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Synthesis, Spectroscopy and Hydrogen/Deuterium Exchange in High-Spin Iron(II) Hydride Complexes

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Abstract

Very few hydride complexes are known in which the metals have a high-spin electronic configuration. We describe the characterization of several high-spin iron(II) hydride/deuteride isotopologues, and their exchange reactions with one another and with H₂/D₂. Though the hydride/ deuteride signal is not observable in NMR spectra, the choice of isotope has an influence on the chemical shifts of distant protons through the PIECS (paramagnetic isotope effect on chemical shift) effect in the dimers. This provides the first way to monitor the exchange of H and D in the bridging positions of these hydride complexes. The rate of exchange depends on the size of the supporting ligand, and this is consistent with the idea that H₂/D₂ exchange into the hydrides occurs through the dimeric complexes rather than a transient monomer. The understanding of H/D exchange mechanisms in these high-spin iron hydride complexes may be relevant to postulated nitrogenase mechanisms.

Keywords

Iron; hydride; exchange; deuterium; nitrogenase

Introduction

Though thousands of transition-metal hydride complexes are known, relatively few of them have unpaired electrons. Even fewer have metals with a high-spin electronic configuration, since hydride is a strong-field ligand, and since hydride complexes are often supported by strong-field ancillary ligands. In a series of publications, we described exceptional di(µhydrido)diiron(II) complexes supported by bulky β-diketiminate ligands (Chart 1).^{2,3,4,5,6} In these complexes, the iron(II) ions had high-spin electronic configurations because of the low

metal coordination number and the π -donor character of the anionic β -diketiminate. All of the iron(II) examples were dimers in the solid state, and the two pseudo-tetrahedral metal centers were bridged by hydride ligands (Chart 1, upper left). With the bulkiest β -diketiminate (L^{tBu}), the dimer dissociated in solution to give a three-coordinate monomer, as shown by a combination of magnetic, spectroscopic, and kinetic studies.

The high-spin electronic configuration of the complexes presents characterization challenges that are distinctive to paramagnetic species. Namely, the resonances in the 1H NMR spectra are broadened and highly shifted, and these chemical shifts do not correlate with structure in the manner that is familiar from diamagnetic complexes. 11 The relaxation of 1H nuclei directly bonded to the paramagnetic metal is particularly rapid, and to our knowledge no metal-bound 1H nuclei have been detected in NMR spectra of hydride complexes with a paramagnetic ground state. 12,13 Another challenge is that the paramagnetic hydride complexes in Chart 1 are highly reactive: for example, they cleave B–C bonds, 14 and reductively eliminate 12,13 Another with added ligands including 13 including 13 Though the high reactivity of the hydrides makes them difficult to handle, their reactivity is also an opportunity to form new C–H bonds, because the M-H bonds undergo rapid [1,2]-addition to practically all multiple bonds in organic molecules. 3 This reactivity can be attributed to Fe-H bond weakening as a result of the partial population of Fe-H * orbitals in the high-spin 14 electronic configuration.

One of the important reactions of coordinatively unsaturated metal-hydride complexes is the exchange of the hydride hydrogens with free $\mathrm{H_2}$. This reaction has biological relevance because of the H/D exchange of $\mathrm{H_2}$ protons with solvent protons in nitrogenase enzymes, which occurs *only* in the presence of $\mathrm{N_2}$. This specificity has been used to suggest that hydride species are key intermediates during $\mathrm{N_2}$ reduction. Wery recently, deuterium atoms from $\mathrm{D_2}$ were incorporated into ethylene produced from acetylene reduction by nitrogenase. However, the mechanisms are difficult to evaluate without "model" studies on well-characterized synthetic hydride complexes, particularly with iron. Unfortunately, the aforementioned inability to observe resonances for H and D bound to a metal has so far prevented the monitoring of H/D exchange in paramagnetic hydride complexes.

In this contribution, we describe synthetic and characterization advances for two high-spin iron(II)-hydride complexes, supported by the β -diketiminate ligands L^{tBu} and L^{Me} (Chart 1). In an interesting twist, these studies benefit from unusually large (up to 5.7 ppm) isotope effects on the 1H NMR chemical shifts of the distant protons upon hydride deuteration, a phenomenon that occurs *only* in the dimeric hydride complexes in our system. This tool for differentiating isotopologues enables the revision of the 1H NMR assignments in two previously reported hydride complexes. In addition, this is the only way to distinguish hydride from deuteride isotopologues, and its discovery enables us to monitor H/D exchange in this system for the first time. The results show that the rates of H/D exchange between hydride complexes, and between these hydride complexes and H2, are greatly influenced by the size of the supporting ligand. Mechanistic considerations lead to new insights into the distinctive reactivity of high-spin hydride complexes.

Results

Spectroscopic Comparison of Protiated and Deuterated L^{tBu}FeH, in Monomeric and Dimeric Forms

The synthesis of $[L^{tBu}Fe(\mu-D)]_2$ (1-D₂) was reported previously, and initial characterization of 1-D₂ by ¹H NMR spectroscopy in C₆D₆ suggested that the deuteride and the hydride complexes had identical ¹H NMR spectra. ⁵ However, closer investigation has shown that the resonances have different chemical shifts in the different isotopologues. A ¹H NMR spectrum of an equimolar mixture of 1 and 1-D₂ in C_6D_6 showed that the differences were not from temperature or medium effects (Figure 1). Close examination of the ¹H NMR spectrum revealed several envelopes of nearby resonances, and the components of each envelope had a 1:2:1 ratio of integrations. For example, resonances at δ –37.2 (resonance assigned to 1), -40.1, and -43.0 (resonance assigned to 1-D₂) ppm were observed in a 1:2:1 ratio, rather than the single resonance at δ –37.2 ppm in 1. When this experiment was repeated, starting from a different mixture of isotopologues that contained more 1 than 1-D₂, the ¹H NMR spectrum showed the same number of resonances, but the integrations were no longer 1:2:1 and favored the hydride resonance at δ –37.2 ppm. The resonances located between 1 and 1-D₂ are most reasonably assigned as $\{L^{tBu}Fe\}_{2}(\mu-H)(\mu-D)$ (1-D). These experiments also indicate that mixing of 1 and 1-D2 rapidly yields an equilibrium mixture of 1, 1-D, and 1-D₂. The difference between the chemical shifts of distant protons in different isotopologues has been described previously in a number of paramagnetic complexes, 21 and Theopold has termed this PIECS (paramagnetic isotope effect on chemical shift).²²

The discovery that **1**, **1-D** and **1-D₂** exhibited PIECS enabled the use of ${}^{1}\text{H}$ NMR spectroscopy to accurately determine the amount of deuterium incorporation into **1-D₂**. This was done by comparing the integrations of the three isotopologues in the ${}^{1}\text{H}$ NMR spectrum. Compound **1-D₂** typically had greater than 90% deuterium incorporation into the hydride positions, which is consistent with the level of deuteration previously reported (in earlier studies this determination was done indirectly, using mass spectrometric analysis of 3-hexene- d_1 generated from treating **1-D₂** with 3-hexyne and then acid).⁵

Note that PIECS is not observed for some of the peaks in the spectrum. Seven of these peaks are assigned to monomeric L^{tBu} FeH, which is in equilibrium with 1 as previously shown.² This is the number of resonances expected for L^{tBu} in an environment having C_{2v} symmetry, when the N-aryl bonds have hindered rotation that makes the two methyl groups of the isopropyl substituents inequivalent. By process of elimination, the PIECS of the peaks of 1 enabled the assignment of 18 resonances each to the 1, 1-D, and 1-D₂ isotopologues of the dimers, giving 54 resonances in total. Therefore, the 1 H NMR spectrum of isotopically pure 1 contains 25 resonances, where 7 may be assigned to L^{tBu} FeH and 18 may be assigned to the dimer.

It is notable that 1H NMR spectra of a mixture of the three isotopologues always showed some additional resonances that neither exhibit PIECS nor can be assigned to the monomer L^{tBu} FeH. These resonances had previously been assigned as resonances of $1.^2$ Comparison with literature 1H NMR spectra indicated that these additional resonances were associated with a persistent impurity, $\{L^{tBu}$ Fe $\}_2(\mu$ -O), which can come from the reaction of 1 with

trace H_2O to give $\{L^{tBu}Fe\}_2(\mu\text{-}O).^{23}$ We were not able to completely avoid or eliminate the oxo impurity, but careful handling gave samples of **1** that were below 10% oxo impurity, with typical samples between 4–8%. The oxo impurity in **1-D₂** was typically higher (around 20%), due to the multiple D_2 additions necessary for full deuterium incorporation.

