



Published in final edited form as:

Am J Cancer Res. 2011 January 1; 1(3): 275–300.

Squamous Cell Carcinoma – Similarities and Differences among Anatomical Sites

Wusheng Yan¹, Ignacio I. Wistuba², Michael R. Emmert-Buck¹, and Heidi S. Erickson³

Wusheng Yan: yanw@mail.nih.gov; Ignacio I. Wistuba: iiwistuba@mdanderson.org; Michael R. Emmert-Buck: mbuck@helix.nih.gov; Heidi S. Erickson: HSErickson@mdanderson.org

¹Pathogenetics Unit, Laboratory of Pathology, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892

²Thoracic Molecular Pathology Lab, Departments of Pathology and Thoracic/Head & Neck Medical Oncology, University of Texas MD Anderson Cancer Center, Houston, TX 77030

³Thoracic Molecular Pathology Lab, Department of Thoracic/Head & Neck Medical Oncology, University of Texas MD Anderson Cancer Center, Houston, TX 77030

Abstract

Squamous cell carcinoma (SCC) is an epithelial malignancy involving many anatomical sites and is the most common cancer capable of metastatic spread. Development of early diagnosis methods and novel therapeutics are important for prevention and mortality reduction. In this effort, numerous molecular alterations have been described in SCCs. SCCs share many phenotypic and molecular characteristics, but they have not been extensively compared. This article reviews SCC as a disease, including: epidemiology, pathology, risk factors, molecular characteristics, prognostic markers, targeted therapy, and a new approach to studying SCCs. Through this comparison, several themes are apparent. For example, HPV infection is a common risk factor among the four major SCCs (NMSC, HNSC, ESCC, and NSCLC) and molecular abnormalities in cell-cycle regulation and signal transduction predominate. These data reveal that the molecular insights, new markers, and drug targets discovered in individual SCCs may shed light on this type of cancer as a whole.

Keywords

Squamous cell carcinoma (SCC); Non-melanoma skin cancer (NMSC); Head and neck squamous cell carcinomas (HNSCC); esophageal squamous cell carcinoma (ESCC); Non-small cell lung cancer (NSCLC); epidemiology; risk factors; molecular characteristics; prognostic markers; targeted therapy

1. Introduction

Squamous cell carcinoma (SCC) is an epithelial malignancy that occurs in organs that are normally covered with squamous epithelium which includes several different anatomic sites, including the skin, lips, mouth, esophagus, urinary tract, prostate, lungs, vagina, and cervix. Of these anatomic sites, there are four which make up the majority of SCC cases: non-melanoma skin cancer, head and neck cancer, esophageal cancer, and non-small cell lung

Corresponding Authors: Dr. Heidi S. Erickson, Department of Thoracic / Head & Neck Medical Oncology, UT MD, Anderson Cancer Center, Houston, Texas 77030. HSErickson@mdanderson.org, Dr. Michael R. Emmert-Buck, Pathogenetics Unit, Laboratory of Pathology, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, Maryland 20892. mbuck@helix.nih.gov.

cancer. Given the range of tissues in which it arises, SCC represents the most common cancer capable of metastatic spread in the US and worldwide [1]. Despite advances in diagnostic methods and combined treatment modalities, the survival rate has not improved significantly over the last 30 years [2] due in part to a lack of reliable early diagnostic biomarkers and a limited number of molecularly targeted therapeutic strategies.

Numerous genetic alterations have been described in SCC sub-types, although the molecular mechanisms contributing to tumor initiation and progression are still poorly understood. SCCs share many phenotypic and molecular characteristics with each other [3–5], thus molecular insights, new markers, or drug targets discovered in individual SCCs may shed light on this type of cancer as a whole. In this article we will review SCC as a disease by describing the most common anatomic types of SCC with regard to their epidemiology, pathology, and risk factors. We will also review the current understanding of the molecular characteristics and prognostic markers. And finally, we will focus on targeted therapy and new approaches to studying SCC.

2. Epidemiology and Pathology

2.1. Non-melanoma skin cancer

Non-melanoma skin cancer (NMSC) is the most common cancer in humans [6], which includes SCC and basal cell carcinoma (BCC), and has shown a dramatic increase in Caucasians in the last few decades. Although BCC is most prevalent, SCC has the higher mortality due to metastases and high incidence [7]. The number of skin cancers diagnosed in the United States outnumbers all other cancers combined, and it is estimated that one in five Americans will develop skin cancer at some point in their life [8]. Most skin SCCs show relatively benign behavior and can be cured by local surgical and dermatologic methods. However, some of these lesions can have a locally invasive and aggressive course. The rate of metastasis is 0.3% to 3.7%, with an overall 5-year survival rate of less than 30% when systemic disease develops [9].

2.2. Head and neck squamous cell carcinomas

Head and neck squamous cell carcinomas (HNSCC) make up the vast majority (more than 90%) of head and neck cancers and rank as the sixth most common cancer worldwide [10], with 45,660 new cases of HNSCC diagnosed in 2007 and 35,720 new cases reported in the US during 2009 [11]. They are a group of tumor entities that arise from squamous mucosal surfaces, including nasal cavities, paranasal sinuses, oral cavity, nasopharynx, oropharynx, hypopharynx, and larynx. In contrast to the declining overall incidence of HNSCC, which is mainly due to smoking prevention and cessation [12], oropharynx carcinoma shows a rising incidence, particularly among individuals less than 45 years of age, suggesting some nontraditional behavioral and environmental factors play a key role in its epidemiology. HNSCC has a 75% overall 5-year survival rate if detected early [13]. Despite advances in detection and treatments over recent decades, most patients present with metastatic disease at the time of diagnosis, reducing the overall 5-year survival rate to 35% [14]. Late diagnosis, formation of additional primary tumors, and metastases largely contribute to this poor survival rate [15].

2.3. Esophageal squamous cell carcinoma

Esophageal cancer (EC) ranks as the eighth most common cancer, with the sixth highest mortality in the world [16, 17]. As the predominant histological subtype of esophageal cancer, esophageal squamous cell carcinoma (ESCC) contributed 80% of all esophageal cancers worldwide. ESCC is characterized by extreme diversity in geographical distribution and high mortality. The "Asian esophageal cancer belt" region shows much higher incidence

than other areas of the world. For example, Linxian and surrounding counties in China [18]. Despite advances in diagnostic methods and combined treatment modalities, the majority of tumors are diagnosed at advanced stages and the overall 5-year survival rate is only 40% [19]. Although relatively less common in the United States than in other countries, there were still 15,560 new cases and 13,940 deaths reported in 2007, which was the sixth leading cause of death from cancers among American men that year [20]. In the US, ESCC occurs more commonly in African American than Caucasian patients and more commonly in men than women, although the prevalence in women has been increasing steadily [21]. The majority of ESCC patients present with advanced metastatic disease, with the overall 5-year survival of these patients being <10% [22].

2.4. Non-small cell lung carcinomas

Lung cancer is the leading cause of cancer death in the United States and most other countries [23], with approximately 30% being SCC [24]. Lung cancers are divided into small cell (SCLC) and non-small cell lung carcinomas (NSCLC) based on their histology and cellular origin. Non-small cell lung cancer (NSCLC) accounts for approximately 80–85% of all cases of lung cancer and is the most common cause of death in men and second only to breast cancer in woman [25]. NSCLC are classified into four histologic subtypes: squamous cell carcinoma (SCC), adenocarcinoma (ADC), large cell carcinoma, and sarcomatoid carcinoma. Anatomically, about 70% of SCC present as central lung tumors [26], whereas adenocarcinomas generally present as peripheral lung tumors [24]. A recent large, randomized phase III trial showed that platinum-based chemotherapy combinations yield a median survival time of only 8–11 months, a 1-year survival rate of 30–45%, and a 2-year survival rate of 10–20% [27, 28]. The overall 5-year survival rate for lung cancer is less than 14% [29].

2.5. Overall comparison

Overall 5-year survival rates for the four major SCCs are among the lowest of the major cancers. NMSC has an advantage over the other SCCs, as it is presented on the skin surface and not an internal organ; therefore, the chance of early detection is much greater and it is often cured by dermatologic and local surgical methods. For all of the major SCCs, including NMSC, a common theme of late diagnosis, formation of additional primary tumors and metastases are associated with the poor survival rates presented above. Regional recurrence after surgical resection is also a contributing factor as is seen more commonly in NSCLC SCC than other histologic subtypes because SCC is able to spread by extending through peribronchial tubes which allow them to directly invade mediastinal lymph nodes and other mediastinal structures [26, 30]. Even though NMSC has an early detection advantage over the other SCCs, repeated exposure to risk factors will influence severity of disease progression and recurrence as equally as observed in the other major SCCs.

3. Risk Factors

The incidence of SCC shows marked variation in its distribution, suggesting that personal habits, environmental exposures, infections, and ethnicity all play a role in the etiology of SCC (Table 1), with several of these risk factors influencing prognosis (Table 2).

3.1. Non-melanoma skin cancer

Unlike other types of squamous cell carcinoma, NMSC is primarily caused by chronic long-term UV solar radiation exposure [31], in conjunction with the patient's skin type. Fair-skinned individuals who always burn and never tan are at a much higher risk for developing skin SCC than those with darker-skin [32], and it has been demonstrated that both sun exposure earlier in life and intense sun exposure appear to heavily predispose the

populations to skin cancer [32]. Furthermore, human papilloma viruses (HPV) may be involved in the multi-step process of skin carcinogenesis as a co-factor with UV-radiation [33], especially in patients with poor immune status such as organ transplant recipients [34]. And, smoking tobacco may double the risk of skin cancer [35], thus although the effect is not as great as in other SCCs, smoking plays a role in the development of NMSC.

3.2. Head and neck squamous cell carcinomas

Contrary to NMSC, alcohol and tobacco use are the most common risk factors for HNSCC in the US, although they have not been associated with survival [36]. Moreover, alcohol and tobacco are likely synergistic in causing cancer of the head and neck [37]. Cigarette smokers have a lifetime increased risk for head and neck cancers which is 5- to 25-fold increased over the general population [38], and smoking cessation does not eliminate the risk of cancer development [39]. In addition, environmental exposure to tobacco smoke also increases the risk of developing HNSCC, even for individuals who have never actively smoked [40]. Heavy alcohol consumption is also an independent risk factor for HNSCC, particularly for cancers of the hypopharynx [41]. Moreover, smokers and alcohol drinkers are at risk for the development of second primary oral cancers [42]. Interestingly, even in the presence of alcohol consumption or tobacco use, a high intake of fruit and vegetables may prevent the development of a quarter of HNSCC and possibly one half of oral and oropharyngeal SCC [43]. Causation has been shown with viral infection for HNSCC and the association varies based on the site of the tumor. For example, human papilloma virus (HPV), in particular HPV16, shows the highest distribution in the tonsils [44], while Epstein-Barr virus (EBV) infection is associated with nasopharyngeal cancer. HPV is associated with 20–25% of HNSCC, and individuals with HPV-positive tumors have a better overall survival compared to those with HPV-negative tumors [45, 46]. Specifically, the presence of HPV-16 is now recognized as a highly favorable prognostic indicator for patients with HNSCC [45]. Betel quid chewing, a common habit in some regions of Asia and some Asian communities in the western world is considered a regional risk factor for cancers with a poorer prognosis [36]. In addition, oral health, acid reflux disease and environmental exposures (nickel refining, textile fibers, and woodworking) are also related to HNSCC tumorigenesis.

3.3. Esophageal squamous cell carcinoma

Similar to HNSCC, smoking and alcohol ingestion are major etiologic factors for the development of ESCC [47]. Studies have shown that ESCC risk is increased approximately three- to seven-fold in current smokers [48–50] and three- to five-fold in heavy alcohol users [50–53], with additional associations between esophageal irritants such as lye ingestion, rapidly consumed high-starch diets without fruits and vegetables, and radiation therapy [47]. There also may be a causal relationship between ESCC and previous diseases such as achalasia, head and neck cancer, and Plummer-Vinson syndrome [54]. Pickled vegetable intake and micronutrient deficiency (such as zinc) may contribute to ESCC formation in some parts of China, especially in light of laboratory experiments demonstrating that high tissue zinc concentration is strongly associated with a reduced risk of developing ESCC in experimental animals [55, 56]. There are also other potential but as yet unsubstantiated risk factors like PAHs and acetaldehyde related to ESCC in China [57], with HVP-16 and HPV-18 reported to be risk factors [55, 58]. In the US, ESCC incidence is highest in African Americans and males. Interestingly, one cohort study of ESCC and esophageal adenocarcinoma (EA) found decreased risk of ESCC, but not EA, was associated with higher intake of both fruit and vegetables [59].

3.4. Non-small cell lung carcinomas

The causal relationship between smoking and lung cancer is well established with a 10- to 20-fold increased risk of lung cancer in smokers compared with never smokers [60] and is

the major risk factor for the development of NSCLC SCC [61, 62]. In the US, smoking is estimated to account for 87% of lung cancer cases (90% in men and 85% in women) [63]. The lifetime risk of developing lung cancer is approximately 17.2% for male smokers and 11.6% for female smokers, and this risk is significantly lower in nonsmokers: 1.3% in men and 1.4% in women [64]. Healthy ex-smokers have been shown to have a similar gene expression pattern in normal bronchial epithelium as non-smokers, indicating that most smoking-induced gene expression changes revert to normal levels after smoking cessation [65]. Aside from smoking, NSCLC is also related to genetic factors [66], radon gas [67], asbestos [68], and air pollution [69]; with Radon exposure reported as the second major cause of lung cancer after smoking [67]. There is a synergistic effect between tobacco smoking and asbestos exposure in the formation of lung cancer [68], and more recently HPV [70], JC virus [71] and cytomegalovirus [72] have been reported as additional potential risk factors for NSCLC.

3.5. Overall comparison

Tobacco smoking and HPV infection appear to be carcinogenic causes for all four sub-types. In addition, several risk factors are shared among the major SCC types. HNSCC and ESCC share the most factors, consistent with their histological relationship, including alcohol consumption dietary factors, and ethnicity. Unlike the other SCCs, UV exposure is the major risk factor for NMSC. Although numerous etiologic factors for SCC are known, the exact roles and molecular mechanisms of action have not been fully elucidated.

4. Molecular Characteristics and Prognostic Markers

The development of clinically evident SCC is a multistep process involving the accumulation of multiple genetic alterations modulated by genetic predisposition, known risk factors, and other unknown environmental influences. The alterations are typically oncogene activation, including recessive oncogenes [73] and tumor suppressor gene (TSG) inactivation via mutations, loss of heterozygosity, deletions, or other mechanisms (e.g. methylation and miRNA modulation of gene expression) [74]. Molecular profiling studies that began with single or relatively small groups of genes or proteins have now progressed to large-scale and high-throughput methods using DNA-, RNA-, and protein-based approaches. These large-scale methods analyze thousands of genes at one time and have led to a better understanding of the complexity of gene abnormality patterns of SCC and have accelerated the discovery of novel genes involved in SCC pathogenesis. In addition to conventional prognostic factors [75], these molecular characteristics are becoming increasingly valuable as biomarkers in adjunct prognostic tools. There are numerous molecular markers that have been identified in SCC, and in this section we compare and contrast the major molecular abnormalities and their prognostic value among the four major SCCs.

4.1. Molecular markers common to NMSC, HNSCC, ESCC, and NSCLC

Different patterns of molecular changes and their role in prognosis have been shown between the four major SCCs; however, several key similarities are present including abnormalities in TP53, p63, Ki-67, CCND1, EGFR, and COX2 (Table 2, Table 3).

