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Priming with very low-affinity peptide ligands gives rise to CD8+ T-cell effectors with enhanced function but with greater susceptibility to transforming growth factor (TGF)β-mediated suppression

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Abstract

While the effects of TCR affinity and TGFβ on $CD8^+$ T-cell function have been studied individually, the manner in which TCR affinity dictates susceptibility to TGFβ-mediated suppression remains unknown. To address this issue, we utilized OVA altered peptide ligands (APLs) of different affinities in the OT-I model. We demonstrate that while decreased TCR ligand affinity initially results in weakened responses, such interactions prime the resultant effector cells to respond more strongly to cognate antigen upon secondary exposure. Despite this, responses by $CD8^+$ T cells primed with lower-affinity TCR ligands are more effectively regulated by TGFβ. Susceptibility to TGFβ-mediated suppression is associated with downregulation of RGS3, a recently recognized negative regulator of TGFβ signaling, but not expression of TGFβ receptors I/

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II. These results suggest a novel tolerance mechanism whereby $CD8^+$ T cells are discriminately regulated by TGFβ according to the affinity of the ligand on which they were initially primed. In addition, because of the major role played by TGFβ in tumor-induced immune suppression, these results identify the affinity of the priming ligand as a primary concern in $CD8^+$ T-cell-mediated cancer immunotherapeutic strategies.

Keywords

Tumor-induced suppression; TGFβ; $CD8^+$; T cells; T-cell receptor affinity; RGS3

Introduction

$CD8^+$ T cells emerge from the thymus bearing T-cell receptors (TCRs) with a wide range of affinities. Key to the understanding of TCR affinity and T-cell function has been the development of altered peptide ligands (APLs). Studies using APLs demonstrate high-affinity interactions between the TCR and peptide-MHC class I complexes (pMHC) result in greater induction of $CD8^+$ T-cell responses [1, 2]. However, the manner in which APLs with differential TCR affinity dictate susceptibility to TGFβ-mediated suppression remains unknown.

Transforming growth factor beta (TGFβ) is an immunoregulatory cytokine with activity affecting T-cell proliferation, differentiation, survival, and self-tolerance [3–7]. TGFβ signals through a heterotetrameric complex of TGFβ receptor (TGFβR)-I and TGFβRII, which phosphorylates the receptor-regulated Smad signaling proteins (R-Smads), including Smad2 and Smad3. These R-Smads then complex with the co-Smad, Smad4, which together translocate to the nucleus to activate transcription of certain TGFβ-responsive genes [8, 9]. The inhibitory Smads, including Smad7, act by preventing phosphorylation of the R-Smads, while the noncanonical inhibitor of TGFβ signaling, the regulator of G-protein signaling (RGS)-3, acts by forming complexes with the R-Smads and co-Smad and prevents the activation of TGFβ-induced gene transcription [10]. Mice with T cells that lack the ability to respond to TGFβ rapidly experience multiorgan, multitarget T-cell-mediated autoimmunity without any prior modification of the T-cell repertoire [11–13]. Studies analyzing these mice have demonstrated that self-reactive T cells exist in the natural repertoire and that TGFβ signaling is required to prevent these responses in the normal physiological state. Additional studies have shown that TGFβ-insensitive polyclonal $CD8^+$ T cells possess enhanced antitumor function and can prevent tumors from developing [14]. However, little is known about natural variations in $CD8^+$ T-cell sensitivity to TGFβ signaling.

TCR affinity and TGFβ-mediated suppression have been individually shown to regulate $CD8^+$ T-cell responses. However, the interplay between these variables remains unknown. In this study, we now demonstrate that while decreased TCR ligand affinity initially results in weakened responses, such interactions prime the resultant effector cells to respond more strongly to cognate antigen upon secondary exposure. In spite of this, responses by $CD8^+$ T cells primed with lower-affinity TCR ligands are more effectively suppressed by TGFβ. These results highlight antigen affinity as an important concern in cancer immunotherapy that may not be addressed by vaccination or increasing the density of the presented antigen.