Solid 1 and 1-D₂ were also evaluated using Mössbauer spectroscopy. The zero-field Mössbauer spectrum of solid 1 at 80 K was previously reported to have $\delta=0.59$ mm/s and | $E_Q|=1.58$ mm/s. However, the published data were reexamined after the discovery of the persistent oxo impurity. The oxo impurity was modeled using parameters that were fixed at literature values of $\delta=0.64$ mm/s and | $E_Q|=1.42$ mm/s, while the amount of the oxo impurity and the parameters of the major component were refined to give the best fit to the data. Complex 1 was determined to be 71% of the earlier sample, and the major component had $\delta=0.57$ mm/s and | $E_Q|=1.63$ mm/s. A new sample of 1 with <5% oxo impurity by NMR spectroscopy yielded a zero-field Mössbauer spectrum at 80 K with parameters $\delta=0.58$ mm/s and | $E_Q|=1.62$ mm/s, which are the same within the experimental uncertainty of ±0.02 mm/s.

The zero-field Mössbauer spectrum of solid **1-D₂** at 80 K is shown in Figure 2. The slightly asymmetric two-line pattern was modeled with a two-component fit, of which the major component was found to have $\delta=0.58$ mm/s and $\mid E_Q\mid=1.74$ mm/s, accounting for 79% of the sample. We assign this subspectrum to compound **1-D₂**, whereas the second doublet was fixed to the properties of the oxo impurity ($\delta=0.64$ mm/s and $\mid E_Q\mid=1.42$ mm/s). The isomer shifts of **1** and **1-D₂** are identical, but there was a variation of 0.12 mm/s in the quadrupole splitting values. The reason for the difference in the quadrupole splitting is unknown at this time. Unfortunately, attempts to fit the variable-field Mössbauer spectra of **1** and of **1-D₂** did not yield a comprehensible model that gave additional insight. The isomer shifts of **1** and **1-D₂** lie in the range of δ 0.47–0.90 mm/s observed in other high-spin Fe(II) diketiminate complexes. ^{23,24}

Synthesis and Characterization of LMeFeBr(THF) (2)

We now shift to iron(II) complexes of the smaller diketiminate ligand L^{Me} , which we have previously derived from the iron chloride precursors $L^{Me}Fe(\mu\text{-Cl})_2Li(THF)$ and $[L^{Me}Fe(\mu\text{-Cl})]_2$. However, the relative ease of removing Br salts encouraged us to prepare $L^{Me}FeBr(THF)$ (2) in 82% yield from $L^{Me}K$ and $FeBr_2$. During the preparation of our work, compound 2 was reported by Tonzetich and Lippard, 26 using a very similar prepatory method with $L^{Me}Na$. Our characterization of 2 by NMR and X-ray diffraction (Supporting Information) is indistinguishable from the literature. 26 However, Mössbauer data have not been reported previously for this compound. The zero-field Mössbauer spectrum of 2 at 80 K (Figure S-1) has a doublet with $\delta = 0.89$ mm/s and $|E_Q| = 2.36$ mm/s, which is nearly identical to that of $L^{Me}Fe(\mu\text{-Cl})_2Li(THF)$.

THF was removed from **2** by dissolving it in non-coordinating solvents; this gave $[L^{Me}Fe(\mu-Br)]_2$, which precipitated from solution as an orange powder. This mirrors the behavior of $[L^{Me}Fe(\mu-Cl)]_2$ which also has low solubility. ^{25b,c} Tonzetich and Lippard also reported this behavior, and they reported that $L^{Me}FeBr(THF)$ had different electronic absorption spectra

in THF versus toluene which resulted from the formation of $[L^{Me}Fe(\mu-Br)]_2$.²⁶ Here, the Mössbauer spectrum of orange $[L^{Me}Fe(\mu-Br)]_2$ derived from **2** was recorded at 80 K (Figure S-2). One quadrupole doublet with $\delta=0.91$ mm/s and $|E_Q|=2.64$ mm/s was observed. The isomer shift of $[L^{Me}Fe(\mu-Br)]_2$ is the same as **2**, but the quadrupole splitting is larger, consistent with a slightly different geometry at iron.

Synthesis of $[L^{Me}Fe(\mu-H)]_2$ (3) Using H₂

The successful synthesis of $[L^{tBu}Fe(\mu-H)]_2$ from addition of H_2 to an iron(I) source⁵ prompted us to use this synthetic method for an improved synthesis of $[L^{Me}Fe(\mu-H)]_2$ and $[L^{Me}Fe(\mu-D)]_2$. However, the order of addition of reagents was important, as reduction of **2** with KC_8 in Et_2O followed by H_2 addition yielded many unidentified resonances in the crude ¹H NMR spectrum. This suggests that the transient iron(I) species formed by KC_8 reduction rapidly decomposes in the absence of a trap.²⁷ Therefore, a degassed solution of **2** in THF was exposed to 14 equiv of H_2 gas *prior to* addition of KC_8 . After 3 h, volatile materials were removed from the brown reaction mixture, and **3** was isolated in 56% yield following workup. The identity of **3** was established by comparing its ¹H NMR spectrum to the spectrum reported in the literature.⁴ Again, the iron-bound hydrogen atoms are not visible by ¹H NMR spectroscopy due to close proximity to the paramagnetic iron atoms.

The deuterated isotopologue $3-D_2$ was synthesized using the above method with D_2 gas in 58% yield. The 1H NMR spectrum of $3-D_2$ (Figure 3) revealed that 3 and $3-D_2$ exhibited PIECS as with the hydride dimers described above. Thus, 1H NMR spectroscopy could similarly be used to determine the amount of deuterium incorporated into the hydride ligands (Figure 6 below shows mixtures of isotopologues). The bridging ligands in samples of $3-D_2$ were typically greater than 90% deuterated, as judged by 1H NMR spectroscopy.

In the ¹H NMR spectrum of 3 at room temperature, only 4 paramagnetically shifted peaks were observed, a number that is well short of the 7 resonances expected for 3 in a dimeric structure with D_{2d} or D_{2h} symmetry. Therefore, we hypothesized that additional resonances were hidden at room temperature. In addition, the integrations of the peaks are inconsistent with the original assignments for the ¹H NMR spectrum of 3,⁴ so further investigations were pursued. ¹H NMR spectra of **3** between 26 °C and 85 °C in C₆D₆ are shown in Figure 4a. The peak at δ 13 ppm at 26 °C corresponds to the backbone methyl and *meta*-aryl protons, two resonances that are only distinct above 60 °C (a close-up is shown in Figure S-3). A previously undetected resonance for isopropyl methyl groups integrating to 24 protons (which had been hidden under the residual benzene at room temperature) became visible above 40 °C. The other resonance for isopropyl methyl protons, a broad peak at δ –25 ppm in the 26 °C spectrum, sharpened at elevated temperatures. Finally, a new broad resonance for the isopropyl methine protons at δ –1.5 ppm was observed above 70 °C. This resonance is broadened into the baseline at room temperature, explaining why it had not been identified in previous studies. The remaining resonances at δ –24 and –56 ppm corresponded to the para-aryl protons and the backbone protons, respectively, completing the catalogue of resonances with the expected integrations (see Experimental Section).

The appearance and sharpening of resonances in the high temperature ^1H NMR spectra of **3** suggested the possibility of a fluxional process in solution. Therefore, low temperature ^1H NMR spectra were measured between $-90\,^{\circ}\text{C}$ and $20\,^{\circ}\text{C}$ in toluene- d_8 (Figure 4b). The broad isopropyl methyl resonance that appeared at $8-25\,$ ppm in the ^1H NMR spectrum of **3** at 20 $^{\circ}\text{C}$ split into two broad resonances at $8-27.6\,$ and $-48.3\,$ ppm in the ^1H NMR spectrum at $-75\,^{\circ}\text{C}$. These resonances moved together and became broader as the sample was warmed, with a coalescence temperature of $0\,^{\circ}\text{C}$. The barrier (G) for this fluxional process, assuming that the resonances at $-75\,^{\circ}\text{C}$ are in the slow-exchange limit, is $G=10.5\,$ kcal/mol. 28 Decoalescence of other peaks was not observed within this temperature range, likely because there was a smaller difference between the frequencies in the slow-exchange limit. The possible nature of this fluxional process is discussed below.

