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Co-Localization of Immediate Early Genes in Catecholamine Cells after Song Exposure in Female Zebra Finches (*Taeniopygia guttata*)

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Abstract

The physiological state of animals in many taxonomic groups can be modified via social interactions, including simply receiving communication signals from conspecifics. Here, we explore whether the catecholaminergic system of female songbirds responds during social interactions that are limited to song reception. We measured the protein product of an immediate early gene (ZENK) within three catecholaminergic brain regions in song-exposed (N = 11) and silent-exposed (N = 6) female zebra finches (*Taeniopygia guttata*). ZENK-ir induction was quantified in catecholamine cells as well as within cells of unknown phenotypes in three brain regions that synthesize catecholamines, the ventral tegmental area (VTA), the periaqueductal gray (PAG) and the locus coeruleus (LoC). Our results reveal that there are no significant differences in the overall number of cells expressing ZENK between song-exposed and silent-exposed females. However, when we limited our measurements to catecholamine-containing cells, we show a greater number of catecholamine-containing cells expressing ZENK within the LoC in the songexposed females as compared to silent-exposed females. Furthermore, we measured five behaviors during the song and silent-exposed period, as behavioral differences between these groups may account for differences in the co-induction of ZENK and TH-ir. Our results reveal that there were no statistically significant differences in the five measured behaviors between song and silentexposed females. Our study demonstrates that noradrenergic cells within the LoC are involved in the neural architecture underlying sound perception and that cells within the catecholaminergic system are modulated by social interactions, particularly the reception of signals used in animal communication.

Keywords

Noradrenaline; Catecholamine; Immediate Early Gene; Animal Communication; Songbird; dopamine; communication

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Introduction

Animal communication requires both the production of a species-typical communication signal as well as the reception of this signal, principally by conspecifics [Bradbury and Vehrencamp, 1998]. The reception of communication signals can trigger a behavioral change in the animal receiving the signal but, as increasingly more studies demonstrate, such behavioral changes only occur when the receiver's physiological state enables them [Wilczynski et. al., 2005; Ball and Balthazart, 2008; Wilczynski and Lynch, 2011]. Furthermore, underlying physiological factors not only modulate the behavioral response during social interactions but are also modulated by social interactions, such as receiving communication signals [Bentley et. al., 2000; Chu and Wilczynski, 2001; Lynch and Wilczynski, 2006; Goymann, 2009; Maruska and Fernald, 2010]. This is the case even when communication signals are limited to a single sensory modality with which to receive the appropriate communication signals [Burmeister and Wilczynski, 2000; Lynch and Wilczynski, 2006]. While reproductive hormones have been repeatedly demonstrated to be a physiological factor that both modulates and is modulated by signal production and signal reception, it is very likely that other physiological systems can also be modified during communication. The catecholamineregic system is one such candidate.

Catecholamines, such as dopamine and noradrenaline, are evolutionary ancient and wellconserved across many taxa [Smeets and Reiner, 1994]. These neuromodulators influence sensory processing, motor responses, and cognitive processes such as attention, arousal, motivation and decision-making in a wide range of vertebrates [Robbins and Everitt, 1995; Berridge and Waterhouse, 2003; Cardin and Schmidt, 2004; Hurley et. al., 2004; Aston-Jones and Cohen, 2005; Solis and Perkel, 2006; Riters and Pawlish, 2007; Gale et. al., 2008; Lynch and Ball, 2008; Winter et. al., 2009; St. Onge et. al., 2009; Mesce and Pierce-Shimomura, 2010; Jocham et. al., 2011]. Here, we examine the responses of catecholaminecontaining cells after exposure to male mate signals (i.e. song). Our aim is to ascertain whether these cell groups are responsive to the reception of sound. In this preliminary investigation, we examine this possibility by exploring whether these cells express an IEG after reception of conspecific songs. Catecholamine cell groups that respond after reception of species-typical communication signals may provide fundamental insights into whether catecholamines act to enable behavior during critical animal communication events, such as sexual selection via mate choice.