Materials and methods

Cells and mice

All cells were cultured in RPMI supplemented with 10% heat-inactivated fetal bovine serum (Atlanta Biologicals, Lawrenceville, GA), 2 mM L-glutamine (Mediatech, Manassas, VA),

and 1% penicillin/streptomycin (Mediatech, Manassas, VA), unless otherwise noted. Six-week-old, specific-pathogen-free C57BL/6-Tg(TcraTcrb)1100Mjb/J (OT-I) mice were purchased from Jackson Laboratories. All mice were housed at The University of Chicago animal facility under conventional conditions, and animal experimentation was conducted in accordance with Institutional Animal Care and Use Committee (IACUC) guidelines.

Peptides

OVA_{257} (SIINFEKL) and, in order of decreasing reported affinity for the OT-I TCR, the APLs Y3 (SIYNFEKL), Q4 (SIIQFEKL), T4 (SIITFEKL), and V4 (SIIVFEKL) were purchased from New England Peptide (Gardner, MA).

In vitro activation and restimulation

Irradiated feeder EL-4 cells were loaded for 2 h with OVA_{257} peptide or OVA APL (1 μg/ml) and washed twice to remove unloaded peptide. OT-I splenocytes were co-cultured with feeder EL-4 cells for two days in media supplemented with 30 U/ml IL-2 (R&D Systems, Minneapolis, MN) prior to the addition of TGFβ1 (EMD Chemicals, Inc., Gibbstown, NJ) at a final concentration of 20 ng/ml. Some cells were washed in PBS and stained for analysis of extracellular and intracellular markers before the addition of TGFβ1. At day 5, cells were restimulated with OVA_{257} peptide (100 ng/ml) overnight in the presence of GolgiPlug (BD Biosciences, San Diego, CA) and TGFβ1 at a final concentration of 20 ng/ml as described. Cells were then washed in PBS and stained for surface and intracellular markers. To examine initial responses to the OVA APLs, OT-I splenocytes were cultured with various concentrations (ranging from 5 μM to 1 pM) of the peptide for a total of 8 h (6 h after the addition of GolgiPlug) without exogenous IL-2. Cells were washed in PBS and stained for analysis by flow cytometry.

Antibodies and flow cytometry

All mouse antibodies against cell surface and intracellular markers were purchased from Ebioscience (San Diego, CA), except APC-Cy7 anti-CD3 (BD Biosciences, San Diego, CA), Pacific Orange anti-CD8 (Invitrogen, Carlsbad, CA), PE anti-TGFβRII (R&D Systems, Minneapolis, MN), Pacific Blue anti-T-bet (BioLegend, San Diego, CA), FITC anti-KLRG1 (Southern Biotech, Birmingham, AL), anti-TGFβRI (Santa Cruz Biotechnology, Santa Cruz, CA), and anti-RGS3. The anti-RGS3 antibody has been described previously [15]. Extracellular and intracellular marker staining was performed as previously described [16].

Statistical analyses

Sigmoidal dose–response and exponential association curves were fit to data using GraphPad Prism (GraphPad Software, San Diego, CA). For the sigmoidal dose–response curves, separate curves for each peptide were accepted only if the extra sum-of-squares *F* test yielded a *P* value of less than 0.05. The goodness of fit for the exponential association curves is indicated by R^2 value. To compare cytokine suppression across multiple experiments, one-way ANOVA with a Tukey HSD post-test was used to calculate *P* values. *P* values below 0.05 were deemed significant.