Mössbauer Spectroscopy of [L^{Me}Fe(μ-H)]₂ and [L^{Me}Fe(μ-D)]₂

The purity and electronic structure of solid 3 and 3-D₂ were evaluated using Mössbauer spectroscopy. The zero-field Mössbauer spectra of solid 3 and 3-D₂ at 80 K are shown in Figure 5. Compound 3 had $\delta=0.51$ mm/s and $\mid E_Q\mid=2.05$ mm/s. The deuterated isotopologue 3-D₂ had an indistinguishable Mössbauer spectrum with $\delta=0.51$ mm/s and $\mid E_Q\mid=2.10$ mm/s. There were impurities of 13 and 6%, respectively, which are discussed in detail in the Supporting Information. One impurity doublet in each case has parameters similar to those in the literature iron(I) benzene compound L^{Me}Fe(η^6 -C₆H₆),²⁷ which has $\delta=0.70$ mm/s and $\mid E_Q\mid=0.74$ mm/s (Figure S-4). We also note that this nearly NMR-silent impurity was the *major* species in the Mössbauer spectrum previously attributed to 3.³

Intermolecular Hydride Exchange in Isotopologues, and Exchange of Hydrides with H₂

As reported above, a 1:2:1 mixture of **1, 1-D**, and **1-D**₂ was obtained upon mixing an equimolar mixture of **1** and **1-D**₂ in C_6D_6 , and PIECS enabled all three isotopologues to be distinguished in the paramagnetic ¹H NMR spectrum. Equilibrium was established within 45 min at room temperature. An analogous experiment was performed using an equimolar solution of **3** and **3-D**₂ maintained at 30 °C in C_6D_6 . A 1:2:1 mixture of **3**, { $L^{Me}Fe$ }₂(μ -H) (μ -D) (**3-D**), and **3-D**₂ was observed after 2 h. This shows that the inability of **3** to form monomeric $L^{Me}FeH$ in solution ¹⁴ does not hinder hydride exchange between isotopologues.

The exchange of hydride and deuteride ligands with D_2 and H_2 was also examined. Compound 3 was treated with an excess of D_2 (1 atm) in C_6D_6 to give a mixture of 3-D and 3-D₂. Equilibrium was established immediately upon mixing (Figure 6A). When this mixture was treated with fresh D_2 , the equilibrium was pushed all the way to the fully deuterated isotopologue, 3-D₂ (Figure 6B). Two treatments with H_2 caused the sample to revert to 3 in quantitative yield (Figures 6C and 6D), showing that the exchange is reversible. In contrast to the immediate exchange in 3, treatment of 1 in C_6D_6 with D_2 (1 atm) produced the deuterated isotopologue only after much longer amounts of time. Compound 1-D₂ was finally observed in quantitative yield after 50 h at room temperature. Hydride exchange between H_2 and 1-D₂ is reversible as treatment of 1-D₂ with H_2 (1 atm) produced 1 in quantitative yield under the same conditions and time. Treatment of 1 with 8 atm of D_2 was faster, but required 24 h to yield 1-D₂ in quantitative yield. A qualitative summary of hydride ligand exchange rates is given in Table 1.

Lack of H/D Exchange in Analogous Cobalt Hydrides

We have also reported the dimeric cobalt(II) hydride complex $[L^{tBu}Co(\mu-H)]_2$. This compound is much less reactive than the iron analogues described above; for example, it does not react with alkenes or Lewis bases. Be The low reactivity was attributed to the greater stability of the dimer, and/or decreased lability of the Co-H bonds. Therefore, it was interesting to evaluate the analogous cobalt compounds for intermolecular H/D exchange.

A sample of $[L^{tBu}Co(\mu-D)]_2$ was prepared from $[L^{tBu}Co(\mu-F)]_2$ and Et_3SiD , using a method analogous to that used to synthesize the protiated analogue. Several peaks were shifted by 0.2–0.5 ppm from those in $[L^{tBu}Co(\mu-H)]_2$, as verified by spiking the sample with an equimolar amount of $[L^{tBu}Co(\mu-H)]_2$ (Figure S-6, S-7, S-8). Thus, this dimer also exhibits PIECS, though the shifts are not as pronounced as in the iron species described above. Heating the mixture of $[L^{tBu}Co(\mu-H)]_2$ and $[L^{tBu}Co(\mu-D)]_2$ to 80 °C for 12 h gave no change in the NMR spectrum, indicating that there is no significant exchange of hydrides between complexes in this time frame. This contrasts with the iron analogues (Table 1), and is consistent with the idea that the cobalt(II) hydride dimer does not break up in solution.

Discussion

Characterization of High-Spin Iron Hydride Complexes, Including PIECS in $^1\mathrm{H}$ NMR Spectra

The Mössbauer spectra of **1** and **3** have similar isomer shifts, $\delta = 0.59$ mm/s and 0.51 mm/s, which are consistent with the values observed in other high-spin iron(II) β -diketiminate complexes. ^{23,24} High-spin, tetrahedral iron(II) sites in iron-sulfide clusters have similar isomer shifts ($\delta = 0.6$ –0.7 mm/s). ³⁰ Low-spin octahedral iron(II) sites have very different isomer shifts in the range $\delta = 0.3$ –0.45 mm/s. ^{30b} The intermediate-spin (S = 1) iron(II) hydride complex [Fe(dppe)₂H]⁺ has $\delta = 0.23$ mm/s. ³¹ The much higher isomer shifts in **1** and **3** strongly support the assignment of **1** and **3** having *high-spin* Fe(II) subsites, and is consistent with the paramagnetic shifts in the ¹H NMR spectra. We note that our assignment of the Mössbauer spectrum of **3** here replaces an incorrect assignment we gave in an earlier paper; ³ the previous spectrum actually corresponds to L^{Me}Fe(arene), which can be formed during the synthesis of **3** when arenes are present.

The 1H NMR spectra of **1** and **3** were found to exhibit significant paramagnetic isotope effects on the chemical shift (PIECS), as all the resonances shifted upon deuterium substitution. The term PIECS was coined by Heintz and Theopold, 22 and has been reported in a number of complexes. 21,32 For example, deuteration of the hydrides in (Cp'') $_4$ Cr $_4$ (μ_3 -H) $_4$ (Cp'' = η^5 -C $_5$ Me $_4$ Et) 22 gives changes in the chemical shifts of the Cp" protons, though they are far removed from the bridging hydride ligands. Most explanations for PIECS are based on the shorter bond lengths to D vs. H. 33 The Heintz/Theopold study is particularly relevant to our complexes because of the presence of bridging hydrides: shorter M-D bonds could decrease the M-M distance and the exchange coupling, which in turn would influence the magnetic susceptibility and thus the chemical shift of the protons. 11 However, other explanations have been advanced for other cases of PIECS: for example, differential M-H/M-D bond energies could influence the ligand-field splitting slightly, which in turn could

influence the paramagnetic shift. ^{21a,c} However, we saw no evidence for any sizable change in the ligand field of iron upon deuteration, because the quadrupole splittings were the same within error between 3 and 3-D₂, and only slightly different ($E_Q = 0.12 \text{ mm/s}$) between 1 and 1-D₂.

The PIECS in the ¹H NMR spectra of **1** and **3** varied from a negligible change in some resonances up to 5.7 ppm in others. Importantly, PIECS was observed *only* for the dimeric form of **1**, as the monomer L^{tBu}Fe(H/D) had the same chemical shifts in the ¹H NMR spectrum for both isotopologues. This observation supports the hypothesis that the PIECS is connected to changes in the Fe-Fe distance.

A significant finding of this study is that variable temperature ¹H NMR spectroscopy together with PIECS were essential for defining the correct ¹H NMR assignments of **1** and **3**.^{2,4,14} Compound **3** was particularly vexing, because almost half of the seven expected resonances were masked. One resonance was hidden under the residual solvent, another apparent resonance was actually two resonances with the same chemical shift, and one was broadened into the baseline at room temperature and was broad even at elevated temperatures. However, variable-temperature studies enabled us to finally assign the resonances for **3** with confidence.

Examination of the variable-temperature 1H NMR data for 3 also revealed a fluxional process in solution with a barrier of G=10.5 kcal/mol at 0 °C. We tentatively assign the low-temperature structure to be similar to the solid-state structure, 4,14 in D_2 symmetry with the diketiminate planes perpendicular to one another. In this case, the barrier would correspond to the energy required to reach the D_{2h} -symmetric conformation in which the N_2C_3Fe planes are coplanar (Scheme 1), and through which two D_2 isomers of different chirality can interconvert.

Meanwhile, PIECS was vital to the assignment of all resonances in the 1 H NMR spectrum of 1, which is complicated by the presence of both monomer and dimer. 2 In addition, the crystallographic symmetry of the dimer in 1 is lower than that in 3, because crowding gives a boat conformation of the $N_{2}C_{3}$ Fe rings that lowers the symmetry to idealized C_{2} . We observed 18 resonances in dimeric 1, which is somewhat less than the 21 predicted from C_{2} symmetry in solution, implying that three of the resonances are lost to overlap or broadness. However, the number of peaks is significantly larger than in 3, indicating that the increased steric bulk of the L^{tBu} ligand in 1 (from a buttressing effect) 3,25 prevented the fluxional process that was observed in 3. The 1 H NMR spectra imply that the hydride ligands are oriented such that there is no plane of symmetry relating the two L^{tBu} ligands. Overall, these studies show the usefulness of variable temperature 1 H NMR spectroscopy and PIECS to decipher paramagnetic spectra and solution structure despite multiple overlapping resonances.