In a wide range of vertebrates, including songbirds, catecholamines are synthesized within the ventral tegmental area (VTA; previously referred to as the area ventralis of Tsai; group A10 in the nomenclature; [Reiner et. al., 2004]), the periaqueductal gray (PAG; previously referred to as midbrain central gray; group A11), and locus coeruleus (LoC; group A6). The brainstem region chiefly responsible for noradrenaline production is the LoC, whereas the midbrain regions (VTA and PAG), primarily synthesize dopamine (see fig. 1a and 1b [Smeets and Reiner, 1994]). These catecholamine-containing cell groups have a profound influence on a large range of higher order processes, including auditory responses to song [Cardin and Schmidt, 2004; Lynch and Ball, 2008]. In female canaries (Serinus canaria) deficits in noradrenaline result in increased errors in behavioral responses to song such that the bird tends to miss the detection of relevant song types [Appeltants et. al., 2002]. This error in behavioral response is mirrored by deficits in auditory responses to song (as measured by immediate early gene induction) in the auditory forebrain, the putative secondary auditory cortex [Lynch and Ball, 2008]). In female European starlings (Sturnus vulgaris), deficits in noradrenaline cause females to increase time spent near nest boxes broadcasting male songs [Riters and Pawlisch, 2007, and noradrenaline reduction decreases estradiol-dependent selectivity in behavioral responses to song in female zebra finches [Vyas et. al., 2008]. These studies indicate a clear role for noradrenaline in regulating female

behavioral and auditory responses to song. Here, we explore whether the reverse is also the case, by exploring whether catecholamines are regulated by the reception of conspecific communication signals. To that end, we use communication signals to probe the song responsive properties of catecholamine-containing cell groups within the female brain.

We investigate whether catecholaminergic cell groups respond during signal reception in female zebra finches (*Taeniopygia guttata*) by co-localizing an immediate early gene within catecholaminergic cells. Immediate early gene expression, specifically zenk mRNA (an acronym for *zif-268, egr-1, ngfi-a*, and *krox 24*) or its protein product, ZENK, are frequently used in studies of animal communication, including studies in songbird communication [Mello et. al., 1992; Mello and Riberio, 1998; Gentner et. al., 2001; Bailey et. al., 2002; Leitner et. al., 2005; Sockman et. al., 2002]. We examined the expression of the ZENK protein product within three catecholamine-producing brain regions, VTA, PAG, LoC in song-exposed and silent-exposed females.

Methods

Female zebra finches were purchased from a local supplier and placed in single-sex housing at Johns Hopkins University, Baltimore, MD, in 49 - 95 - 51 cm cages with no more than six birds per cage. Birds were fed finch food, provided water *ad libitum* and kept on a light schedule of 14L:10D. Females were randomly placed into one of two groups; a negative control (silence) and treatment (conspecific song exposure). Song stimuli were recorded from five unfamiliar males at 20 kHz using a Realistic microphone (Iotech DaqBoard 2000) and custom software written by Amish Dave. Male zebra finches were placed into acoustic isolation chambers to record song, such that songs were recorded with no background noise. However, because males were isolated, the recorded songs were undirected song, which can be less attractive to female zebra finches as compared to directed song (Woolley and Doupe, 2008). We captured hundreds of examples of song from each male. No single song appeared more than once within the 30 min exposure time. Song stimuli were assembled using Adobe Audition 1.5 with approximately 20 seconds of song per minute. The total duration of song presentation was roughly 10 min whereas the total silence was approximately 20 min. Songs were broadcast into the acoustic chamber using RCA speakers at 82 dB SPL, as measured 30 cm from the speaker. Song presentation took place in a 42.5 cm - 60 - 57.5 cm controlled acoustic environment (Industrial Acoustic Company, Bronx N.Y) with acoustic foam lining the interior to reduce reverberation. Females were visually and acoustically isolated from all other birds for approximately 24 hours prior to testing. This allowed the females to habituate to the sound chamber prior to the onset of the experiment. We chose this time period to ensure that females habituated to their new surroundings while simultaneously minimizing time spent in social isolation. Song-exposed females heard 30 min of conspecific zebra finch song (n = 11) whereas the silent-exposed females (n = 6) did not. Ninety minutes after the onset of song playback (or silence), females were sacrificed via rapid decapitation. All procedures were approved by Johns Hopkins University Animal Care and Use Committee.