Results

Priming with low-affinity peptide ligands gives rise to effectors with enhanced function

In order to investigate the effect of TCR signaling strength at priming on effector function, OT-I $CD8^+$ T cells were primed with EL4 cells loaded with wild-type (wt) OVA_{257} or one of four single residue-substituted versions of the peptide, referred to collectively as APLs

[1]. These APLs bear substitutions only at TCR-facing residues and not anchor residues and accordingly have been shown to affect OT-I TCR-binding affinity without affecting affinity of binding to the class I MHC molecule, H-2K^b. Restimulation with 100 ng/ml wt OVA_{257} was performed at day 5 after priming. While single cytokines are often used as measures of effector CD8$^+$ T-cell function, a number of studies have shown that the simultaneous expression of several cytokines correlates far better with protective immunity than the magnitude of any one single cytokine [17–20]. Therefore, antigen-specific production of the cytokines IFN-γ, TNF-α and IL-2 were used as measures of effector function. In contrast to our expectations, a greater proportion of those cells primed with the lower-affinity APLs produced all three cytokines than those primed with higher-affinity ligands, including wt OVA_{257} (Fig. 1a, b, Table 1). Previous studies have reported that decreased antigen density or availability can lead to enhanced responses at restimulation by maintaining increased levels of TCR and the coreceptor CD8 at the cell surface [21, 22]. However, in response to decreased ligand affinity at the same concentration in our system, no differences were observed with respect to CD3ε or CD8 expression at the cell surface (Supplemental Fig. 1). Furthermore, the effect of lower-affinity ligands was not mediated by activating fewer cells during the 5-day priming period, as the expression of CD44 and CD62L in CD8$^+$ T cells remain unchanged between stimulatory cultures regardless of ligand affinity (Supplemental Fig. 2).

Priming with low-affinity peptide ligands gives rise to effectors with increased susceptibility to TGFβ signaling

As peptide ligand affinity was found to control the quality and magnitude of the secondary response, we endeavored to also investigate the role of ligand affinity during priming on later sensitivity to immune regulation by TGFβ. This was accomplished by priming the OT-I CD8$^+$ T cells in the same manner as before but incubating half of the cells in media containing physiological concentrations (20 ng/ml) of TGFβ1 [23] beginning at day 2 after priming and during restimulation at day 5. As the magnitude of the secondary effector response was enhanced by lower-affinity ligands, we expected to find that these cells were also more resistant to TGFβ-mediated immune regulation. However, while there was little apparent effect of TGFβ on cells primed with higher-affinity ligands, there were large apparent suppressive effects on those primed with the lower-affinity ligands (Fig. 1a, b, Table 1). Furthermore, this effect was observed with respect to production of all three cytokines (Fig. 2a, Table 1).

Ligand affinity at priming correlates with susceptibility to suppression

To further gain insight into the functional effects of APLs with lower affinities, we stimulated naive OT-I CD8$^+$ T cells with various concentrations of OVA_{257} or the APLs and measured cytokine output. As expected, higher concentrations of the lower-affinity APLs were necessary to reach half-maximal stimulation (EC_{50}) and the maximum proportion of cells producing cytokines with stimulation from lower-affinity ligands was also lower than those stimulated with higher-affinity ligands (Fig. 2b and Supplemental Fig. 3). Plotting these experimentally determined relative EC_{50} values, which are similar to previously published values for these peptides [1], against the degree of suppression with each peptide ligand reveals a clear relationship between peptide ligand affinity during priming and later susceptibility to TGFβ-mediated suppression during the effector phase (Fig. 2c). The curves that best correlate with the data are defined by exponential association equations, suggesting that as the affinity of the TCR for the priming ligand decreases, the degree of suppression will eventually approach an asymptotic maximum. In addition, all of these equations include y-intercepts that fall near zero (some are negative), which corresponds to the suppressive effect of 20 ng/ml TGFβ on cells originally primed with wt OVA_{257}. Hence, lower-affinity peptide priming leads to increased susceptibility to TGFβ-mediated suppression. Statistically

significant differences in the degree of TGFβ-mediated suppression were evident with respect to each cytokine or concurrent production of all three cytokines (Fig. 2d).