TP53—TP53 is one of the most important tumor suppressor genes in humans [76] and functions as a transcriptional regulator that controls the expression of genes involved in the cell cycle, DNA repair, apoptosis, and senescence. Under stress conditions, p53 is activated and triggers a variety of cellular responses needed to maintain the integrity of the genome, thus the protein has been designated a guardian of the genome. p53 mutations can lead to inactivation of p53 and have been found in a broad spectrum of human cancers [77], including NMSC, HNSCC, ESCC, and NSCLC. Inactivation of p53 is considered a critical

step in the development of NMSC [78]; however, TP53 mutation has not been correlated with aggressiveness of SCC, indicating the involvement of subsequent molecular events that determine tumor behavior [79]. TP53 alterations and its loss of function is a characteristic early change in HNSCC [80]. In HNSCC, it has been observed that TP53 mutations that occur within the core domain completely blocking DNA binding are linked to accelerated tumor progression, reduced therapeutic responsiveness, and decreased patient survival compared to tumors that harbor less disruptive TP53 mutations [81, 82]. In ESCC, p53 mutation has been frequently identified [83, 84] and its function positively correlated with MDM2 and p14(ARF) expression [85]. Overexpression p53 has also been significantly correlated with poorer prognosis for ESCC [86]. In NSCLCs, TP53 mutations have been detected in 40–90% of resected tumors [87–89], with disruption of the TP53 pathway frequently observed in SCC [73]. TP53 mutations can produce chemotherapy resistance [90]; however, the specific type of mutation and sensitivity to chemotherapy agents has not been identified [88, 91]. As described, TP53/p53 abnormalities are present in all four major SCCs, but to date is only considered a prognostic factor for HNSCC and ESCC.

p63—p63, a member of the p53 family, is critical for the development of stratified epithelial tissues such as epidermis [92] and is usually limited to the proliferative (basal layer) compartment of the epithelium [93]. p63 expression has been reported to be a strong predictor of poorly differentiated NMSC [94]. Whereas, expression of p63 is frequently (>95%) observed in HNSCC and associated with increased survival [95–98]. Hence, p63 may be involved in squamous cell carcinoma formation through various paths. p63 alteration is also seen in ESCC and reduced expression of p63 has prognostic implications for patients [99]. In NSCLC SCC, p63 is used to differentiate SCC from adenocarcinoma because a diffuse strong p63 and CK5/6 immunoreexpression is essentially restricted to SCC [24]. Interestingly, in NSCLC high expression of CCND1 [100] and CD24 [101] are associated with a worse clinical outcome, while expression of p63 [102] and BCL-2 [103] have a positive prognostic value. In contrast to TP53, p63 abnormalities in the four major SCCs are considered prognostic for SCC.

Ki-67 (MKI67)—Ki-67 is a cell proliferation index marker typically increased in tumors is related to rapid growth and recurrence in NMSC [104]. In HNSCC, co-expression of p21/Ki-67 is a strong negative prognostic factor [105]. In patients with stage II and III advanced ESCC, a significant correlation has been identified between Ser392 phosphorylation of p53 and high levels of Ki-67, lymphatic invasion, and poorer prognosis [106]. Similarly, one cohort study found 97% of NSCLC samples expressed Ki-67 and overexpression was associated with significantly shorter survival [107]. Similar to p63, Ki-67 abnormalities are prognostic for the four major SCCs. But in contrast to p63, Ki-67 abnormalities all appear to be indicative of poor prognosis in all four major SCCs.

CCND1—CCND1, a cell cycle regulator, acts by phosphorylating and inactivating the retinoblastoma protein [108]. In NMSC, CCND1 is involved in the early development of SCC via abnormal tissue organization and differentiation [109], with overexpression frequently seen in keratinocyte carcinogenesis [110–112]. Interestingly, CCND1 overexpression coupled with TP53 mutation has been correlated with poorer prognosis in NMSC [110–112]. In HNSCC, CCND1 polymorphism (GG) in exon 4 has been shown to be an independent prognostic indicator of disease-free interval [113]. In ESCC, CCND1 is overexpressed in 23 to 73% of tumor samples [55, 114] and is significantly correlated with poorer prognosis [115, 116]. Contrary to CCND1 in ESCC, the absence of CCND1 immunoreexpression in NSCLC is associated with worse prognosis [117]. Similarly to p63, CCND1 is associated with poorer prognosis in NMSC, ESCC, and NSCLC, but is conversely associated with improved prognosis in HNSCC.

EGFR—EGFR is present in the cell membrane as a monomer and is activated by ligand binding to the extracellular domain [118]. Mutations that lead to EGFR overexpression have been associated with a number of cancers. For example, some studies identified EGFR overexpression in metastatic SCC of the skin [119–121]. EGFR was also shown to be up-regulated/overexpressed in 90% of HNSCCs and associated with local recurrence and poor survival [122, 123]. Overexpression of EGFR [124, 125], is significantly correlated with poorer prognosis for ESCC. SCCs demonstrate most of the genetic abnormalities commonly present in NSCLCs, except for *KRAS* and *EGFR* gene mutations, which are more frequent in adenocarcinomas [73, 126]. Overexpression of EGFR, p53 [127–129] and Her2 [130, 131] has shown conflicting results in NSCLC and their use as prognostic markers require further study [132–136]. However, two prognostic proteins in NSCLC have been identified and are used together. NSCLCs having both IGFR-1- and EGFR-positive immunoreactivity represent a subpopulation capable of developing aggressive clinical behavior [137]. Again, similarly to p63, EGFR is associated with poorer prognosis in NSCM, HNSCC and ESCC. But due to conflicting reports, the use of p63 as prognostic marker in NSCLC needs to be investigated further.

COX2—COX2 (PTGS2) is an enzyme that functions in protein metabolism by increasing prostaglandin synthesis and plays a role in tumorigenesis. The expression of COX-2 is upregulated in many cancers and its product, PGH₂, is converted by prostaglandin E2 synthetase into PGE₂ which in turn can stimulate cancer progression [138]. For example, in NMSC, high expression of COX-2 has been found in AK, SCC and BCC [139] and COX-2 expression increases during progression from AK to SCC [140]. Overexpression of COX2 has also been shown in HNSCC, and some *ex vivo* studies have demonstrated that COX-2 is overexpressed in ESCC and premalignant lesions [141, 142]. Statistically significant COX-2 overexpression has also been found in 28.9 % of NSCLC SCC [143]. Overexpression of COX-2 and upregulation of the prostaglandin pathway plays a significant role in SCC and blockade of the process has strong potential for cancer prevention and therapy. However, to date, COX2 has not been identified as a prognostic marker of SCCs.

4.2. Other key molecular markers shared in several SCCs

In addition to the shared cell cycle regulation (TP53, p63, Ki-67, and CCND1), signal transduction (EGFR), and protein metabolism (COX2) molecular abnormalities, several other molecular abnormalities of gene expression, protein expression, gene mutation, and epigenetic regulation and their respective prognostic values have been characterized in SCCs (Table 2, Table 3). Key markers shared in two or three of the major SCCs involve several classes of genes including signal transduction (VEGF), transcription factor (SOX2), cell adhesion (CDH1), and extracellular matrix degradation (MMPs). Additional markers affecting only one of the SCCs can be found in Table 2 and Table 3.

VEGF—VEGF is an important signal transduction protein involved in both vasculogenesis and angiogenesis. Four subtypes have been described (A, B, C, and D). VEGF alteration is involved in HNSCC and ESCC. In oral SCC, it was reported that overexpression as measured by immunohistochemistry of subtypes A and B were correlated with tumor angiogenesis and subtypes C and D with metastases and poor prognosis [144]. In ESCC, VEGF (VEGF A) expression has historically ranged between 24–93% [145], and recently elevated immunoreactivity has been reported in 55% of ESCC tissues (n= 108) [146]. Overexpression of VEGF has been significantly correlated with poorer prognosis of ESCC [145].

SOX2—A recently discovered lineage-survival oncogene, SOX2, has been shown to be important in HNSCC, ESCC, and NSCLC. Lineage survival oncogenes are activated by

somatic mutations and thus may play an important role in carcinogenesis. The genomic amplification of the SOX 2 embryonic stem cell transcription factor located on chromosome 3q26.33 was first reported in ESCC and NSCLC [147]. SOX2 has been associated with poor prognosis in ESCC [148], but its use as a prognostic marker in NSCLC has not yet been elucidated. In HNSCC (oral), high protein expression of SOX2 has been found to correspond to copy number gain in 52% of oral SCC tumors [149]. miR-145 is involved in regulating SOX2 [150]; however, its role as a prognostic factor in SCCs is still unknown.

Our group has recently characterized SOX-2 protein expression related to the pathogenesis of NSCLC SCC [151]. By assessing SOX2 mRNA expression in various published datasets against the previously characterized OCT4/SOX2/NANOG signature, we were able to effectively separate SCCs from adenocarcinomas. In this study, we further characterized SOX immunoeexpression of NSCLC tissues and identified SOX2 protein expression pattern in SCC development (hyperplasia, dysplasia, and carcinoma *in situ*) [151].

CDH1 (E-cadherin)—CDH1 belongs to the cadherin family of Ca²⁺-dependent cell-cell adhesion molecules which induce and maintain intercellular connections. CDH1 has been implicated in carcinogenesis due to reduced expression [152] and promoter hypermethylation [153]. In NMSC, down-regulation of CDH1 is linked to increased potential for tumor invasiveness and distant metastasis and the frequencies of CDH1 promoter hypermethylation appear to be correlated with a more advanced stage of squamous carcinogenesis in skin [153]. In ESCC, tumors with reduced CDH1 expression invade deeper, have more lymph node metastasis, and have more lymphatic invasion than tumors with preserved CDH1 expression [154]. Disorganized CDH1 expression was also reported to be a feature of advanced ESCC [152].

MMPs—Matrix metalloproteinases (MMPs) are a family of zinc-containing endopeptidases that degrade various components of the extracellular matrix, and have been strongly implicated in multiple stages of cancer progression including the acquisition of invasive and metastatic properties. MMPs are altered in NMSC, HNSCC and ESCC. In NMSC, MMP-2 and MMP-9 immunoeexpression is associated with NMSC pathogenesis and is an indicator of cutaneous cancer invasion and progression [155]. Expression of MMPs has been identified in both the epithelial and stromal elements of HNSCC [156], with MMP2 and MMP9 showed an association with invasive potential [157, 158] and poor outcome [159]. Gu *et al.* evaluated the expression of MMP1, MMP7, MMP9, and MMP13 in ESCC tissues from 208 patients and found that MMP7, MMP9, and MMP13 may be involved with early stage ESCC, and their coexpression predicted poor outcome for ESCC patients [160]. These data are consistent with the previous finding that expression of MMP-7 and MMP-9 may be a good marker for the presence of lymphatic metastasis in ESCC [161]. Moreover, our group identified MMP3 and MMP10 as potential early diagnosis biomarkers for ESCC. Both enzymes showed a tumor increase at both the transcriptional and proteomic levels [162].

Gene mutations—Gene mutations other than those in TP53, EGFR, and CCND1 have also been shown to be important in SCCs, with a large proportion of mutations having been identified in NSCLC and to a less extent HNSCC. Functional inactivation of p16 through deletion frequently occurs in HNSCC [163]. Chromosome 3p contains numerous TSGs (e.g. ALS2CL, EPHA3 and CMYA1) and the loss of heterozygosity in this region may contribute to SCCs, including HNSCC [164] and NSCLC [165]. Activating RAS mutations occur in approximately 15% to 20% of NSCLC, with a majority of them being KRAS [166], however these mutations are rare in SCC [167]. Although, c-MET, TTF-1, LKB1, BRAF and PIK3CA are often mutated or amplified in NSCLC [167].

Methylation—The epigenetic mechanism of methylation has been shown to modulate of gene expression in SCCs. Since its discovery as a cyclin-dependent kinase inhibitor in 1993, the tumor suppressor p16 (INK4A/MTS-1/CDKN2A) has gained widespread importance in cancer [168]. Like NMSC, gene methylation has also been identified in HNSCC, ESCC, and NSCLC to varying degrees. p16 inactivation often occurs via methylation in both HNSCC [163] and ESCC [169]. This silencing of p16 in ESCC has been shown to lead to deregulation of cell proliferation and consequent genomic instability [169]. Different patterns of gene methylation have been found in the major histological types of NSCLCs, with LKB1 and RASSF1 being the most important in NSCLC SCC. Inactivation of the TSG LKB1 by mutation and deletion is a relatively frequent event in SCC (19%) of the lung [170]. But, RAS association domain family 1 gene (RASSF1) is the most frequently hypermethylated gene in NSCLC. RASSF1A methylation has been correlated with worse prognosis in surgically resected NSCLC patients and was confirmed as an independent prognostic factor by multivariate analysis [171].

miRNAs—miRNAs are a class of small (~18–24 mer) nucleic acids that negatively regulate gene expression. Through their targets, miRNAs are known to play important roles in cell differentiation, proliferation, and apoptosis, and altered miRNA levels result in the aberrant expression of gene products that may contribute to cancer biology [172, 173]. An emerging number of studies have shown that miRNAs can act as oncogenes, as tumor suppressor genes, or sometimes as both [174]. High-throughput analyses have demonstrated that miRNA expression is commonly dysregulated in human cancer [173]. However, considerable disagreement remains with respect to the miRNA signature for specific cancer cell types, which appears to depend largely on the analytical platform [173].

Dysregulation of miRNAs have been identified in HNSCC, ESCC, and NSCLC and not NMSC. However, Drosha, an important enzyme in the miRNA machinery, has been found to be overexpressed in NMSC; thus giving strength to the hypothesis of miRNA involvement in NMSC carcinogenesis [175]. In HNSCC, overexpression of miR-211 has been associated with invasive potential [176]. In addition to miR-211, miRNA profiling has revealed four upregulated miRNAs (miR-21, -31, -18, and -221) and 13 down-regulated miRNAs (miRNA-133a, -133b, -125a, -138, -139, -200c, -26b, -302b, -302c, -342, -371, -373, -375) associated with HNSCC [177]. Moreover, the ratio of miR-221:miR-375 showed a high discriminatory potential, with a sensitivity of 92% and specificity of 93% in distinguishing tumor from normal tissue, suggesting that this simple molecular marker may hold significant clinical potential as a diagnostic tool [178]. miR-21 has also been reported in ESCC and found to induce cell proliferation and invasion [179]. Our group evaluated ESCC related miRNAs by comparing microdissected cells involved in normal differentiation and tumorigenesis and confirmed that miR-21 was overexpressed in tumors (Zhu, *et al*, submitted). miRNAs have been shown to regulate several important pathways in NSCLC and have been correlated with disease outcome in NSCLC [180–182]. Of interest, a five miRNA signature (let-7a, miR-221, miR-137, miR-372, and miR-182) has been identified in NSCLC that predicts treatment outcome [182]. In that study, patients with high risk scores in their miRNA signatures showed poor overall and disease-free survivals compared with patients with low risk scores [182]. Loss of expression of miRNA-128b, putative regulator of EGFR, correlated with response to targeted EGFR inhibition in primary NSCLC [183]. miRNA is an area of very active research that will have an impact on pathogenesis and therapy as more is learned about the role of miRNAs in SCC.

Numerous molecular abnormalities in gene expression, protein expression, gene mutation, and epigenetic regulation have been characterized in SCC (Table 3), with several of these markers associated with disease prognosis (Table 2). Commonalities in molecular changes present in the four major SCCs are predominantly found in cell cycle regulation and signal

transduction. Although not all of the described molecular abnormalities are shared in all four of the SCCs, many are shared in at least two or three of the SCCs. The comparison of molecular characteristic similarities and differences in SCC provide insight not only into the relationships between NMSC, HNSCC, ESCC, and NSCLC, but to SCC as a whole. And this insight into SCC can putatively be translated to improved disease control and treatment. Currently, drugs targeting several of these gene and pathway abnormalities are being used in the treatment of SCC.