Tissue Processing

Brain tissue was fixed in 5 % acrolein and cryoprotected in 30% sucrose until cut into three series of 40µm coronal free-floating sections. We conducted double label immunocytochemistry (ICC) using a tyrosine-hydroxylase (TH) antibody and a ZENK antibody. Tyrosine-hydroxylase is a rate-limiting enzyme necessary for the synthesis of catecholamines, and is commonly used to detect catecholaminergic neurons [Ball et. al., 2003; Bharati and Goodson, 2006; Lynch et. al., 2008]. We conducted the double label ICC by performing two sequential free-floating reactions following standard avidin-biotin (ABC) complex procedures, as described in Lynch et. al., [2008]. An abbreviated description is as

follows. We incubated the tissue in ZENK antibody (1:2000 dilution of polyclonal Egr-1 antibody raised in rabbit; Santa Cruz Biotechnology, Santa Cruz, CA; cat. # sc-189) at 4° C for 48 h followed by incubation in biotinylated goat anti-rabbit secondary antibody (1: 250 dilution Vector Laboratories, Burlingame, Ca.) for 1 h at room temperature. Antibody bound to ZENK protein was then visualized using a 1: 200 dilution of Vectastain ABC Elite kit (Vector Laboratories) and nickel enhanced 3,3'-diaminobezidene tetrahydrochloride chromagen, yielding black reaction product (DAB; Polysciences, Warrington, Pa.). We continued by incubating the tissue in a TH antibody (1:10,000 dilution of TH monoclonal antibody raised in mouse; Immunostar, Hudson, WI.; cat. # 22941). This antibody is isolated and purified from rat cells and has wide species cross reactivity because it recognizes a midregion of the TH molecule where extensive homology exists. Primary antibody incubation was done at 4° C for 48 h. The final incubation was a biotinylated horse anti-mouse secondary antibody (1:250 dilution Vector Laboratories) for 1 h at room temperature. Antibody bound to TH was visualized as described above except this DAB reaction lacked nickel enhancement, which yielded brown reaction product (see fig. 1c and 1d). Tissue was then mounted onto slides and coverslipped using Permount.

Quantification

We examined two immunoreactive patterns: co-localization of TH and ZENK-ir (double labeled cells), and cells expressing only ZENK-ir. A Zeiss Axioskop microscope (Carl Zeiss, Thornwood, NY) with a 100x objective was used to count double-labeled cells. ZENK-ir alone and TH-ir cells were counted using a 40x objective. Openlab 3.1.7 software (Improvision, Lexington, Ma.) captured photomicrographs of the whole region for each region of interest (see fig. 1a and 1b). These were used for a manual count of each immunoreactive pattern of interest (see fig. 2 for immunoreactive patterns). The number of sections quantified depended on the region itself; LoC was the smallest region, ranging from two to four sections. The PAG and VTA were larger regions, ranging from four to six sections. Tissue sections that were folded or damaged were excluded. We calculated the mean number of cells expressing either ZENK- ir or ZENK and TH –ir for each subject.

Because there was no *a priori* reason to predict that the number of TH-ir cells would rapidly change over a short period due to song or silent exposure, we randomly chose a subset of subjects in which to estimate overall TH cell number (song-exposed; n = 8; silent-exposed; n = 4). As described above, the mean number of TH-ir cells was calculated for each region.

Behavioral Measurements

We recorded the behavior of female zebra finches using a camera and a VHS recorder. We measured five behaviors during the first 10 minutes of song to determine if song onset caused a behavioral change in the song-exposed group as compared to the silent-exposed group. These behaviors include: time spent near speaker, time spent feeding /drinking, number of movements across the perch, number of grooming behaviors (preening, bill wipes) and number of flights (either to a new perch or to the ground). The side in which the speaker was placed was switched to control for possible side biases. A bird was considered near the speaker if she moved within 15 cm of the speaker. The perch was deliberately placed outside of this range so that the female would need to move off the perch in order to be near the speaker. Time spent near the speaker and time spent feeding/ drinking is expressed as total number of seconds whereas all other variables were measured by counting the number of occurrences. The time spent near the speaker provides a rough estimate of the female's interest in the song or speaker and time spent feeding/drinking may provide an estimate of lack of interest in the song or speaker. Number of perch movements, grooming behaviors and flights can provide an estimate of overall motor responses during the experiment. Because responses of catecholaminergic cells can be related to motor activity,

we used these measures to estimate motor activity. The behavior of one female in the silence group was not recorded due to video tape failure during behavioral observations. Therefore, the final sample sizes for behavioral measurements was n = 11 for the song-exposed group and n = 5 for the silence-exposed group. Lights went on at 8 a.m. and experiments (song playback or silence along with behavioral recordings) started at approximately 10 a.m. every day. Four females were tested on each day. The start of the experiment was staggered by a maximum of 15 minutes so that at the end of song playback (or silence) there was sufficient time to sacrifice each bird and place the brain into fixative. Therefore, all behavior was nearly simultaneously recorded for both the silent and song-exposed females.