Intermolecular Hydride Exchange

Compounds 1 and 3 undergo intermolecular hydride exchange with their deuterated isotopologues. Dissolving 1 and $1-D_2$ in C_6D_6 produced an equilibrium mixture containing dimeric 1, 1-D, and $1-D_2$, along with monomeric L^{tBu} FeH and L^{tBu} FeD. In the iron(II)

complexes, formation of **1-D** most likely results from equilibrium between monomeric and dimeric **1**, which is slow on the NMR timescale but rapid on the chemical manipulation timescale (the rate of dissociation has been estimated as 5×10^{-4} s⁻¹ at 288 K based on kinetic studies of the reaction of **1** with alkynes²). Consistent with this rate regime, equilibrium between the isotopologues of **3** is reached within 45 minutes at room temperature as judged by ¹H NMR spectroscopy.

The hydride isotope exchange between 3 and 3-D₂ cannot follow this process because 3 does not interconvert with monomer in solution, as previously shown using kinetic studies on the reaction with boranes. ¹⁴ Two possible mechanisms for hydride isotope exchange between 3 and 3-D₂ are shown in Scheme 2. The first is a concerted process, while the second requires rate-limiting cleavage of one of the bridging hydrides to give a single terminal hydride. The hydride ligands in the terminal position could attack another dimer to give exchange. Partial breaking of the dimer of 3 to give a terminal hydride, as in Scheme 2b, was previously found to be the most reasonable mechanism for the reaction of 3 with trialkylboranes, ¹⁴ and so we consider this to be the most likely possibility that is consistent with the combined studies on 3.

It is also notable that the cobalt analogue, $[L^{tBu}Co(\mu-H)]_2$, did not undergo H/D exchange even with heating to 80 °C. Its low reactivity in general may be attributable to its inability to open one of the bridges, as in Scheme 2b for 3, or both bridges, as in 1.

Hydride Exchange with H₂ and D₂

The exchange of hydride complexes with H_2 and D_2 has been studied in detail in the literature. ¹⁶ It is well-established in mononuclear iron-hydride complexes. ³⁴ In each of these cases, an open coordination site is required to bind H_2 in an η^2 binding mode. The oxidative addition of the H_2 is not necessary, because there can be direct H transfer from coordinated H_2 to the hydride without changing the oxidation state at the metal.

It is interesting that the inability of $\bf 3$ to form monomeric L^{Me} FeH in solution did not hinder hydride ligand exchange with D_2 . Equilibrium was established in less than a minute after treatment of $\bf 3$ with D_2 . Though we cannot determine the mechanism unambiguously, we can advance two possible mechanisms. First, the "opened" form of the dimer has an open coordination site on one iron that could coordinate H_2 or D_2 to make a transient side-on D_2 complex that is well-situated to exchange with the hydride on the other metal (Scheme 3a). Another potential mechanism involves oxidative addition of H_2 , either to one metal (giving a transient iron(IV) on one side) or to both metals (giving a diiron(III) species). The latter possibility is shown in Scheme 3b.

It is also relevant that compound 1 does not undergo facile hydride exchange with D_2 , despite its ability to form monomer in solution. This required over 2 d at room temperature under 1 atm of D_2 and 1 d under 8 atm of D_2 . The hindered reaction of 1 with D_2 supports the contention that H_2/D_2 exchange in these hydride species does not proceed through a *transient monomer*. It is possible that the reaction of 1 with D_2 proceeded via the monomer at a significantly slower rate. Another possibility is that the exchange goes through the

dimeric form of 1, but that the bulkier ligands hinder its ability to access the reactive conformation.

Dinuclear iron sites with bridging hydride ligands have been studied extensively as [FeFe]-hydrogenase models. These diiron hydride complexes have been reported to undergo H/D hydride ligand exchange with D_2/H_2 via photolysis. In these systems, photolysis opens a coordination site by dissociating CO or cleaving a hydride bridge, and the H/D hydride exchange requires days (which contrasts with exchange in 3/3- D_2 that occurs in seconds). H/D hydride ligand exchange has also been reported using D_2/H_2O for hydrogenases, diiron complexes, diiron complexes, other metal complexes, along with D^+ as a deuterium source. In addition, nitrogenase can exchange D from D_2O into H_2 , but only does so in the presence of N_2 . The suggest that the mechanisms advanced above for hydride D_2 exchange in D_2 should be considered in nitrogenase: specifically, bridging hydride species may play key roles in D_2/D_2 exchange in the FeMoco cluster.

Conclusions

 D_2 was utilized as a deuterium source to enable the isolation and characterization of the low-coordinate iron deuteride complexes [L^{tBu}Fe(μ -D)]₂ and [L^{Me}Fe(μ -D)]₂. The Mössbauer spectra of these hydride complexes indicated that the metal centers are high-spin iron(II). The ¹H NMR spectra of the hydride isotopologues exhibited a paramagnetic isotope effect on chemical shift (PIECS). This effect was observed only in the dimeric complexes, implicating the slightly smaller size of the $M_2(\mu$ -D)₂ core as the main cause of PIECS.

PIECS also enabled the correct ¹H NMR assignments of the hydride complexes, as well as the study of intermolecular hydride exchange. The exchange of hydrides between hydride complexes of the largest supporting ligand is likely to occur through dissociation of the dimers into monomers. However, exchange of the hydrides with added H₂ occurs most rapidly with the smaller supporting ligand, implicating diiron(II) hydrides as the key intermediates. More generally, these studies show that ¹H NMR spectroscopy can be a powerful tool for the study of paramagnetic iron hydride complexes: not *despite* the paramagnetism, but *because* of the paramagnetism through the PIECS effect.

Experimental Section

General Considerations

All manipulations were performed under a nitrogen atmosphere (or argon atmosphere where specified) by Schlenk techniques or in an M. Braun glovebox maintained at or below 1 ppm of O_2 and O_2 and O_3 Glassware was dried at 150 °C overnight, and Celite was dried overnight at 200 °C under vacuum. Pentane, hexane, benzene, diethyl ether, and toluene were purified by passage through activated alumina and O_3 columns from Glass Contour Co. (Laguna Beach, O_3). THF was distilled under O_3 from a potassium benzophenone ketyl solution. All solvents were degassed by removing a small amount of solvent under reduced pressure prior to argon glovebox entry. All solvents were stored over 3 Å molecular sieves. Benzene- O_3 0 was dried and stored over flame-activated alumina. Toluene- O_3 1 and THF- O_3 2 were vacuum transferred from sodium benzophenone ketyl solutions and were stored over 3 Å molecular

sieves. Before use, an aliquot of each solvent was tested with a drop of sodium benzophenone ketyl in THF solution. Ultra-high purity H_2 was purchased from Air Products, and D_2 (99.8% D) was purchased from Sigma-Aldrich or Cambridge Isotope Laboratories. $L^{tBu}FeCl$, 25a potassium graphite, 15 [$L^{tBu}Fe(\mu-H)$]₂, 5 [$L^{tBu}Co(\mu-H)$]₂, 8 and $L^{Me}H^{41}$ were prepared by published procedures. $L^{Me}K$ was prepared using the published procedure 42 except Et_2O was used as the solvent instead of toluene.

¹H NMR data were recorded on a Bruker Avance spectrometer at 500 MHz. All resonances in the ¹H NMR spectra are referenced to residual protiated solvents: benzene (7.16 ppm), toluene (2.09 ppm), or THF (3.58 or 1.73 ppm). Resonances were singlets unless otherwise noted. The NMR probe temperature was calibrated using either ethylene glycol or methanol. ⁴³ IR data were recorded on a Shimadzu FTIR spectrophotometer (FTIR-8400S) using a KBr pellet. UV-vis spectra were recorded on a Cary 50 spectrophotometer using Schlenk-adapted quartz cuvettes with a 1 mm optical path length. GC-MS was performed using a Shimadzu QP2010 system with electron impact ionization. Solution magnetic susceptibilities were determined by the Evans method. ⁴⁴ Elemental analyses were obtained from the CENTC Elemental Analysis Facility at the University of Rochester. Microanalysis samples were weighed with a PerkinElmer Model AD-6 Autobalance in a VAC Atmospheres glove box under argon, and their compositions were determined with a PerkinElmer 2400 Series II Analyzer.

¹H NMR data for 1

¹H NMR (C_6D_6 , 25 °C): see Figure 1. Dimeric [L^{tBu}Fe(μ-H)]₂: δ 67.3, 21.8, 19.8, 14.3 (18H, ^tBu), 12.0 (18H, ^tBu), 5.4, -2.6, -7.5, -9.7, -10.6, -14.5, -15.4, -22.0, -27.8, -31.6, -37.2, -51.8, -57.5 ppm. Resonances in the dimers could not be assigned to specific proton environments, because of overlap that prevented accurate integration. Monomeric L^{tBu}FeH: δ 115 (1H, α-H), 41.7 (18H, ^tBu), 11.7 (4H, ⁱPr-CH or aryl *m*-H), -26.5 (12H, ⁱPr CH₃), -109 (4H, ⁱPr-CH or aryl *m*-H), -113 (2H, aryl *p*-H), -122 (12H, ⁱPr CH₃) ppm.

