or Alexa Fluor® 568 for 45 min at room temperature. The coverslips were mounted in 10% glycerol and photomicrographs were recorded with a Zeiss AxioCam HR attached to a Zeiss Axioskop (Carl Zeiss Vision GmbH, Germany).

Cell adhesion assay

Cell adhesion assays were performed as described [15,17]. Briefly, cells were seeded into CCN2- or FN-coated 96-well tissue culture plates in the presence or absence of CCN2 or FN for 30 min. In some experiments, cells were pre-treated with anti-integrin antibody, NaClO₃ and/or Na₂SO₄ for 30 min or 24 hr; specific details are provided in the figure legends. Wells were then rinsed with PBS to remove non-adherent cells and cell adhesion was subsequently measured by fluorescent emission from the wells at 520 nm following addition of 50 μ l CyQuant reagent in lysis buffer.

siRNA transfection

Day 3 or Day 7 cells were transfected with 50 nM siRNA targeting mouse CCN2 gene (siCCN2) (sense, 5'-UGA CAA UAC CUU CUG CAG Att-3'; antisense, 5'-UCU GCA GAA GGU AUU GUC Att-3') or scrambled siRNA using siPORT™ NeoFX™ Transfection Agent according to the manufacturer's directions, and total cellular RNA were harvested 48 hr later. Preliminary siRNA transfection efficiency was determined using siRNA targeting GAPDH gene according to the manufacturer's directions, with siRNA CCN2 transfection producing a knock-down in endogenous CCN2 mRNA expression of about 90%. Preliminary experiments with another siCCN2 (sense, 5'-AGA AUA UGA UGU UCA UCA Att-3'; antisense, 5'-UUG AUG AAC AUC AUA UUC Utt-3') demonstrated similar results regarding CCN2 knockdown efficiency and regulation of integrin subunits gene expression (data not shown), showing that the effects observed were authentic rather than non-specific or off-target in nature.

Statistical analysis

Data are presented as mean \pm SD and ANOVA one-way test was performed with $p < 0.05$ as statistical significance.

RESULTS

CCN2 mRNA expression in primary cultures of mHSC

As assessed by reverse transcription PCR, immunostaining, or quantitative real-time PCR, mHSC underwent autonomous activation over the first 7 days of culture as shown by increased expression of mRNA for α -SMA, a well-characterized HSC activation marker (Fig. 1). During the same time period, CCN2 gene expression increased as well (Fig. 2), consistent with previous studies which documented activation-associated expression of CCN2 in primary HSC from rats [10,12].

mRNA expression of the α v, β 3, α 5, or β 1 integrin subunits CCN2 in primary cultures of mHSC

During the first 7 days of culture, expression in mHSC of mRNA for the α v, β 3, α 5, or β 1 integrin subunits increased gradually, with a variable expression pattern for each subunit (Fig. 3). Maximal expression levels were observed on Day 5 for the α 5 subunit, on Days 5–6 for the β 1 subunit, and on Day 7 for the α v and β 3 subunits (Fig. 3).

Differential modulation of the α 5, α v, β 1 and β 3 integrin subunits by CCN2 in primary cultures of mHSC

When added to transitional mHSC on Day 3, recombinant human CCN2₁₋₄ (200 ng/ml) caused increased expression of the α 5, β 1, or α v integrin subunits by 4.19-, 1.36- or 1.14-fold respectively, while expression of the integrin β 3 subunit was reduced by 48% (Fig. 4A). On the other hand, in activated mHSC on Day 7, CCN2 caused about an 11-fold increased

expression of the $\beta 1$, αv , or $\beta 3$ subunits while expression of the $\alpha 5$ subunit was reduced by 48% (Fig. 4B).