Statistical Analysis

Separate two-way analyses of variance were used to determine if the number of cells with either co-localized TH/ZENK-ir or ZENK-ir alone was significantly different among the brain regions (main effect) or between song-exposure groups (main effect). These tests were followed by different post-hoc analyses. A Tukey's *post hoc* test explored whether the brain regions exhibited a statistical difference in the number of cells co-localizing TH and ZENK-ir or ZENK-ir alone. However, this *post-hoc* analysis revealed that the noradrenergic region (LoC) was statistically dissimilar from the other two regions that were primarily dopaminergic in nature (VTA and PAG). Therefore, pairwise comparisons were done using independent t-tests so that we could examine dopaminergic regions separate from the noradrenergic regions. These comparisons determined whether any of these brain regions exhibited significantly different double label as a consequence of treatment group. We used a Bonferroni correction for multiple comparisons when examining the dopaminergic brain regions as they appeared to be statistically similar to one another. The alpha value for the post-hoc pairwise comparison of dopaminergic brain regions was p 0.025, whereas the alpha value for the single noradrenergic region was set at 0.05.

Independent t-tests were also used to compare the behavioral variables between songexposed and silent-exposed females. We used Kolmogorov-Smirnov tests to assess normality and a Levene's test to assess equality of variances. Where variances were unequal, the Levene's corrected statistics were reported for all pairwise comparisons. The means \pm standard errors are reported throughout these analyses.

Results

ZENK-ir

There was a significant difference in the number of cells expressing ZENK-ir as a consequence of brain region (df = 2, 51; F = 5.93; p = 0.005; fig. 3a) but not as a consequence of treatment group (df = 1, 51; F = 1.17; p = 0.284; fig. 3a). There was no significant interaction between brain region and treatment (df = 2, 51; F = 0.86; p = 0.92). Tukey's *post hoc* comparison revealed a significant difference in the number of cells expressing ZENK-ir between VTA and PAG (p = 0.047), and between the LoC and VTA (p = 0.002) whereas LoC exhibited no significant in the number of ZENK-ir cells compared to PAG (p = 0.45).

TH-ir cells expressing ZENK-ir

There was a significant difference in the number of cells with co-localized TH / ZENK-ir among the brain regions (df = 2, 51; F = 6.14; p = 0.004; fig. 3b) as well as a significant effect of song-exposure treatment (df = 1, 51; F = 4.64; p = 0.037; fig. 3b). There was no significant interaction between brain region and treatment (df = 2, 51; F = 1.84; p = 0.171). Tukey's *post hoc* comparison revealed no significant difference in the number of cells with co-localized TH/ZENK-ir between the dopaminergic brain regions (VTA and PAG; p =

0.42), whereas LoC exhibited a significantly greater number of co-localized TH/ZENK-ir cells compared to both of the dopaminergic regions (VTA; p = 0.0001; PAG; p = 0.02). Independent t-tests revealed there was no significant difference in the number of cells expressing TH / ZENK –ir as a consequence of treatment within either of the dopaminergic brain regions (PAG; t = 0.29; df = 15; p = 0.78; VTA; t = 0.8, df = 15, p = 0.43). However, there were significantly more cells with co-localized TH/ZENK-ir within the LoC in song-exposed females as compared to silent-exposed females (LoC; t = 2.46, df = 15, p = 0.026; fig.3b).

TH-ir

The differences in co-localized TH/ZENK-ir expression were not related to possible differences in overall number of TH cell numbers present between the song-exposure treatments. While there was a significant difference in the number of cells expressing TH-ir as a consequence of brain region (df = 2, 51; F = 37.4; p = 0.00), there was no difference as a consequence of song-exposure treatment (df = 1, 51; F = 2.47; p = 0.13). Tukey's *post hoc* comparison revealed a significant difference in the number of cells expressing TH-ir between VTA and PAG (p = 0.00), and between the LoC and VTA (p = 0.000) whereas LoC exhibited no significant difference in the number of TH-ir cells as compared to PAG (p = 0.11).

Behavior

There were no significant correlations between the number of double labeled cells and behavior for any of the five behaviors we measured. In addition, we also examined whether there was a difference in any of the five behaviors between treatment groups. There were no significant differences in any of the behavioral measurements between the treatments (fig. 4). The time spent near the speaker and the time spent feeding / drinking were not significantly different between song-exposed and silence-exposed females (t = 0.18, df = 16, p = 0.85; t = 0.30, df = 16, p = 0.76 respectively). The frequency of short flights, grooming behaviors or perch movements were not significantly different between song-exposed and silence-exposed females (t = 1.3, df = 16, p = 0.21; t = 0.38, df = 16, p = 0.71; t = 0.65, df = 16, p = 0.53 respectively).