Improved synthesis of 1-D₂ from D₂

The synthesis of $[L^{tBu}Fe(\mu-D)]_2$ relied on the same procedure as the synthesis of **1** from H_2 .⁵ After 16 h, the headspace gases were removed and fresh D_2 was added; this process was repeated twice. **1-D₂** was isolated in 51% yield. ¹H NMR (C_6D_6 , 25 °C): $[L^{tBu}Fe(\mu-D)]_2$: δ 73.0, 23.0, 21.0, 14.3 (18H, ^tBu), 13.0 (18H, ^tBu), 6.4, -1.7, -3.9, -8.0, -10.6, -16.9, -17.5, -25.1, -29.6, -33.0, -43.0, -50.2, -55.0 ppm. $L^{tBu}FeD$: δ 115 (1H, a-H), 41.7 (18H, ^tBu), 11.7 (4H, ⁱPr-CH or aryl m-H), -26.5 (12H, ⁱPr CH₃), -109 (4H, ⁱPr-CH or aryl m-H), -113 (2H, aryl p-H), -122 (12H, ⁱPr CH₃) ppm. The mono-deuterated hydride dimer, **1-D**, was also observed in solution as described above. ¹H NMR (C_6D_6 , 25 °C): δ 70.2, 22.4, 20.5, 14.3 (18H, ^tBu), 12.5 (18H, ^tBu), 5.9, -2.1, -6.6, -7.7, -10.6, -15.7, -16.5, -23.6, -28.7, -32.3, -40.1, -51.0, -56.2 ppm.

Synthesis of L^{Me}FeBr(THF) (2)

 $L^{Me}K$ (3.215 g, 7.039 mmol) was added to a flask with a Teflon pin closure and was dissolved in THF (75 mL) to give a light yellow solution. Anhydrous FeBr₂ (1.532 g, 7.104 mmol, 1.01 equiv) was added to the solution which produced a red reaction mixture. The

flask was sealed, and the mixture was heated at 70 °C for 16 h. The reaction mixture turned yellow in color upon heating. The reaction mixture was cooled to room temperature and filtered through Celite. The yellow solution was concentrated to 20 mL and pentane (100 mL) was added to precipitate additional insoluble material (presumably KBr) which was removed by filtration through Celite. The yellow solution was concentrated to 5 mL which resulted in the formation of a large amount of yellow crystalline solid. The supernatant was decanted and the crystalline yellow solid was washed with pentane (12 mL). The solid was dried under reduced pressure to give 2.282 g of product. Additional product (1.328 g) was collected from subsequent crystallizations of the supernatant by layering with pentane and cooling to -45 °C. The total yield was 3.610 g (82.0%). ¹H NMR (THF-d₈, 22 °C): δ 18.6 (4H, aryl m-H), 4.9 (12H, ⁱPr CH₃), -8.7 (12H, ⁱPr CH₃), -12.3 (br, 4H, ⁱPr-CH), -39.9 (2H, aryl p-H), -67.3 (6H, backbone CH₃), -78.7 (1H, α -H) ppm. $\mu_{\rm eff}$ (THF- d_8 , 22 °C) 5.5(1) $\mu_{\rm B}$. IR (KBr): 3058 (w), 2964 (s), 2928 (s), 1529 (s), 1459 (s), 1437 (s), 1388 (vs), 1316 (s), 1261 (m), 1176 (m), 1100 (m), 1057 (w), 1022 (m), 935 (m), 899 (w), 872 (m), 855 (m), 795 (s), 758 (s) cm⁻¹. UV-vis (THF): 333 (21.2 mM⁻¹cm⁻¹), 433 (sh, \sim 0.9 mM⁻¹cm⁻¹) nm. Anal. Calcd for C₃₃H₄₉N₂FeBrO: C, 63.36; H, 7.91; N, 4.48. Found: C, 63.13; H, 8.10; N, 4.31.

Modified Synthesis of 3

In an Ar-filled glove box, L^{Me}FeBr(THF) (703 mg, 1.12 mmol) was dissolved in THF (20 mL) to give a yellow solution which was added to a small 3-neck round bottom flask with a stir bar. On a vacuum line, H₂ (1 atm) was added to a bulb (297.5 mL, 12.4 mmol, 11 equiv), and the bulb was brought into the glove box. KC₈ (182 mg, 1.35 mmol, 1.2 equiv) was added to a solid addition tube. The 3-neck flask was attached to the volume bulb, a vacuum adapter, and the solid addition piece. The reaction apparatus was degassed under reduced pressure until a small amount of THF had been removed. Then, the apparatus was backfilled with H₂ by slowly opening the stopcock of the volume bulb. KC₈ was added to the stirring solution by inverting the solid addition piece which immediately produced a dark green reaction mixture. The mixture was stirred and turned brown in color after 20 min. After 3 h, the mixture was filtered through Celite, and the volatile components were removed under reduced pressure. The resulting brown residue was dissolved in toluene (35 mL) and was filtered through Celite to remove additional insoluble material. Toluene was removed under reduced pressure to give a brown powder, which was washed with cold pentane (-45 °C, 10 mL). The solid was dried under reduced pressure to give 263 mg of brown powder. The pentane wash was concentrated to 3 mL and was layered with hexamethyldisiloxane (4 mL). Cooling to -45 °C yielded an additional 35 mg of product. The total yield was 298 mg (56.1%). ${}^{1}H$ NMR (C₆D₆, 25 °C): **6** 13.0 (12H + 8H, backbone CH₃ and aryl m-H), 7.1 (24H, iPr CH₃), -24.0 (4H, aryl p-H), -24.8 (br, 24H, iPr CH₃), -55.9 (2H, a-H) ppm. The iPr-CH protons were not observed at this temperature (see text). The deuterated isotopologue of $3, 3-D_2$, was synthesized using the same method with D_2 , in 58.1% yield.

Details of high pressure gas addition apparatus

A Wilmad 522-PV-7 pressure NMR tube with a 5 mm outer diameter (OD) and a maximum pressure rating of 200 psi was used for all high pressure gas experiments. The tube comes

equipped with a PV-ANV valve that is capable of accepting a Swagelok® 1/8" tubing nut and ferrule. PTFE tubing (OD = 1/8") and Swagelok® 1/8" tubing nuts and ferrules were used for all the connections. PTFE tubing was used to connect the gas regulator to a T-shaped splitter which provided two paths. One path connected through PTFE tubing a high pressure gas gauge and the PV-ANV valve on the NMR tube. The second path connected the PTFE tubing to a Swagelok® valve, which was connected to a metal O-ring joint. The other part of the O-ring joint was equipped with a metal to glass flange that had a glass 14/20 female joint. This allowed the apparatus to be attached to the Schlenk line for evacuation.

Mössbauer spectroscopy

Mössbauer data were recorded on a spectrometer with alternating constant acceleration. The minimum experimental line width was 0.24 mm/s (full width at half-height). The sample temperature was maintained constant in an Oxford Instruments Variox cryostat. The γ -ray source was ca. 0.6 GBq 57 Co/Rh. Isomer shifts are quoted relative to iron metal at 300 K. The zero-field spectra were simulated by using Lorentzian doublets.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References and Notes

- 1. Peruzzini, M.; Poli, R., editors. Recent Advances in Hydride Chemistry. Elsevier; New York: 2001.
- 2. Smith JM, Lachicotte RJ, Holland PL. J. Am. Chem. Soc. 2003; 125:15752. [PubMed: 14677959]
- 3. Yu Y, Sadique AR, Smith JM, Dugan TR, Cowley RE, Brennessel WW, Flaschenriem CJ, Bill E, Cundari TR, Holland PL. J. Am. Chem. Soc. 2008; 130:6624. [PubMed: 18444648]
- 4. Vela J, Smith JM, Yu Y, Ketterer NA, Flaschenriem CJ, Lachicotte RJ, Holland PL. J. Am. Chem. Soc. 2005; 127:7857. [PubMed: 15913376]
- 5. Dugan TR, Holland PL. J. Organomet. Chem. 2009; 694:2825.
- 6. Rodriguez MM, Bill E, Brennessel WW, Holland PL. Science. 2011; 334:780. [PubMed: 22076372]
- 7. Holland PL. Acc. Chem. Res. 2008; 41:905. [PubMed: 18646779]
- 8. High-spin diketiminate-cobalt-hydride complexes Ding K, Brennessel WW, Holland PL. J. Am. Chem. Soc. 2009; 131:10804. [PubMed: 19621923] Dugan TR, Goldberg JM, Brennessel WW, Holland PL. Organometallics. 2012; 31:1349.
- Related high-spin diketiminate-nickel-hydride complexes Pfirrmann S, Limberg C, Ziemer B.
 Dalton Trans. 2008:6689. [PubMed: 19153616] Pfirrmann S, Yao S, Ziemer B, Stosser R, Driess
 M, Limberg C. Organometallics. 2009; 28:6855. Gehring H, Metzinger R, Herwig C, Intemann J,
 Harder S, Limberg C. Chem. Eur. J. 2013; 19:1629. [PubMed: 23292919]
- 10. Upon reducing the complex to the iron(I) oxidation state, the bridging hydrides became terminal, and it was possible to crystallographically characterize three-coordinate iron-hydride species Chiang KP, Scarborough CC, Horitani M, Lees NS, Ding K, Dugan TR, Brennessel WW, Bill E, Hoffman BM, Holland PL. Angew. Chem. Int. Ed. 2012; 51:3658.
- 11. (a) Mar La, GN.; Horrocks, WD.; Holm, RH., editors. NMR of Paramagnetic Molecules. Academic Press; New York: 1973. (b) Ming, L-J. "Nuclear Magnetic Resonance of