5. Targeted Therapy

Targeted therapy is a type of treatment that uses drugs that identify and attack specific cancer cells without harming normal cells and has been extensively investigated in recent decades, both as a single modality therapy and in combination with cytotoxic treatments such as radiotherapy or chemotherapy. With the growing understanding of molecular genetics of SCC, targeted therapies now offer expanded treatment options for patients that have clinically significant benefits. The targets that are currently considered the most relevant in SCCs fall into one of the following categories: cell-cycle regulation, signal transduction, growth factor receptors, angiogenesis, and protein degradation.

Similar to the fact that there are few prognostic markers for NMSC, there are currently no targeted drugs developed for NMSC. Although not developed as a targeted therapy, Diclofenac, a dual inhibitor of COX-1 and COX-2 with higher selectivity for COX-2, has been investigated and reported to be effective for patients with AK, a precancerous syndrome of skin [184].

Several targeted therapies are being investigated for HNSC, ESCC, and NSCLC with many of the molecular targets being shared among the three SCCs (Table 4). Targeted therapies have been extensively investigated in HNSC and NSCLC, especially EGFR TKIs and their mechanisms of resistance.

5.1. Growth Factor Receptor Antagonists

EGFR—EGFR is a member of the ERBB family of transmembrane tyrosine kinase receptors [132]. EGFR and their receptors are involved in signal transduction and tumor growth, thus blockade of these systems provides a therapeutic approach, through neutralizing ligands, inhibiting ligand binding, or blocking the tyrosine kinases of the receptors. Examples of EGFR inhibitors include monoclonal antibodies against the extracellular domain of the receptor (e.g., cetuximab and panitumumab) and receptor tyrosine kinase inhibitors (TKIs) that target the intracellular domain (e.g., gefitinib and erlotinib). EGFR inhibitors have been applied in HNSCC, ESCC and extensively in NSCLC clinical trials. In HNSCC, patients with locally advanced disease have been shown to benefit from the addition of EGFR inhibition (for example, cetuximab) to radiotherapy [185]. But, EGFR targeted therapy trials conducted in HNSCC to date still show various disadvantages such as low efficacy and significant toxicity [186]. In ESCC, inhibition of EGFR-TK by erlotinib is promising through inducing growth inhibition and cell cycle arrest in human esophageal cancer cells and enhancing the antineoplastic effects of other targeted agents [187]. Cetuximab and panitumumab are currently being investigated in NSCLC. Phase II trials showed that cetuximab improved survival in-chemo-naïve patients with advanced cancer [188, 189] and panitumumab is currently in phase II trial [190].

EGFR TKIs gefitinib and erlotinib were the first two targeted agents recently approved for the treatment of NSCLC in the United States and several markers have been identified that predict response in NSCLC patients. These EGFR TKIs produce responses in approximately 10% of NSCLC patients having progressed with prior chemotherapy [191–193]. But in those

patients who benefit from gefitinib or erlotinib the responses can be dramatic and may last for longer than a year, with favorable response being associated with activating mutations in the EGFR tyrosine kinase domain (exons 18 to 21), increased gene copy number, and increased protein expression [191–194]. Even though targeting EGFR mutated NSCLCs with gefitinib or erlotinib has been effective, most of these patients acquire resistance to the EGFR TKI therapy [195, 196] in an average of 6–12 months [197]. Interestingly, smokers presenting with NSCLCs are generally resistant to EGFR-TKIs [198, 199], which may have implications in other targeted therapies of this class because SCC is the predominant histologic subtype associated with smokers. In an effort to counteract EGFR TKI resistance mechanisms, an initial study has shown TKI resistant NSCLC cell lines can be treated by administering PI3K-mTOR and MEK signaling inhibitors simultaneously with EGFR TKIs [200]; but, to date this has not been tested in NSCLC patient populations.

Other growth factor receptor antagonists—IGF-I acts through the IGF receptor 1 (IGF- R1) to promote cell survival and cell proliferation [201]. It plays an important role in normal growth and has anabolic effects in adults normally. But IGF-I has been implicated in cancers [202] as its anti-apoptotic properties allow cancerous cells to resist the cytotoxic effects of chemotherapeutic drugs or radiotherapy. A recent clinical trial reported that combining the fully humanized anti-IGF-IR monoclonal antibody A12 with radiation to treat HNSCC resulted in more pronounced antitumor activity than either agent alone [203]. In NSCLC, early clinical trials showed an acceptable safety profile together with pharmacodynamic evidence that IGF-RI can be successfully targeted [204]. And, biomarkers of the IGF-IR pathway were shown to be key elements in development and monitoring of anti-IGF-RI therapy in NSCLC [205]. Recently, phase II study data has suggested that co-administration of an anti-IGF-IR antibody with chemotherapy improves the response rate and progression-free survival [204].

HER3 (ERBB3), a gene encoding a member of the epidermal growth factor receptor (EGFR) family of receptor tyrosine kinases has emerged for HNSCC targeted therapy within the past four years [206, 207]. Several laboratories are designing antibodies that will block HER3 heterodimerization with EGFR or HER2 to prevent signaling to PI3K/Akt [208].

5.2. Cell-Cycle Regulation

Although cell-cycle regulation contains the most molecular abnormalities in SCC (Table 3) and subsequently numerous potential therapeutic biomarkers, currently only one marker, CCND1, has been exploited. Mutations, amplification and overexpression of CCND1 are frequently observed in a variety of tumors, including SCCs, and may contribute to tumorigenesis. Flavopiridol, the first cyclin-dependent kinase inhibitor in human clinical trials, was reported as a targeting drug for HNSCC [209]. Flavopiridol has also been shown to decrease CCND1 expression in ESCC cell lines and was subsequently found to induce radiosensitivity [210] and may have application to the other SCCs.

5.3. Signal Transduction

Signal transduction was the second most common group of molecular abnormalities in the four major SCCs (Table 3). Of which, both of mTOR and MET have been identified in HNSCC and NSCLC as molecular targets. mTOR is a serine/threonine protein kinase that regulates cell growth, cell proliferation, cell motility, cell survival, protein synthesis, and transcription. It is activated by Akt and blocks apoptosis to increase the proliferative potential of cancer cells. Several mTOR inhibitors are currently under investigation in HNSCC [211]. In an ongoing phase II trial, combined inhibition of mTOR with everolimus and gefitinib was evaluated in patients with stage IIIB/IV NSCLC [212]. Bortezomib is a small-molecule proteasome inhibitor that has shown encouraging results in a phase II trial,

and a phase III trial of gemcitabine/carboplatin ± bortezomib in advanced stage NSCLC is in progress [213].

C-MET is the cell surface receptor for hepatocyte growth factor (HGF), also known as scatter factor [214]. Binding of the receptor to its ligand, hepatocyte growth factor, induces receptor dimerization that triggers conformational changes that activate MET tyrosine kinase activity which then have profound effects on cell growth, survival, motility, invasion and angiogenesis [215]. Dysregulation of MET signalling has been shown to contribute to tumorigenesis in a number of malignancies and hence can serve as a potential drug target. Zucali, et al. found that activated cMET appeared to be a marker of primary gefitinib resistance in NSCLC patients and suggested cMET may be a target for treatment [216]. In NSCLC, several phase II/III trial with PF-02341066 either as monotherapy or in combination with EGF-R inhibitors are currently underway [217].

In addition, preliminary data from a phase II trial testing of sorafenib, a potent inhibitor of the Raf-1, B-Raf, VEGFR-2-3, and PDGFR-B pathways, in metastatic or recurrent HNSCC were recently reported [218]. Sorafenib treatment for NSCLC is being evaluated in several phase III studies [217].

5.4. Protein Degradation

Cox-2 has been implicated in apoptosis resistance, angiogenesis, decreased host immunity and enhanced invasion and metastasis, and, thus is involved in critical aspects of carcinogenesis [219]. COX-2 selective inhibitors, a form of non-steroidal anti-inflammatory drug (NSAID) that directly targets COX-2, has been shown to reduce the occurrence of cancers and pre-cancerous growths [220] and is in clinical trials for both ESCC and NSCLC [219]. Histone deacetylase inhibitors (HDIs) have been shown to block the activation of COX2 transcription [221]. In recent years, there has been an effort to develop HDIs for cancer therapy and they have demonstrated activity in patients with advanced solid tumors in phase I trials as well as in patients with relapsed NSCLC [222]. Since COX2 dysregulation was also shown in all of the four major SCCs, HDIs should be investigated for SCC as a whole.

5.5. Angiogenesis

Although VEGF alteration has not yet been shown to be directly involved in NMSC and NSCLC, VEGF expression in generally is commonly seen in tumors due to its involvement in vasculogenesis and angiogenesis. Furthermore, VEGF may cause a cell to survive, move, or further differentiate through various molecular mechanisms, thus VEGF is a potential target for the treatment of cancer, including SCCs. Anti-VEGF therapies have capitalized upon this potential and have proven to be important in the treatment of certain cancers using monoclonal antibodies (bevacizumab) and orally-available small molecule VEGF TKIs (sorafenib). In SCC, sorafenib has been used in clinical trials of HNSCC and NSCLC [223, 224]. Bevacizumab, the first commercially available angiogenesis inhibitor, has been tested in HNSCC and NSCLC [225, 226]. Interestingly, improved outcome has been shown using bevacizumab in combination with the combination chemotherapy of paclitaxel carboplatin in patients with advanced NSCLC, but is contraindicated for SCC due to safety risks [226]. This contraindication for SCC is due to grade 5 hemoptysis in SCC patients and identification via multivariate analysis of SCC as an individual significant risk factor [226]. As such, bevacizumab in combination with paclitaxel carboplatin is now only used with non-squamous NSCLCs.

Advances in understanding the molecular pathogenesis of SCC has provided a unique opportunity to attack SCC by targeted therapy. Examination of molecular abnormalities in

tumors has become increasingly important. Similarly, the development of molecular “signatures” (e.g. mRNA expression profiles) from tumors that provide information on the prognosis and predict the response of individual patient’s tumors to specified therapy would be a major step forward. For a targeted therapy to truly be effective, we must also have biomarkers to precisely predict or monitor tumor response or resistance to cytotoxic and targeted agents [227]. Unfortunately, to date, no good clinical or biological markers to predict outcomes of targeted therapy have been identified.

6. New Approach to Studying Molecular Abnormalities in SCC

In an effort to advance the molecular profiling and subsequent understanding of SCC as a whole, new methodologies are being developed using specific anatomic site SCCs with the goal of applying these novel approaches to the characterization of molecular abnormalities in other SCCs. Our group utilized a microdissected normal basal-tumor gene expression comparison to identify pathways and genes that could be putative therapeutic targets for ESCC (Yan *et al.* submitted). In other words, we contrasted the expression profile of a normal dividing cell population against its counterpart transformed cell population in a search for growth-related genes that are unique to cancer and not part of the standard cell growth machinery *per se* [228]. The data showed that gene expression in normal differentiated cells was markedly different from normal basal cells and tumor; whereas, tumor and normal basal cells were more closely related. Tumor cells showed a general decrease in differentially expressed genes relative to normal basal cells as opposed to differentiated cells that exhibited the opposite trend. The results identified two highly dysregulated networks in normal differentiation and tumorigenesis; DNA repair pathways were involved in normal and pathological growth; and some individual cell differentiation related pathway and genes were uniquely expressed in basal cells compared to differentiated cells. Furthermore, using our ‘biologic filter’, 12 genes were identified as being unique to the normal basal-tumor comparison and could potentially be therapeutic targets for treating ESCC.

In a separate study, we have focused on characterizing targeted-therapy related molecular biomarkers from NSCLC ever-smokers versus never-smokers, using microdissected paired tumor/normal cells and a novel qRT-PCR with pre-amplification method developed by our group (Yan *et al.* submitted). The data provided potentially useful information in guiding an individual treatment approach for lung cancer.

Although these strategies have been developed in ESCC, these novel methodologies can be applied to other SCCs to identify potential therapeutic targets directly related to tumorigenesis. We hope that these new approaches to studying SCC will also elucidate markers for prognosis and lead to effective therapies for SCCs of all anatomical sites.

7. Conclusion

Despite improvements in diagnosis and therapy, mortality and morbidity rates for some forms of SCC remain high. Early diagnosis is of course important in preventing this cancer and reducing mortality, and in parallel to improving screening and diagnostic efforts, there is a significant need to develop novel therapeutic agents for patients with advanced disease. SCCs demonstrate a wide range of epithelial tumors that vary in their anatomic sites. These tumors show varying degrees of relationship to risk factors, with HPV showing the greatest relationship.

Modern molecular genetic analysis allows us to probe beneath the phenotypic surface to the underlying etiologic molecular abnormalities of SCC and a number of molecules that contribute to the complex events of carcinogenesis and cancer progression in these cancers

have been identified. The molecular lesions found in SCC tumors share common elements and characteristic changes, with molecular abnormalities of cell-cycle regulation and signal transduction predominating SCCs as a whole.

Encouraged by the development of methodologies for isolation of cells from small histologic lesions, such as laser microdissection, combined with techniques to perform genomic studies from minute amount of DNA, RNA and protein, several groups, including ours, have made substantial progress on unveiling the molecular and genetic abnormalities of SCCs. The development and application of new molecular genetic methods for analysis of SCC tissue specimens will help delineate the significant molecular abnormalities responsible for SCC development and progression. Additional studies are needed to further improve our understanding of the similarities and differences among the various SCCs, toward improvements in diagnosis, prognosis and therapy.

Acknowledgments

This work was supported by the Cohen-Reinauch BATTLE-2 Fund and the Intramural Program of the Center for Cancer Research, National Cancer Institute, NIH.