TCR affinity differentially regulates RGS3 expression in the presence of TGFβ

To determine the molecular basis for the increased susceptibility to TGFβ-mediated suppression, we similarly primed cells with wt OVA_{257} and each of the APLs and analyzed expression levels of proteins in the TGFβ signaling pathway, namely TGFβRI and RII. Yet for neither of these proteins did expression levels correlate with susceptibility to TGFβ-mediated suppression. There were no significant changes in expression for TGFβRI, TGFβRII, or the inhibitory Smad7 (Fig. 3 and data not shown). Interestingly, expression of RGS3, a recently defined noncanonical inhibitor of TGFβ signaling [10], was found to be higher in cells primed with lower-affinity peptide ligands (Fig. 3). In order to more completely recreate the suppressive conditions and thus accurately recreate the susceptible phenotype, the original priming scheme was followed. However, at day 5 after priming, the cells were analyzed for expression levels of each of the TGFβ pathway proteins. Once again, expression levels for the TGFβ receptors I and II were stable across the different peptide ligands (Fig. 3). However, while RGS3 once again was more highly expressed in cells that were primed with lower-affinity ligands in the absence of TGFβ, the presence of TGFβ reversed these phenotypes: RGS3 was upregulated in cells primed with high-affinity ligands and down-regulated in cells primed with low-affinity ligands (Fig. 3). Thus, in the suppressive environment, cells primed with lower-affinity ligands adopted a more TGFβ-responsive phenotype. Interestingly, this change was not accompanied by any significant changes in activation status markers, such as CD44, CD62L, KLRG1, CD127, T-bet, or Eomes (Supplemental Fig. 2). In addition, no changes were observed with respect to CD3, CD44, CD62L, RGS3, TGFβRI, TGFβRII, KLRG1, CD127, or T-bet between priming cultures at day two, prior to the addition of TGFβ (Supplemental Fig. 4).

Discussion

While previous studies have defined the role of TCR affinity in central tolerance, our study highlights the potential interplay between TCR affinity and TGFβ in peripheral tolerance. The outcome of low-affinity TCR ligand stimulation on $CD8^+$ T-cell cytokine production during the primary response is known; however, the effects of such priming on secondary activation and effector function have not yet been defined. Through the use of OVA_{257} peptide and residue-substituted peptide OVA_{257} analogs for which the OT-I TCR has reduced affinity, we have shown that OT-I $CD8^+$ T cells primed with lower-affinity ligands are better suited to respond to the cognate antigen upon restimulation. Furthermore, while it has been reported that functional avidity may be modulated by antigen dose through mechanisms involving altering surface expression levels of the TCR and the CD8 coreceptor [21, 22], we find that the levels of CD3ε and CD8 are unchanged at the surface regardless of the affinity of the priming ligand. This effect may partially underlie the importance of low-affinity self-reactivity in the periphery, where $CD8^+$ T cells may be better suited to recognize foreign, high-affinity antigens and thus resolve infection because of earlier recognition of self-peptide ligands.

One of the central issues in immunology is the way in which autoimmunity is prevented or controlled. This is especially significant for T-cell-mediated autoimmunity, as T cells are positively selected on the basis of self-antigen recognition during thymic development yet are not generally self-reactive in the periphery. The prevailing explanation for this phenomenon is that because T cells that emerge from thymic development have only a low-affinity interaction with self-antigen due to negative selection, such an interaction could not normally lead to a productive immune response [24, 25]. Our results demonstrate an additional mechanism to avert autoimmunity, whereby $CD8^+$ T cells that have been primed

with such a lowaffinity interaction become more sensitive to regulation by TGFβ while those primed with high-affinity interactions are minimally affected. This mechanism allows the host to selectively suppress those $CD8^+$ T-cell clones that may be deleterious while maintaining those that, by virtue of TCR affinity for pMHC, may be considered strictly foreign antigen-specific, even while both are responding to the same antigen in the same microenvironment. The same principle can be applied to tumor-reactive $CD8^+$ T cells, as they generally recognize tumor-associated antigens (most of which are unaltered self-proteins) with very low affinity. This mechanism to avoid autoimmunity would then allow the tumor to more effectively suppress $CD8^+$ T-cell-mediated immune responses, particularly against the tumor itself, via TGFβ.