- ParamagneticMetal Centers in Proteins and Synthetic Complexes,". In: Que, L., editor. Physical Methods in BioinorganicChemistry. University Science Books; Sausalito, CA: 2000. p. 375
- 12. Limberg has recently reported interesting shifts in the hydride signal of a nickel-hydridecomplex with a low-lying paramagnetic excited state Pfirrmann S, Limberg C, Herwig C, Knispel C, Braun B, Bill E, Stösser R. J. Am. Chem. Soc. 2010; 132:13684. [PubMed: 20828129]
- Electron-nuclear double-resonance (ENDOR) spectroscopy can detect NMR transitions inhydrides using EPR. See Brecht M, van Gastel M, Buhrke T, Friedrich B, Lubitz W. J.Am. Chem. Soc. 2003; 125:13075. [PubMed: 14570480] Igarashi RY, Laryukhin M, Dos Santos PC, Lee HI, Dean DR, Seefeldt LC, Hoffman BM. J. Am. Chem. Soc. 2005; 127:6231. [PubMed: 15853328] Kinney RA, Hetterscheid DGH, Hanna BS, Schrock RR, Hoffman BM. Inorg. Chem. 2010; 49:704. [PubMed: 20000748] Jablonskyte A, Wright JA, Fairhurst SA, Peck JNT, Ibrahim SK, Oganesyan VS, Pickett CJ. J. Am. Chem. Soc. 2011; 133:18606. [PubMed: 22035325] Pandelia M-E, Infossi P, Stein M, Giudici-Orticoni M-T, Lubitz W. Chem. Commun. 2012; 48:823. Kinney RA, Saouma CT, Peters JC, Hoffman BM. J. Am. Chem. Soc. 2012; 134:12637. See also ref. 8. [PubMed: 22823933]
- 14. Yu Y, Brennessel WW, Holland PL. Organometallics. 2007; 26:3217. [PubMed: 18725998]
- 15. Smith JM, Sadique AR, Cundari TR, Rodgers KR, Lukat-Rodgers G, Lachicotte RJ, Flaschenriem CJ, Vela J, Holland PL. J. Am. Chem. Soc. 2006; 128:756. [PubMed: 16417365]
- 16. Selected reviews and recent examples Maseras F, Lledós A, Clot E, Eisenstein O. Chem. Rev. 2000; 100:601. [PubMed: 11749246] Pons V, Conway SLJ, Green MLH, Green JC, Herbert BJ, Heinekey DM. Inorg. Chem. 2004; 43:3475. [PubMed: 15154811] Trovitch RJ, Lobkovsky E, Chirik PJ. Inorg. Chem. 2006; 45:7252. [PubMed: 16933926] Weller AS, McIndoe JS. Eur. J. Inorg. Chem. 2007:4411.
- 17. Burgess BK, Lowe DJ. Chem. Rev. 1996; 96:2983. [PubMed: 11848849]
- (a) Leigh GJ. Eur. J. Biochem. 1995; 229:14. [PubMed: 7744024] (b) Sellmann D, Fürsattel A, Sutter J. Coord. Chem. Rev. 2000; 200–202:545.(c) Hoffman BM, Lukoyanov D, Dean DR, Seefeldt LC. Acc. Chem. Res. 2013; 46:587. [PubMed: 23289741]
- 19. Yang Z-Y, Khadka N, Lukoyanov D, Hoffman BM, Dean DR, Seefeldt LC. Proc. Natl. Acad. Sci. USA. 2013; 110:16327. [PubMed: 24062454]
- 20. Iron-hydride complexes are important because the binding site of N₂ to the FeMoco ofnitrogenases is most likely to be iron Seefeldt LC, Dance IG, Dean DR. Biochemistry. 2004; 43:1401. [PubMed: 14769015] Igarashi RY, Dos Santos PC, Niehaus WG, Dance IG, Dean DR, Seefeldt LC. J. Biol. Chem. 2004; 279:34770. [PubMed: 15181010] Hoffman BM, Dean DR, Seefeldt LC. Acc. Chem. Res. 2009; 42:609. [PubMed: 19267458]
- 21. (a) Hebendanz N, Köhler FH, Scherbaum F, Schlesinger B. Magn. Reson. Chem. 1989; 27:798.(b) Theopold KH, Silvestre J, Byrne EK, Richeson DS. Organometallics. 1989; 8:2001.(c) Medforth CJ, Shiau F-Y, La Mar GN, Smith KM. J. Chem. Soc. Chem. Commun. 1991:590.(d) Evans B, Smith KN, La Mar GN, Viscio DB. J. Am. Chem. Soc. 1977; 99:7070. [PubMed: 903538] (e) Horn RR, Everett GW Jr. J. Am. Chem. Soc. 1971; 93:7173.(f) Johnson A, Everett GW Jr. J. Am. Chem. Soc. 1972; 94:1419.(g) Johnson A, Everett GW Jr. J. Am. Chem. Soc. 1972; 94:6397.(h) Wieder NL, Gallagher M, Carroll PJ, Berry DH. J. Am. Chem. Soc. 2010; 132:4107. [PubMed: 20199024] (i) Hetterscheid DGH, Hanna BS, Schrock RR. Inorg. Chem. 2009; 48:8569. [PubMed: 19639973] (j) Beck R, Shoshani M, Johnson SA. There can also be influences on 31P NMR spectra. Angew.Chem. Int. Ed. Engl. 2012; 51:11753. [PubMed: 23086708]
- 22. Heintz RA, Neiss TG, Theopold KH. Angew. Chem. Int. Ed. Engl. 1994; 33:2326.
- 23. Eckert NA, Stoian S, Smith JM, Bominaar EL, Münck E, Holland PL. J. Am. Chem.Soc. 2005; 127:9344. [PubMed: 15984842]
- 24. (a) Andres H, Bominaar EL, Smith JM, Eckert NA, Holland PL, Münck E. J. Am. Chem. Soc. 2002; 124:3012. [PubMed: 11902893] (b) Vela J, Stoian S, Flaschenriem CJ, Münck E, Holland PL. J. Am. Chem. Soc. 2004; 126:4522. [PubMed: 15070362] (c) Cowley RE, Bill E, Neese F, Brennessel WW, Holland PL. Inorg. Chem. 2009; 48:4828. [PubMed: 19397284] (d) Cowley RE, Elhaïk J, Eckert NA, Brennessel WW, Bill E, Holland PL. J. Am. Chem. Soc. 2008; 130:6074. [PubMed: 18419120] (e) Stoian SA, Smith JM, Holland PL, Munck E, Bominaar EL. Inorg. Chem. 2008; 47:8687. [PubMed: 18781730] (f) Eckert NA, Vaddadi S, Stoian S, Lachicotte RJ, Cundari TR, Holland PL. Angew. Chem. Int. Ed. 2006; 45:6868.

25. (a) Smith JM, Lachicotte RJ, Holland PL. Chem. Commun. 2001:1542.(b) Eckert NA, Smith JM, Lachicotte RJ, Holland PL. Inorg. Chem. 2004; 43:3306. [PubMed: 15132641] (c) Stubbert BD, Holland PL, Adhikari D, Mindiola DJ. Inorg. Synth. 2010; 35:38.