References

1. Marinkovich MP. Tumour microenvironment: laminin 332 in squamous-cell carcinoma. *Nat Rev Cancer*. 2007; 7:370–380. [PubMed: 17457303]
2. Agada FO, Patmore H, Alhamarneh O, Stafford ND, Greenman J. Genetic profile of head and neck squamous cell carcinoma: clinical implications. *J Laryngol Otol*. 2009; 123:266–272. [PubMed: 18694533]
3. Farhadieh RD, Salardini A, Yang JL, Russell P, Smee R. Diagnosis of second head and neck tumors in primary laryngeal SCC is an indicator of overall survival and not associated with poorer overall survival: a single centre study in 987 patients. *J Surg Oncol*. 101:72–77. [PubMed: 19798688]
4. Travis WD. Pathology of lung cancer. *Clin Chest Med*. 2002; 23:65–81. viii. [PubMed: 11901921]
5. Pisani P, Bray F, Parkin DM. Estimates of the world-wide prevalence of cancer for 25 sites in the adult population. *Int J Cancer*. 2002; 97:72–81. [PubMed: 11774246]
6. Christenson LJ, Borrowman TA, Vachon CM, Tollefson MM, Otley CC, Weaver AL, Roenigk RK. Incidence of basal cell and squamous cell carcinomas in a population younger than 40 years. *JAMA*. 2005; 294:681–690. [PubMed: 16091570]
7. Clayman GL, Lee JJ, Holsinger FC, Zhou X, Duvic M, El-Naggar AK, Prieto VG, Altamirano E, Tucker SL, Strom SS, Kripke ML, Lippman SM. Mortality risk from squamous cell skin cancer. *J Clin Oncol*. 2005; 23:759–765. [PubMed: 15681519]
8. Rigel DS, Friedman RJ, Kopf AW. Lifetime risk for development of skin cancer in the U.S. population: current estimate is now 1 in 5. *J Am Acad Dermatol*. 1996; 35:1012–1013. [PubMed: 8959974]
9. Kwa RE, Campana K, Moy RL. Biology of cutaneous squamous cell carcinoma. *J Am Acad Dermatol*. 1992; 26:1–26. [PubMed: 1732313]
10. Parkin DM. Global cancer statistics in the year 2000. *Lancet Oncol*. 2001; 2:533–543. [PubMed: 11905707]
11. Webb JL, Burns RE, Brown HM, Leroy BE, Kosarek CE. Squamous cell carcinoma. *Compend Contin Educ Vet*. 2009; 31:133–142. [PubMed: 19412903]
12. Sturgis EM, Cinciripini PM. Trends in head and neck cancer incidence in relation to smoking prevalence: an emerging epidemic of human papillomavirus-associated cancers? *Cancer*. 2007; 110:1429–1435. [PubMed: 17724670]
13. Silveira NJ, Varuzza L, Machado-Lima A, Lauretto MS, Pinheiro DG, Rodrigues RV, Severino P, Nobrega FG, Silva WA Jr, de BPCA, Tajara EH. Searching for molecular markers in head and neck squamous cell carcinomas (HNSCC) by statistical and bioinformatic analysis of larynx-derived SAGE libraries. *BMC Med Genomics*. 2008; 1:56. [PubMed: 19014460]

14. Chin D, Boyle GM, Williams RM, Ferguson K, Pandeya N, Pedley J, Campbell CM, Theile DR, Parsons PG, Coman WB. Novel markers for poor prognosis in head and neck cancer. *Int J Cancer*. 2005; 113:789–797. [PubMed: 15499618]
15. Ragin CC, Modugno F, Gollin SM. The epidemiology and risk factors of head and neck cancer: a focus on human papillomavirus. *J Dent Res*. 2007; 86:104–114. [PubMed: 17251508]
16. Kamangar F, Dores GM, Anderson WF. Patterns of cancer incidence, mortality, and prevalence across five continents: defining priorities to reduce cancer disparities in different geographic regions of the world. *J Clin Oncol*. 2006; 24:2137–2150. [PubMed: 16682732]
17. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin*. 2005; 55:74–108. [PubMed: 15761078]
18. Szumilo J. Epidemiology and risk factors of the esophageal squamous cell carcinoma. *Pol Merkuri Lekarski*. 2009; 26:82–85. [PubMed: 19391515]
19. Carneiro A, Isinger A, Karlsson A, Johansson J, Jonsson G, Bendahl PO, Falkenback D, Halvarsson B, Nilbert M. Prognostic impact of array-based genomic profiles in esophageal squamous cell cancer. *BMC Cancer*. 2008; 8:98. [PubMed: 18405350]
20. Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ. Cancer statistics, 2007. *CA Cancer J Clin*. 2007; 57:43–66. [PubMed: 17237035]
21. Blot WJ, McLaughlin JK. The changing epidemiology of esophageal cancer. *Semin Oncol*. 1999; 26:2–8. [PubMed: 10566604]
22. WHO WHO/TWHR. , editor. Food, Nutrition and the Prevention of cancer. A Global Perspective. 1997.
23. Edwards BK, Brown ML, Wingo PA, Howe HL, Ward E, Ries LA, Schrag D, Jamison PM, Jemal A, Wu XC, Frieman C, Harlan L, Warren J, Anderson RN, Pickle LW. Annual report to the nation on the status of cancer, 1975-2002, featuring population-based trends in cancer treatment. *J Natl Cancer Inst*. 2005; 97:1407–1427. [PubMed: 16204691]
24. Erickson, Heidi S.; Wistuba, Ignacio I. Pathology of Lung Cancer. In: Kernstine, Kemp, MD, Ph.D; Reckamp, Karen, M.D., M.S; Thomas, Charles, J.R., M.D, editors. Lung Cancer: A Multidisciplinary Approach to Diagnosis and Management. New York, NY: DemosMedical, Inc.; 2010.
25. Ramalingam S, Belani C. Systemic chemotherapy for advanced non-small cell lung cancer: recent advances and future directions. *Oncologist*. 2008; 13 Suppl 1:5–13. [PubMed: 18263769]
26. Hammar, SP.; Brambilla, C.; Pugatch, B., et al. Tumours of the lung. Squamous cell carcinoma. In: Travis, WDBE.; Muller-Hermelink, HK., et al., editors. Pathology and genetics: Tumours of the lung, pleura, thymus and heart. Lyon: IARC Press; 2004. p. 26-34.
27. Rudd RM, Gower NH, Spiro SG, Eisen TG, Harper PG, Littler JA, Hatton M, Johnson PW, Martin WM, Rankin EM, James LE, Gregory WM, Qian W, Lee SM. Gemcitabine plus carboplatin versus mitomycin, ifosfamide, and cisplatin in patients with stage IIIB or IV non-small-cell lung cancer: a phase III randomized study of the London Lung Cancer Group. *J Clin Oncol*. 2005; 23:142–153. [PubMed: 15625369]
28. Martoni A, Marino A, Sperandi F, Giaquinta S, Di Fabio F, Melotti B, Guaraldi M, Palomba G, Preti P, Petralia A, Artioli F, Picece V, Farris A, Mantovani L. Multicentre randomised phase III study comparing the same dose and schedule of cisplatin plus the same schedule of vinorelbine or gemcitabine in advanced non-small cell lung cancer. *Eur J Cancer*. 2005; 41:81–92. [PubMed: 15617993]
29. Travis WD, Travis LB, Devesa SS. Lung cancer. *Cancer*. 1995; 75:191–202. [PubMed: 8000996]
30. Jang KM, Lee KS, Shim YM, Han D, Kim H, Kwon OJ, Kim J, Kim TS. The rates and CT patterns of locoregional recurrence after resection surgery of lung cancer: correlation with histopathology and tumor staging. *J Thorac Imaging*. 2003; 18:225–230. [PubMed: 14561907]
31. Armstrong BK, Krickler A. The epidemiology of UV induced skin cancer. *J Photochem Photobiol B*. 2001; 63:8–18. [PubMed: 11684447]
32. Lee DA, Miller SJ. Nonmelanoma skin cancer. *Facial Plast Surg Clin North Am*. 2009; 17:309–324. [PubMed: 19698913]
33. Akgul B, Cooke JC, Storey A. HPV-associated skin disease. *J Pathol*. 2006; 208:165–175. [PubMed: 16362995]

34. Moloney FJ, Comber H, O'Lorcain P, O'Kelly P, Conlon PJ, Murphy GM. A population-based study of skin cancer incidence and prevalence in renal transplant recipients. *Br J Dermatol.* 2006; 154:498–504. [PubMed: 16445782]
35. Morita A. Tobacco smoke causes premature skin aging. *J Dermatol Sci.* 2007; 48:169–175. [PubMed: 17951030]
36. Lo WL, Kao SY, Chi LY, Wong YK, Chang RC. Outcomes of oral squamous cell carcinoma in Taiwan after surgical therapy: factors affecting survival. *J Oral Maxillofac Surg.* 2003; 61:751–758. [PubMed: 12856245]
37. Talamini R, Bosetti C, La Vecchia C, Dal Maso L, Levi F, Bidoli E, Negri E, Pasche C, Vaccarella S, Barzan L, Franceschi S. Combined effect of tobacco and alcohol on laryngeal cancer risk: a case-control study. *Cancer Causes Control.* 2002; 13:957–964. [PubMed: 12588092]
38. Andre K, Schraub S, Mercier M, Bontemps P. Role of alcohol and tobacco in the aetiology of head and neck cancer: a case-control study in the Doubs region of France. *Eur J Cancer B Oral Oncol.* 1995; 31B:301–309. [PubMed: 8704646]
39. Schlecht NF, Franco EL, Pintos J, Kowalski LP. Effect of smoking cessation and tobacco type on the risk of cancers of the upper aero-digestive tract in Brazil. *Epidemiology.* 1999; 10:412–418. [PubMed: 10401876]
40. Zhang ZF, Morgenstern H, Spitz MR, Tashkin DP, Yu GP, Hsu TC, Schantz SP. Environmental tobacco smoking, mutagen sensitivity, and head and neck squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev.* 2000; 9:1043–1049. [PubMed: 11045786]
41. Sturgis EM, Wei Q, Spitz MR. Descriptive epidemiology and risk factors for head and neck cancer. *Semin Oncol.* 2004; 31:726–733. [PubMed: 15599850]
42. Wynder EL, Mushinski MH, Spivak JC. Tobacco and alcohol consumption in relation to the development of multiple primary cancers. *Cancer.* 1977; 40:1872–1878. [PubMed: 332333]
43. Pavia M, Pileggi C, Nobile CG, Angelillo IF. Association between fruit and vegetable consumption and oral cancer: a meta-analysis of observational studies. *Am J Clin Nutr.* 2006; 83:1126–1134. [PubMed: 16685056]
44. Perez-Ordóñez B, Beauchemin M, Jordan RC. Molecular biology of squamous cell carcinoma of the head and neck. *J Clin Pathol.* 2006; 59:445–453. [PubMed: 16644882]
45. Chung CH, Gillison ML. Human papillomavirus in head and neck cancer: its role in pathogenesis and clinical implications. *Clin Cancer Res.* 2009; 15:6758–6762. [PubMed: 19861444]
46. Duvvuri U, Myers JN. Cancer of the head and neck is the sixth most common cancer worldwide. *Curr Probl Surg.* 2009; 46:114–117. [PubMed: 19111678]
47. Martin IG, Young S, Sue-Ling H, Johnston D. Delays in the diagnosis of oesophagogastric cancer: a consecutive case series. *BMJ.* 1997; 314:467–470. [PubMed: 9056794]
48. Carstensen JM, Pershagen G, Eklund G. Mortality in relation to cigarette and pipe smoking: 16 years' observation of 25,000 Swedish men. *J Epidemiol Community Health.* 1987; 41:166–172. [PubMed: 3655638]
49. Ishikawa A, Kuriyama S, Tsubono Y, Fukao A, Takahashi H, Tachiya H, Tsuji I. Smoking, alcohol drinking, green tea consumption and the risk of esophageal cancer in Japanese men. *J Epidemiol.* 2006; 16:185–192. [PubMed: 16951537]
50. Freedman ND, Abnet CC, Leitzmann MF, Mouw T, Subar AF, Hollenbeck AR, Schatzkin A. A prospective study of tobacco, alcohol, and the risk of esophageal and gastric cancer subtypes. *Am J Epidemiol.* 2007; 165:1424–1433. [PubMed: 17420181]
51. Boffetta P, Garfinkel L. Alcohol drinking and mortality among men enrolled in an American Cancer Society prospective study. *Epidemiology.* 1990; 1:342–348. [PubMed: 2078609]
52. Brown LM, Hoover R, Silverman D, Baris D, Hayes R, Swanson GM, Schoenberg J, Greenberg R, Liff J, Schwartz A, Dosemeci M, Pottern L, Fraumeni JF. Excess incidence of squamous cell esophageal cancer among US Black men: role of social class and other risk factors. *Am J Epidemiol.* 2001; 153:114–122. [PubMed: 11159155]
53. Brown LM, Hoover RN, Greenberg RS, Schoenberg JB, Schwartz AG, Swanson GM, Liff JM, Silverman DT, Hayes RB, Pottern LM. Are racial differences in squamous cell esophageal cancer explained by alcohol and tobacco use? *J Natl Cancer Inst.* 1994; 86:1340–1345. [PubMed: 8064893]

54. Layke JC, Lopez PP. Esophageal cancer: a review and update. *Am Fam Physician*. 2006; 73:2187–2194. [PubMed: 16836035]
55. Stoner GD, Gupta A. Etiology and chemoprevention of esophageal squamous cell carcinoma. *Carcinogenesis*. 2001; 22:1737–1746. [PubMed: 11698334]
56. Abnet CC, Lai B, Qiao YL, Vogt S, Luo XM, Taylor PR, Dong ZW, Mark SD, Dawsey SM. Zinc concentration in esophageal biopsy specimens measured by x-ray fluorescence and esophageal cancer risk. *J Natl Cancer Inst*. 2005; 97:301–306. [PubMed: 15713965]
57. Kamangar F, Chow WH, Abnet CC, Dawsey SM. Environmental causes of esophageal cancer. *Gastroenterol Clin North Am*. 2009; 38:27–57. vii. [PubMed: 19327566]
58. Castillo A, Aguayo F, Koriyama C, Torres M, Carrascal E, Corvalan A, Roblero JP, Naquira C, Palma M, Backhouse C, Argandona J, Itoh T, Shuyama K, Eizuru Y, Akiba S. Human papillomavirus in esophageal squamous cell carcinoma in Colombia and Chile. *World J Gastroenterol*. 2006; 12:6188–6192. [PubMed: 17036393]
59. Freedman ND, Park Y, Subar AF, Hollenbeck AR, Leitzmann MF, Schatzkin A, Abnet CC. Fruit and vegetable intake and esophageal cancer in a large prospective cohort study. *Int J Cancer*. 2007; 121:2753–2760. [PubMed: 17691111]
60. Brownson RC, Alavanja MC, Caporaso N, Simoes EJ, Chang JC. Epidemiology and prevention of lung cancer in nonsmokers. *Epidemiol Rev*. 1998; 20:218–236. [PubMed: 9919440]
61. Wu-Williams, AHSJ. *Epidemiology of Lung Cancer*. Lung Biology in Health and Disease. New York: Dekker; 1994.
62. Wistuba II. Genetics of preneoplasia: lessons from lung cancer. *Curr Mol Med*. 2007; 7:3–14. [PubMed: 17311529]
63. Samet JM, Wiggins CL, Humble CG, Pathak DR. Cigarette smoking and lung cancer in New Mexico. *Am Rev Respir Dis*. 1988; 137:1110–1113. [PubMed: 3264122]
64. Villeneuve PMY. Lifetime probability of developing lung cancer, by smoking status, Canada. *Canadian Journal of Public Health*. 1994; 85:4.
65. Spira A, Beane J, Shah V, Liu G, Schembri F, Yang X, Palma J, Brody JS. Effects of cigarette smoke on the human airway epithelial cell transcriptome. *Proc Natl Acad Sci U S A*. 2004; 101:10143–10148. [PubMed: 15210990]
66. Gorlova OY, Weng SF, Zhang Y, Amos CI, Spitz MR. Aggregation of cancer among relatives of never-smoking lung cancer patients. *Int J Cancer*. 2007; 121:111–118. [PubMed: 17304511]
67. Catelinois O, Rogel A, Laurier D, Billon S, Hemon D, Verger P, Tirmarche M. Lung cancer attributable to indoor radon exposure in france: impact of the risk models and uncertainty analysis. *Environ Health Perspect*. 2006; 114:1361–1366. [PubMed: 16966089]
68. O'Reilly KM, McLaughlin AM, Beckett WS, Sime PJ. Asbestos-related lung disease. *Am Fam Physician*. 2007; 75:683–688. [PubMed: 17375514]
69. Coyle YM, Minahjuddin AT, Hynan LS, Minna JD. An ecological study of the association of metal air pollutants with lung cancer incidence in Texas. *J Thorac Oncol*. 2006; 1:654–661. [PubMed: 17409932]
70. Cheng YW, Chiou HL, Sheu GT, Hsieh LL, Chen JT, Chen CY, Su JM, Lee H. The association of human papillomavirus 16/18 infection with lung cancer among nonsmoking Taiwanese women. *Cancer Res*. 2001; 61:2799–2803. [PubMed: 11306446]
71. Zheng H, Abdel Aziz HO, Nakanishi Y, Masuda S, Saito H, Tsuneyama K, Takano Y. Oncogenic role of JC virus in lung cancer. *J Pathol*. 2007; 212:306–315. [PubMed: 17534844]
72. Giuliani L, Jaxmar T, Casadio C, Gariglio M, Manna A, D'Antonio D, Syrjanen K, Favalli C, Ciotti M. Detection of oncogenic viruses SV40, BKV, JCV, HCMV, HPV and p53 codon 72 polymorphism in lung carcinoma. *Lung Cancer*. 2007; 57:273–281. [PubMed: 17400331]
73. Minna JD, Roth JA, Gazdar AF. Focus on lung cancer. *Cancer Cell*. 2002; 1:49–52. [PubMed: 12086887]
74. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell*. 1990; 61:759–767. [PubMed: 2188735]
75. Chansky K, Sculier JP, Crowley JJ, Giroux D, Van Meerbeek J, Goldstraw P. The International Association for the Study of Lung Cancer Staging Project: prognostic factors and pathologic TNM

