



Published in final edited form as:

J Neurosci. 2012 March 7; 32(10): 3516–3528. doi:10.1523/JNEUROSCI.3905-11.2012.

Identification of Modulators of Hair Cell Regeneration in the Zebrafish Lateral Line

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Abstract

The external location of the zebrafish lateral line makes it a powerful model for studying mechanosensory hair cell regeneration. We have developed a chemical screen to identify FDA-approved drugs and biologically active compounds that modulate hair cell regeneration in zebrafish. Of the 1,680 compounds evaluated, we identified 2 enhancers and 6 inhibitors of regeneration. The two enhancers, dexamethasone and prednisolone, are synthetic glucocorticoids that potentiated hair cell numbers during regeneration and also induced hair cell addition in the absence of damage. BrdU analysis confirmed that the extra hair cells arose from mitotic activity. We found that dexamethasone and prednisolone, like other glucocorticoids, suppress zebrafish caudal fin regeneration, indicating that hair cell regeneration occurs by a distinctly different process. Further analyses of the regeneration inhibitors revealed that two of the six, flubendazole and topotecan, significantly suppress hair cell regeneration by preventing proliferation of hair cell precursors. Flubendazole halted support cell division in M-phase, possibly by interfering with normal microtubule activity. Topotecan, a topoisomerase inhibitor, killed both hair cells and proliferating hair cell precursors. A third inhibitor, fulvestrant, moderately delays hair cell regeneration by reducing support cell proliferation. Our observation that hair cells do not regenerate when support cell proliferation is impeded confirms previous observations that cell division is the primary route for hair cell regeneration after neomycin treatment in zebrafish.

INTRODUCTION

Mechanosensory hair cells of the inner ear allow vertebrates to sense sound and orient in space. Normal aging, genetic predispositions, prolonged or excessive exposure to high sound levels, environmental toxins and some medications can readily induce damage or loss of hair cells, often leading to permanent hearing deficits and balance disorders. Mammals, including humans, have a full complement of hair cells at birth, but lack the capacity to regenerate lost hair cells (Roberson and Rubel, 1994; Oesterle and Stone, 2008; Warchol, 2010).

Replacement of damaged and dying sensory hair cells is robust in both developing and mature non-mammalian vertebrates (Cruz et al., 1987; Cotanche, 1987; Corwin and

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The authors declare no conflicts of interest.

Cotanche, 1988; Jorgensen and Matheson, 1988; Ryals and Rubel, 1988; Stone and Cotanche, 2007; Edge and Chen, 2008; Brignull et al., 2009). Candidate molecules have been assessed for their role in hair cell regeneration, including *Atoh1*, members of the Notch signaling pathway, *p27^{Kip1}*, *Rb*, and *TGF α* (Chen and Segil, 1999; Lowenheim et al., 1999; Stone and Rubel, 1999; Zheng and Gao, 2000; Li et al., 2003; Izumikawa et al., 2005; Sage et al., 2005, 2006; Cafaro et al., 2007; Breuskin et al., 2008; Collado et al., 2008; Oesterle and Stone, 2008; Brigande and Heller, 2009; Groves, 2010). However, many questions remain about the precise pathways responsible for initiating, maintaining and limiting hair cell regeneration in non-mammalian vertebrates, and correspondingly, what molecular pathways prevent regeneration of hair cells in the mammalian inner ear.

To identify novel and unexplored pathways that can regulate hair cell regeneration, we undertook a chemical screen for modulators of hair cell regeneration using zebrafish lateral line neuromasts as a platform. Zebrafish larvae have been utilized in many high-throughput drug-screening assays for studying cell death and regeneration (Fleming et al., 2005; Ton and Parnig, 2005; Mathew et al., 2007; Coffin et al., 2010; Oppedal and Goldsmith, 2010; Ou et al., 2010). The hair cells located within neuromasts of the zebrafish lateral line system share structural and molecular similarities with mammalian inner ear hair cells (Whitfield, 2002; Nicolson, 2005; Brignull et al., 2009). Like mammalian hair cells, zebrafish hair cells are sensitive to damage from ototoxic drugs such as aminoglycoside antibiotics. Unlike mammals, zebrafish have the capacity to renew the full complement of hair cells within 72 hrs (Williams and Holder, 2000; Harris et al., 2003; Hernandez, et al., 2006; Lopez-Schier and Hudspeth, 2006; Ma et al., 2008). The ability to directly visualize lateral line hair cells and manipulate environmental exposure in concert with an array of genetic tools and high fecundity makes the zebrafish lateral line a useful system for understanding hair cell regeneration.

We have developed a chemical screen that takes advantage of neuromasts in the larval zebrafish lateral line system to evaluate FDA-approved drugs and bioactives for modulators of hair cell regeneration. Two drugs were found to enhance hair cell regeneration while six others retarded regeneration. We report dose-response relationships and experiments to assess the targets and the mechanisms by which these drugs modulate hair cell regeneration.

MATERIALS AND METHODS

Zebrafish

Adult zebrafish were housed and maintained at 28.5°C in the University of Washington zebrafish facility (Westerfield, 1995). All zebrafish procedures and protocols were approved by the University of Washington Animal Care and Use Committee. Zebrafish embryos were obtained from paired matings and raised at 28.5°C in fish embryo medium (hereafter abbreviated EM; 1 mM MgSO₄, 120 μ M KH₂PO₄, 74 μ M Na₂HPO₄, 1 mM CaCl₂, 500 μ M KCl, 15 μ M NaCl, and 500 μ M NaHCO₃ in dH₂O). Larvae were kept at a density of 50 animals per 100-mm² Petri dish in approximately 30 mL EM. At 4 days post-fertilization (dpf), larvae were fed live rotifers and dry food, and then transferred into fresh EM. For our experiments, in addition to the wildtype *AB strain, we used *Tg(pou4f3:gap43-GFP)* zebrafish, a transgenic line that endogenously expresses green fluorescent protein (GFP) in both immature and mature hair cells of the lateral line and inner ear under the control of the *pou4f3* (*brn3c*) promoter (courtesy of H. Baier). In these fish, GFP is targeted to the plasma membrane with a GAP-43 membrane targeting sequence (Xiao et al., 2005).

Drug Libraries

Two drug libraries were used: the NINDS Custom Collection II (Microsource, Gaylordsville, CT) and the FDA-approved drug library (Enzo LifeSciences, formerly BioMol, Farmingdale, NY). The NINDS library consists of 1,040 drugs and bioactive compounds dissolved in dimethyl sulfoxide (DMSO) at 10 mM concentration. Some drugs in this library are not FDA-approved, but all have known biological activity. The Enzo library contains 640 FDA-approved compounds dissolved in DMSO at 2 mg/mL, all of which have confirmed and well-characterized bioactivity. The identities of the individual compounds were blinded during the primary and confirmatory screens.

Primary screening protocol

Larvae from *Tg(pou4f3:gap43-GFP)* heterozygote crosses were collected and initially screened for GFP expression in hair cells after 2 dpf. At 5–6 dpf, all free-swimming larvae (except untreated controls; see below) were treated with 400 μ M neomycin sulfate (Sigma, catalog #N1142, St. Louis, MO) in EM for 1 hr and immediately rinsed four times in fresh EM (Harris et al., 2003; Murakami et al., 2003; Santos et al., 2006). Larvae were distributed using a wide-bore glass pipette into 48-well plates (Corning, catalog #3338, Lowell, MA) at a density of 3 fish per well. The volume in each well was adjusted to 1 mL with fresh EM. Each 48-well plate contained 8 control wells and 40 experimental wells (Figure 1A). The control wells included: fish that received no neomycin exposure and were subsequently held in drug-free EM (mock treatment; 2 wells); neomycin-treated fish, subsequently held in drug-free EM (neomycin-only treatment; 2 wells); neomycin-treated fish, subsequently held in either 0.1% (control for NINDS library) or 0.2% (control for Enzo library) DMSO (neomycin + drug vehicle treatment; 2 wells); and neomycin-treated fish subsequently held in 50 μ M DAPT (N-[N-(3,5-Difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester), a γ -secretase inhibitor (positive control; 2 wells). DAPT has been previously shown to diminish Notch signaling by preventing cleavage and release of the Notch intracellular domain (Geling et al., 2002), which results in an increase in the number of zebrafish lateral line hair cells regenerated after neomycin exposure (Ma et al., 2008). Experimental drugs from the NINDS Custom Collection II library were diluted in EM to a final concentration of 10 μ M drug in 0.1% DMSO. Drugs from the Enzo FDA-approved drug library were diluted to a final concentration of 4 μ g/mL in 0.2% DMSO. While concentrations of drugs varied, 93% were between 1–20 μ M concentration and none had a concentration greater than 35 μ M. Larvae were fed live rotifers 24 hrs after the neomycin treatment and monitored daily for general drug toxicity. After 48 hrs in the experimental drug, larvae were anesthetized in buffered 0.001% MS-222 (3-aminobenzoic acid ethyl ester methanesulfonate; Sigma, catalog #E10521) prior to observation and scoring.

Larvae were examined and scored under epifluorescent illumination using a 40 \times objective (NA = 0.75) on a Zeiss Axioplan 2ie epifluorescence microscope (Thornwood, NY). Each fish was scored for degree of neuromast hair cell regeneration on a scale from 1 to 5 (Figure 1B), with 1 being greatly reduced or absent regeneration, 3 indicating normal regeneration as seen in drug-free and DMSO controls, and 5 being excessive regeneration, such as that seen with DAPT treatment. At least 8 neuromasts per fish were evaluated prior to assigning a score and 3 or more fish were scored per drug. To control for variation between experiments, scores for each round of screening experiments were calibrated to the DMSO controls in that experiment. All scoring was performed by the same investigator.

Drugs (hereafter termed “regeneration modulators”) with average scores of ≥ 3.5 were considered putative enhancers of regeneration while drugs with average scores of ≤ 2.5 were considered putative inhibitors of regeneration. All regeneration modulators were retested twice following an identical protocol to verify the effect of the modulator on regeneration.

Drugs that consistently had high or low scores across the three replicates (initial screen + two rescreens) were established as putative “hits” and further tested, as described below.

Regeneration modulators that caused a reduction in the number of regenerated hair cells (inhibitors) were tested for hair cell toxicity. To discriminate between inhibition of hair cell regeneration and toxicity, 3 larvae were treated with only the inhibitor for 48 hrs (no prior neomycin-induced hair cell death) at the same concentrations as in the primary screen. Fish were then anesthetized and scored as before.

Dose-Response Functions

We next assessed the dose-response relationships of all putative regeneration modulators to determine the lowest concentration that produced the maximum modulation effect. Wildtype *AB larval zebrafish (5–6 dpf) were treated with 400 μ M neomycin for 1 hr and rinsed 4 times in fresh EM. Groups of 10–12 neomycin-treated larvae were then transferred by Netwell baskets (Corning, catalog #3480) into separate wells of a 6-well plate containing concentrations of experimental drug, ranging from 0.1 to 100 μ M, for 48 hrs at 28.5°C. In order to standardize the DMSO concentration between the different test drug concentrations, DMSO concentrations were (with 1 exception) adjusted to 1.0% for all conditions. The exception was topotecan, which is not soluble in DMSO; hence topotecan was dissolved in water. DAPT treatment and no drug treatment controls were also evaluated in each experiment.

For rapid assessment, lateral line hair cells were labeled with the fluorescent vital dye DASPEI (2-(4-(dimethylamino)styryl)-N-ethylpyridinium iodide; 0.005% final concentration in EM; Molecular Probes, catalog #D426, Eugene, OR) for 15 min (Harris et al., 2003; Murakami et al., 2003; Owens et al., 2008, 2009; Coffin et al., 2009). Larvae were then rinsed twice in fresh EM, anesthetized and visualized using a Leica MZFI111 epifluorescent dissecting microscope (Wetzlar, Germany) equipped with a DASPEI filter set (excitation 450–490 nm and barrier 515; Chroma Technologies, Brattleboro, VT). Ten neuromasts were evaluated per fish: supraorbital (SO1, SO2), infraorbital (IO1–4), mandibular (M2), middle (MI1, MI2) and otic (O2) (Raible and Kruse, 2000). Each was assigned a score of 0–2: 0 (little/no staining), 1 (reduced staining) and 2 (normal staining) for a combined score between 0 and 20 per fish. 8–12 fish were assessed for each condition and scores were averaged for each group (\pm one standard deviation).