- 26. Tonzetich ZJ, Heroguel F, Do LH, Lippard SJ. Inorg. Chem. 2011; 50:1570. [PubMed: 21244036]
- 27. It is particularly important to protect the reaction mixture from arenes. Even traces of benzene in the vapor phase in the glove box led to undesired formation of L^{Me}Fe(C₆H₆). This benzene complexwas previously reported Smith JM, Sadique AR, Cundari TR, Rodgers KR, Lukat-Rodgers G, Lachicotte RJ, Flaschenriem CJ, Vela J, Holland PL. J. Am. Chem. Soc. 2006; 128:756. [PubMed: 16417365]
- 28. (a) Legzdins P, Phillips EC, Yee VC, Trotter J, Einstein FWB, Jones RH. Organometallics. 1991; 10:986.(b) Thomas WA. Annu. Rev. NMR Spectrosc. 1968; 1:43.
- 29. In an independent experiment, a mixture of Et₃SiH and Et₃SiD was used to make a statistical mixture of isotopologues of [L^{tBu}Co(μ-H)]₂, and the peaks for the singly deuterated isotopologue was distinguishable by ¹H NMR spectroscopy. See Supporting Information.
- (a) Beinert H, Holm RH, Münck E. Science. 1997; 277:653. [PubMed: 9235882] (b) Münck, E. "Aspects of ⁵⁷FeMössbauer Spectroscopy," in. In: Que, L., editor. Physical Methods in Bioinorganic Chemistry. UniversityScience Books; Sausalito, CA: 2000. p. 287
- 31. Franke O, Wiesler BE, Lehnert N, Peters G, Burger P, Tuczek F. Z. Anorg. Allg. Chem. 2006; 632:1247.
- 32. This phenomenon is distinct from IPR (isotopic perturbation of resonance) effects indiamagnetic complexes. For an example of IPR in a hydride-H₂ complex: Heinekey DM, Oldham WJ. J. Am. Chem. Soc. 1994; 116:3137.
- 33. Bartell LS, Roskos RR. J. Chem. Phys. 1966; 44:457.
- 34. See ref 16c and references therein.
- 35. (a) Darensbourg MY, Lyon EJ, Smee JJ. Coord. Chem. Rev. 2000; 206–207:533.(b) Tard C, Pickett CJ. Chem. Rev. 2009; 109:2245. [PubMed: 19438209]
- 36. (a) Zhao X, Georgakaki IP, Miller ML, Yarbrough JC, Darensbourg MY. J. Am.Chem. Soc. 2001; 123:9710. [PubMed: 11572707] (b) Zhao X, Georgakaki IP, Miller ML, Mejia-Rodriguez R, Chiang C-Y, Darensbourg MY. Inorg. Chem. 2002; 41:3917. [PubMed: 12132916]
- 37. Vignais PM. Coord. Chem. Rev. 2005; 249:1677.
- 38. Georgakaki IP, Miller ML, Darensbourg MY. Inorg. Chem. 2003; 42:2489. [PubMed: 12691553]
- (a) Carriker JL, Wagenknecht PS, Hosseini MA, Fleming PE. J. Mol. Catal. A: Chem. 2007;
 267:218.(b) Kovács G, Nádasdi L, Joó F, Laurenczy GCR. Acad. Sci. Paris, Sér. IIc: Chim. 2000;
 3:601.(c) Kovács G, Nádasdi L, Laurenczy G, Joó F. Green Chem. 2003;
 5:213.(d) Kovács G, Schubert G, Joó F, Pápai I. Organometallics. 2005;
 24:3059.
- 40. Wang N, Wang M, Liu J, Chen L, Sun L. Inorg. Chem. 2009; 48:11551. [PubMed: 20000647]
- 41. Budzelaar PHM, Moonen NNP, De Gelder R, Smits JMM, Gal AW. Eur. J.Inorg. Chem. 2000:753.
- 42. Clegg W, Cope EK, Edwards AJ, Mair FS. Inorg. Chem. 1998; 37:2317. [PubMed: 11670390]
- (a) Kaplan ML, Bovey FA, Cheng HN. Anal. Chem. 1975; 47:1703.(b) Ammann C, Meier P, Merbach AE. J. Magn. Reson. 1982; 46:319.
- 44. (a) Baker MV, Field LD, Hambley TW. Inorg. Chem. 1988; 27:2872.(b) Schubert EM. J. Chem. Educ. 1992; 69:62.

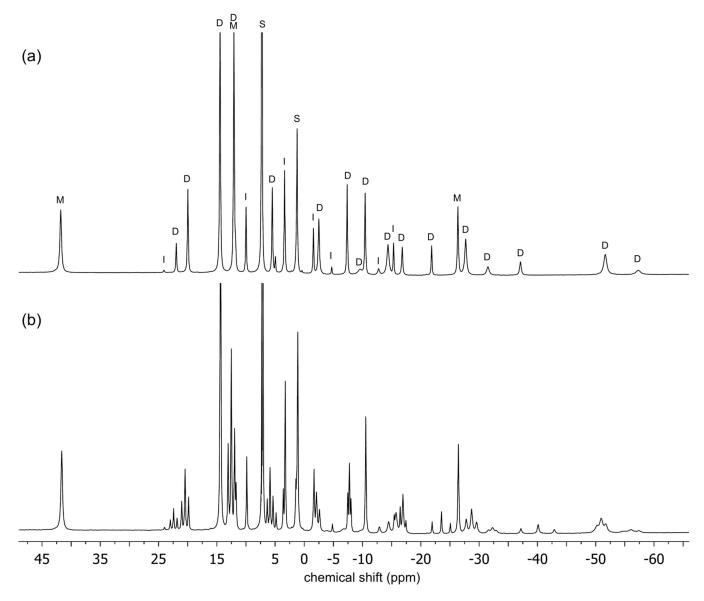


Figure 1. (a) 1H NMR spectrum of $[L^{tBu}Fe(\mu-H)]_2$ (1) in C_6D_6 . This particular sample has a 7% impurity of the oxo complex $\{L^{tBu}Fe\}_2(\mu-O)$. Peaks for the dimer are marked with D, for monomers with M, for the oxo impurity with I, and for solvent and solvent impurities with S. (b) 1H NMR spectrum from mixing equimolar amounts of $[L^{tBu}Fe(\mu-H)]_2$ (1) and $[L^{tBu}Fe(\mu-D)]_2$ (1-D₂) in C_6D_6 for 45 min. All three isotopologues of the dimer (H/H, H/D, and D/D) are visible in (b), as several groups of three nearby peaks in a statistical 1:2:1 ratio. Only the parts of the spectra from δ 45 to -70 ppm are shown for clarity.

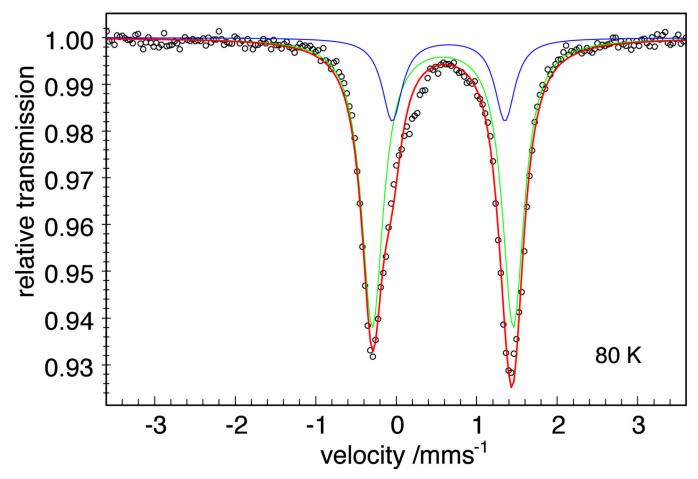
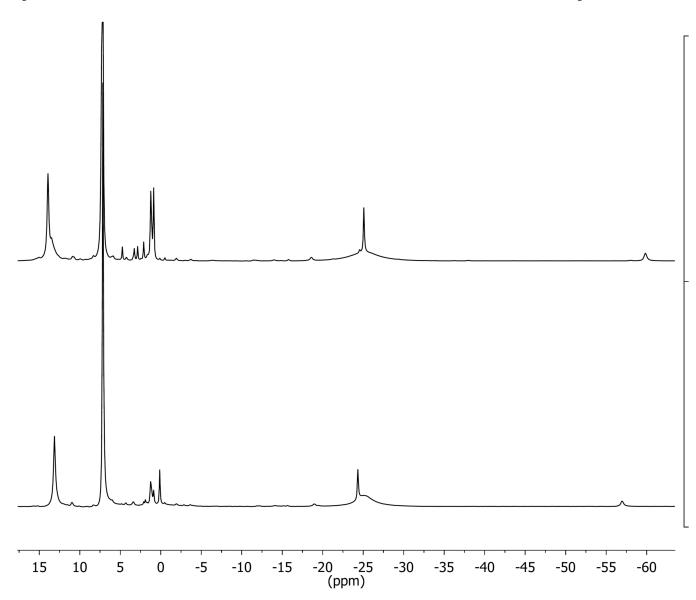


Figure 2. Zero-field Mössbauer spectrum of $[L^{tBu}Fe(\mu-D)]_2$ (1-D₂) recorded at 80 K. The signal with $\delta=0.58$ mm/s and $|E_Q|=1.74$ mm/s accounted for 79% of the sample. The blue line represents the contribution of the oxo impurity, the green line represents the contribution of 1-D₂, the red line represents the sum, and the black circles are the data.