These results thus show that CCN2 modulates gene expression of its cognate receptor integrin subunits - integrins αv , $\beta 3$, $\alpha 5$, and $\beta 1$ - in primary cultures of mHSC isolated from normal mice. Meanwhile, CCN2 differentially regulates expression of the above individual integrin subunit gene expression in early and activated primary cultures of mHSC. A role for endogenous CCN2 in integrin expression in mHSC was demonstrated by treating the cells with 50 nM siCCN2. In Day 3 transitional cells, this treatment down-regulated the $\alpha 5$ subunit by 95% but up-regulated the $\beta 3$ or $\beta 1$ subunits by 1.85- or 3.85-fold respectively (Fig. 5A) whereas in Day 7 activated cells it caused a down-regulation of the $\alpha 5$, αv , or $\beta 3$ subunits by 95%, 26% or 47% respectively but 6.73-fold up-regulation of the $\beta 1$ subunit (Fig. 5B). The efficacy of the siCCN2 was confirmed by immunostaining of mHSC which showed decreased in CCN2 protein on Day 3 or Day 7 (Fig. 5C). These results thus confirm that CCN2 differentially modulates gene expression of the above integrin subunits in primary cultured mHSC.

Integrin- or HSPG-dependency of CCN2-mediated mHSC adhesion

As compared to plates coated with albumin alone, adhesion of Day 3 mHSC was 2-, 2.5- or 2.3-fold higher in plates coated with, respectively, CCN2₁₋₄, CCN2₄, or FN (Fig. 6). Pre-treatment of the cells with 10 mM sodium chlorate to block sulfation of HSPGs inhibited mHSC adhesion to CCN2₁₋₄ or CCN2₄ by 63% and 43%, respectively, but concurrent incubation with 10 mM sodium sulfate reversed this effect (Fig. 6). Pre-incubation of the cells with anti-integrin $\alpha 5\beta 1$ antibody (10 μ g/ml) decreased their adhesion to either CCN2₁₋₄ or CCN2₄ by 68% and 57%, respectively (Fig. 6). These data demonstrate that adhesion to CCN2 by transitional mHSC involves engagement by cell surface HSPG and/or integrin $\alpha 5\beta 1$ with one or more binding sites in the C-terminal 103 residues of CCN2.

Co-operative interaction between CCN2 and FN in promoting mHSC adhesion

CCN2₁₋₄ or CCN2₄ (200 ng/ml) promoted mHSC adhesion to FN by 2.2- and 1.5-fold, respectively, but their effects were decreased 62% or 50% respectively by anti-integrin $\alpha 5\beta 1$ antibody (10 μ g/ml) (Fig. 7). Similarly, FN (200 ng/ml) promoted mHSC adhesion to CCN2₁₋₄ by 1.9-fold, but pre-treatment of the cells with heparinase I (2 units/ml) decreased FN-enhanced cell adhesion by 53%, and prior occupancy of heparin-binding site by pre-incubation with exogenously added heparin (2 μ g/ml) decreased the cell binding to FN by 59% (Fig. 8). On the other hand, FN promoted mHSC adhesion to CCN2₄ by 1.2-fold, an effect that was inhibited by 37% with heparin or by 29% after pre-treatment of the cells with heparinase I (Fig. 8). The effect of heparinase I was unaffected by exposure to Protease Inhibitor Cocktail, showing that contaminating proteolytic enzymes did not account for the decreased cell adhesion (Fig. 8).

DISCUSSION

Detailed investigations over the last decade have firmly established that integrins are a principal class of cell-surface receptors for CCN2 and related molecules [23]. The binding interactions between these proteins and specific integrins are complex and involve engagement of unique non-Arg-Gly-Asp motifs in the ligands that in some cases are functionally regulated by HSPG co-receptors [15,17,19,23-27]. Engagement of cell surface integrins and HSPG by CCN2 likely represents a major mechanism for modulating the responses of the target cell to a diverse variety of molecular stimuli and environmental cues.