Discussion

These results reveal that only one group of catecholaminergic cells responds (as measured by IEG expression within TH labeled cells) after exposure to conspecific songs in female songbirds. These cells are primarily noradrenergic, reside within the LoC, and exhibit greater ZENK-ir expression in song-exposed females as compared to silent-exposed females. In contrast, there is no song-dependent increase in ZENK-ir within non-catecholaminergic cells in any of the brain regions, nor is there elevated song-dependent ZENK-ir in brain regions that are primarily dopaminergic in nature. Moreover, elevated IEG induction in catecholamine cells within LoC following song- or silence-exposure is not related to behavioral responses to song or to the overall number of TH cells. This lack of behavioral differences indicates that ZENK expression within the LoC in song-exposed females is likely a consequence of song reception rather than behavioral responses to song.

Our results indicate that a group of primarily noradrenergic cells tend to respond when a female zebra finch hears sound. However, because we only presented females with conspecific songs (many different examples of conspecific song, yet still only conspecific songs), it is not clear whether catecholamine neurons in LoC respond selectively to conspecific songs. It is possible that these neurons are not song-selective. In which case, they may express ZENK to sound in general, including heterospecific communication

signals. However, these results do indicate that a group of cells that are primarily noradrenergic tend to respond when a female zebra finch hears sound. Future studies will examine the selectivity of this response to ascertain whether noradrenaline cells express ZENK in response to differences within conspecific species (i.e. less attractive, undirected songs vs. attractive, directed songs) as well as differences between species (i.e. conspecific song vs. heterospecific song).

It has been repeatedly demonstrated that ZENK-ir induction in the auditory forebrain, specifically the caudal medial nidopallium (NCM) and the caudal medial mesopallium (CMM), exhibits selective responses to song (Mello et. al., 1992). Such selectivity in these auditory regions is modified by catecholamine deficits (Lynch et. al., 2008) and moreover, catecholamine innervation to these auditory regions is estradiol-dependent (Matragrano et. al., 2011). Interestingly, noradrenergic-specific lesions stimulate female European starlings to approach nest boxes with broadcasted song, which may indicate a decrease in song selectivity if these females are broadening the range of songs to which they would attend as compared to untreated females (Riters and Pawlish, 2007). In addition, noradrenergic deficits decrease estradiol-dependent selectivity in behavioral responses to song in female zebra finches (Vyas et. al., 2008). Consequently, it is possible that estrogen-dependent changes in catecholamine innervation to some regions of the auditory forebrain result in modified responses in these auditory regions, which may in turn influence the female's behavioral responses to song. Thus, future studies will examine the relationship between ZENK-ir induction in these higher auditory regions and ZENK-ir induction within the midbrain and brainstem regions examined here. Overall, however, our results are consistent with the hypothesis that cells within LoC are involved in the neural circuitry underlying conspecific song or sound reception, at least in female zebra finches.

Noradrenaline regulates a wide range of effects from sensory and motor processes to cognitive processes such as attention, arousal, motivation and decision-making among many taxa [Robbins and Everitt, 1995; Berridge and Waterhouse, 2003; Cardin and Schmidt, 2004; Aston-Jones and Cohen, 2005; Solis and Perkel, 2006; Lynch et. al., 2008]. Thus, alternative possibilities exist for explaining sound-dependent regulation of noradrenergic cells within the LoC. For instance, it is possible that these sound-related responses may be a consequence of arousal due to the sudden onset of sound or the onset of attention-related mechanisms [Robbins and Everitt, 1995]. Alternatively, it may be possible that sound reception induces responses from these cells in order to modulate sensory systems. There are a number of studies providing evidence for the latter possibility. For instance, in songbirds, auditory forebrain regions receive robust noradrenergic innervation [Reiner et. al., 1994; Mello et. al., 1998], and chemical lesions of noradrenergic cells remove the differential ZENK induction in forebrain auditory regions that typically occurs in response to hearing conspecific vs. heterospecific songs in female zebra finches [Lynch and Ball, 2008]. Moreover, noradrenaline release into a single region of the auditory forebrain, NIf (nucleus interfacialis of the nidopallium), modifies auditory processing in a state-dependent manner [Cardin and Schmidt, 2004]. In mammals, depletion of noradrenergic innervation reduces sensory-induced expression of IEGs [Cirelli and Tononi, 2004]. It is distinctly possible that sound-related regulation of these cells within the LoC is due to attention and arousal processes that ultimately influence auditory neurons within the forebrain. Distinguishing between these alternative hypotheses may provide more insight into the exact role noradrenaline plays in regulating neural responses to song.