- stage in surgically managed non-small cell lung cancer. *J Thorac Oncol.* 2009; 4:792–801. [PubMed: 19458556]
76. Kozomara RJ, Brankovic-Magic MV, Jovic NR, Stosic SM, Magic ZM. Prognostic significance of TP53 mutations in oral squamous cell carcinoma with human papilloma virus infection. *Int J Biol Markers.* 2007; 22:252–257. [PubMed: 18161655]
 77. Harris SL, Levine AJ. The p53 pathway: positive and negative feedback loops. *Oncogene.* 2005; 24:2899–2908. [PubMed: 15838523]
 78. Rodust PM, Stockfleth E, Ulrich C, Leverkus M, Eberle J. UV-induced squamous cell carcinoma-- a role for antiapoptotic signalling pathways. *Br J Dermatol.* 2009; 161 Suppl 3:107–115. [PubMed: 19775366]
 79. El-Deiry WS. Targeting mutant p53 shows promise for sunscreens and skin cancer. *J Clin Invest.* 2007; 117:3658–3660. [PubMed: 18060027]
 80. Bonner JA, Harari PM, Giralt J, Azarnia N, Shin DM, Cohen RB, Jones CU, Sur R, Raben D, Jassem J, Ove R, Kies MS, Baselga J, Yousoufian H, Amellal N, Rowinsky EK, Ang KK. Radiotherapy plus cetuximab for squamous-cell carcinoma of the head and neck. *N Engl J Med.* 2006; 354:567–578. [PubMed: 16467544]
 81. Delfino V, Casartelli G, Garzoglio B, Scala M, Mereu P, Bonatti S, Margarino G, Abbondandolo A. Micronuclei and p53 accumulation in preneoplastic and malignant lesions of the head and neck. *Mutagenesis.* 2002; 17:73–77. [PubMed: 11752237]
 82. Shin DM, Charuruks N, Lippman SM, Lee JJ, Ro JY, Hong WK, Hittelman WN. p53 protein accumulation and genomic instability in head and neck multistep tumorigenesis. *Cancer Epidemiol Biomarkers Prev.* 2001; 10:603–609. [PubMed: 11401909]
 83. Hollstein M, Sidransky D, Vogelstein B, Harris CC. p53 mutations in human cancers. *Science.* 1991; 253:49–53. [PubMed: 1905840]
 84. Reid BJ, Blount PL, Rabinovitch PS. Biomarkers in Barrett's esophagus. *Gastrointest Endosc Clin N Am.* 2003; 13:369–397. [PubMed: 12916666]
 85. Cheng TH, Hsu PK, Li AF, Hung IC, Huang MH, Hsu HS. Correlation of p53, MDM2 and p14(ARF) protein expression in human esophageal squamous cell carcinoma. *J Cancer Res Clin Oncol.* 2009; 135:1577–1582. [PubMed: 19488782]
 86. Ikeda G, Isaji S, Chandra B, Watanabe M, Kawarada Y. Prognostic significance of biologic factors in squamous cell carcinoma of the esophagus. *Cancer.* 1999; 86:1396–1405. [PubMed: 10526265]
 87. Brattstrom D, Bergqvist M, Lamberg K, Kraaz W, Scheibenflug L, Gustafsson G, Inganas M, Wagenius G, Brodin O. Complete sequence of p53 gene in 20 patients with lung cancer: comparison with chemosensitivity and immunohistochemistry. *Med Oncol.* 1998; 15:255–261. [PubMed: 9951689]
 88. Tsai CM, Chang KT, Wu LH, Chen JY, Gazdar AF, Mitsudomi T, Chen MH, Perng RP. Correlations between intrinsic chemoresistance and HER-2/neu gene expression, p53 gene mutations, and cell proliferation characteristics in non-small cell lung cancer cell lines. *Cancer Res.* 1996; 56:206–209. [PubMed: 8548764]
 89. Safran H, King T, Choy H, Gollerkeri A, Kwakwa H, Lopez F, Cole B, Myers J, Tarpey J, Rosmarin A. p53 mutations do not predict response to paclitaxel/radiation for nonsmall cell lung carcinoma. *Cancer.* 1996; 78:1203–1210. [PubMed: 8826941]
 90. Blandino G, Levine AJ, Oren M. Mutant p53 gain of function: differential effects of different p53 mutants on resistance of cultured cells to chemotherapy. *Oncogene.* 1999; 18:477–485. [PubMed: 9927204]
 91. Bergqvist M, Brattstrom D, Gullbo J, Hesselius P, Brodin O, Wagenius G. p53 status and its in vitro relationship to radiosensitivity and chemosensitivity in lung cancer. *Anticancer Res.* 2003; 23:1207–1212. [PubMed: 12820372]
 92. Barbieri CE, Pietenpol JA. p63 and epithelial biology. *Exp Cell Res.* 2006; 312:695–706. [PubMed: 16406339]
 93. Koster MI, Kim S, Roop DR. P63 deficiency: a failure of lineage commitment or stem cell maintenance? *J Investig Dermatol Symp Proc.* 2005; 10:118–123.

94. Zhang H, Liu J, Cagle PT, Allen TC, Laga AC, Zander DS. Distinction of pulmonary small cell carcinoma from poorly differentiated squamous cell carcinoma: an immunohistochemical approach. *Mod Pathol.* 2005; 18:111–118. [PubMed: 15309021]
95. Takeda T, Sugihara K, Hirayama Y, Hirano M, Tanuma JI, Semba I. Immunohistological evaluation of Ki-67, p63, CK19 and p53 expression in oral epithelial dysplasias. *J Oral Pathol Med.* 2006; 35:369–375. [PubMed: 16762018]
96. Bortoluzzi MC, Yurgel LS, Dekker NP, Jordan RC, Regezi JA. Assessment of p63 expression in oral squamous cell carcinomas and dysplasias. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2004; 98:698–704. [PubMed: 15583543]
97. Zangen R, Ratovitski E, Sidransky D. DeltaNp63alpha levels correlate with clinical tumor response to cisplatin. *Cell Cycle.* 2005; 4:1313–1315. [PubMed: 16123597]
98. Rocco JW, Leong CO, Kuperwasser N, DeYoung MP, Ellisen LW. p63 mediates survival in squamous cell carcinoma by suppression of p73-dependent apoptosis. *Cancer Cell.* 2006; 9:45–56. [PubMed: 16413471]
99. Takahashi Y, Noguchi T, Takeno S, Kimura Y, Okubo M, Kawahara K. Reduced expression of p63 has prognostic implications for patients with esophageal squamous cell carcinoma. *Oncol Rep.* 2006; 15:323–328. [PubMed: 16391849]
100. Caputi M, Groeger AM, Esposito V, Dean C, De Luca A, Pacilio C, Muller MR, Giordano GG, Baldi F, Wolner E, Giordano A. Prognostic role of cyclin D1 in lung cancer. Relationship to proliferating cell nuclear antigen. *Am J Respir Cell Mol Biol.* 1999; 20:746–750. [PubMed: 10101007]
101. Lee HJ, Choe G, Jheon S, Sung SW, Lee CT, Chung JH. CD24, a novel cancer biomarker, predicting disease-free survival of non-small cell lung carcinomas: a retrospective study of prognostic factor analysis from the viewpoint of forthcoming (seventh) new TNM classification. *J Thorac Oncol.* 5:649–657. [PubMed: 20354454]
102. Massion PP, Taflan PM, Jamshedur Rahman SM, Yildiz P, Shyr Y, Edgerton ME, Westfall MD, Roberts JR, Pietenpol JA, Carbone DP, Gonzalez AL. Significance of p63 amplification and overexpression in lung cancer development and prognosis. *Cancer Res.* 2003; 63:7113–7121. [PubMed: 14612504]
103. Martin B, Paesmans M, Berghmans T, Branle F, Ghisdal L, Mascaux C, Meert AP, Steels E, Vallot F, Verdebout JM, Lafitte JJ, Sculier JP. Role of Bcl-2 as a prognostic factor for survival in lung cancer: a systematic review of the literature with meta-analysis. *Br J Cancer.* 2003; 89:55–64. [PubMed: 12838300]
104. Skalova A, Michal M. Patterns of cell proliferation in actinic keratoacanthomas and squamous cell carcinomas of the skin. Immunohistochemical study using the MIB 1 antibody in formalin-fixed paraffin sections. *Am J Dermatopathol.* 1995; 17:332–334. [PubMed: 8600794]
105. Fischer CA, Jung M, Zlobec I, Green E, Storck C, Tornilo L, Lugli A, Wolfensberg M, Terracciano LM. Co-overexpression of p21 and Ki-67 in head and neck squamous cell carcinoma relative to a significantly poor prognosis. *Head & Neck.* 2010
106. Matsumoto M, Furihata M, Kurabayashi A, Sasaguri S, Araki K, Hayashi H, Ohtsuki Y. Prognostic significance of serine 392 phosphorylation in overexpressed p53 protein in human esophageal squamous cell carcinoma. *Oncology.* 2004; 67:143–150. [PubMed: 15539919]
107. Meert AP, Martin B, Verdebout JM, Feoli F, Mascaux C, Ninane V, Sculier JP. EGFR, c-erbB-2 and ki-67 in NSCLC and preneoplastic bronchial lesions. *Anticancer Res.* 2006; 26:135–138. [PubMed: 16475689]
108. Alao JP. The regulation of cyclin D1 degradation: roles in cancer development and the potential for therapeutic invention. *Mol Cancer.* 2007; 6:24. [PubMed: 17407548]
109. Burnworth B, Popp S, Stark HJ, Steinkraus V, Brocker EB, Hartschuh W, Birek C, Boukamp P. Gain of 11q/cyclin D1 overexpression is an essential early step in skin cancer development and causes abnormal tissue organization and differentiation. *Oncogene.* 2006; 25:4399–4412. [PubMed: 16547504]
110. Mansoor A, McKee PH, Simpson JA, McGuire B, Hobbs C. Prognostic significance of Ki-67 and p53 immunoreactivity in cutaneous squamous cell carcinomas. *Am J Dermatopathol.* 1996; 18:351–357. [PubMed: 8879297]

111. Jensen V, Prasad AR, Smith A, Raju M, Wendel CS, Schmelz M, Leyva W, Warneke J, Krouse RS. Prognostic criteria for squamous cell cancer of the skin. *J Surg Res.* 159:509–516. [PubMed: 19375720]
112. Bito T, Ueda M, Ahmed NU, Nagano T, Ichihashi M. Cyclin D and retinoblastoma gene product expression in actinic keratosis and cutaneous squamous cell carcinoma in relation to p53 expression. *J Cutan Pathol.* 1995; 22:427–434. [PubMed: 8594075]
113. Matthias C, Branigan K, Jahnke V, Leder K, Haas J, Heighway J, Jones PW, Strange RC, Fryer AA, Hoban PR. Polymorphism within the cyclin D1 gene is associated with prognosis in patients with squamous cell carcinoma of the head and neck. *Clin Cancer Res.* 1998; 4:2411–2418. [PubMed: 9796972]
114. Mandard AM, Hainaut P, Hollstein M. Genetic steps in the development of squamous cell carcinoma of the esophagus. *Mutat Res.* 2000; 462:335–342. [PubMed: 10767643]
115. Kuwahara M, Hirai T, Yoshida K, Yamashita Y, Hihara J, Inoue H, Toge T. p53, p21(Waf1/Cip1) and cyclin D1 protein expression and prognosis in esophageal cancer. *Dis Esophagus.* 1999; 12:116–119. [PubMed: 10466043]
116. Itami A, Shimada Y, Watanabe G, Imamura M. Prognostic value of p27(Kip1) and CyclinD1 expression in esophageal cancer. *Oncology.* 1999; 57:311–317. [PubMed: 10575318]
117. Anton RC, Coffey DM, Gondo MM, Stephenson MA, Brown RW, Cagle PT. The expression of cyclins D1 and E in predicting short-term survival in squamous cell carcinoma of the lung. *Mod Pathol.* 2000; 13:1167–1172. [PubMed: 11106072]
118. Herbst RS. Review of epidermal growth factor receptor biology. *Int J Radiat Oncol Biol Phys.* 2004; 59:21–26. [PubMed: 15142631]
119. Shimizu T, Izumi H, Oga A, Furumoto H, Murakami T, Ofuji R, Muto M, Sasaki K. Epidermal growth factor receptor overexpression and genetic aberrations in meta-static squamous-cell carcinoma of the skin. *Dermatology.* 2001; 202:203–206. [PubMed: 11385224]
120. Maubec E, Duvillard P, Velasco V, Crickx B, Avril MF. Immunohistochemical analysis of EGFR and HER-2 in patients with metastatic squamous cell carcinoma of the skin. *Anticancer Res.* 2005; 25:1205–1210. [PubMed: 15865067]
121. Krahn G, Leiter U, Kaskel P, Udart M, Utikal J, Bezold G, Peter RU. Coexpression patterns of EGFR, HER2, HER3 and HER4 in non-melanoma skin cancer. *Eur J Cancer.* 2001; 37:251–259. [PubMed: 11166154]
122. Ang KK, Berkey BA, Tu X, Zhang HZ, Katz R, Hammond EH, Fu KK, Milas L. Impact of epidermal growth factor receptor expression on survival and pattern of relapse in patients with advanced head and neck carcinoma. *Cancer Res.* 2002; 62:7350–7356. [PubMed: 12499279]
123. Maurizi M, Almadori G, Ferrandina G, Distefano M, Romanini ME, Cadoni G, Benedetti-Panici P, Paludetti G, Scambia G, Mancuso S. Prognostic significance of epidermal growth factor receptor in laryngeal squamous cell carcinoma. *Br J Cancer.* 1996; 74:1253–1257. [PubMed: 8883413]
124. Hanawa M, Suzuki S, Dobashi Y, Yamane T, Kono K, Enomoto N, Ooi A. EGFR protein overexpression and gene amplification in squamous cell carcinomas of the esophagus. *Int J Cancer.* 2006; 118:1173–1180. [PubMed: 16161046]
125. Kitagawa Y, Ueda M, Ando N, Ozawa S, Shimizu N, Kitajima M. Further evidence for prognostic significance of epidermal growth factor receptor gene amplification in patients with esophageal squamous cell carcinoma. *Clin Cancer Res.* 1996; 2:909–914. [PubMed: 9816249]
126. Shigematsu H, Lin L, Takahashi T, Nomura M, Suzuki M, Wistuba II, Fong KM, Lee H, Toyooka S, Shimizu N, Fujisawa T, Feng Z, Roth JA, Herz J, Minna JD, Gazdar AF. Clinical and biological features associated with epidermal growth factor receptor gene mutations in lung cancers. *J Natl Cancer Inst.* 2005; 97:339–346. [PubMed: 15741570]
127. Steels E, Paesmans M, Berghmans T, Branle F, Lemaitre F, Mascaux C, Meert AP, Vallot F, Lafitte JJ, Sculier JP. Role of p53 as a prognostic factor for survival in lung cancer: a systematic review of the literature with a meta-analysis. *Eur Respir J.* 2001; 18:705–719. [PubMed: 11716177]