Hair cell immunohistochemistry

To count hair cells, *Tg(pou4f3:gap43-GFP)* transgenic larvae were euthanized and fixed in 4% paraformaldehyde (PFA) in 0.1 M phosphate-buffered saline (PBS) at pH 7.2, overnight at 4°C. After fixation, larvae were rinsed 3 times for 20 min in PBS with 0.1% Triton X-100 (PBS-T), washed for 30 min in distilled water, and incubated for 1 hr with blocking solution (1% bovine serum albumin, 1% DMSO, and 0.02% sodium azide in PBS-T) plus 5% normal goat serum to prevent nonspecific antibody binding. Larvae were subsequently held overnight in rabbit anti-GFP antibody (1:500 in blocking solution; Invitrogen, catalog #G10362, Eugene, OR), rinsed 3 times for 20 min with PBS-T, and incubated for 5 hrs in a goat anti-rabbit IgG antibody conjugated with Alexa-488 (1:500 in blocking solution; Invitrogen, catalog #A11008). Larvae were rinsed 3 times in PBS-T, mounted in 50% glycerol/PBS on bridged cover slips and visualized with a Zeiss Axioplan 2ie epifluorescence microscope under a 40 \times objective (NA = 0.75). GFP-positive hair cells were counted in 7 neuromasts per fish (MI1–2, O1–2, M2, IO4, OP1; Raible and Kruse, 2000) for 10 fish per group. Data are presented as mean summed hair cells in these 7 neuromasts (\pm one standard deviation), as compared to control fish maintained in the same conditions within the same experiment.

Cell proliferation labeling

To assess mitotic events in neuromasts of the lateral line during the first 24 hrs of the regeneration period, fish were co-treated with 5'-bromo-2'-deoxyuridine (BrdU; Sigma, catalog #B5002) with and without each modulatory drug. Following neomycin exposure, larvae were simultaneously incubated with the optimal concentration of drug as determined in dose-response tests and 5 mM BrdU in EM (adjusted to a 1% DMSO concentration) for 24 hrs (for inhibitors) or 48 hrs (for enhancers) at 28.5°C. The fish were then euthanized with MS-222, fixed in 4% PFA overnight at 4°C and rinsed several times in PBS-T. In order to visualize hair cells as well as BrdU incorporation, fish were first immunostained with rabbit anti-GFP and Alexa 488-conjugated goat anti-rabbit. BrdU immunohistochemical processing was then performed as described previously (Harris, et al., 2003; Ma et al., 2008), with several modifications. Fixed larvae were rinsed three times for 20 min in PBDT (PBS, 1% DMSO, 0.1% Tween 20). Due to the superficial nature of hair cells and neuromasts, methanol dehydration/rehydration and proteinase K were not used. Instead, samples were incubated with 1 N hydrochloric acid for 1 hr at room temperature and rinsed 3 times in PBDT for 5 min. Before the addition of antibodies, larvae were placed in blocking solution (10% normal goat serum in PBDT) for 1 hr at room temperature. Mouse anti-BrdU (BD Pharmingen, catalog #555627, San Jose, CA) was used at a 1:250 dilution in blocking solution. Samples were then incubated in goat anti-mouse IgG secondary antibody conjugated with Alexa-568 (1:500 dilution in blocking solution; Invitrogen, catalog #A11004). Larvae were finally rinsed several times in PBS-T and stored in 50% glycerol/PBS at 4°C prior to visualization.

To assess cells in M-phase, whole-mount immunohistochemistry was performed to detect phospho-histone H3 (Ma et al., 2008). Fish were placed in 400 µM neomycin for 1 hr, rinsed 4 times in fresh EM, and treated with the optimal concentration of inhibitor drug (1% DMSO adjusted) in EM for 24 hrs at 28.5°C. Larvae were then anesthetized with MS-222, fixed in 4% PFA overnight at 4°C and rinsed several times for 20 min in PBS-T. Hair cell labeling was first performed using the rabbit anti-GFP antibody and Alexa 488-conjugated goat anti-rabbit IgG before immunostaining for phospho-histone H3. Larvae were then washed in distilled water for 30 min, treated with 5% normal goat serum in blocking solution and incubated overnight with rat anti-phospho-Histone H3 (ser10) primary antibodies (1:500 dilution in blocking solution; Millipore, catalog #06-570, Billerica, MA). Following three washes with PBS-T for 20 min, samples were labeled for 5 hrs with Alexa-568 conjugated goat anti-rat IgG (1:500 dilution in blocking solution). Larvae were finally rinsed three more times in PBS-T and stored in 50% glycerol/PBS at 4°C prior to visualization.

For evaluating total cell numbers within neuromasts, *AB wildtype zebrafish larvae were euthanized and fixed in 4% PFA overnight at 4°C. After several rinses in PBS-T, fish were incubated in the pan-nuclear dye, SYTOX Green (1:10,000 in PBS-T; Invitrogen; catalog #S7020), which stains nucleic acids with high-affinity, and therefore both hair cells and support cells. After 5 minutes, larvae were rinsed several times in PBS-T and stored in 50% glycerol/PBS at 4°C prior to imaging.

Samples were mounted in 50% glycerol/PBS on bridged coverslips and observed on a Zeiss LSM5 Pascal confocal microscope under a 40× objective (NA = 1.3). Double-labeled cells were counted in 7 neuromasts per fish (MI1-2, O1-2, M2, IO4, OP1) and compared to values from vehicle-only control fish. Images were processed using ImageJ, Pascal, and Adobe Photoshop CS4 software. Counts are presented as mean total cell numbers per fish (+/- one standard deviation). We calculated *t*-tests and ANOVA (one- and two-way) to evaluate statistical significance ($\alpha=0.05$).

Caudal fin amputation

Wildtype *AB zebrafish embryos were dechorionated at 2–3 dpf, anesthetized with MS-222 and transferred to a glass depression slide. Using a surgical blade (Fine Science Tools, catalog #10015-00, 10007-12, Foster City, CA), the caudal fin primordia was amputated posterior to the notochord (Mathew et al., 2007). Larvae were then photographed under a Zeiss Axioplan 2 bright field microscope (Thornwood, NY) with a differential interference contrast filter using a 10× objective (NA = 0.17) to document the point of amputation. Fish was then placed in individual wells of a 48-well plate containing the optimal concentration of a modulator drug in 1 mL EM (1% DMSO adjusted) and allowed to recover at 28.5°C for 72 hrs. Each fish was anesthetized with MS-222 and the caudal fin was re-imaged. Caudal fin regeneration was assessed by subtracting the distance from the caudal fin tip to the notochord immediately after amputation from that measurement at 72 hrs post-amputation. The caudal fin tip was defined as the point at the end of the fin directly along the midline from the notochord.

RESULTS

Rapid screening for modulators of hair cell regeneration

Using the zebrafish lateral line system, we screened for novel drugs and small drug-like compounds that affect the rate or amount of hair cell regeneration (Figure 1A). To induce regeneration, we exposed zebrafish larvae at 5 days post-fertilization (5 dpf) to 400 μM neomycin for 1 hr, which kills > 90% of mature lateral line hair cells. Regeneration occurs rapidly in wildtype animals, reaching control levels within approximately 72 hrs after aminoglycoside exposure (Williams and Holder, 2000; Harris et al., 2003; Murakami et al., 2003; Lopez-Schier and Hudspeth, 2006; Hernandez et al., 2007; Ma et al., 2008). To facilitate visualization and assessment of regeneration, we used *Tg(pou4f3:gap43-GFP)* zebrafish, a transgenic line that expresses membrane-bound green fluorescent protein (GFP) in hair cells under the control of the *pou4f3* promoter. These animals are otherwise wildtype and show aminoglycoside-induced hair cell death (Figure 2A) and regeneration comparable to control animals (Figure 2B). To test whether a drug modulates regeneration, neomycin-treated animals were placed in 48-well plates and immediately exposed to test drug for a 48 hr period. At 48 hrs post-treatment, we assessed the regeneration of hair cells of several anterior lateral line neuromasts. Larvae were assigned a score on a scale from 1 to 5 based on the degree of neuromast hair cell regeneration, with a score of 1 reflecting attenuated or absent regeneration, 3 representing normal regeneration as seen in drug-free and DMSO controls, and 5 indicating excessive regeneration (Figure 1B).

Results from initial screening of the 1,680 drugs in two custom libraries are shown in Table 1. We identified 176 drugs as potential inhibitors of regeneration and 115 drugs as potential enhancers of regeneration. Similar “hit” rates were observed in the two libraries, with 18.2% (189/1040) and 15.9% (102/640) identified from the NINDS Custom Collection II and the Enzo FDA-approved library, respectively. During testing, 243 compounds from both libraries were determined to be lethal to fish. We did not assess whether lower concentrations of these drugs would allow animals to survive but affect hair cell regeneration. It should be noted that none of the compounds identified potentiated regeneration to the extent of that routinely seen with exposure to DAPT; most of the putative enhancers were given a score between 3 and 4 based on our screening method, while the DAPT-treated fish reliably produced scores of 5.

Our initial screen was designed to maximize sensitivity and thus we expected to have many false positives and ideally few false negatives for the drugs and concentrations tested. Each potential modulator of regeneration was tested twice more under the same conditions as

described in the initial screen to assess reliability and thereby eliminate false positives. We confirmed 10 compounds that putatively enhanced hair cell regeneration and 40 that putatively inhibited regeneration (Table 1). All putative modulatory drugs were then tested for ototoxicity in the absence of neomycin and a dose-response function was determined using a range of concentrations of each modulatory drug. From these validation experiments, we confirmed that 2 drugs reliably enhanced hair cell regeneration and 6 drugs that consistently inhibited regeneration (Table 2).

The synthetic glucocorticoids, dexamethasone and prednisolone, enhance hair cell regeneration in a dose-dependent manner

To find the lowest dose that produced the maximal modulation effect, we exposed larvae to varying concentrations of each enhancer drug for 48 hrs after 400 μM neomycin treatment. Both dexamethasone and prednisolone induced a significant increase in regenerated hair cells relative to DMSO controls (Figure 3). Analysis of these data with two-way ANOVA showed significant main effects of drug and concentration (control vs. drug exposure), as well as an interaction effect (p 's < 0.001). Bonferroni post-hoc tests indicated that the lowest concentration of prednisolone and dexamethasone that induced a significant increase in hair cells was 1 μM ($p < 0.01$ and $p < 0.001$ respectively). Beyond 5 μM , there was no additional increase in the number of hair cells with increasing dosage.

Each compound was also assessed for any noticeable effect in the absence of aminoglycoside-induced hair cell death. We found that both dexamethasone and prednisolone promoted hair cell addition in the absence of neomycin damage (Figure 3B, C; squares). Larvae treated in dexamethasone or prednisolone for 48 hrs had more hair cells relative to DMSO vehicle controls at drug concentrations of 1 μM or greater. The relative increase in the number of hair cells with exposure to either enhancer was similar to that seen after regeneration was induced by neomycin. A two-way ANOVA revealed significant main effects of drug and concentration (control vs. drug exposure), as well as an interaction effect ($p < 0.0001$). The lowest concentration of prednisolone and dexamethasone inducing a significant increase in hair cell numbers was again 1 μM ($p < 0.001$ and $p < 0.001$ respectively; Bonferroni post-hoc). Neither enhancer was toxic to hair cells at the doses tested (0.1–50 μM).