We have previously shown that adhesion of primary rat HSC to CCN2 employs an integrin $\alpha v \beta 3$ binding site located in residues 247–239 [15] and that adhesion of the rat HSC-T6 cell line to CCN2 involves an integrin $\alpha 6 \beta 1$ binding site in residues 199–243 [28]. In other studies we showed that integrin $\alpha 5 \beta 1$ supported CCN2-mediated adhesion of primary rat pancreatic stellate cells [17,18]. In view of the emerging importance of mouse models in probing CCN2 action in the liver *in vivo* [29,30], studies of integrin-CCN2 interactions in primary cultures of mHSC are important so that underlying mechanisms can be explored. Therefore, in this investigation, we examined the role of integrins in CCN2-dependent mHSC adhesion. Since quiescent HSC typically exhibit low expression of CCN2 and integrins, our study was focused on a relatively early stage of HSC culture to establish which integrins are functional as CCN2 receptors during the period of transitional activation. Using Day 3 cells we showed that CCN2 is indeed an adhesive substrate and that this is attributable to engagement by CCN2 of integrin $\alpha 5 \beta 1$ and HSPG. Additionally, since the binding of CCN2₄ was both HSPG- and integrin $\alpha 5 \beta 1$ -dependent, the data demonstrate the importance of binding sites for these moieties in the C-terminal 103 residues of CCN2. Although the precise location of the integrin $\alpha 5 \beta 1$ binding site for mHSC has yet to be mapped in CCN2, we have previously identified an integrin $\alpha 5 \beta 1$ binding domain in residues 285–291 of CCN2 that supports adhesion of rat pancreatic stellate cells [17]. Moreover, heparin-binding domains in CCN2 have been localized to stretches of highly basic residues in the C-terminal portion of the CCN2 molecule, including residues 247–260 and residues 274–286 [31].

Since blocking antibodies to the mouse form of integrin $\alpha v \beta 3$ are not available, we cannot exclude the possibility that integrin $\alpha v \beta 3$ also plays a role in CCN2-induced adhesion of mHSC, as we have shown previously for rat HSC [15]. Indeed incubation of Day 3 mHSC with echistatin, a snake venom disintegrin with high specificity for integrin $\beta 1$ and $\beta 3$, was somewhat more effective than anti-integrin $\alpha 5 \beta 1$, anti-integrin $\beta 1$ or anti-integrin $\beta 3$ in blocking CCN2-induced cell adhesion (data not shown) suggesting that additional integrins likely function as CCN2 adhesion receptors in transitional mHSC.

In this study we showed that CCN2 expression increases as a function of culture-induced activation, consistent with previous data from other systems [10]. While it has been documented that certain integrins are up-regulated in fibrotic liver disease or during HSC activation [32–36], previous studies have not addressed whether this phenomenon may be related to the action of CCN2 itself, leading us to investigate if integrin expression by mHSC was CCN2-dependent. Previously in human mesangial cells, CCN2 was shown to stimulate the production of integrin $\alpha 5 \beta 1$ and to mediate, at least partially, TGF- $\beta 1$ -induced gene expression of integrin $\alpha 5 \beta 1$ [27]. The results from our study show that, when treated with recombinant CCN2, mHSC demonstrated enhanced levels of the $\alpha 5$, $\beta 1$, or αv integrin subunits in Day 3 cells and of the $\beta 1$, αv , or $\beta 3$ subunits in Day 7 cells. On the other hand, the $\beta 3$ or $\alpha 5$ subunits were reduced by CCN2 treatment on, respectively Days 3 or 7, suggesting that integrin expression in response to CCN2 is highly dependent on the state of activation of the cells. This conclusion is supported by the siRNA data which showed that down-regulation of endogenous CCN2 expression led to variable changes in integrin expression depending on the subunit in question and the stage of culture. Therefore, while these data show clearly that integrin subunit expression in primary mHSC is CCN2-dependent, the precise role played by CCN2 is especially influenced by the particular microenvironment at a specific point in time. This contextual action is fully consistent with the emerging matricellular action of CCN2 [37]. We have further shown that the integrin $\alpha 6$ subunit, which also functions as a CCN2 receptor [28], is also produced by primary mHSC on Day 3 and Day 7 but its expression appears to be independent of CCN2 stimulation (data not shown).