128. Mitsudomi T, Hamajima N, Ogawa M, Takahashi T. Prognostic significance of p53 alterations in patients with non-small cell lung cancer: a meta-analysis. *Clin Cancer Res*. 2000; 6:4055–4063. [PubMed: 11051256]
129. Tammemagi MC, McLaughlin JR, Bull SB. Meta-analyses of p53 tumor suppressor gene alterations and clinicopathological features in resected lung cancers. *Cancer Epidemiol Biomarkers Prev*. 1999; 8:625–634. [PubMed: 10428201]
130. Nakamura H, Kawasaki N, Taguchi M, Kabasawa K. Association of HER-2 overexpression with prognosis in nonsmall cell lung carcinoma: a metaanalysis. *Cancer*. 2005; 103:1865–1873. [PubMed: 15770690]
131. Pelosi G, Del Curto B, Dell'Orto P, Pasini F, Veronesi G, Spaggiari L, Maisonneuve P, Iannucci A, Terzi A, Lonardoni A, Viale G. Lack of prognostic implications of HER-2/neu abnormalities in 345 stage I non-small cell carcinomas (NSCLC) and 207 stage I–III neuroendocrine tumours (NET) of the lung. *Int J Cancer*. 2005; 113:101–108. [PubMed: 15386424]
132. Mendelsohn J. Targeting the epidermal growth factor receptor for cancer therapy. *J Clin Oncol*. 2002; 20:1S–13S. [PubMed: 12235219]
133. Meert AP, Martin B, Delmotte P, Berghmans T, Lafitte JJ, Mascaux C, Paesmans M, Steels E, Verdebout JM, Sculier JP. The role of EGF-R expression on patient survival in lung cancer: a systematic review with metaanalysis. *Eur Respir J*. 2002; 20:975–981. [PubMed: 12412692]
134. Niemiec J, Kolodziejwski L, Dyczek S. EGFR LI and Ki-67 LI are independent prognostic parameters influencing survivals of surgically treated squamous cell lung cancer patients. *Neoplasma*. 2005; 52:231–237. [PubMed: 15875085]
135. Hirsch FR, Varella-Garcia M, Bunn PA Jr, Di Maria MV, Veve R, Bremmes RM, Baron AE, Zeng C, Franklin WA. Epidermal growth factor receptor in non-small-cell lung carcinomas: correlation between gene copy number and protein expression and impact on prognosis. *J Clin Oncol*. 2003; 21:3798–3807. [PubMed: 12953099]
136. Nakamura H, Kawasaki N, Taguchi M, Kabasawa K. Survival impact of epidermal growth factor receptor overexpression in patients with non-small cell lung cancer: a meta-analysis. *Thorax*. 2006; 61:140–145. [PubMed: 16284218]
137. Ludovini V, Bellezza G, Pistola L, Bianconi F, Di Carlo L, Sidoni A, Semeraro A, Del Sordo R, Tofanetti FR, Mameli MG, Daddi G, Cavaliere A, Tonato M, Crino L. High coexpression of both insulin-like growth factor receptor-1 (IGFR-1) and epidermal growth factor receptor (EGFR) is associated with shorter disease-free survival in resected non-small-cell lung cancer patients. *Ann Oncol*. 2009; 20:842–849. [PubMed: 19153117]
138. Menter DG, Schilsky RL, DuBois RN. Cyclooxygenase-2 and cancer treatment: understanding the risk should be worth the reward. *Clin Cancer Res*. 16:1384–1390. [PubMed: 20179228]
139. Higashi Y, Kanekura T, Kanzaki T. Enhanced expression of cyclooxygenase (COX)-2 in human skin epidermal cancer cells: evidence for growth suppression by inhibiting COX-2 expression. *Int J Cancer*. 2000; 86:667–671. [PubMed: 10797288]
140. An KP, Athar M, Tang X, Katiyar SK, Russo J, Beech J, Aszterbaum M, Kopelovich L, Epstein EH Jr, Mukhtar H, Bickers DR. Cyclooxygenase-2 expression in murine and human nonmelanoma skin cancers: implications for therapeutic approaches. *Photochem Photobiol*. 2002; 76:73–80. [PubMed: 12126310]
141. Zimmermann KC, Sarbia M, Weber AA, Borchard F, Gabbert HE, Schror K. Cyclooxygenase-2 expression in human esophageal carcinoma. *Cancer Res*. 1999; 59:198–204. [PubMed: 9892207]
142. Xu XC. COX-2 inhibitors in cancer treatment and prevention, a recent development. *Anticancer Drugs*. 2002; 13:127–137. [PubMed: 11901304]
143. Ko YHI, Soon Ja, Kim Jeong Oh, Byun Jae Ho, Jung Chan Kwon, Lee Myung Ah, Hong Yeong Seon, Park Jae Kil, Wang Young Pil, Kang Jin Hyoung. Correlation of CD44, VEGF-C and COX-2 with clinicopathologic parameters and clinical outcomes. *Journal of Thoracic Oncology*. 2007; 2:779.
144. Uehara M, Sano K, Ikeda H, Sekine J, Irie A, Yokota T, Tobita T, Ohba S, Inokuchi T. Expression of vascular endothelial growth factor and prognosis of oral squamous cell carcinoma. *Oral Oncol*. 2004; 40:321–325. [PubMed: 14747064]

145. Kleespies A, Guba M, Jauch KW, Bruns CJ. Vascular endothelial growth factor in esophageal cancer. *J Surg Oncol.* 2004; 87:95–104. [PubMed: 15282704]
146. Boone J, van Hillegersberg R, Offerhaus GJ, van Diest PJ, Borel Rinkes IH, Ten Kate FJ. Targets for molecular therapy in esophageal squamous cell carcinoma: an immunohistochemical analysis. *Dis Esophagus.* 2009; 22:496–504. [PubMed: 19302210]
147. Bass AJ, Watanabe H, Mermel CH, Yu S, Perner S, Verhaak RG, Kim SY, Wardwell L, Tamayo P, Gat-Viks I, Ramos AH, Woo MS, Weir BA, Getz G, Beroukhi R, O'Kelly M, Dutt A, Rozenblatt-Rosen O, Dziunycz P, Komisarof J, Chirieac LR, Lafague CJ, Scheble V, Wilbertz T, Ma C, Rao S, Nakagawa H, Stairs DB, Lin L, Giordano TJ, Wagner P, Minna JD, Gazdar AF, Zhu CQ, Brose MS, Ceconello L, Jr UR, Marie SK, Dahl O, Shivdasani RA, Tsao MS, Rubin MA, Wong KK, Regev A, Hahn WC, Beer DG, Rustgi AK, Meyerson M. SOX2 is an amplified lineage-survival oncogene in lung and esophageal squamous cell carcinomas. *Nat Genet.* 2009; 41:1238–1242. [PubMed: 19801978]
148. Wang Q, He W, Lu C, Wang Z, Wang J, Giercksky KE, Nesland JM, Suo Z. Oct3/4 and Sox2 are significantly associated with an unfavorable clinical outcome in human esophageal squamous cell carcinoma. *Anticancer Res.* 2009; 29:1233–1241. [PubMed: 19414369]
149. Freier K, Knoepfle K, Flechtenmacher C, Pungs S, Devens F, Toedt G, Hofele C, Joos S, Lichter P, Radlwimmer B. Recurrent copy number gain of transcription factor SOX2 and corresponding high protein expression in oral squamous cell carcinoma. *Genes Chromosomes Cancer.* 49:9–16. [PubMed: 19787784]
150. Xu N, Papagiannakopoulos T, Pan G, Thomson JA, Kosik KS. MicroRNA-145 regulates OCT4, SOX2, and KLF4 and represses pluripotency in human embryonic stem cells. *Cell.* 2009; 137:647–658. [PubMed: 19409607]
151. Yuan P, Kadara H, Behrens C, Tang X, Woods D, Solis LM, Huang J, Spinola M, Dong W, Yin G, Fujimoto J, Kim E, Xie Y, Girard L, Moran C, Hong WK, Minna JD, Wistuba II. Sex determining region Y-Box 2 (SOX2) is a potential cell-lineage gene highly expressed in the pathogenesis of squamous cell carcinomas of the lung. *PLoS One.* 5:e9112. [PubMed: 20161759]
152. Bongiorno PF, al-Kasspoles M, Lee SW, Rachwal WJ, Moore JH, Whyte RI, Orringer MB, Beer DG. E-cadherin expression in primary and metastatic thoracic neoplasms and in Barrett's oesophagus. *Br J Cancer.* 1995; 71:166–172. [PubMed: 7819034]
153. Chiles MC, Ai L, Zuo C, Fan CY, Smoller BR. E-cadherin promoter hypermethylation in preneoplastic and neoplastic skin lesions. *Mod Pathol.* 2003; 16:1014–1018. [PubMed: 14559984]
154. Uchikado Y, Natsugoe S, Okumura H, Setoyama T, Matsumoto M, Ishigami S, Aikou T. Slug Expression in the E-cadherin preserved tumors is related to prognosis in patients with esophageal squamous cell carcinoma. *Clin Cancer Res.* 2005; 11:1174–1180. [PubMed: 15709186]
155. O'Grady A, Dunne C, O'Kelly P, Murphy GM, Leader M, Kay E. Differential expression of matrix metalloproteinase (MMP)-2, MMP-9 and tissue inhibitor of metalloproteinase (TIMP)-1 and TIMP-2 in non-melanoma skin cancer: implications for tumour progression. *Histopathology.* 2007; 51:793–804. [PubMed: 18042068]
156. Kurahara S, Shinohara M, Ikebe T, Nakamura S, Beppu M, Hiraki A, Takeuchi H, Shirasuna K. Expression of MMPs, MT-MMP, and TIMPs in squamous cell carcinoma of the oral cavity: correlations with tumor invasion and metastasis. *Head Neck.* 1999; 21:627–638. [PubMed: 10487950]
157. Gorogh T, Beier UH, Baumken J, Meyer JE, Hoffmann M, Gottschlich S, Maune S. Metalloproteinases and their inhibitors: influence on tumor invasiveness and metastasis formation in head and neck squamous cell carcinomas. *Head Neck.* 2006; 28:31–39. [PubMed: 16265652]
158. Jeon YK, Lee BY, Kim JE, Lee SS, Kim CW. Molecular characterization of Epstein-Barr virus and oncoprotein expression in nasopharyngeal carcinoma in Korea. *Head Neck.* 2004; 26:573–583. [PubMed: 15229899]
159. Ondruschka C, Buhtz P, Motsch C, Freigang B, Schneider-Stock R, Roessner A, Boltze C. Prognostic value of MMP-2, -9 and TIMP-1,-2 immunoreactive protein at the invasive front in advanced head and neck squamous cell carcinomas. *Pathol Res Pract.* 2002; 198:509–515. [PubMed: 12389993]

160. Gu ZD, Li JY, Li M, Gu J, Shi XT, Ke Y, Chen KN. Matrix metalloproteinases expression correlates with survival in patients with esophageal squamous cell carcinoma. *Am J Gastroenterol.* 2005; 100:1835–1843. [PubMed: 16086722]
161. Tanioka Y, Yoshida T, Yagawa T, Saiki Y, Takeo S, Harada T, Okazawa T, Yanai H, Okita K. Matrix metalloproteinase-7 and matrix metalloproteinase-9 are associated with unfavourable prognosis in superficial oesophageal cancer. *Br J Cancer.* 2003; 89:2116–2121. [PubMed: 14647147]
162. Mukherjee S, Roth MJ, Dawsey SM, Yan W, Rodriguez-Canales J, Erickson HS, Hu N, Goldstein AM, Taylor PR, Richardson AM, Tangrea MA, Chuaqui RF, Emmert-Buck MR. Increased matrix metalloproteinase activation in esophageal squamous cell carcinoma. *J Transl Med.* 8:91. [PubMed: 20920372]
163. O'Regan EM, Toner ME, Finn SP, Fan CY, Ring M, Hagmar B, Timon C, Smyth P, Cahill S, Flavin R, Sheils OM, O'Leary JJ. p16(INK4A) genetic and epigenetic profiles differ in relation to age and site in head and neck squamous cell carcinomas. *Hum Pathol.* 2008; 39:452–458. [PubMed: 18261630]
164. Lee DJSF, Banuchi VE, Qiu W, Close LG, Assaad AM, Su GH. Multiple tumor-suppressor genes on chromosome 3p contribute to head and neck squamous cell carcinoma tumorigenesis. *Cancer Biol Ther.* 2010:10.
165. Wistuba II, Behrens C, Virmani AK, Mele G, Milchgrub S, Girard L, Fondon JW 3rd, Garner HR, McKay B, Latif F, Lerman MI, Lam S, Gazdar AF, Minna JD. High resolution chromosome 3p allelotyping of human lung cancer and preneoplastic/preinvasive bronchial epithelium reveals multiple, discontinuous sites of 3p allele loss and three regions of frequent breakpoints. *Cancer Res.* 2000; 60:1949–1960. [PubMed: 10766185]
166. Aviel-Ronen S, Blackhall FH, Shepherd FA, Tsao MS. K-ras mutations in non-small-cell lung carcinoma: a review. *Clin Lung Cancer.* 2006; 8:30–38. [PubMed: 16870043]
167. Herbst RS, Heymach JV, Lippman SM. Lung cancer. *N Engl J Med.* 2008; 359:1367–1380. [PubMed: 18815398]
168. Liggett WH Jr, Sidransky D. Role of the p16 tumor suppressor gene in cancer. *J Clin Oncol.* 1998; 16:1197–1206. [PubMed: 9508208]
169. Bian YS, Osterheld MC, Fontollet C, Bosman FT, Benhattar J. p16 inactivation by methylation of the CDKN2A promoter occurs early during neoplastic progression in Barrett's esophagus. *Gastroenterology.* 2002; 122:1113–1121. [PubMed: 11910361]
170. Ji H, Ramsey MR, Hayes DN, Fan C, McNamara K, Kozlowski P, Torrice C, Wu MC, Shimamura T, Perera SA, Liang MC, Cai D, Naumov GN, Bao L, Contreras CM, Li D, Chen L, Krishnamurthy J, Koivunen J, Chirieac LR, Padera RF, Bronson RT, Lindeman NI, Christiani DC, Lin X, Shapiro GI, Janne PA, Johnson BE, Meyerson M, Kwiatkowski DJ, Castrillon DH, Bardeesy N, Sharpless NE, Wong KK. LKB1 modulates lung cancer differentiation and metastasis. *Nature.* 2007; 448:807–810. [PubMed: 17676035]
171. Yanagawa N, Tamura G, Oizumi H, Kanauchi N, Endoh M, Sadahiro M, Motoyama T. Promoter hypermethylation of RASSF1A and RUNX3 genes as an independent prognostic prediction marker in surgically resected non-small cell lung cancers. *Lung Cancer.* 2007; 58:131–138. [PubMed: 17606310]
172. Miska EA. How microRNAs control cell division, differentiation and death. *Curr Opin Genet Dev.* 2005; 15:563–568. [PubMed: 16099643]
173. Jay C, Nemunaitis J, Chen P, Fulgham P, Tong AW. miRNA profiling for diagnosis and prognosis of human cancer. *DNA Cell Biol.* 2007; 26:293–300. [PubMed: 17504025]
174. Fabbri M. MicroRNAs and cancer epigenetics. *Curr Opin Investig Drugs.* 2008; 9:583–590.
175. Sand M, Gambichler T, Skrygan M, Sand D, Scola N, Altmeyer P, Bechara FG. Expression levels of the microRNA processing enzymes Drosha and Dicer in epithelial skin cancer. *Cancer Invest.* 28:649–653. [PubMed: 20210522]
176. Chang KW, Liu CJ, Chu TH, Cheng HW, Hung PS, Hu WY, Lin SC. Association between high miR-211 microRNA expression and the poor prognosis of oral carcinoma. *J Dent Res.* 2008; 87:1063–1068. [PubMed: 18946016]