Dexamethasone- and prednisolone-induced hair cells arise mitotically

Increased numbers of regenerated hair cells can arise from enhanced hair cell precursor proliferation, promotion of differentiation of post-mitotic precursors, transdifferentiation of other cells into new hair cells, or prevention of hair cell death. To help determine which of these possibilities occur, we assessed the incorporation of the thymidine analog, BrdU, during the 48 hr regeneration period following neomycin-induced hair cell loss in the presence of dexamethasone or prednisolone. As Figure 4 illustrates, groups treated with 5 μM dexamethasone or prednisolone showed an increase in the number of GFP+ hair cells as compared to the DMSO vehicle control fish (t -test; $p = 0.011$ and $p = 0.021$, respectively). There was a similar increase in the number of BrdU+ hair cells, suggesting that most of the regenerated hair cells arose from dividing precursors (t -test; p 's < 0.001). In samples treated with BrdU and glucocorticoids only (no prior neomycin treatment), a significant increase in BrdU+ hair cells was noted in both dexamethasone and prednisolone-treated groups (t -test; $p = 0.0019$ and $p = 0.0202$, respectively). These observations indicate that dexamethasone and prednisolone act by stimulating presumptive hair cell precursors to divide. We observed an increase of about 1–4 additional hair cells per neuromast, indicating that these glucocorticoids stimulate a 10–25% increase in the number of hair cells in just 48 hrs. These new hair cells may arise from either additional support cells entering the cell cycle, an

interference with signaling that limits the number of hair cells produced during regeneration, or a faster cell cycle that allows a daughter cell to divide multiple times.

Glucocorticoids inhibit zebrafish caudal fin regeneration

Studies on another zebrafish tissue capable of regeneration, the caudal fin, have shown that activation of the glucocorticoid receptor (GR) using exogenous glucocorticoids was sufficient to block regeneration following fin amputation (Mathew et al., 2007). Specifically, glucocorticoids were found to impair wound epithelium and blastema formation, as well as reduce proliferation of cells just beneath the plane of amputation in an inflammation-independent manner. To determine whether either dexamethasone or prednisolone have a similar effect, we performed a fin regeneration assay similar to that described by Mathew et al., (2007). Larvae were dechorionated at 2 dpf and anesthetized. We then amputated their caudal fins and photographed them prior to exposure to glucocorticoids (Figure 5A). After 3 days with or without glucocorticoid treatment, the caudal fins were re-imaged to assess regeneration (Figure 5B). Tail fin regeneration was quantified as additional distance from the fin tip directly midline to the notochord of the regenerated fin compared to fin distance directly after amputation. In our hands, two of the strongest inhibitors from Mathew et al. (2007), beclomethasone and clobetasol, severely inhibited regeneration at a concentration of 5 μ M (Figure 5C, E). A characteristic V-shaped fin was always observed. Dexamethasone and prednisolone also produced a strong inhibitory affect at 5 μ M (Figure 5D, E), though to a somewhat lesser degree than beclomethasone and clobetasol. These results suggest but do not prove that dexamethasone and prednisolone inhibit fin regeneration via the GR pathway. Interestingly, in one regenerative tissue, the caudal fin, dexamethasone and prednisolone inhibit regeneration, while in another, the lateral line, they enhance regeneration. This hints that there may be mechanistic differences in regeneration and may reflect a difference in the cell types responsible for regeneration of these two tissues.

Testing putative inhibitors of regeneration for hair cell toxicity

A decrease in the number of regenerated hair cells might occur from impairment of proliferation, delay or inhibition of hair cell differentiation, or death of newly generated hair cells. To differentiate between inhibition of hair cell regeneration and toxicity, we tested each putative inhibitor in the absence of neomycin. All three inhibitors identified in the NINDS library showed significant hair cell toxicity in the absence of neomycin treatment (Table 1). Of the 37 inhibitors identified from the Enzo FDA-approved drug library, 21 were toxic. Since these drugs do not affect regeneration, *per se*, no further tests were conducted using these compounds. However, these drugs may provide new tools to evaluate hair cell death.

Dose-response relationships of regeneration inhibitors

Drugs identified as hair cell regeneration inhibitors that were not overtly toxic to hair cells were more thoroughly tested for dose-dependent effects. Six of these drugs significantly inhibit hair cell regeneration in a concentration-dependent manner (Figure 6). We observed two general dose-response profiles: moderate (e.g. fenoprofen) and severe inhibition (e.g. flubendazole). Several drugs showed additional hair cell toxicity or overall lethality at concentrations above 50 μ M. The lowest concentration of drug that induced the maximal inhibition of regeneration without direct toxicity was used for further experiments (Table 2). Of the 6 regeneration inhibitors identified, the two strongest were flubendazole and topotecan. These drugs reduced hair cell regeneration by 77% and 64% of controls at 5 and 50 μ M concentrations, respectively. Fulvestrant, a moderate inhibitor, reduced hair cell regeneration by 46% at a concentration of 10 μ M. Flubendazole, topotecan and fulvestrant, two strong and one moderate inhibitor of regeneration, were further pursued for more thorough characterization.

Hair cell regeneration is halted by topotecan and flubendazole and delayed by fulvestrant

Drugs that inhibit hair cell regeneration may alter regeneration by preventing proliferation, delaying parts of the process, preventing cell maturation, or promoting hair cell or hair cell progenitor death. During our screening and dose-response experiments, we analyzed hair cell regeneration only at 48 hrs post-treatment. To more precisely define when regeneration inhibitors act, we evaluated the effects of inhibitors on hair cell replacement over a 72 hr time-span. As demonstrated in Figure 7, both 5 μ M flubendazole and 10 μ M topotecan were strong inhibitors of regeneration throughout the 72 hr period tested; few to no GFP+ hair cells were observed in neuromasts at any of the time points evaluated. In the absence of neomycin treatment, flubendazole showed little hair cell toxicity prior to 72 hrs, after which there was a slight but significant decrease in hair cell numbers ($p < 0.01$; Figure 7A). Topotecan alone began noticeably killing hair cells after 48 hrs of treatment, with complete hair cell death by 72 hrs (Figure 7B). The inhibitory effects of topotecan are noticeable within 24 hrs of regeneration after neomycin treatment, while the overall toxicity of the drug does not take noticeable effect until 48 hrs, suggesting that this drug does indeed inhibit the machinery involved in regeneration. Thus, flubendazole seems to act predominantly on the process of regeneration while topotecan may block regeneration through two modes, one that occurs early in the regeneration process and perhaps later on as well, affecting hair cell differentiation or mature hair cells. In samples treated with 5 μ M fulvestrant, hair cell regeneration does occur but there is a slight decrease or delay in the number of hair cells produced at 48 and 72 hrs (Figure 7C). Analysis of these data by two-way ANOVA yielded significant main effects of group and time, and a significant interaction (all p 's < 0.0001). Pairwise comparisons (Bonferroni) yielded highly significant decreases in hair cell production by fulvestrant at 48 and 72 hrs of recovery following neomycin exposure (regeneration; filled and open triangles in Figure 7C; p 's < 0.001) and a marginally significant decrease in hair cell numbers due to fulvestrant treatment at 72 hrs in groups not receiving prior neomycin treatment (normal neuromast growth; filled and open squares in Figure 7C; $p < 0.05$).

Impact of inhibitors on proliferation

We next determined whether flubendazole, topotecan, and fulvestrant affect proliferation during regeneration. After neomycin exposure, fish were allowed to recover for 24 hrs in EM containing 5 μ M BrdU and the inhibitor. Previous studies indicate that the majority of support cell proliferation begins soon after exposure to neomycin and the majority of new hair cells arise from mitotic events within 24 hrs of neomycin exposure (Harris et al., 2003; Ma et al., 2008). After the 24 hr incubation period, we assessed the number of GFP+ cells, GFP+/BrdU+ cells and GFP-/BrdU+ cells in 7 neuromasts of 10 fish per group (Figure 8). GFP-/BrdU+ cells were presumed to be dividing support cells. Hair cell counts in the absence of neomycin-induced regeneration were not affected by 5 μ M flubendazole or fulvestrant, but significantly diminished with 10 μ M topotecan (t -test; $p < 0.001$; Figure 8B, compare white bars between no neo groups with and without inhibitor). Notably, some GFP- cells incorporated BrdU in and around the neuromast of mock-treated controls, indicating that ongoing cell division is occurring in the absence of damage (Figure 8, black bars in no neo groups without inhibitor). However, there is a 6-fold decrease in the number of GFP-/BrdU+ cells with either flubendazole or topotecan, showing that both drugs dramatically inhibit division of at least a subset of the support cells in the absence of damage to hair cells (t -test; $p < 0.0001$; compare black bars in no neo groups with and without inhibitor). Samples treated with fulvestrant without prior neomycin-induced damage showed no significant change in the number of GFP-/BrdU+ cells when compared to controls.

In fish treated with neomycin and allowed to recover in vehicle only (DMSO or embryo media) for 24 hrs, we see many newly formed GFP+ hair cells (Figure 8, white bars between

neo groups). Of these cells, the majority are BrdU+, indicating they arose from proliferating support cells (Figure 8, compare white bars to striped bars). In flubendazole and topotecan-treated groups, fewer hair cells arise and virtually none of these hair cells are labeled with BrdU. The number of these GFP+/BrdU- hair cells is consistent with the number of immature, neomycin-insensitive hair cells expected (Murakami et al., 2003; Santos et al., 2006). Notably, there is a substantial decrease in GFP-/BrdU+ support cells. We do not observe a repopulation of GFP+ hair cells that are BrdU- in the presence of these mitotic inhibitors after neomycin treatment.

In fulvestrant-treated groups, most of the GFP+ hair cells that regenerated after 24 hrs are also BrdU+, similar to control groups (Figure 8C, compare white bars to striped bars in neo groups). There is a slight decrease in support cell proliferation, seen as fewer GFP-/BrdU+ cells (*t*-test; *p* < 0.01). Thus, the mild decrease or delay in hair cell regeneration in larvae treated with fulvestrant may be attributed to a moderate decrease in support cell division. The effects of fulvestrant on hair cell numbers may not occur until beyond the 24 hr time-point.

To assess whether the inhibitors are overtly toxic to non-dividing support cells within the neuromast, total cell counts were performed for each neuromast. Fish were exposed to inhibitor drugs for 48 hrs without prior neomycin treatment and subsequently labeled using the SYTOX Green nucleic acid stain. Examination with high magnification confocal microscopy revealed no obvious DNA condensation or fragmentation within cells of the neuromast in any treatment or control group, nor was there any evidence of large gaps of missing cells. In particular, groups that were treated with inhibitor drugs in the absence of initial neomycin exposure did not exhibit significantly fewer cells than their mock-treated vehicle controls (data not shown). This suggests that topotecan, flubendazole and fulvestrant do not cause death of non-dividing support cells either alone at the concentrations used or during the 48 hours after neomycin treatment.

Recovery from inhibition

While flubendazole and topotecan appear to prevent hair cell regeneration by affecting the mitotically active precursor cells destined to repopulate the neuromast with hair cells, it is not clear whether these drugs are halting cells in the cell cycle or actually killing precursors. To determine whether precursors can recover from inhibitor treatment, we exposed fish to inhibitor drugs for 24 hrs following neomycin treatment. We then rinsed the fish thoroughly and allowed recovery for up to an additional 72 hrs. Hair cells counts were performed at 0, 24, 48, 72 and 96 hrs after neomycin treatment (Figure 9). Control larvae exhibit robust regeneration of hair cells with numbers that return to pre-treatment levels between 72 and 96 hrs post-treatment. Incubation with flubendazole for 24 hrs initially blocks hair cell production, but robust regeneration resumes after inhibitor washout at a rate similar to control fish, though delayed (Figure 9A). By contrast, topotecan has lasting effects on regeneration (Figure 9B). As expected, no new hair cells were produced in fish treated with topotecan at 24 hrs after neomycin treatment. However after topotecan was removed, hair cell regeneration was greatly attenuated, even 72 hrs after washout of the inhibitor (Figure 9B, compare black and white triangles). Interestingly, treatment with topotecan alone for 24 hrs resulted in delayed loss of hair cells (Figure 9B, white squares), suggesting that topotecan may kill maturing hair cells. It has yet to be determined whether topotecan kills dividing cells, newly formed hair cells or both.