The immune system has evolved the capacity to mount responses to foreign pathogens while avoiding responses to self-antigens. From studies showing that loss of TGFβ signaling in T cells leads rapidly to multitarget autoimmunity, it can be inferred that self-reactive conventional T cells exist in the natural repertoire and are kept in check through normal levels of TGFβ [11, 12]. In addition, as mice reconstituted with TGFβ-insensitive $CD8^+$ T cells prevent EL4 thymoma or B16 melanoma tumors from developing without any therapeutic intervention, these tumors appear to require the suppressive function of TGFβ to evade immune destruction [14, 26]. However, very little is known about differences in TGFβ sensitivity in T-cell populations. Sanjabi et al. have shown that during the contraction phase, short-lived effector cells respond to TGFβ by undergoing apoptosis, whereas memory progenitor effector cells are preferentially maintained despite elevated TGFβ levels [27]. There is evidence that both antigen affinity and availability during priming control the size of the resulting memory population [1, 28, 29]. It can be hypothesized that in our model, those $CD8^+$ T cells primed with high-affinity ligands are more predisposed to becoming memory cells than those primed with lowaffinity ligands.

A noncanonical function of RGS3 in regulating TGFβ signaling has recently been defined [10]. RGS3 has been shown to bind Smad2, Smad3, and Smad4, thereby impeding heteromerization of R-Smads and Smad4 and preventing TGFβ-induced, Smad-mediated transcriptional activation [10]. While no differences were observed with respect to expression levels of the type I or II TGFβ receptors or the inhibitory Smad, Smad 7, RGS3 expression correlated with sensitivity to TGFβ-mediated suppression of effector function in the presence of TGFβ1. Previous studies have shown that RGS protein expression may be modulated by TLR signaling in DCs [30] and as a result of activation in B cells [31]. However, our data represent the first evidence that RGS3 expression can be modulated in response to both TGFβ signaling and a program resulting from TCR affinity during priming.

Our data demonstrate that while lower TCR ligand affinity results in less intense initial responses, such interactions during priming lead effector cells to become better able to respond to the cognate antigen upon secondary antigen exposure. Despite this, these cells are also more effectively suppressed by TGFβ. Collectively, these results suggest a novel tolerance mechanism, whereby $CD8^+$ T cells are discriminately regulated by TGFβ according to the affinity of the ligand on which they were initially primed. Furthermore, these findings suggest that the low-affinity TCR ligands expressed by tumors may render responding $CD8^+$ T cells more sensitive to TGFβ-mediated suppression and that this programming may be avoided by initially priming $CD8^+$ T cells with higher-affinity ligands.

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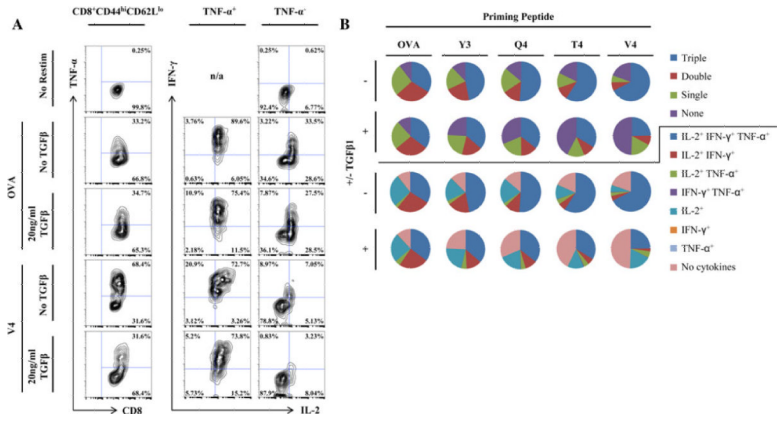

Fig. 1.