177. Masashi Shiiba * KUaHT: MicroRNAs in Head and Neck Squamous Cell Carcinoma (HNSCC) and Oral Squamous Cell Carcinoma (OSCC). *Cancers*. 2010; 2:16.
178. Avissar M, Christensen BC, Kelsey KT, Marsit CJ. MicroRNA expression ratio is predictive of head and neck squamous cell carcinoma. *Clin Cancer Res*. 2009; 15:2850–2855. [PubMed: 19351747]
179. Hiyoshi Y, Kamohara H, Karashima R, Sato N, Imamura Y, Nagai Y, Yoshida N, Toyama E, Hayashi N, Watanabe M, Baba H. MicroRNA-21 regulates the proliferation and invasion in esophageal squamous cell carcinoma. *Clin Cancer Res*. 2009; 15:1915–1922. [PubMed: 19276261]
180. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, Yi M, Stephens RM, Okamoto A, Yokota J, Tanaka T, Calin GA, Liu CG, Croce CM, Harris CC. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell*. 2006; 9:189–198. [PubMed: 16530703]
181. Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M, Prueitt RL, Yanaihara N, Lanza G, Scarpa A, Vecchione A, Negrini M, Harris CC, Croce CM. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci U S A*. 2006; 103:2257–2261. [PubMed: 16461460]
182. Yu SL, Chen HY, Chang GC, Chen CY, Chen HW, Singh S, Cheng CL, Yu CJ, Lee YC, Chen HS, Su TJ, Chiang CC, Li HN, Hong QS, Su HY, Chen CC, Chen WJ, Liu CC, Chan WK, Li KC, Chen JJ, Yang PC. MicroRNA signature predicts survival and relapse in lung cancer. *Cancer Cell*. 2008; 13:48–57. [PubMed: 18167339]
183. Weiss GJ, Bemis LT, Nakajima E, Sugita M, Birks DK, Robinson WA, Varella-Garcia M, Bunn PA Jr, Haney J, Helfrich BA, Kato H, Hirsch FR, Franklin WA. EGFR regulation by microRNA in lung cancer: correlation with clinical response and survival to gefitinib and EGFR expression in cell lines. *Ann Oncol*. 2008; 19:1053–1059. [PubMed: 18304967]
184. Stockfleth E, Kerl H. Guidelines for the management of actinic keratoses. *Eur J Dermatol*. 2006; 16:599–606. [PubMed: 17229598]
185. Bernier J, Bentzen SM, Vermorken JB. Molecular therapy in head and neck oncology. *Nat Rev Clin Oncol*. 2009; 6:266–277. [PubMed: 19390553]
186. Razak AR, Siu LL, Le Tourneau C. Molecular targeted therapies in all histologies of head and neck cancers: an update. *Curr Opin Oncol*. 22::212–220.
187. Sutter AP, Hopfner M, Huether A, Maaser K, Scherubl H. Targeting the epidermal growth factor receptor by erlotinib (Tarceva) for the treatment of esophageal cancer. *Int J Cancer*. 2006; 118:1814–1822. [PubMed: 16217753]
188. Butts CA, Bodkin D, Middleman EL, Englund CW, Ellison D, Alam Y, Kreisman H, Graze P, Maher J, Ross HJ, Elis PM, McNulty W, Kaplan E, Pautret V, Weber MR, Shepherd FA. Randomized phase II study of gemcitabine plus cisplatin or carboplatin [corrected], with or without cetuximab, as first-line therapy for patients with advanced or metastatic non small-cell lung cancer. *J Clin Oncol*. 2007; 25:5777–5784. [PubMed: 18089875]
189. Rosell R, Robinet G, Szczesna A, Ramlau R, Constenla M, Mennecier BC, Pfeifer W, O'Byrne KJ, Welte T, Kolb R, Pirker R, Chemaissani A, Perol M, Ranson MR, Ellis PA, Pilz K, Reck M. Randomized phase II study of cetuximab plus cisplatin/vinorelbine compared with cisplatin/vinorelbine alone as first-line therapy in EGFR-expressing advanced non-small-cell lung cancer. *Ann Oncol*. 2008; 19:362–369. [PubMed: 17947225]
190. Crawford J, Sandler AB, Hammond LA. ABX-EGF in combination with paclitaxel and carboplatin for advanced non-small lung cancer (NSCLC). *J Clin Oncol*. 2004; 22:7083. (abstr).
191. Shepherd FA, Rosell R. Weighing tumor biology in treatment decisions for patients with non-small cell lung cancer. *J Thorac Oncol*. 2007; 2 Suppl 2:S68–S76. [PubMed: 17589302]
192. Sequist LVL. EGFR tyrosine kinase inhibitors in lung cancer: An evolving story. *Annu Rev Med*. 2007
193. Sequist LV, Bell DW, Lynch TJ, Haber DA. Molecular predictors of response to epidermal growth factor receptor antagonists in non-small-cell lung cancer. *J Clin Oncol*. 2007; 25:587–595. [PubMed: 17290067]

194. John T, Liu G, Tsao MS. Overview of molecular testing in non-small-cell lung cancer: mutational analysis, gene copy number, protein expression and other biomarkers of EGFR for the prediction of response to tyrosine kinase inhibitors. *Oncogene*. 2009; 28 Suppl 1:S14–S23. [PubMed: 19680292]
195. Jackman D, Pao W, Riely GJ, Engelman JA, Kris MG, Janne PA, Lynch T, Johnson BE, Miller VA. Clinical definition of acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancer. *J Clin Oncol*. 2009; 28:357–360. [PubMed: 19949011]
196. Linardou H, Dahabreh IJ, Bafaloukos D, Kosmidis P, Murray S. Somatic EGFR mutations and efficacy of tyrosine kinase inhibitors in NSCLC. *Nat Rev Clin Oncol*. 2009; 6:352–366. [PubMed: 19483740]
197. Nguyen KS, Kobayashi S, Costa DB. Acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancers dependent on the epidermal growth factor receptor pathway. *Clin Lung Cancer*. 2009; 10:281–289. [PubMed: 19632948]
198. Pao W, Miller V, Zakowski M, Doherty J, Politi K, Sarkaria I, Singh B, Heelan R, Rusch V, Fulton L, Mardis E, Kupfer D, Wilson R, Kris M, Varmus H. EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc Natl Acad Sci U S A*. 2004; 101:13306–13311. [PubMed: 15329413]
199. Kobayashi K, Nishioka M, Kohno T, Nakamoto M, Maeshima A, Aoyagi K, Sasaki H, Takenoshita S, Sugimura H, Yokota J. Identification of genes whose expression is upregulated in lung adenocarcinoma cells in comparison with type II alveolar cells and bronchiolar epithelial cells in vivo. *Oncogene*. 2004; 23:3089–3096. [PubMed: 14755238]
200. Faber AC, Li D, Song Y, Liang MC, Yeap BY, Bronson RT, Lifshits E, Chen Z, Maira SM, Garcia-Echeverria C, Wong KK, Engelman JA. Differential induction of apoptosis in HER2 and EGFR addicted cancers following PI3K inhibition. *Proc Natl Acad Sci U S A*. 2009; 106:19503–19508. [PubMed: 19850869]
201. LeRoith D, Werner H, Beitner-Johnson D, Roberts CT Jr. Molecular and cellular aspects of the insulin-like growth factor I receptor. *Endocr Rev*. 1995; 16:143–163. [PubMed: 7540132]
202. Warshamana-Greene GS, Litz J, Buchdunger E, Garcia-Echeverria C, Hofmann F, Krystal GW. The insulin-like growth factor-I receptor kinase inhibitor, NVP-ADW742, sensitizes small cell lung cancer cell lines to the effects of chemotherapy. *Clin Cancer Res*. 2005; 11:1563–1571. [PubMed: 15746061]
203. Allen GW, Saba C, Armstrong EA, Huang SM, Benavente S, Ludwig DL, Hicklin DJ, Harari PM. Insulin-like growth factor-I receptor signaling blockade combined with radiation. *Cancer Res*. 2007; 67:1155–1162. [PubMed: 17283150]
204. Gualberto A, Pollak M. Emerging role of insulin-like growth factor receptor inhibitors in oncology: early clinical trial results and future directions. *Oncogene*. 2009; 28:3009–3021. [PubMed: 19581933]
205. Gualberto CLM A, Dean A, Ang AL, Reynolds JM, Lee AV, Terstappen LW, Haluska P, Lipton A, Karp DD. Characterization of NSCLC patients responding to anti-IGF-IR therapy. *Journal of Clinical Oncology, 2008 ASCO Annual Meeting Proceedings*. 2008:26.
206. Wheeler DL, Huang S, Kruser TJ, Nechrebecki MM, Armstrong EA, Benavente S, Gondi V, Hsu KT, Harari PM. Mechanisms of acquired resistance to cetuximab: role of HER (ErbB) family members. *Oncogene*. 2008; 27:3944–3956. [PubMed: 18297114]
207. Sergina NV, Rausch M, Wang D, Blair J, Hann B, Shokat KM, Moasser MM. Escape from HER-family tyrosine kinase inhibitor therapy by the kinase-inactive HER3. *Nature*. 2007; 445:437–441. [PubMed: 17206155]
208. Harari PM, Wheeler DL, Grandis JR. Molecular target approaches in head and neck cancer: epidermal growth factor receptor and beyond. *Semin Radiat Oncol*. 2009; 19:63–68. [PubMed: 19028347]
209. Patel V, Senderowicz AM, Pinto D Jr, Igishi T, Raffeld M, Quintanilla-Martinez L, Ensley JF, Sausville EA, Gutkind JS. Flavopiridol, a novel cyclin-dependent kinase inhibitor, suppresses the growth of head and neck squamous cell carcinomas by inducing apoptosis. *J Clin Invest*. 1998; 102:1674–1681. [PubMed: 9802881]

210. Sato S, Kajiyama Y, Sugano M, Iwanuma Y, Tsurumaru M. Flavopiridol as a radio-sensitizer for esophageal cancer cell lines. *Dis Esophagus*. 2004; 17:338–344. [PubMed: 15569374]
211. Hay N, Sonenberg N. Upstream and downstream of mTOR. *Genes Dev*. 2004; 18:1926–1945. [PubMed: 15314020]
212. Kris MG, Riely GJ, Azzoli CG, Heelan RT, Krug LM, Pao W. Combined inhibition of mTOR and EGFR with everolimus (RAD001) and gefitinib in patients with non-small cell lung cancer who have smoked cigarettes: A phase II trial. *J Clin Oncol*. 2007; 25 abstr 7575.
213. Davies AM, McCoy J, Lara PN, Gumerlock PH, Crowley J, Gandara DR. Bortezomib + gemcitabine (Gem)/carboplatin (Carbo) results in encouraging survival in advanced non-small cell lung cancer (NSCLC): Results of a phase II Southwest Oncology Group (SWOG) trial (S0339). *J Clin Oncol*. 2006; 24 abstr 7017.
214. Eder JP, Vande Woude GF, Boerner SA, LoRusso PM. Novel therapeutic inhibitors of the c-Met signaling pathway in cancer. *Clin Cancer Res*. 2009; 15:2207–2214. [PubMed: 19318488]
215. Puri N, Salgia R. Synergism of EGFR and c-Met pathways, cross-talk and inhibition, in non-small cell lung cancer. *J Carcinog*. 2008; 7:9. [PubMed: 19240370]
216. Zucali PA, Ruiz MG, Giovannetti E, Destro A, Varella-Garcia M, Floor K, Ceresoli GL, Rodriguez JA, Garassino I, Comoglio P, Roncalli M, Santoro A, Giaccone G. Role of cMET expression in non-small-cell lung cancer patients treated with EGFR tyrosine kinase inhibitors. *Ann Oncol*. 2008; 19:1605–1612. [PubMed: 18467317]
217. Dempke WC, Suto T, Reck M. Targeted therapies for non-small cell lung cancer. *Lung Cancer*. 2007; 67:257–274. [PubMed: 19914732]
218. Thariat J, Milas L, Ang KK. Integrating radiotherapy with epidermal growth factor receptor antagonists and other molecular therapeutics for the treatment of head and neck cancer. *Int J Radiat Oncol Biol Phys*. 2007; 69:974–984. [PubMed: 17967298]
219. Lee JM, Mao JT, Krysan K, Dubinett SM. Significance of cyclooxygenase-2 in prognosis, targeted therapy and chemoprevention of NSCLC. *Future Oncol*. 2007; 3:149–153. [PubMed: 17381414]
220. Chow LW, Loo WT, Toi M. Current directions for COX-2 inhibition in breast cancer. *Biomed Pharmacother*. 2005; 59 Suppl 2:S281–S284. [PubMed: 16507393]
221. Kentaro Yamaguchi AL, Andrew J. Dannenberg, and Kotha Subbaramaiah1: Histone Deacetylase Inhibitors Suppress the Induction of c-Jun and Its Target Genes Including COX-2*. *THE JOURNAL OF BIOLOGICAL CHEMISTRY VOL*. 2005; 280:9.
222. Traynor AM, Dubey S, Eickhoff JC, Kolesar JM, Schell K, Huie MS, Groteluschen DL, Marcotte SM, Hallahan CM, Weeks HR, Wilding G, Espinoza-Delgado I, Schiller JH. Vorinostat (NSC# 701852) in patients with relapsed non-small cell lung cancer: a Wisconsin Oncology Network phase II study. *J Thorac Oncol*. 2009; 4:522–526. [PubMed: 19347984]
223. Williamson JM SK, Huang CH, Guaglianone P, Wolf GT, Urba SG. A phase II trial of sorafenib in patients with recurrent and/or metastatic head and neck squamous cell carcinoma (HNSCC): A Southwest Oncology Group (SWOG) trial. *Journal of Clinical Oncology, 2008 ASCO Annual Meeting Proceedings*. 2007; 25:1.
224. Gridelli C, Maione P, Del Gaizo F, Colantuoni G, Guerriero C, Ferrara C, Nicoletta D, Comunale D, De Vita A, Rossi A. Sorafenib and sunitinib in the treatment of advanced non-small cell lung cancer. *Oncologist*. 2007; 12:191–200. [PubMed: 17296815]
225. Karamouzis DF MV, Johnson R, Rajasenan K, Branstetter B, Argiris A. Phase II trial of pemetrexed (P) and bevacizumab (B) in patients (pts) with recurrent or metastatic head and neck squamous cell carcinoma (HNSCC): An interim analysis. *Journal of Clinical Oncology, 2008 ASCO Annual Meeting Proceedings*. 2007; 25:1.
226. Johnson DH, Fehrenbacher L, Novotny WF, Herbst RS, Nemunaitis JJ, Jablons DM, Langer CJ, DeVore RF 3rd, Gaudreault J, Damico LA, Holmgren E, Kabbinavar F. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J Clin Oncol*. 2004; 22:2184–2191. [PubMed: 15169807]
227. Fong KM, Sekido Y, Gazdar AF, Minna JD. Lung cancer. 9: Molecular biology of lung cancer: clinical implications. *Thorax*. 2003; 58:892–900. [PubMed: 14514947]