Flubendazole arrests dividing cells in M-phase

Previous literature suggests that flubendazole may inhibit microtubule assembly and function, resulting in cell cycle arrest (Cumino et al., 2009; Spagnuolo et al., 2010). To test

this idea, fish from control and neomycin-treated groups were exposed to flubendazole for 24 hrs and then immunolabeled with phospho-Histone H3 antibody (hereafter PH3), a marker of cells in mitosis. While cells labeled with PH3 were rare in control groups, we found a notable increase in the number of cells in M-phase with flubendazole (*t*-test; $p < 0.0001$; Figure 10A). Fish exposed to flubendazole in the absence of neomycin-induced hair cell death also exhibited significantly more cells labeled with PH3 (*t*-test; $p < 0.0001$), presumably halting those cells dividing in the process of normal neuromast growth. It should be noted that this effect of flubendazole was not limited to the neuromasts, as there were more PH3-labeled cells along the entire fish body after flubendazole treatment.

If flubendazole blocks hair cell regeneration primarily through cell cycle arrest, we hypothesized that it would only effectively block regeneration if larvae were treated during the peak of proliferation. Characterization of proliferative events during zebrafish hair cell regeneration has revealed a transient increase in support cells entering S-phase, peaking between 12–21 hrs after neomycin-induced hair cell death, and a peak in M-phase cells occurring approximately 3–6 hrs later (Harris et al., 2003; Ma et al., 2008). We therefore exposed groups of neomycin-treated fish with flubendazole or DMSO vehicle alone for either 0–14, 14–28 or 28–42 hrs after neomycin treatment. All larvae were collected 48 hrs after the exposure to neomycin and immunolabeled for hair cells. As shown in Figure 10B, hair cell numbers were reduced only in fish treated with flubendazole between 14–28 hrs after neomycin treatment (*t*-test; $p < 0.0001$), further supporting the idea that flubendazole delays hair cell regeneration by arresting cells in M-phase.

DISCUSSION

We used the inherent regenerative capacity of zebrafish lateral line hair cells to develop an efficient assay to screen through drug libraries for novel modulators of hair cell regeneration. While others have surveyed selected compounds that have potential to influence proliferation in mammalian vestibular epithelia (Montcouquiol and Corwin, 2001), this study represents the first broad chemical screen directed towards identifying compounds that alter hair cell regeneration. Two drug libraries composed of 1,680 compounds yielded 2 enhancers and 6 inhibitors of regeneration, a 0.48% modulator compound “hit rate.” Additionally, we identified 34 chemicals that were ototoxic over a 48 hr incubation period (Table 1). Drugs were tested at concentrations between 1–35 μ M. Our experience has shown that there is little toxicity at these concentrations (Ou et al., 2009). We anticipate that there are false negatives among the drugs tested, that we overlooked compounds that would have altered hair cell regeneration at higher concentrations and that compounds originally identified as lethal or toxic to hair cells may have modulated regeneration at lower concentrations.

We identified two compounds that enhanced regeneration. Dexamethasone and prednisolone are both synthetic glucocorticoids that regulate several physiological processes, including the immune response. Both drugs are thought to act primarily as anti-inflammatory agents, inhibiting macrophage activation and cytokine production (e.g., Isobe and Lillehoj, 1992; Hubner et al., 1996; Rhen and Cidlowski, 2005; Schaaf et al., 2009). Alteration of immune response might underlie their effects on hair cell regeneration. During tissue damage, leukocytes are recruited to sites of injury and play an important part in tissue repair. In birds, the resident population of leukocytes in the inner ear sensory epithelia increases after trauma, prior to the proliferation of hair cell progenitors (Warchol, 1997, 1999; Bhave et al., 1998; Warchol et al., 2001; Oesterle et al., 2003; O’Halloran and Oesterle, 2004). Secretion of TGF- α and TNF- α by macrophages has been implicated in promoting the proliferation of avian utricular supporting cells after damage (Warchol et al., 2001). In contrast to our findings, the use of glucocorticoids was shown to reduce regenerative

proliferation after hair cell lesions in birds (Warchol, 1999). In zebrafish fin regeneration is severely inhibited by glucocorticoid receptor (GR) activation and expression of GR target genes (Mathew et al., 2007).

Our data suggest that dexamethasone and prednisolone may enhance hair cell regeneration in zebrafish by mechanisms other than immunosuppression. We observed that glucocorticoids promote an increase in hair cell numbers in the absence of neomycin exposure, demonstrating that they also affect hair cell addition independent of damage. Furthermore, other classes of anti-inflammatory compounds in our drug libraries, including non-steroidal anti-inflammatory drugs, had no effect on hair cell regeneration. Dexamethasone and prednisolone may instead be acting on hair cell precursors directly. Since we see only a modest increase in hair cell numbers, it is possible that they act on a subset of precursors that are “primed” to divide. Alternatively, glucocorticoids may interfere with cessation of the regeneration process. While it is possible that glucocorticoids may act on immune cells to maintain hair cell numbers as a normal part of regeneration, this has not been observed in time-lapse imaging of hair cell regeneration after neomycin exposure (Wibowo et al., 2011; Lopez-Schier and Hudspeth, 2006). Nevertheless, our data does not directly address the issue of immunosuppression. Whether the effects of dexamethasone and prednisolone on hair cells are due to GR activation is also unknown. Further studies will be needed to carefully explore these possibilities.

We find it surprising that so few enhancers of regeneration were identified in our screen. To date, the gamma-secretase inhibitor DAPT has been the only drug in zebrafish that is capable of producing excessively large increases in regenerated hair cells. DAPT works by interfering with Notch signaling, a pathway found to be important in regulating the number of hair cells regenerated in both fish and birds (Stone and Rubel, 1999; Lanford et al., 1999; Ma et al., 2008; Brigande and Heller, 2009; Daudet et al., 2009). The relative paucity of regeneration enhancers may reflect the composition of the libraries screened; interrogating a more diverse set of small molecules might reveal new compounds capable of promoting regeneration.

The inhibitors of hair cell regeneration identified from the primary and secondary screens are diverse in structure and function. We further characterized the inhibitors flubendazole, topotecan and fulvestrant. Flubendazole and topotecan produced the strongest inhibitory profile. Fulvestrant was selected as an example of one of the moderate inhibitors because of the drug’s previously established pharmacological profile (Osborne et al., 2004).

Flubendazole is a benzimidazole antihelmintic that acts by binding to the colchicine-sensitive site of tubulin, thus inhibiting its polymerization or assembly into microtubules (Cumino et al., 2009; Spagnuolo et al., 2010). Flubendazole treatment of zebrafish larvae appears to arrest cells in M-phase, as indicated by increased staining with the PH3 antibody. When flubendazole is removed, hair cell regeneration resumes, suggesting that its effects on microtubules are reversible. Since flubendazole reversibly arrests cell division, it appears that the signals controlling the hair cell regeneration process are maintained and promote full recovery after the delay. Flubendazole is very similar in structure to fenbendazole and albendazole, two drugs identified as ototoxic in our screen of the NINDS Custom Collection II library. It is interesting that subtle changes in structure or side-groups produce markedly different effects. The other strong inhibitor, topotecan blocks regeneration almost completely, and repopulation of the neuromasts remains severely depressed for the entire period studied. Topotecan is a topoisomerase I inhibitor and is most commonly used as a chemotherapeutic agent. This drug is thought to act on tumor cells by stabilizing the covalent complex of topoisomerase I enzyme and strand-cleaved DNA, eventually inducing cell death (Kollmannsberger et al., 1999; Staker et al., 2002; Koster et al., 2007; Lorusso et

al., 2010). This suggests that topotecan may act by killing the dividing support cells responsible for repopulating hair cells. Several authors have suggested that while a subset of support cells divide to give rise to new hair cells, another subset divide to replenish the depleted hair cell precursors (Jones and Corwin, 1996; Lopez-Schier and Hudspeth, 2006; Ma et al., 2008). The delay in hair cell regeneration after topotecan is removed may reflect the time needed to restore the hair cell precursor population eliminated by topotecan treatment. It is also possible that topotecan does not wash out of cells readily, resulting in the observed delay.

Both flubendazole and topotecan suppress regeneration by inhibiting support cell division. This observation is in agreement with other studies suggesting that hair cell regeneration in the zebrafish lateral line is accomplished solely through mitotic replacement of lost hair cells (Harris et al., 2003; Lopez-Schier and Hudspeth, 2006; Ma et al., 2008; Wibowo et al., 2011). This is in sharp contrast to regeneration in the inner ear of birds and amphibians, where substantial transdifferentiation normally occurs and is accentuated by blocking the mitotic response (Adler and Raphael, 1996; Baird et al., 1996; Roberson et al., 1996; Baird et al., 2000; Roberson et al., 2004; Taylor and Forge, 2005).

We observe 1–3 GFP+/BrdU– hair cells within neuromasts of fish treated with flubendazole after neomycin treatment. These hair cells are likely to have been immature hair cells not susceptible to neomycin-induced death at the time of treatment (Santos et al., 2006). Nevertheless, we currently lack markers to distinguish between support cells and immature post-mitotic hair cells, thus we cannot rule out the possibility that some direct transdifferentiation occurs in zebrafish after neomycin treatment.

The more moderate inhibitor of regeneration, fulvestrant, is an estrogen receptor (ER) antagonist. Fulvestrant is thought to act by down-regulating and degrading estrogen and progesterone receptors (Wakeling et al., 1991; Howell et al., 2000; Osborne et al., 2004). Support cell divisions in the neuromast are reduced in the presence of fulvestrant, suggesting that the drug may be affecting the signaling pathways involved in cellular proliferation. It is unknown whether fulvestrant acts on ER signaling in hair cells and if so, which ERs are affected. Estrogen signaling has been implicated in proliferative signaling during avian hair cell regeneration (McCullar and Oesterle, 2009) and a microarray study detected significant up-regulation of estrogen receptors after either aminoglycoside antibiotic or laser damage of chicken auditory and vestibular epithelia (Hawkins et al., 2007). The *esr2a* and *esr2b* estrogen receptor transcripts are abundant in both hair cells and support cells within zebrafish lateral line neuromasts during development (Tinguad et al., 2004), and blocking *esr2a* expression prevents hair cell development, without affecting support cells (Froehlicher et al., 2009). Hence, fulvestrant may be suppressing hair cell regeneration by interfering with ER signaling.

The predominant cause of hearing loss in humans is the death of the mechanosensory hair cells of the inner ear. In 1987, it was discovered that birds regenerate damaged and lost inner ear hair cells (Cotanche, 1987; Cruz et al., 1987). Since that time considerable progress has been made (Collado et al., 2008; Lowenheim et al., 2008; Brigande and Heller, 2009; Groves, 2010), but robust regeneration is yet to be achieved in the inner ear of mature mammals. Our results demonstrate that the zebrafish lateral line system provides a platform for high throughput screening to identify new modulators of hair cell regeneration. Extension of the methods described here to new compound libraries may provide additional tools to dissect regeneration in systems that can normally undergo this process and perhaps promote regeneration in those that do not.

Acknowledgments

This work was supported by Action on Hearing Loss, and NIDCD Grants DC04661 and DC011269.