In this study we demonstrated that FN, which is also an integrin $\alpha 5 \beta 1$ ligand in HSC [38], interacts co-operatively with CCN2 to promote mHSC. Similar interactions have been reported for fibroblasts [24], chondrocytes [25], and oval cells [19], and FN has been shown to regulate the cellular response to CCN2 [39]. In the mHSC system, incubation with anti-integrin $\alpha 5 \beta 1$ antibody or heparinase decreased CCN2₁₋₄- or CCN2₄-induced cell adhesion to FN, demonstrating that residues 247–349 were involved in this modulatory effect. Consistent with this observation are previous reports which showed that residues 249–349 of CCN2 were involved in a direct binding interaction with the FN protein which enhanced chondrocyte adhesion via integrin $\alpha 5 \beta 1$ [25]. From studies of hepatic oval cells, an additional FN-binding site has been localized to residues 21–107 of CCN2 [19] and therefore a role for this site in co-operative binding interactions between CCN2₁₋₄ and FN in our mHSC studies cannot currently be excluded. Whatever the nature of the underlying molecular mechanisms, the data are important in the context of fibrotic liver disease in which the co-operative interactions between CCN2 and FN are likely manifested within the provisional matrix which contains enriched levels of each molecule.

In summary, the current study demonstrates that during primary culture, mHSC increasingly express CCN2, as they become autonomously activated *in vitro*, and that CCN2 differently modulates its cognate integrin receptor subunits ($\alpha 5$, αv , $\beta 1$, $\beta 3$) in transitional or activated mHSC. Moreover, CCN2 promotes heparin-dependent adhesion of early primary culture mHSC through its C-terminal domain and it interacts co-operatively with FN through a common cell surface receptor, integrin $\alpha 5 \beta 1$. Collectively these data demonstrate the presence of complex interacting pathways between CCN2, its integrin receptors, and other binding moieties (e.g. HSPG, FN) in regulating HSC function.

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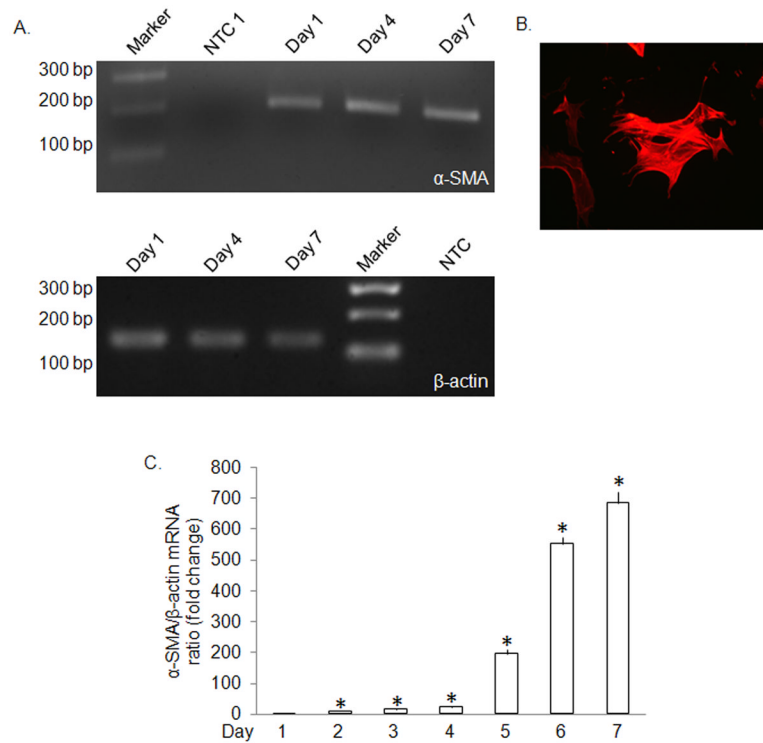