Priming with lower-affinity peptide ligands leads to enhanced secondary responses but also increased sensitivity to TGFβ-mediated suppression. OT-I splenocytes were primed on peptide-loaded, irradiated EL4 cells for 5 days in vitro prior to restimulation with 100 ng/ml OVA_{257} for all groups. Half of all samples were incubated in 20 ng/ml TGFβ starting at day 2 and during restimulation. Cells were analyzed by flow cytometry for polycytokine production in the $CD3^+CD8^+CD44^{hi}$ antigen-experienced $CD8^+$ T-cell gate. **a** *Contour plots* demonstrating differential cytokine output by cells primed with OVA_{257} and the V4 APL in the presence and absence of TGFβ. Cytokine output for cells primed for 5 days with OVA257 but not restimulated is also shown. **b** *Pie charts* representing the proportion of effector $CD8^+$ T cells producing all three cytokines (triple), a set of only two cytokines (double), only a single cytokine (single), or no cytokines (none), or the proportion of cells producing a precise combination of the three cytokines. Data shown are representative of at least three individual experiments with similar results

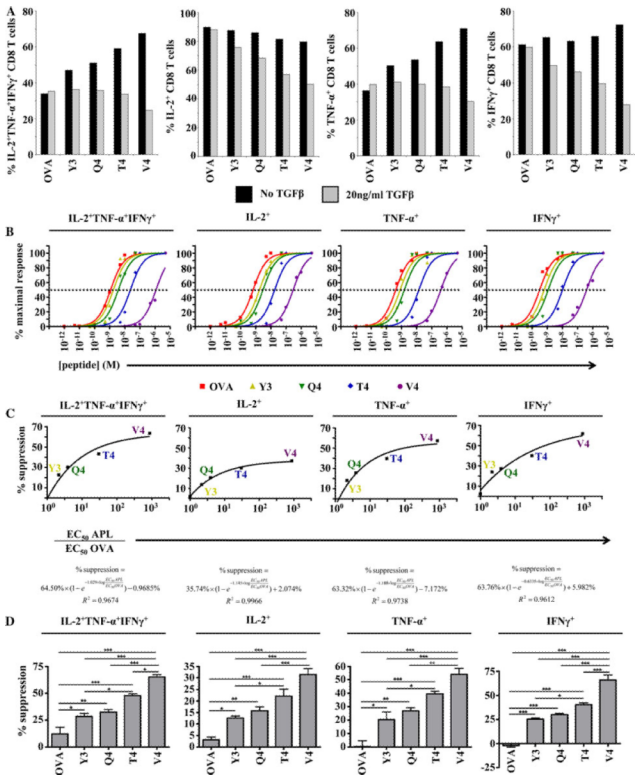

Fig. 2.

Degree of TGFβ-mediated suppression is dependent on the affinity of the ligand on which $CD8^+$ T cells are primed. **a** $CD8^+$ T cells were primed and restimulated as shown in Fig. 1. *Bar graphs* indicate the percentages of $CD3^+CD8^+CD44^{hi}$ cells producing all three cytokines or producing each cytokine, regardless of whether these cells produced any other cytokines. **b** OT-I splenocytes were stimulated in vitro for 8 h with decreasing concentrations of OVA_{257} or the APLs. Following stimulation, cells were analyzed for polycytokine production. For each peptide, responses were normalized to the peak response. The data were fit with sigmoidal dose–response *curves*. Separate *curves* were accepted for each peptide as the extra sum-of-squares *F* test yielded *P* values less than 0.0001 for each plot. A dotted line denotes 50% maximal stimulation. EC_{50} values were derived from the intersection between the sigmoidal dose–response *curves* and the *line* denoting 50% maximal stimulation. **c** The degree to which effector responses were suppressed by TGFβ was plotted against the derived EC_{50} values for each peptide relative to the derived EC_{50} value for OVA_{257}. The equation defining the best-fit curve and its R^2 value is shown for each plot. **d** The degree of suppression of cytokine production by TGFβ was measured across four individual experiments. *Columns* represent mean suppression. *Error bars* represent standard deviation. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$. Suppression was defined by $\left(\%\,\text{cytokine}^+_{\text{noTGF}\beta} - \%\,\text{cytokine}^+_{\text{TGF}\beta}\right)/\%\,\text{cytokine}^+_{\text{noTGF}\beta}$. Data shown are representative of at least three individual experiments with similar results