 $\label{eq:Figure 3.1} \begin{subarray}{l} Figure 3. \\ {}^{l}H \ NMR \ spectra \ of \ [L^{Me}Fe(\mu-H)]_2 \ (3) \ (bottom) \ and \ [L^{Me}Fe(\mu-D)]_2 \ (3-\textbf{D_2}) \ (top) \ in \ C_6D_6. \end{subarray}$

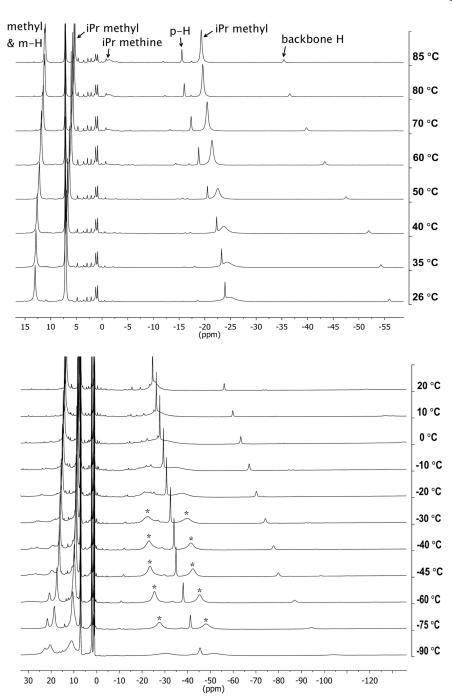


Figure 4.(a) Variable-temperature ${}^{1}H$ NMR spectra of $[L^{Me}Fe(\mu-H)]_{2}$ (3) between 26 °C and 85 °C in $C_{6}D_{6}$. (b) Variable temperature ${}^{1}H$ NMR spectra of 3 between -90 °C and 20 °C in toluene- d_{8} . The asterisks indicate the resonance that splits with a coalescence temperature of 0 °C.

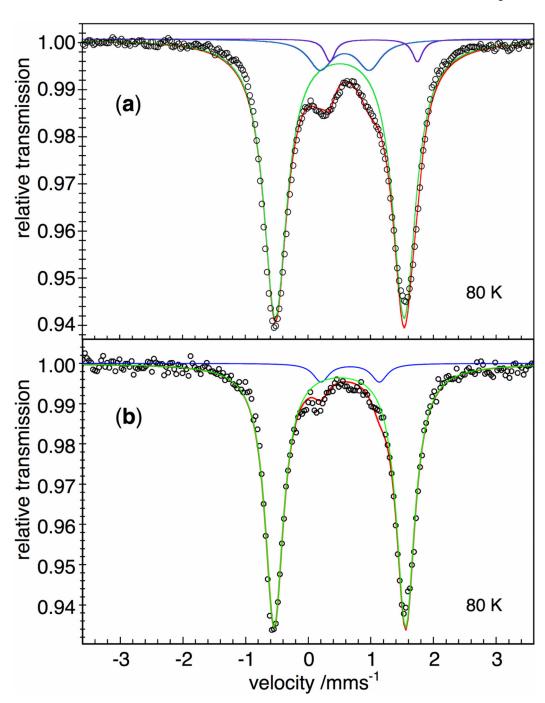


Figure 5. (a) Mössbauer spectrum of $[L^{Me}Fe(\mu-H)]_2$ (3) (b) Mössbauer spectrum of $[L^{Me}Fe(\mu-D)]_2$ (3- $\mathbf{D_2}$). Both spectra were recorded at 80 K, with zero field. The black circles are the data and the red lines represent the sums of a major doublet for 3 (green) and impurities (blue, purple) that are discussed in the Supporting Information.

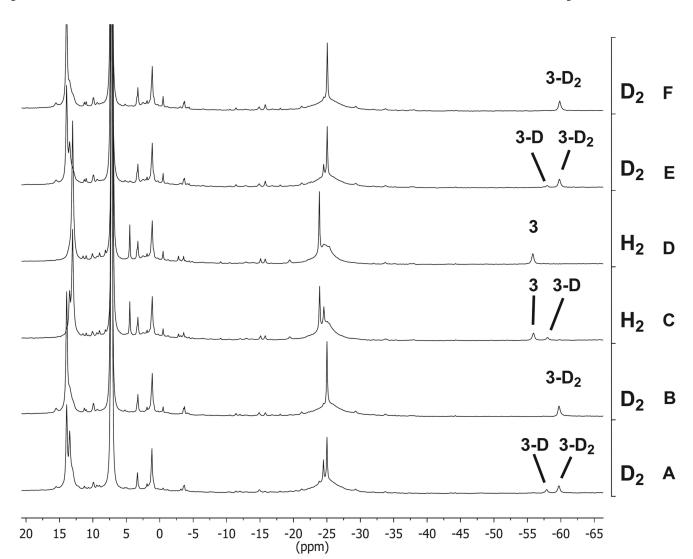


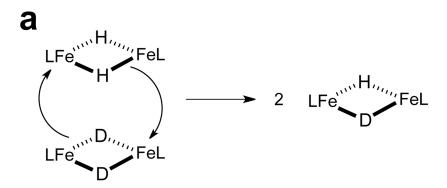
Figure 6. 1 H NMR spectra of $[L^{Me}Fe(\mu-H)]_{2}$ (3), $\{L^{Me}Fe\}_{2}(\mu-H)(\mu-D)$ (3-D), and $[L^{Me}Fe(\mu-D)]_{2}$ (3-D₂) isotopologues in $C_{6}D_{6}$ during gas exchange. The columns on the right indicate the order and type of gas that was added to give the observed spectrum.

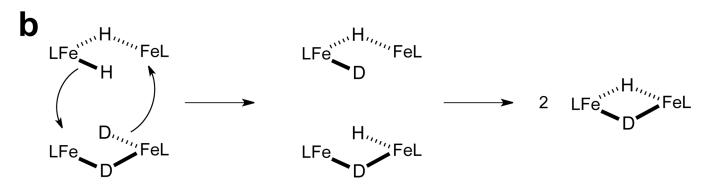
D₂ symmetry crystallographically observed

D_{2h} symmetry transition state for fluxional process

Scheme 1.

In the X-ray crystal structure of $[L^{Me}Fe(\mu-H)]_2$ (3), the hydride ligands are oriented such that the geometry at each iron is between tetrahedral and square planar, and the N_2C_3Fe planes are perpendicular.^{4,14} The idealized D_2 symmetry is consistent with the low-temperature 1H NMR spectrum, and its fluxional behavior could involve interconversions of two D_2 structures through a D_{2h} transition state.

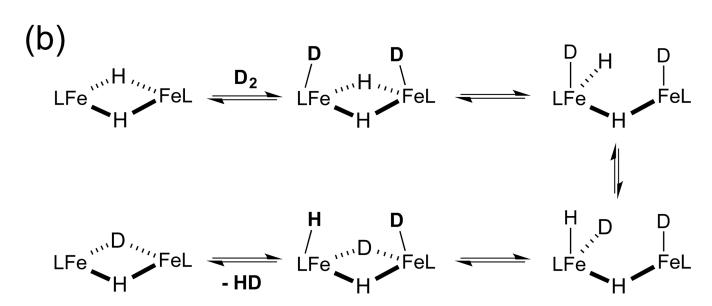




Scheme 2.

Possible mechanisms for hydride ligand exchange between isotopologues in 3. Dissociation of 3 into monomers is inconsistent with earlier kinetic studies on the reaction of boranes with 3.

LFe
$$\xrightarrow{H}$$
 FeL $\xrightarrow{D_2}$ LFe \xrightarrow{H} FeL $\xrightarrow{D_2}$ LFe \xrightarrow{H} FeL $\xrightarrow{D_1}$ FeL \xrightarrow{H} FeL



Scheme 3. Proposed mechanisms for hydride ligand exchange with D_2 in 3.

Chart 1. Diagram of the Fe $_2(\mu$ -H) $_2$ core, and three β -diketiminate ligands that form crystallographically characterized complexes with this core.

Table 1

Times for exchange of hydride ligands between isotopologues, and for exchange of hydride with 1 atm of H_2/D_2 gas, giving qualitative times to reach equilibrium. Each solution was shaken for the duration of the experiment.

| Compound | Isotopologue exchange | Gas exchange |
|--|--------------------------|--------------|
| $[L^{tBu}Fe(\mu-H)]_2$ (1) | < 45 min | 2 d |
| $[L^{Me}Fe(\mu\text{-}H)]_2(\textbf{3})$ | 2 h | < 1 min |
| $[L^{tBu}Co(\mu\text{-}H)]_2$ | none | N/A |