Fig. 1. α -SMA production by primary culture mHSC. **(A)** Reverse-transcriptase PCR of total cellular RNA extracted from primary mHSC for determination of α -SMA gene expression on Day 1, Day 4 or Day 7 of primary culture. A DNA marker ladder and non-cDNA template (NTC) are also shown. **(B)** Immunofluorescent detection of α -SMA protein in Day 3 mHSC (magnification X100). This staining pattern was seen in about 40% of the cells on Day 3. **(C)** Quantitative real-time PCR detection of α -SMA mRNA on Days 1–7. Data are the means \pm SD of triplicate determinations and are representative of three independent experiments. * $p < 0.05$ vs Day 1.

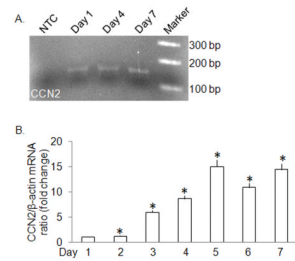


Fig. 2. CCN2 expression by primary culture mHSC. **(A)** Reverse-transcriptase PCR of total cellular RNA extracted from primary mHSC for determination of CCN2 gene expression on Day 1, Day 4 or Day 7 of primary culture. A DNA marker ladder and non-cDNA template (NTC) are also shown. **(B)** Quantitative real-time PCR detection of CCN2 mRNA on Days 1–7. Data are the means \pm SD of triplicate determinations and are representative of three independent experiments. * $p < 0.05$ vs Day 1.

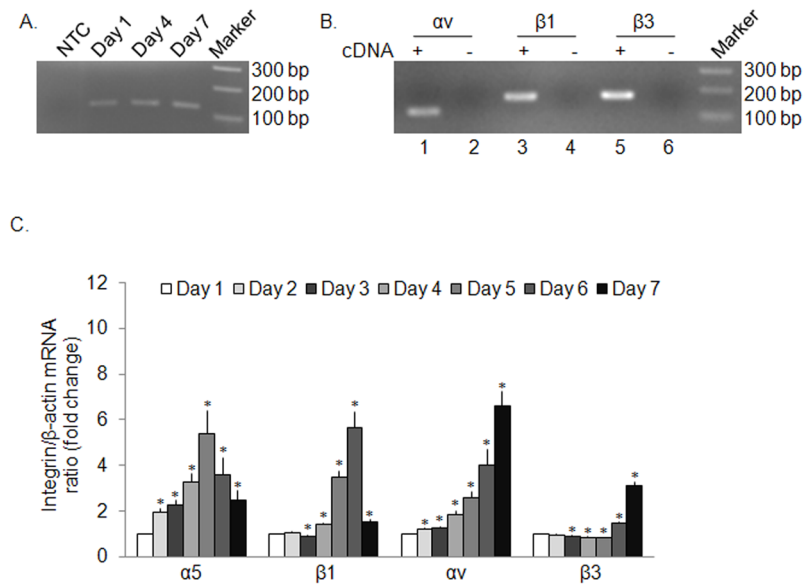


Fig. 3. Integrin expression by primary culture mHSC. Total cellular RNA extracted from primary cultures of mHSC was subjected to reverse transcription, followed by PCR (A, B) or real-time PCR (C) to determine gene expression of the $\alpha 5$, αv , $\beta 1$ or $\beta 3$ integrin subunits. (A) RT-PCR for integrin $\alpha 5$ assessed on Day 1, Day 4 or Day 7. A DNA marker ladder and non-cDNA template (NTC) are also shown. (B) RT-PCR for integrin αv (lane 1), $\beta 1$ (lane 3) or $\beta 3$ (lane 5) and their non-cDNA template controls (lanes 2, 4 and 6 respectively). A DNA marker ladder is also shown. (C) Quantitative real-time PCR detection of integrins $\alpha 5$, αv , $\beta 1$ or $\beta 3$ on Days 1–7. Data are the means \pm SD of triplicate determinations and are representative of three independent experiments. * $p < 0.05$ vs Day 1.