The overall number of LoC cells expressing ZENK-ir within TH labeled cells suggests that it may only require a few cells to produce a biological effect. Furthermore, this effect may be species-specific. For instance, while we saw only a few ZENK-labeled cells within LoC, we were, nonetheless, able to clearly detect cells with co-localized expression. In contrast,

there were no ZENK positive cells identified in TH labeled cells after song exposure in LoC of female white-throated sparrows (Zonotrichia albicollis [LeBlanc et al., 2007]). Differences in co-induction of ZENK-ir within TH cells in LoC between zebra finches and white-throated sparrows may be a consequence of species differences or possibly experience-related differences as LeBlanc et. al. (2007) tested wild-caught birds whereas the birds in the present study were born and raised in captivity. Importantly, although the overall number of LoC cells co-expressing ZENK and TH in the current study is small, the difference between the song-exposure groups is significant. Thus, it is possible that activity in only a few noradrenaline cells may produce robust neuromodulatory activity. Following this reasoning, tract tracing studies of catecholaminergic projections from LoC to HVC (a telencephalic nucleus in the song control system in the songbird brain) report only a small number of cells projecting from the brainstem to the telencephalon [Appeltants et. al., 2000], yet there are high noradrenaline concentrations [Barclay and Harding, 1988; Barclay and Harding, 1990], high rates of noradrenaline turnover [Harding et. al., 1998], and high densities of a-2adrenergic receptors in HVC [Riters and Ball, 2002; Riters et. al., 2002]. Thus, it is possible that responses of only a few catecholamine cells are needed to produce a significant biological effect.

Modulation of physiology via social interaction, particularly reception of communication signals, has been demonstrated in a variety of taxa, especially with respect to hormones [Bentley et. al., 2000; Burmeister and Wilczynski, 2000; Chu and Wilczynski, 2001; Lynch and Wilczynski, 2006]. Here, we demonstrate that modulation of catecholamine-containing cells in the LoC is not related to differences in motor behaviors between song and silent-exposed birds. Rather, the responses of these cells are related to the reception of song. Our study supports the hypothesis that noradrenergic cells within the LoC are involved in the neural architecture underlying sound perception. Overall, the results of this study reveal that catecholamine-containing cells within the LoC are responsive to conspecific sociosexual signals and consequently, it is likely that these cells are part of the neural architecture underlying the reception of signals used in animal communication and possibly the decision to respond.

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Figure 1.

Panels A and B are photomicrographs of the three catecholaminergic brain regions that were investigated in this study. Panel A includes the ventral tegmental area (VTA) and the periaqueductal gray (PAG) in a coronal plane of section. Panel B includes the locus coeruleus (LoC) in a coronal plane of section. Scale bar = 1mm.



Figure 2.

Panel A illustrates VTA cells stained for tyrosine hydroxylase (TH). Scale bar = $10\mu m$. Panel B is a photomicrograph of TH cells with no ZENK-ir induction within the nucleus. Scale bar = $.1\mu m$. Panels C and D show cells with ZENK-ir within the nucleus of a TH cell. Scale bar = $1\mu m$.



Figure 3.

a. Mean number of ZENK-ir cells within three catecholaminergic nuclei of song-exposed (N = 11) and silent-exposed female zebra finches (N = 6). 3b. Mean number of cells with colocalized TH and ZENK-ir within three catecholaminergic nuclei of song-exposed and silent-exposed female zebra finches. All means are reported \pm S.E.M. Asterisks denote significant differences between groups.





Figure 4.

Panel A shows the comparison between the amount of time spent near the speaker and the time spent feeding or drinking between song-exposed (N = 11) and silent-exposed (N = 5) female zebra finches. A bird was considered to be spending time near the speaker if she moved to the same side and corner as the speaker and within approximately 15 cm. Panel B shows the comparison of the number of flights (either to a new perch or to the ground), grooming behaviors (bill wipes, preening) and movements across the perch between song-exposed and silent-exposed female zebra finches.