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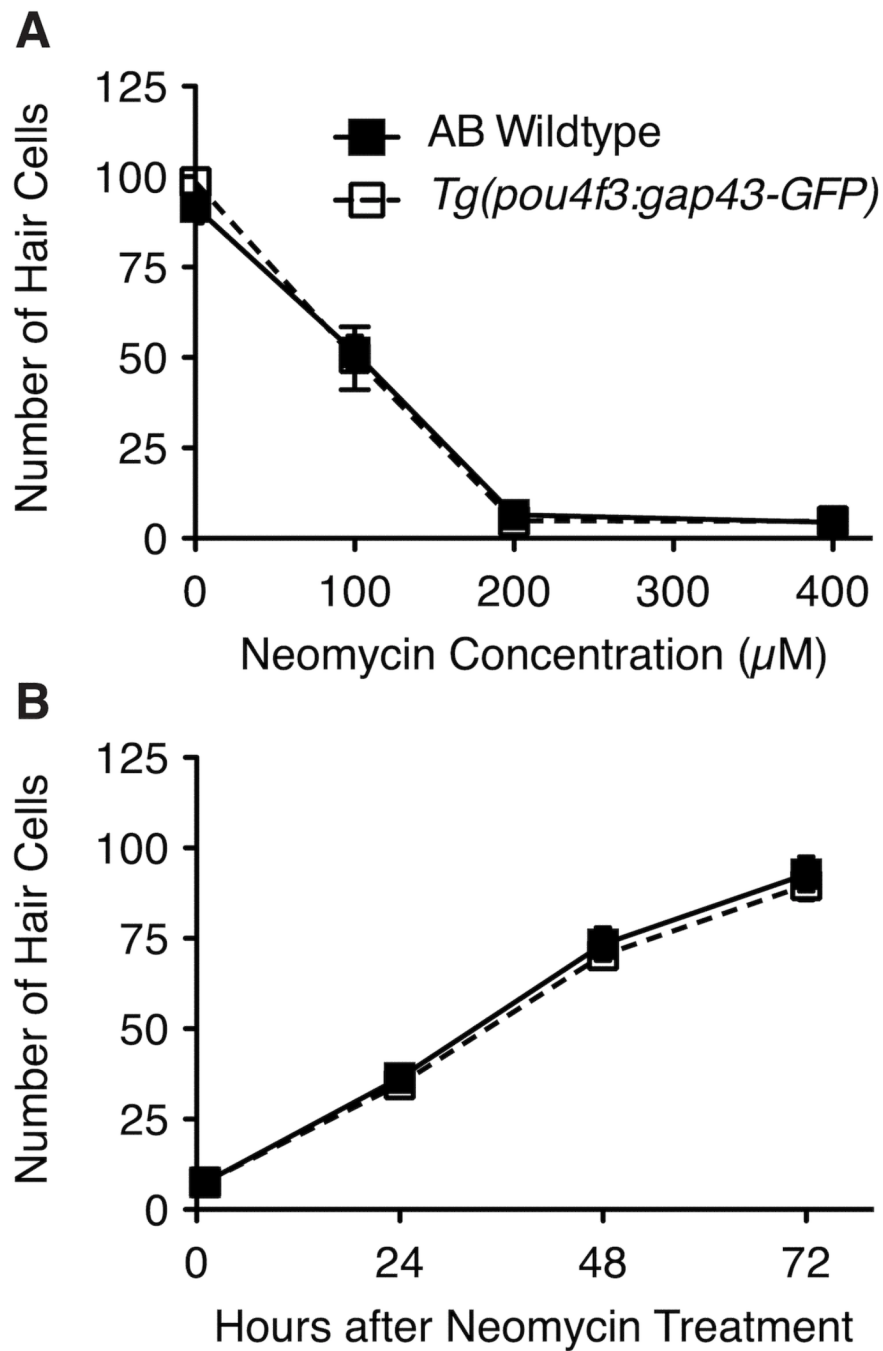


Figure 1. Chemical screening protocol for modulators of hair cell regeneration. **A.** Schematic diagramming the regeneration screen. 5–6 dpf *Tg(pou4f3:gap43-GFP)* zebrafish expressing membrane-bound green fluorescent protein under the *pou4f3* promoter were treated with 400 µM neomycin for 1 hr, rinsed and transferred into 48-well plates. Fish were then either treated for 48 hrs with a test drug, DMSO vehicle, or DAPT (a positive control for enhanced hair cell regeneration) and subsequently scored for GFP+ hair cells on a 1–5 scale, with a score of 3 representing normal regeneration. **B.** Fluorescent images of the MII neuromasts illustrate the scoring scale used in the screen for regeneration modulators. Examples from

Tg(pou4f3:gap43-GFP) larval zebrafish that would have received a score of 1, 3 and 5 are shown. Scale = 10 μ m.

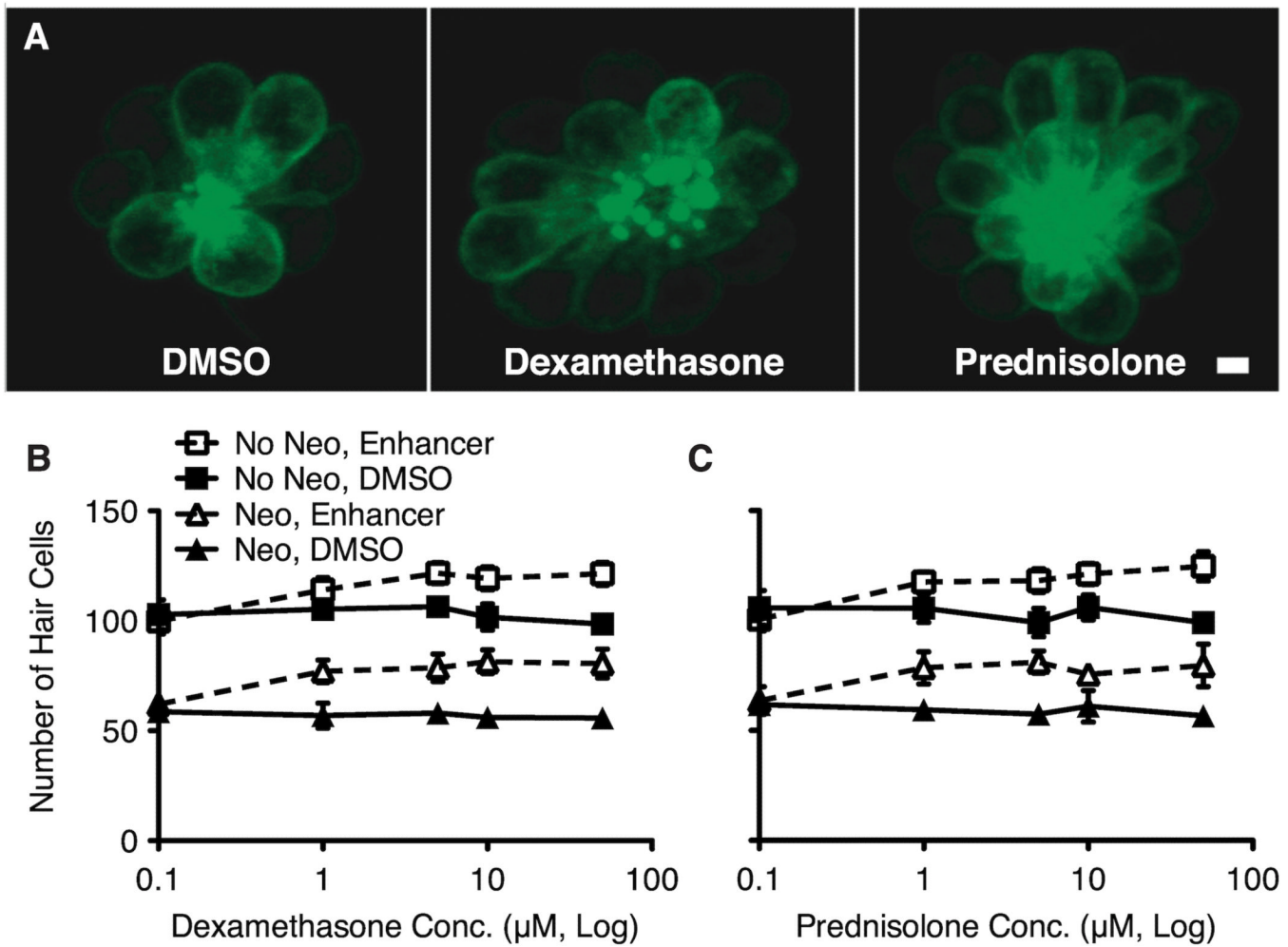


Figure 2.

A. Hair cell death in *Tg(pou4f3:gap43-GFP)* fish is similar to that of *AB wildtype fish. 5 dpf zebrafish larvae were treated with 0, 100, 200 or 400 μM neomycin for 1 hr. Graph displays the mean total number of parvalbumin-labeled hair cells in 7 neuromasts from each fish (n = 10 fish/group). **B.** Hair cell regeneration in *Tg(pou4f3:gap43-GFP)* fish is similar to that of *AB wildtype fish. 5 dpf zebrafish larvae were treated with 400 μM neomycin for 1 hr and then held in EM for 0, 24, 48 or 72 hrs. Graph displays the mean total number of parvalbumin-labeled hair cells in 7 neuromasts of 10 fish for each group. Error bars indicate \pm 1 SD.

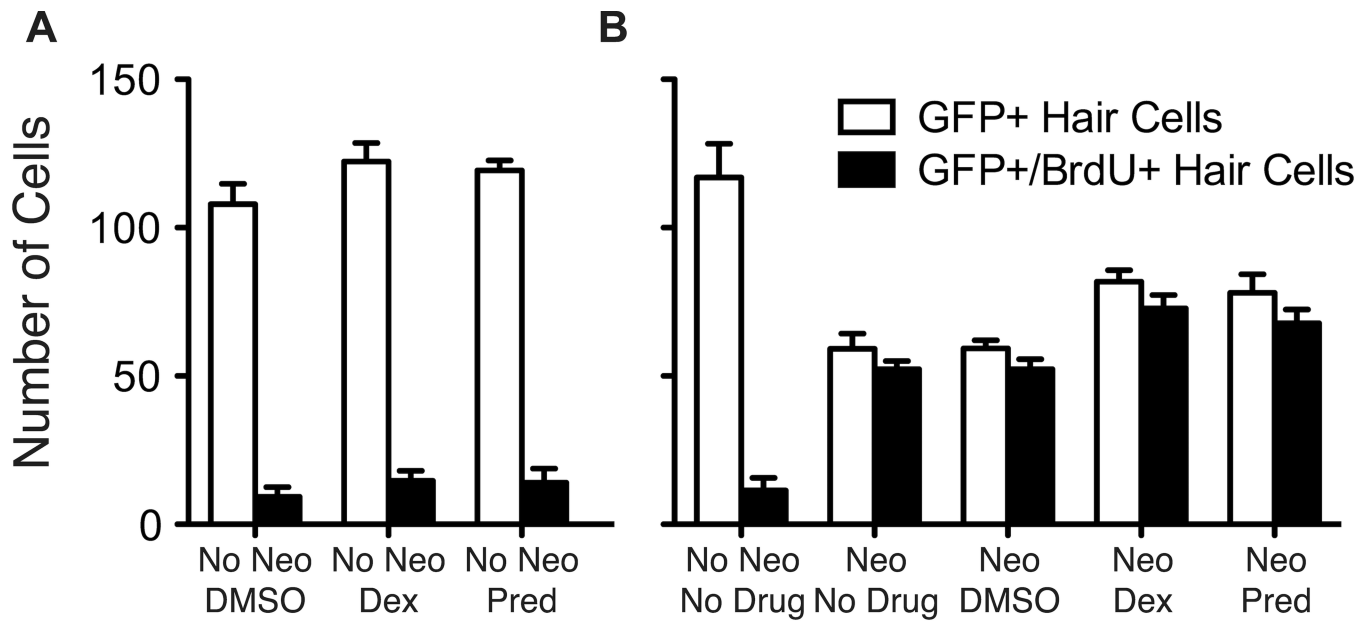


Figure 3.

The glucocorticoids dexamethasone and prednisolone enhance hair cell regeneration. **A.** Fluorescent confocal images of GFP+ hair cells treated continuously for 48 hrs with DMSO vehicle (left), 5 μ M dexamethasone (middle) or 5 μ M prednisolone (right), after 400 μ M neomycin treatment for 1 hr. Scale = 2.5 μ m. **B, C.** An increase in total number of hair cells occurs following exposure to either dexamethasone (**B**) or prednisolone (**C**). Graphs indicate the mean total number of GFP+ hair cells among seven neuromasts from 10 fish after neomycin damage (triangles) or mock treatment (squares) and subsequent 48 hr exposure to dexamethasone or prednisolone (denoted by open symbols and dotted lines in comparison to DMSO vehicle-only denoted with filled symbols and solid lines). Analysis by 2-way ANOVA showed significant main effects of drug, concentration and an interaction for dexamethasone and prednisolone as compared to DMSO control fish in both neomycin and mock-treated groups. Error bars indicate +/- 1 SD.