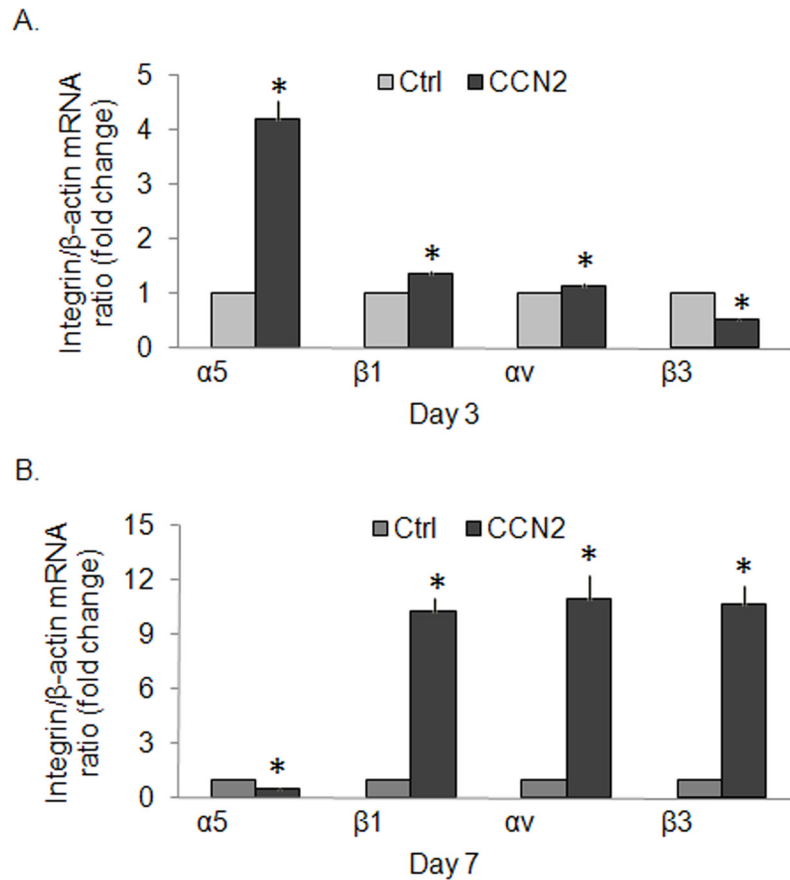


Fig. 4. Regulation of integrin subunit expression by recombinant CCN2. mHSC on (A) Day 3 or (B) Day 7, both of which were pre-incubated in DMEM/F12 containing 0.5% fetal bovine serum for 24 hr, were treated with or without CCN2 (200 ng/ml) for an additional 24 hr prior to analysis of integrin gene expression by quantitative real-time PCR. The data are the means \pm SD of triplicate determinations and are representative of three independent experiments. Ctrl, non-treated cells. * $p < 0.05$ vs Ctrl.