228. Cole KA, Krizman DB, Emmert-Buck MR. The genetics of cancer--a 3D model. *Nat Genet.* 1999; 21:38–41. [PubMed: 9915499]
229. Osawa H, Nakajima M, Kato H, Fukuchi M, Kuwano H. Prognostic value of the expression of Smad6 and Smad7, as inhibitory Smads of the TGF-beta superfamily, in esophageal squamous cell carcinoma. *Anticancer Res.* 2004; 24:3703–3709. [PubMed: 15736400]
230. Orian-Rousseau V. CD44, a therapeutic target for metastasising tumours. *Eur J Cancer.* 46:1271–1277. [PubMed: 20303742]
231. Fong D, Spizzo G, Gostner JM, Gastl G, Moser P, Krammel C, Gerhard S, Rasse M, Laimer K. TROP2: a novel prognostic marker in squamous cell carcinoma of the oral cavity. *Mod Pathol.* 2008; 21:186–191. [PubMed: 18084248]
232. Yanamoto S, Kawasaki G, Yoshitomi I, Iwamoto T, Hirata K, Mizuno A. Clinicopathologic significance of EpCAM expression in squamous cell carcinoma of the tongue and its possibility as a potential target for tongue cancer gene therapy. *Oral Oncol.* 2007; 43:869–877. [PubMed: 17207659]
233. Tse GM, Yu KH, Chan AW, King AD, Chen GG, Wong KT, Tsang RK, Chan AB. HER2 expression predicts improved survival in patients with cervical node-positive head and neck squamous cell carcinoma. *Otolaryngol Head Neck Surg.* 2009; 141:467–473. [PubMed: 19786214]
234. Polkowski W, van Sandick JW, Offerhaus GJ, ten Kate FJ, Mulder J, Obertop H, van Lanschoot JJ. Prognostic value of Lauren classification and c-erbB-2 oncogene overexpression in adenocarcinoma of the esophagus and gastroesophageal junction. *Ann Surg Oncol.* 1999; 6:290–297. [PubMed: 10340889]
235. Takayama T, Nagao M, Sawada H, Yamada Y, Emoto K, Fujimoto H, Ueno M, Hirao S, Nakajima Y. Bcl-X expression in esophageal squamous cell carcinoma: association with tumor progression and prognosis. *J Surg Oncol.* 2001; 78:116–123. [PubMed: 11579389]
236. Ikeguchi M, Maeta M, Kaibara N. Bax expression as a prognostic marker of postoperative chemoradiotherapy for patients with esophageal cancer. *Int J Mol Med.* 2001; 7:413–417. [PubMed: 11254884]
237. Pai SI, Westra WH. Molecular pathology of head and neck cancer: implications for diagnosis, prognosis, and treatment. *Annu Rev Pathol.* 2009; 4:49–70. [PubMed: 18729723]
238. Ohashi Y, Sasano H, Yamaki H, Shizawa S, Shineha R, Akaishi T, Satomi S, Nagura H. Cell cycle inhibitory protein p27 in esophageal squamous cell carcinoma. *Anticancer Res.* 1999; 19:1843–1848. [PubMed: 10470125]
239. Filipowicz E, Adegboyega P, Sanchez RL, Gatalica Z. Expression of CD95 (Fas) in sun-exposed human skin and cutaneous carcinomas. *Cancer.* 2002; 94:814–819. [PubMed: 11857317]
240. Kitamura A, Yashima K, Okamoto E, Andachi H, Hosoda A, Kishimoto Y, Shiota G, Ito H, Kaibara N, Kawasaki H. Reduced Fhit expression occurs in the early stage of esophageal tumorigenesis: no correlation with p53 expression and apoptosis. *Oncology.* 2001; 61:205–211. [PubMed: 11574776]
241. Khademi B, Shirazi FM, Vasei M, Doroudchi M, Gandomi B, Modjtahedi H, Pezeshki AM, Ghaderi A. The expression of p53, c-erbB-1 and c-erbB-2 molecules and their correlation with prognostic markers in patients with head and neck tumors. *Cancer Lett.* 2002; 184:223–230. [PubMed: 12127695]
242. Paweletz CP, Ornstein DK, Roth MJ, Bichsel VE, Gillespie JW, Calvert VS, Vocke CD, Hewitt SM, Duray PH, Herring J, Wang QH, Hu N, Linehan WM, Taylor PR, Liotta LA, Emmert-Buck MR, Petricoin EF 3rd. Loss of annexin 1 correlates with early onset of tumorigenesis in esophageal and prostate carcinoma. *Cancer Res.* 2000; 60:6293–6297. [PubMed: 11103786]
243. Sharma R, Sud N, Chattopadhyay TK, Ralhan R. TC21/R-Ras2 upregulation in esophageal tumorigenesis: potential diagnostic implications. *Oncology.* 2005; 69:10–18. [PubMed: 16088230]
244. Rodriguez-Pineiro AM, Blanco-Prieto S, Sanchez-Otero N, Rodriguez-Berrocal FJ, de la Cadena MP. On the identification of biomarkers for non-small cell lung cancer in serum and pleural effusion. *J Proteomics.* 73:1511–1522. [PubMed: 20230924]

245. Verma A, Shukla NK, Deo SV, Gupta SD, Ralhan R. MEMD/ALCAM: a potential marker for tumor invasion and nodal metastasis in esophageal squamous cell carcinoma. *Oncology*. 2005; 68:462–470. [PubMed: 16024937]
246. Cooper SJ, Bowden GT. Ultraviolet B regulation of transcription factor families: roles of nuclear factor-kappa B (NF-kappaB) and activator protein-1 (AP-1) in UVB-induced skin carcinogenesis. *Curr Cancer Drug Targets*. 2007; 7:325–334. [PubMed: 17979627]
247. Kim DJ, Murray IA, Burns AM, Gonzalez FJ, Perdew GH, Peters JM. Peroxisome proliferator-activated receptor-beta/delta inhibits epidermal cell proliferation by down-regulation of kinase activity. *J Biol Chem*. 2005; 280:9519–9527. [PubMed: 15632134]
248. Chung JY, Braunschweig T, Hu N, Roth M, Traicoff JL, Wang QH, Knezevic V, Taylor PR, Hewitt SM. A multiplex tissue immunoblotting assay for proteomic profiling: a pilot study of the normal to tumor transition of esophageal squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev*. 2006; 15:1403–1408. [PubMed: 16835344]
249. Ridge, JA.; Glisson, BS.; Lango, MN., et al. "Head and Neck Tumors". In: Pazdur, RWL.; Camphausen, KA.; Hoskins, WJ., editors. *Cancer Management: A Multidisciplinary Approach*. Vol. 11. 2008. (Series Editor)
250. Garon EB, Finn RS, Hosmer W, Dering J, Ginther C, Adhami S, Kamranpour N, Pitts S, Desai A, Elashoff D, French T, Smith P, Slamon DJ. Identification of common predictive markers of in vitro response to the Mek inhibitor selumetinib (AZD6244; ARRY-142886) in human breast cancer and non-small cell lung cancer cell lines. *Mol Cancer Ther*. 9:1985–1994. [PubMed: 20587667]
251. Von Pawel J, Reck M, Digel W, Kortsik C, Thomas M, Frickhofen N. 1 inhibitor, in patients with relapsed advanced or metastatic non-small cell lung cancer (NSCLC). *J Clin Oncol*. 2008; 26 Suppl:8030.
252. Lapenna S, Giordano A. Cell cycle kinases as therapeutic targets for cancer. *Nat Rev Drug Discov*. 2009; 8:547–566. [PubMed: 19568282]
253. Ashkenazi A, Herbst RS. To kill a tumor cell: the potential of proapoptotic receptor agonists. *J Clin Invest*. 2008; 118:1979–1990. [PubMed: 18523647]
254. Paz-Ares L, Sanchez Torres JM, Diaz-Padilla I. Safety and efficacy of AMG 655 in combination with paclitaxel and carboplatin (PC) in patients with advanced non-small cell lung cancer. *J Clin Oncol*. 2009; 27:19048.
255. Heider KH, Kuthan H, Stehle G, Munzert G. CD44v6: a target for antibody-based cancer therapy. *Cancer Immunol Immunother*. 2004; 53:567–579. [PubMed: 14762695]

Table 1

Risk factors of SCC.

Risk factor	NMSC	HNSCC	ESCC	NSCLC
Personal habits				
Tobacco smoking	•	•••	•••	•••••
Alcohol consumption		•••	•••	
Dietary factors		• ^a	• ^b	
Oral health		•		
Environmental exposures				
UV light	•••••	•		
Occupational exposure (nickel refining, textile fibers, woodworking)		•		
Radon gas				•
Particulate matter				•
Asbestos				•
Viral infections				
HPV	•	•	•	•
EBV		•		
JCV				•
Other diseases				
Acid reflux disease		•		
Achalasia			•	
Plummer-Vinson Syndrome			•	
Ethnicity				
		• ^c	• ^d	

- = common factor
- = major factor
- = predominant factor

^a Betel nut chewing

^b High-starch diet without fruits and vegetables, or pickled vegetables

^c African American

^d African American, male

Table 2

Prognostic indicators and markers of SCC.

	NMSC	HNSCC	ESCC	NSCLC
Risk factors				
HPV		•		
Molecular markers				
TP53/p53		•	•	
EGFR	•	•	•	
Ki-67	•	•	•	•
p63	•	•	•	•
VEGF		•	•	
SOX2			•	
Smad6/7			• ^[229]	
CDH1	•		•	
CD44v6		• ^[230]		
MMPs	•	•	•	
Trop2		• ^[231]		
EpCAM		• ^[232]		
HER2		• ^[233]	• ^[234]	
CCND1	•	•	•	•
CCND1 + p53 mutation	•			
Bcl-x			• ^[235]	
Bcl-2			•	•
Bax			• ^[236]	
p16			•	
CD24				•
IGFR-1 + EGFR				•
RASSF1A				•
miR-21			•	
miR-211		•		

Table 3

Molecular abnormalities in SCC.

Marker	Class	Function	Abnormality	NMSC	HNSCC	ESCC	NSCLC
TP53	CCR	Tumor-suppressor gene regulating cell-cycle progression and cell survival	GE, GM	•	•	•	•
pRB	CCR	Tumor-suppressor gene regulating cell-cycle progression and apoptosis	GE	• ^[237]			
p16	CCR	Tumor-suppressor gene regulating senescence and cell-cycle progression	GM, M	•	•		
PTEN	CCR	Tumor-suppressor gene controlling cell proliferation and apoptosis	GE	• ^[237]			
MK167 (Ki-67)	CCR	Proto-oncogene regulating cell proliferation, increased from normal to pre-cancer	GE	•	•	•	•
CCND1	CCR	Proto-oncogene regulating cell-cycle progression	GE	•	•	•	•
p63	CCR	A member of the p53 family of transcription factors	GE	•	•	•	•
p27	CCR	G1 arrest	GE		• ^[238]		
CD95	CCR	Death ligands	GE	• ^[239]			
FHIT	CCR	Negative regulation of progression through cell cycle	GE		• ^[240]		
Bcl-x	CCR	regulate programmed cell death	GE		• ^[235]		
Bcl-2	CCR	regulate programmed cell death	GE		• ^[103]		
Bax	CCR	regulate programmed cell death	GE		• ^[236]		
EGFR	ST	Transmembrane TK acting as a central transducer of multiple signaling pathways	GE, GM	•	•	•	•
VEGF	ST	Transmembrane TK that promotes the proliferation, migration, and survival of endothelial cells during tumor growth	GE		•	•	
HER2	ST	Cell membrane surface-bound receptor tyrosine kinase leading to cell growth and differentiation	GE		• ^[241]	• ^[234]	
Annexin I	ST	Cell surface receptor linked signal transduction	GE		• ^[242]		
RRas2	ST	Small GTPase mediated signal transduction	GE, GM		• ^[243]		
Smad6/7	ST	TGF beta receptor signaling pathway	GE		• ^[229]		
NKX2-1 (TTF-1)	ST	Regulates gene transcription	GE, GM				•
BRAF	ST	Protein kinase is involved in sending signals in cells and in cell growth	GE, GM				•
c-MET	ST	Membrane receptor that is essential for embryonic development and wound healing	GE, GM				•
PEDF	ST	Serine protease inhibitor	GE				• ^[244]
PIK3CA	ST	Phosphatidylinositol 3-kinase	GE, GM				•
LKB1	ST	Protein kinase regulates cell polarity and functions as a tumor suppressor	GE, GM				•
RASSF1A	ST	Encord protein mediated signal transduction	GE, M				•
CD24	ST	Encord protein mediated signal transduction	GE				• ^[101]

Marker	Class	Function	Abnormality	NMSC	HNSCC	ESCC	NSCLC
CD44v6	CA	Transmembrane glycoproteins that maintain tissue integrity by mediating contact between cells or between cells and the extracellular matrix	GE		● ^[230]		
CDH1	CA	Ca 2+ -dependent cell-cell adhesion molecule, induce and maintain intercellular connections	GE, M	●		●	
Trop2	CA	Cell adhesion, signal transduction	GE		● ^[231]		
EpCAM	CA	Cell adhesion, signal transduction	GE, GM		● ^[232]		
ALCAM	CA	Cell adhesion, signal transduction	GE			● ^[245]	
MMPs	EMD	A family of zinc-dependent proteolytic enzymes that degrade the basement membrane and other components of the extracellular matrix	GE	●	●	●	
COX-2	PM	Prostaglandin metabolism	GE	●	●	●	●
SOX-2	TF	Transcription factor	GE		●	●	●
AP-1	TF	Transcript factor	GE	● ^[246]			
PPARb/d	TF	Transcript factor	GE	● ^[247]			
CK-14	CK	Epidermis development	GE			● ^[248]	
CK-4	CK	Cytoskeleton organization and biogenesis	GE			● ^[248]	

CCR = cell-cycle regulation

ST = signal transduction

CA = cell adhesion

EMD = extracellular matrix degradation

PM = protein metabolism

TF = transcript factor

CK = cytokeratin

PK = protein kinase

GE = gene expression

GM = gene mutation

M = methylation

Table 4

Molecular targets for therapy in SCC.

Target category	Target	Drug example	Construction	NMSC	HNSCC	ESCC	NSCLC
Growth factor receptor	EGFR	cetuximab/panitumumab	Monoclonal Antibody		•	•	•
		erlotinib/gefitinib	tyrosine kinase inhibitors		•	•	•
Signal transduction	IGF-IR	IGF-IR antibody	Monoclonal Antibody		•		•
	HER3	HER3 antibody	Monoclonal Antibody		•		•
	mTOR	everolimus	mTOR inhibitor		•		•
Raf/VEGFR	MET	SU11274/PF-02341066	Small molecule		•		•
	Sorafenib	Sorafenib	Small molecule		•		•
	Ras	reovirus	Virus		• ^[249]		
Angiogenesis	MEK1/2	AZD6244	Small molecule		• ^[250]		
	VEGF	bevacizumab	Monoclonal Antibody		•		•
Cell cycle	Raf/VEGFR	Sorafenib	Small molecule		•		•
	CCND1	Flavopiridol	Kinase inhibitor		•		•
Protein degradation	Cox-2	Celecoxib/Meloxicam	Kinase inhibitor		• ^a		•
	Histone deacetylases	vorinostat	HDAC inhibitors				•
Mitosis	Polo-like kinases	BI 2536	PLK1 inhibitor				• ^[251]
	Aurora Kinase A/B	Aurora Kinase inhibitor	serine/threonine kinases inhibitor		• ^[252]		
Inducers of apoptosis	APO2	AMG655/conatumumab	Monoclonal Antibody		• ^[253]		• ^[254]
	CD44v6	CD44v6 antibody	Monoclonal Antibody		• ^[230, 255]		

^aDiclofenace