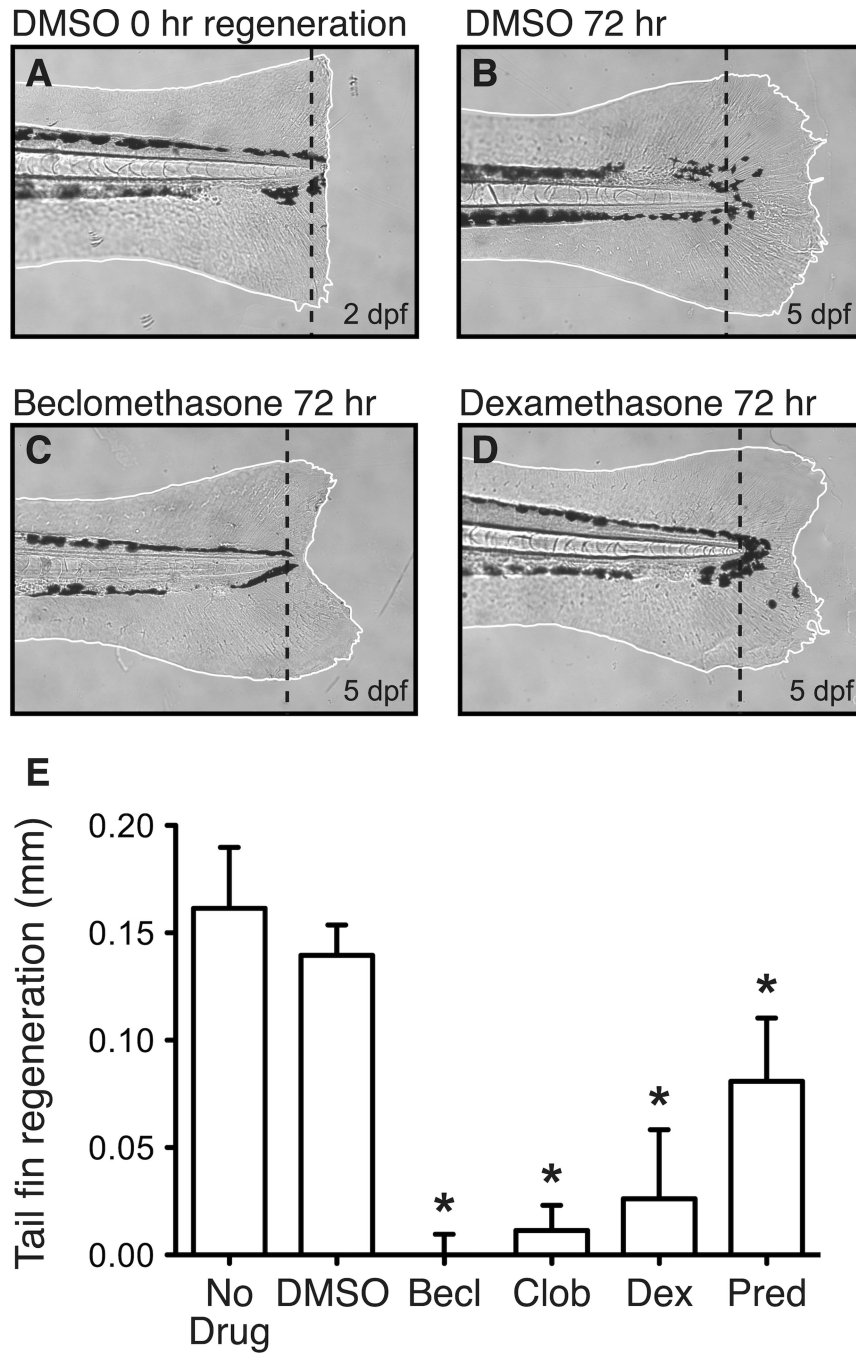


Figure 4. Dexamethasone and prednisolone potentiate support cell proliferation. **A.** In fish co-treated with 5 μ M dexamethasone (Dex) or prednisolone (Pred) and BrdU without prior neomycin treatment (No Neo), there is an increase in the number of GFP-labeled cells (p 's < 0.0001) and a comparable increase in the number of GFP+/BrdU+ hair cells (p 's < 0.01), suggesting that the increase in hair cells arose from support cell proliferation and differentiation of new hair cells. **B.** In larvae treated with 400 μ M neomycin (Neo) and then allowed to recover in 5 μ M dexamethasone or prednisolone for 48 hrs in the presence of BrdU, there was a significant increase of GFP-labeled hair cells (p 's < 0.0001) and a comparable increase in BrdU-labeled hair cells (p 's < 0.0001) for dexamethasone and prednisolone-treated groups

compared to DMSO vehicle controls. Means of total hair cells in 7 neuromasts per fish (n = 10 fish/group) are presented. Error bars indicate 1 SD.

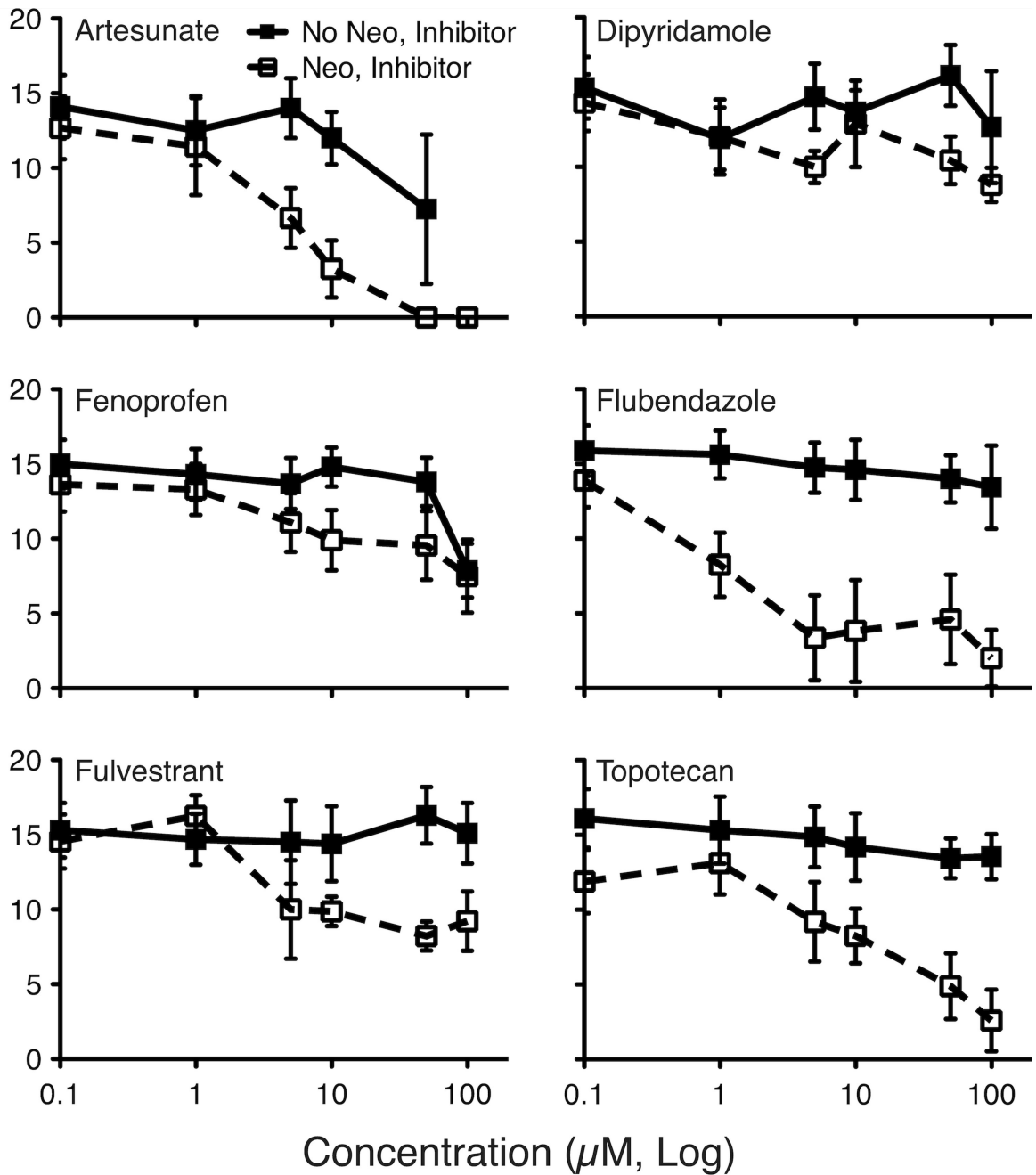


Figure 5.

Dexamethasone and prednisolone suppress fin regeneration in zebrafish. Caudal fins of 2 dpf zebrafish larvae were amputated and then exposed to glucocorticoids or left untreated in EM (or EM + DMSO) for 72 hrs. **A.** Brightfield image displaying the caudal fin of a fish immediately after amputation. **B–D.** Brightfield images of caudal fin 72 hrs post-amputation when exposed to DMSO only, 5 µM beclomethasone or 5 µM dexamethasone in DMSO. **E.** All four glucocorticoids, each tested at a 5 µM concentration, significantly inhibit fin regeneration compared to vehicle controls (*t*-test; Becl: $p < 0.0001$; Clob: $p < 0.0001$; Dex: $p < 0.0001$; Pred: $p = 0.0014$). Five fins were examined per condition. Bars indicate 1 SD.

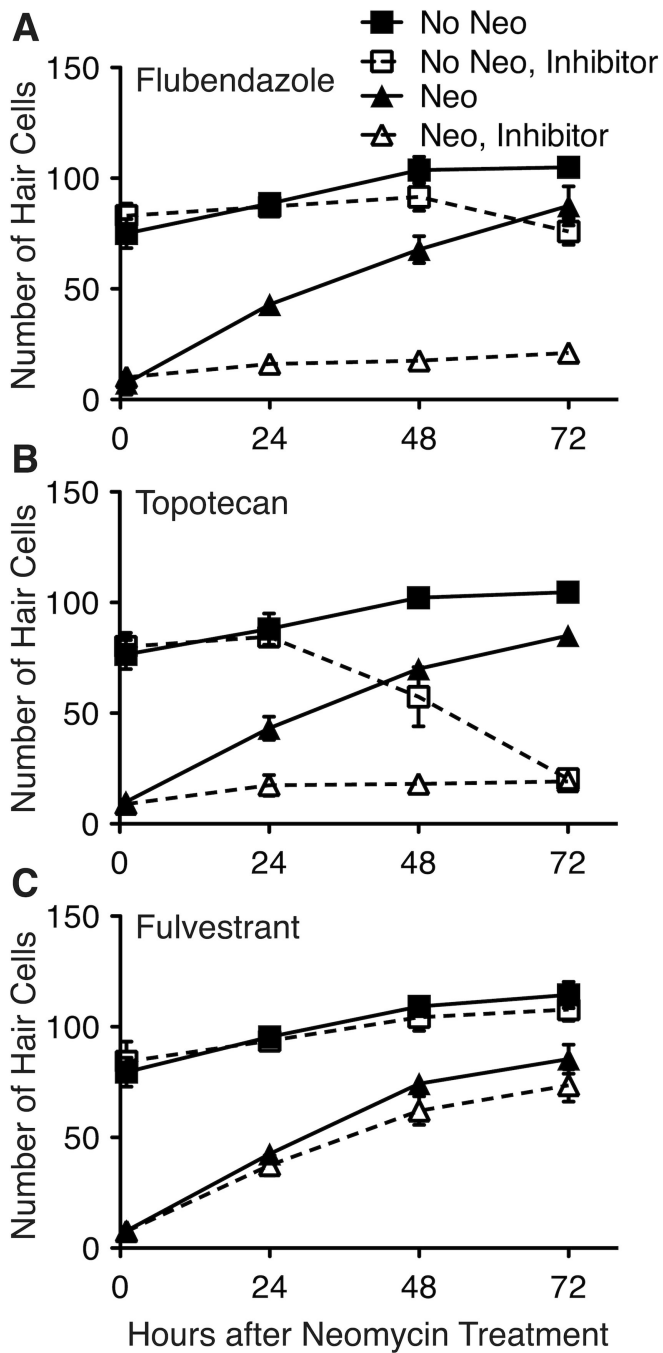


Figure 6. Dose-response functions for six confirmed regeneration inhibitors. Fish were acutely exposed to 400 μ M neomycin (Neo) in embryo media for 1 hr (or mock treated; No Neo), and then incubated in 0, 0.1, 1, 5, 10, 50 and 100 μ M of the noted inhibitor drug in EM for 48 hrs during the regeneration period. Hair cell regeneration was assessed using the DASPEI scoring system. For each fish ($n = 10-12$ fish/group), 10 neuromasts were scored, and total DASPEI score was recorded. Means \pm 1 SD are shown. All six inhibitor drugs show significant suppression of hair cell regeneration over samples that received no neomycin treatment (p 's < 0.01). For both artesunate and fenoprofen, significant drug toxicity was noted in the mock-treated controls at high concentrations.