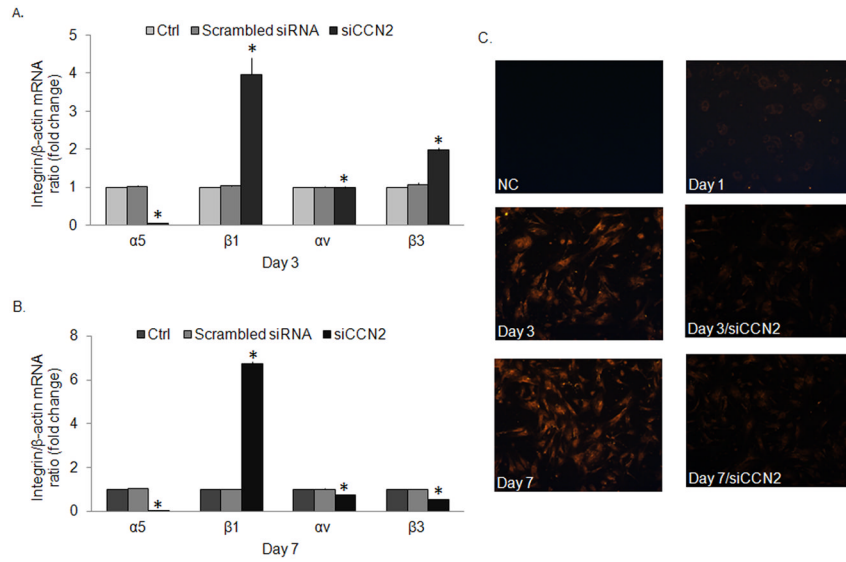


Fig. 5. CCN2 siRNA-mediated down regulation of integrin expression. mHSC on (A) Day 3 or (B) Day 7 were treated for 48 hr with siCCN2 (50 nM) prior to analysis of mRNA expression of the integrin $\alpha 5$, αv , $\beta 1$ or $\beta 3$ subunits by quantitative real-time PCR. Data are the means \pm SD of triplicate determinations and are representative of three independent experiments. * $p < 0.05$ vs Ctrl. (C) Immunofluorescent detection of CCN2 in Day 1 (without siCCN2 treatment), Day 3 or Day 7 mHSC treated with or without siCCN2 (magnification X100). NC, negative control.

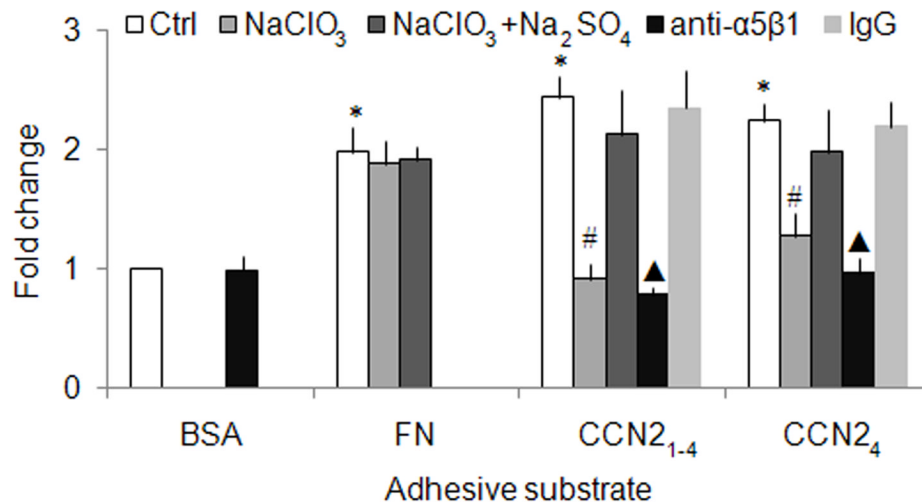


Fig. 6. CCN2 or FN promotes mHSC adhesion. Day 3 mHSC were pre-incubated with 10 mM NaClO₃ in the presence or absence of 10 mM Na₂SO₄, for 24 hours or with 10 μg/ml anti-integrin α5β1 antibody for 30 mins and then plated for 30 minutes into 96-well tissue culture plates (10⁵ cells/well in 100 μl of medium) that had been pre-coated with 2 μg/ml CCN2₁₋₄, CCN2₄ or FN. Wells were then washed and the frequency of adherent cells was evaluated using a CyQuant NF Assay Kit. The data are the means ± SD of quadruplicate determinations and are representative of three independent experiments. **p*<0.05 vs BSA-coated group; #*p*<0.05 vs Ctrl or NaClO₃+Na₂SO₄ group; ▲*p*<0.05 vs Ctrl group.