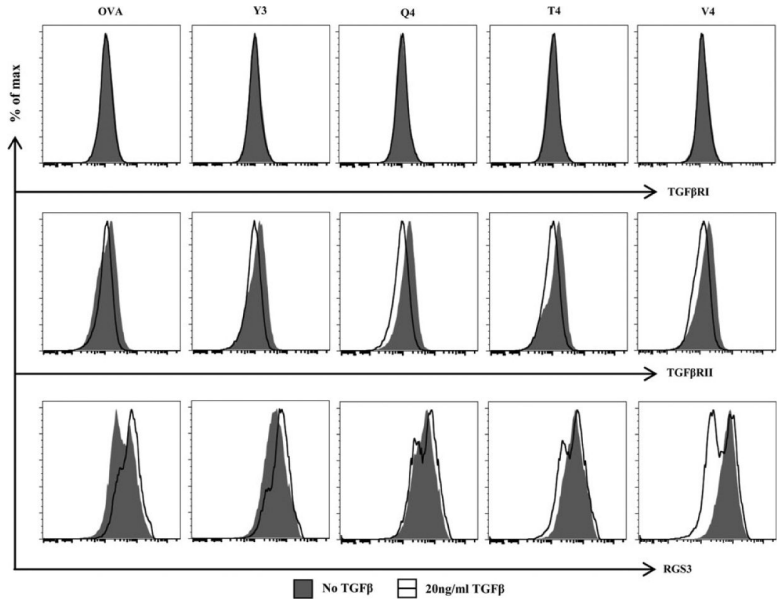

Fig. 3.

Ligand affinity during priming dictates later expression levels of RGS3. OT-I cells were primed by the various ligands and incubated as shown in Fig. 1. At day 5, the cells were analyzed for expression levels of TGFβRI, TGFβRII, and RGS3. Each histogram indicates the expression level of TGFβRI, TGFβRII, or RGS3 in $CD3^{+}CD8^{+}CD44^{hi}$ cells incubated with or without 20 ng/ml TGFβ. Data shown are representative of at least three individual experiments with similar results

Table 1

Cytokine profile of cells primed on OVA or APLs in the presence and absence of TGFβ

Priming peptide	± TGF/β1	Triple	Double	IL−2+, IFN-γ+	IL−2+, TNF-α+	IFN-γ+, TNF-α+	Single	IL−2+	IFN-γ+	TNF-α+	No cytokines	Cumulative		
												IL−2+	TFN-γ+	TNF-α+
OVA	−	33.98	29.29	27.02	2.27	0.00	26.58	26.34	0.18	0.06	10.24	89.61	36.31	61.18
	+	35.28	28.59	24.12	4.35	0.13	24.51	24.25	0.26	0.00	11.60	88.00	39.76	59.79
Y3	−	46.99	21.14	17.91	3.14	0.09	19.73	19.46	0.27	0.00	12.10	87.50	50.22	65.26
	+	36.38	17.63	12.92	4.47	0.24	22.01	21.83	0.12	0.06	24.05	75.60	41.15	49.66
Q4	−	51.03	14.45	11.97	2.48	0.00	20.74	20.52	0.18	0.05	13.76	86.00	53.56	63.18
	+	35.66	14.11	9.94	3.90	0.26	19.00	18.80	0.17	0.03	31.27	68.30	39.86	46.03
T4	−	59.10	10.52	6.17	4.07	0.28	12.45	12.03	0.28	0.14	17.86	81.37	63.59	65.83
	+	33.56	9.94	5.24	4.38	0.32	14.22	13.72	0.40	0.11	42.27	56.90	38.37	39.52
V4	−	67.47	7.26	3.93	2.62	0.71	5.95	5.55	0.20	0.20	19.31	79.58	71.01	72.32
		24.67	8.19	2.54	5.22	0.44	17.69	17.58	0.07	0.04	49.45	50.01	30.37	27.72

OT-I splenocytes were primed and restimulated as shown in Fig. 1. The percentages of the resulting effector $CD8^+$ T cells producing a particular set of cytokines or all of those producing an individual cytokine (Cumulative) are shown. Data shown are representative of at least three individual experiments with similar results

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