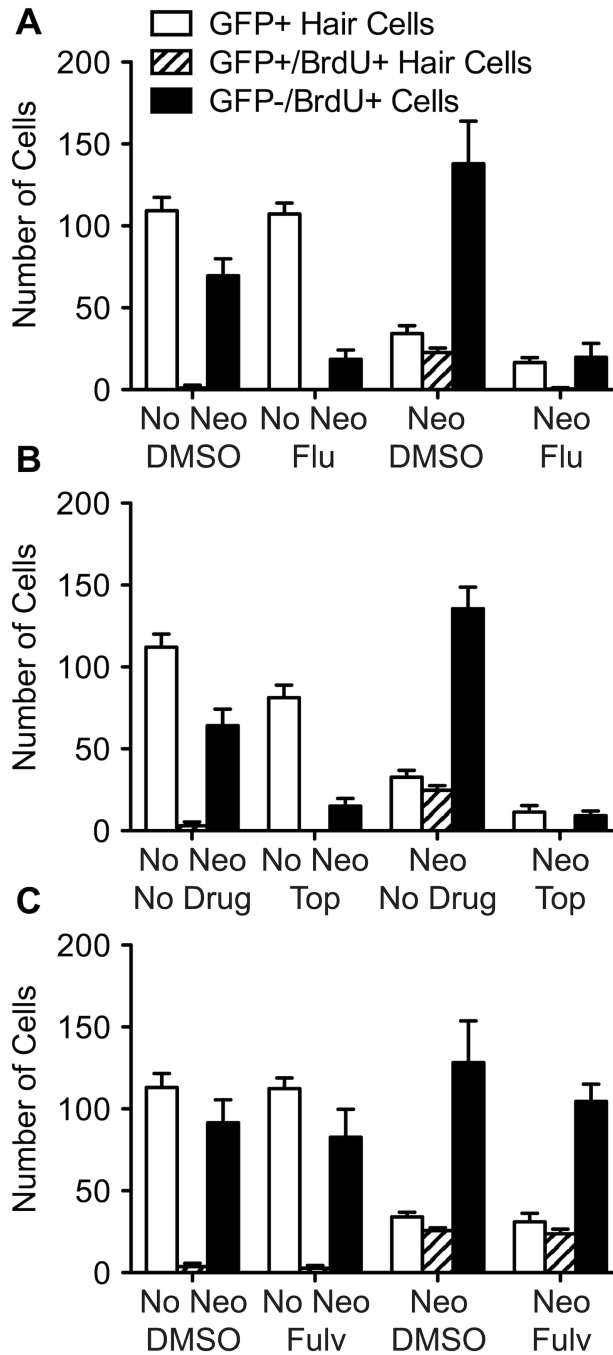


Figure 7. Hair cell regeneration is differentially suppressed during 72 hr inhibitor exposure. Fish were treated with 400 μ M neomycin for 1 hr (triangles) or mock treated (squares), and allowed to recover in each inhibitor for 72 hrs (open symbols, dotted lines) or in vehicle (filled symbols, solid lines). GFP+ hair cell counts were performed at 1, 24, 48 and 72 hrs following treatment. **A.** Fish treated with neomycin and subsequently with 5 μ M flubendazole (open triangles) displayed essentially no hair cell regeneration, while controls (filled triangles) showed complete recovery by 72 hrs. Flubendazole alone (open squares) produce a small (and significant, $p < 0.001$) decrease in total hair cell number at 72 hrs (filled squares). **B.** Hair cell regeneration in fish treated with neomycin and then exposed to

10 μ M topotecan (open triangles) is also virtually totally suppressed relative to controls (filled triangles). In addition, treatment with topotecan in control fish causes toxicity at 48 hrs and beyond (open squares vs. filled squares). **C.** 5 μ M fluvestrant (open triangles) modestly reduced the number of regenerated hair cells versus controls (fills triangles) following neomycin pretreatment ($p < 0.01$ at 48 and 72 hrs post-neomycin treatment). Fish treated with fulvestrant alone (open squares) did not show any significant hair cell loss relative to mock-treated controls (filled squares). For each group, 7 neuromasts were examined ($n = 10$ fish). Error bars indicate ± 1 SD.

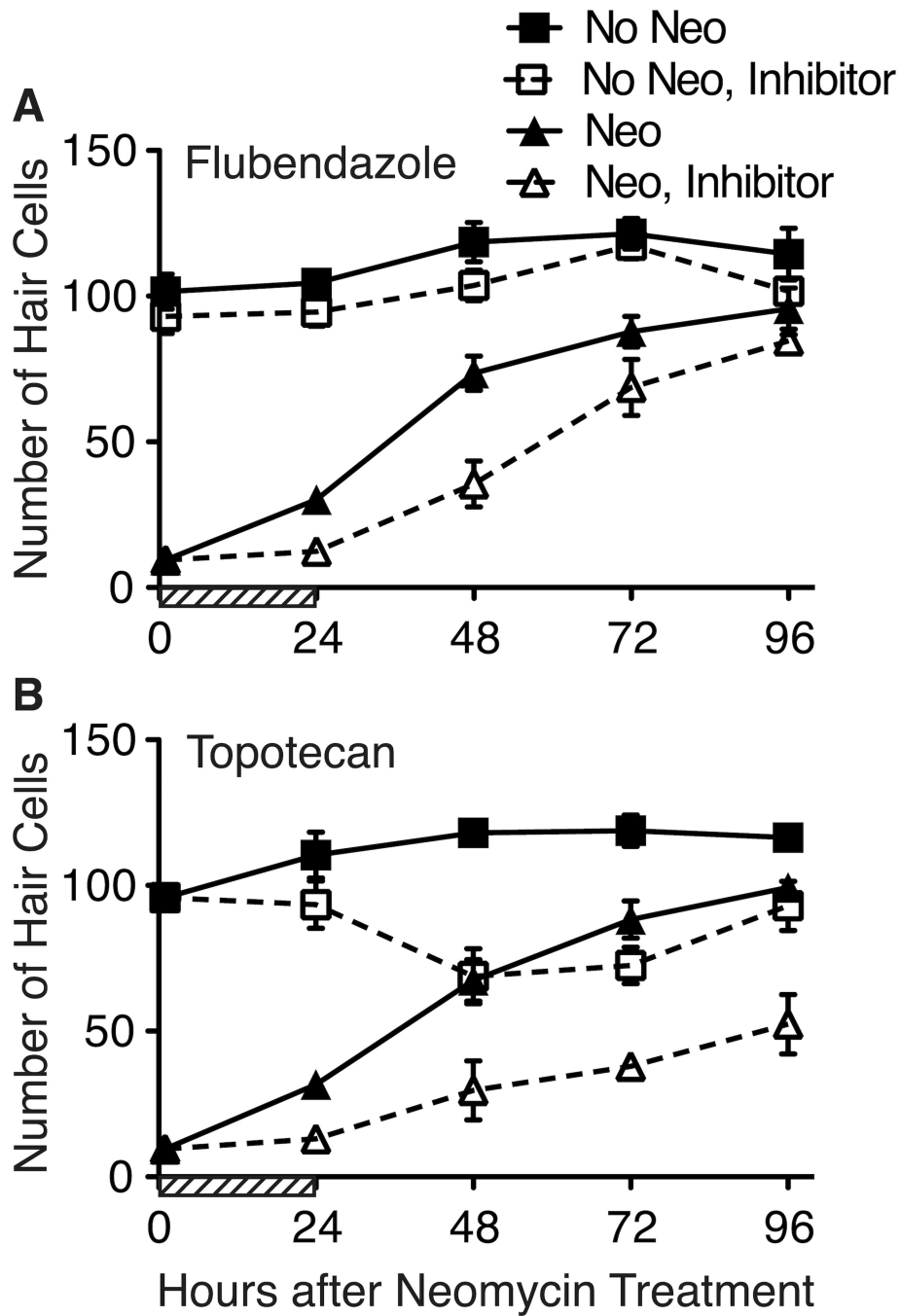


Figure 8. Inhibitor compounds prevent support cell division. Fish were treated with 400 μ M neomycin (or mock-treated) for 1 hr, and then exposed to 5 μ M BrdU plus inhibitor (5 μ M flubendazole, 10 μ M topotecan, or 5 μ M fulvestrant) for 24 hrs. At 24 hrs post-neomycin treatment, animals were fixed and immunolabeled for hair cells and BrdU. **A.** Fish mock-treated (No Neo) and then incubated in DMSO have virtually no GFP⁺/BrdU⁺ hair cells, as is observed in fish mock-treated and exposed to flubendazole. The number of GFP⁻/BrdU⁺ cells is greatly diminished between these groups suggesting that flubendazole is suppressing cell division in the absence of hair cell death. In fish treated with neomycin (Neo) and DMSO, significant regeneration is observed after 24 hrs (open bar), most of which are co-

labeled with BrdU (striped bar). GFP⁻/BrdU⁺ cell counts are substantially elevated (black bar), due to the fact that support cell division is occurring at a rapid pace to replace lost hair cells. In fish treated with neomycin and subsequently with flubendazole (Flu), significantly fewer hair cells are produced, and of those hair cells visible, almost none are labeled with BrdU. The number of BrdU-labeled support cells is dramatically reduced as well. **B.** An almost identical pattern of GFP and BrdU labeling is observed in samples treated with topotecan (Top) as seen above. GFP⁻/BrdU⁺ labeling is dramatically reduced in fish treated with topotecan, regardless of neomycin treatment, indicating that the drug is suppressing all cell divisions. After neomycin treatment and subsequent incubation in topotecan for 24 hrs, there are significantly fewer hair cells regenerated, and of those few hair cells, none are co-labeled with BrdU. In mock-treated fish (No Neo), there is a noticeable decrease in the number of hair cells in fish exposed to topotecan, consistent with the idea that topotecan also kills mature hair cells. **C.** Mock-treated fish (No Neo) that were incubated in 5 μ M fulvestrant (Fulv) display an essentially identical pattern of GFP and BrdU labeling to mock-treated fish incubated in DMSO. There is no significant GFP⁺ hair cell or GFP⁻/BrdU⁺ support cell loss between these groups. In fish treated with neomycin and later incubated in fulvestrant, GFP⁺ hair cell and GFP⁺/BrdU⁺ hair cell counts are similar to controls. However, GFP⁻/BrdU⁺ cell counts in fulvestrant-treated groups were slightly diminished ($p < 0.01$) compared to DMSO controls. For each fish, 7 neuromasts were examined ($n = 10$ fish/group). Error bars indicate ± 1 SD.