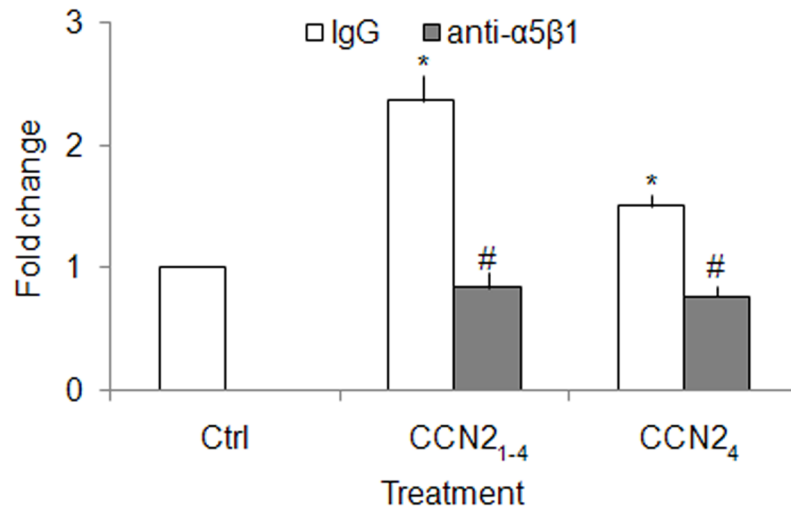


Fig. 7.

CCN2 promotes mHSC cell adhesion to FN. Day 3 primary mHSC were pre-incubated with or without 10 $\mu\text{g/ml}$ anti-integrin $\alpha 5\beta 1$ for 30 min and then placed for 30 mins in 96-well tissue culture plates (10^5 cells/well in 100 μl of medium) pre-coated with FN (2 $\mu\text{g/ml}$) in the presence or absence of 200 ng/ml CCN2₁₋₄ or CCN2₄. Wells were washed and the frequency of adherent cells was evaluated using a CyQuant NF Assay Kit. Data are representative of 3 separate experiments. Values represent mean \pm SD of quadruplicate determinations from 3 experiments. * $p < 0.05$ vs Ctrl group; # $p < 0.05$ vs IgG group.

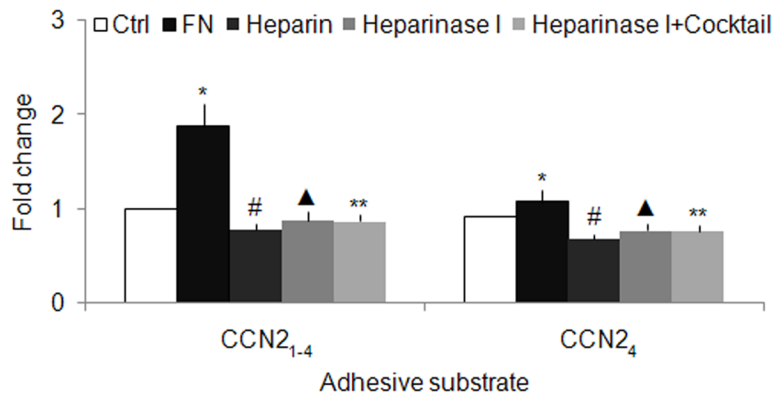


Fig. 8.

FN promotes mHSC adhesion to CCN2. Day 3 primary mHSC were pre-incubated with or without 2 $\mu\text{g/ml}$ heparin or 2 units/ml heparinase I in the presence or absence of 1X Protease Inhibitor Cocktail for 30 min, and then placed for 30 mins in 96-well tissue culture plates (10⁵ cells/well in 100 μl of medium) that had been pre-coated with 2 $\mu\text{g/ml}$ CCN2₁₋₄ or CCN2₄, in the presence or absence of 200ng/ml FN. Wells were washed and frequency of adherent cells was evaluated using a CyQuant NF Assay Kit. Data are representative of 3 separate experiments. Values represent mean \pm SD of quadruplicate determinations from 3 experiments. Cocktail, Protease Inhibitor Cocktail. * $p < 0.05$ vs Ctrl group; # $p < 0.05$ vs FN group; ▲ $p < 0.05$ vs FN group; ** $p > 0.05$ vs Heparinase I group.