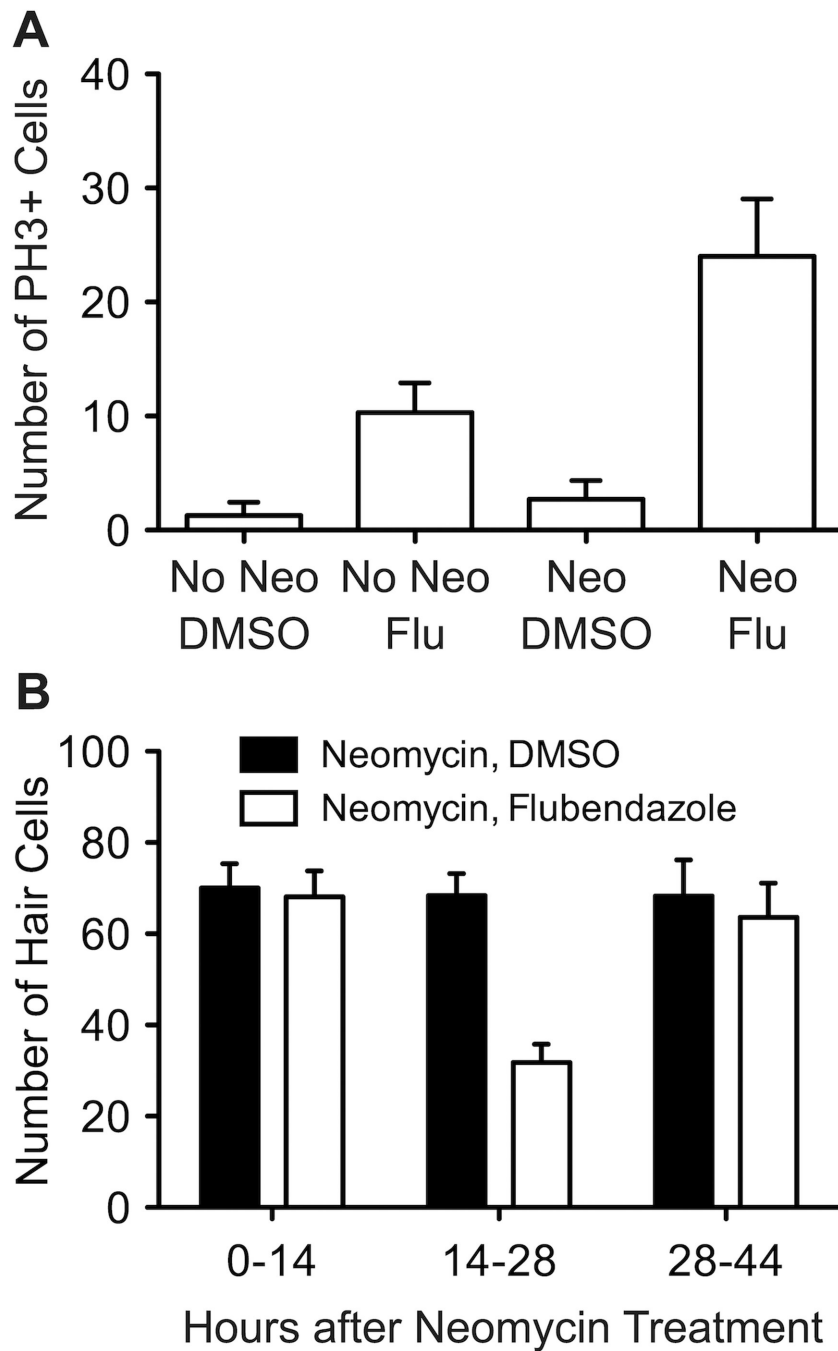


Figure 9. Flubendazole and topotecan arrest, but do not eliminate, hair cell regeneration. Fish were treated with neomycin, exposed to flubendazole or topotecan for 24 hrs (or mock-treated with DMSO; cross-hatched area on abscissa), and then placed in fresh embryo media for an additional 72 hrs. **A.** Hair cell numbers in fish treated with flubendazole remain low while flubendazole is present (through 24 hrs). When flubendazole was removed, hair cell regeneration resumes, and by 96 hrs, hair cell numbers in flubendazole-treated and DMSO-treated controls are not significantly different. **B.** Similar suppression of hair cell regeneration is seen in fish treated with topotecan for 24 hrs. However, when the inhibitor

drug was removed, hair cell regeneration was greatly attenuated. Hair cells were counted in 7 neuromasts per fish (n = 10 fish per group). Error bars indicate \pm 1 SD.

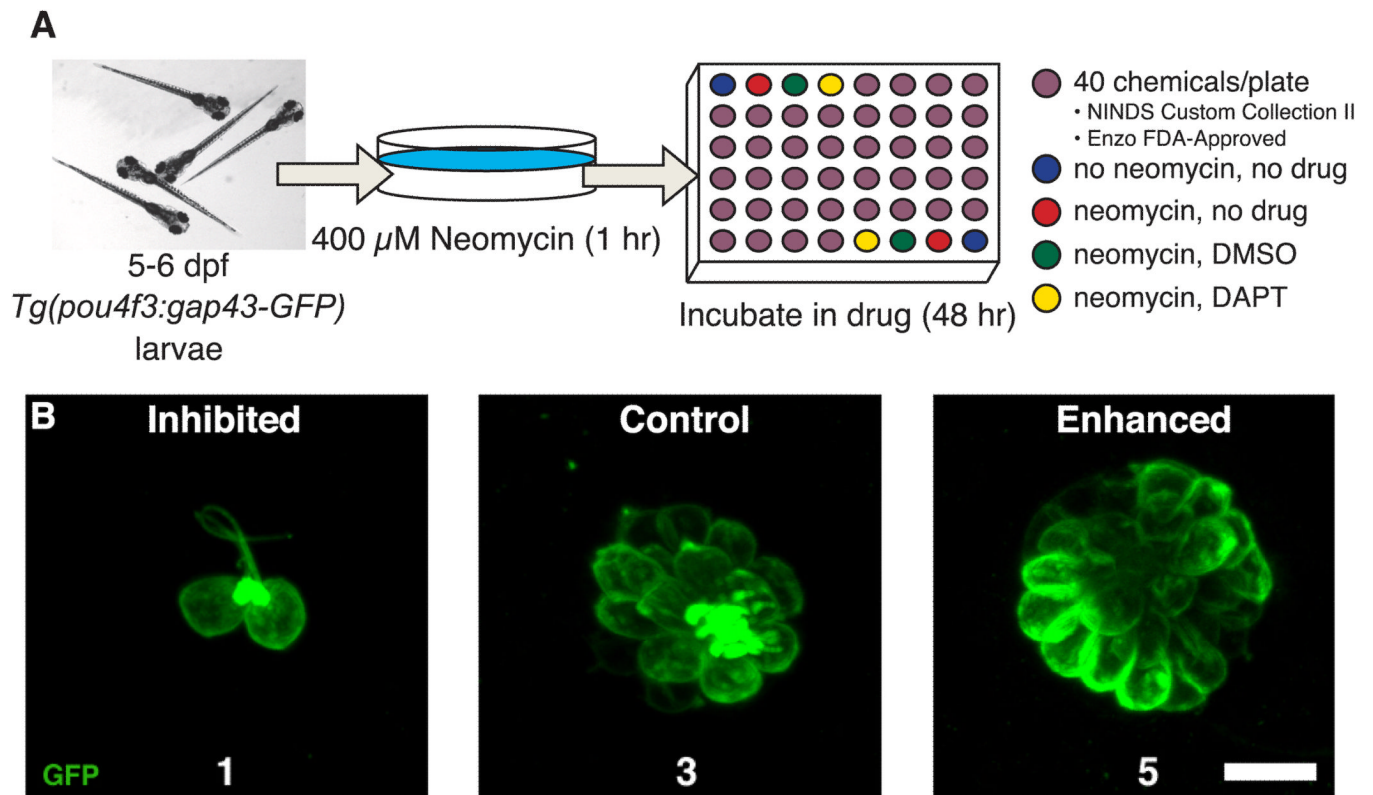


Figure 10.

Flubendazole induces increased number of cells in M-phase. **A.** 5 dpf fish were either treated with 400 μ M neomycin or mock-treated in vehicle for 1 hr, and then exposed to flubendazole for 48 hrs. Fish were immunostained with an antibody to phospho-Histone H3 to assess the number of cells in M-phase at the time of death. Increased numbers of PH3+ cells are present in flubendazole-treated fish, regardless of whether native hair cells were killed by neomycin or not (p 's < 0.001) and more labeled cells were seen in neuromasts after neomycin treatment than controls after flubendazole exposure. **B.** Flubendazole reduces hair cell regeneration primarily between 14–28 hrs post-damage. Fish were treated with 400 μ M neomycin for 1 hr, and then treated with DMSO vehicle or 5 μ M flubendazole for one of three time periods (0–14 hrs, 14–28 hrs or 28–44 hrs post-neomycin). Larvae were then held in EM only for the remainder of time prior to fixation at 48 hrs and immunostained for GFP + hair cells. There is a marked decrease in the number of hair cells when proliferating cells are suspended in M-phase specifically during the 14–28 hr incubation period after neomycin treatment, but not during other periods. For each fish, 7 neuromasts were examined ($n = 10$ fish/group). Error bars indicate ± 1 SD.

Table 1

A matrix of the numerical results from the regeneration screen and subsequent rescreens. Listed are the numbers of putative enhancers and inhibitors identified in both the primary screen and subsequent rescreens.

Numerical results (and percentages) from the regeneration screen and subsequent rescreens			
<i>Library</i>	<i>Initial screen</i>	<i>Rescreen</i>	<i>Confirmed by dose-response analysis</i>
NINDS Custom Collection II			
Number of drugs evaluated	1040	189	5
Number of putative enhancers *	86 (8.3%)	2 (0.2%)	2 (0.2%)
Number of putative inhibitors **	103 (9.9%)	3 (0.3%)	0
BioMol FDA-approved I			
Number of drugs evaluated	640	102	45
Number of putative enhancers *	29 (4.5%)	8 (1.3%)	0
Number of putative inhibitors **	73 (11.4%)	37 (5.8%)	6 (0.9%)

* Putative enhancers average scored ≥ 3.5

** Putative inhibitors average scored ≤ 2.5

Table 2

Confirmed modulatory drugs from our screening protocol that alter hair cell regeneration in zebrafish. Listed are the candidate modulatory drugs, their EC_{50} and optimal drug range, based on dose-response testing, and previously characterized drug activity.

Drugs that modulate hair cell regeneration, identified in screen and confirmed upon retesting				
<i>Candidate drug</i>	<i>Direction of modulation</i>	<i>EC₅₀</i> [*]	<i>Optimal range</i> ^{**}	<i>Drug category; known activity</i>
Artesunate	Inhibitor	5.39	5–10 μ M	Derivative of artemisinin; anti-malarial treatment.
Dexamethasone	Enhancer	0.13	5–10 μ M	Synthetic glucocorticoid; anti-inflammatory/ immunosuppressant; treatment for autoimmune and inflammatory conditions.
Dipyridamole	Inhibitor	1.07	5–50 μ M	Thromboxane synthase inhibitor; inhibits thrombus formation and induces vasodilation.
Fenoprofen	Inhibitor	5.81	10–50 μ M	Non-steroidal anti-inflammatory drug; inhibits cyclooxygenase activity and prostaglandin synthesis; treatment for pain, swelling, and stiffness caused by osteoarthritis.
Flubendazole	Inhibitor	0.65	1–5 μ M	Benzimidazole anthelmintic; treatment of gastrointestinal parasites.
Fulvestrant	Inhibitor	3.25	5–10 μ M	Selective estrogen receptor down-regulator; treatment for hormone receptor-positive metastatic breast cancer.
Prednisolone	Enhancer	0.58	5–10 μ M	Synthetic glucocorticoid; anti-inflammatory/ immunosuppressant; treatment of autoimmune and inflammatory conditions.
Topotecan	Inhibitor	4.93	10–50 μ M	Topoisomerase I inhibitor; treatment of ovarian and lung cancer.

* Calculated from dose-response functions

** Optimal range is the drug dose that confers maximal change in regeneration (inhibition or enhancement) without noticeable toxicity to the fish