

## **HHS Public Access**

Author manuscript

Cartilage. Author manuscript; available in PMC 2016 January 01.

Published in final edited form as:

Cartilage. 2015 January; 6(1): 30-44. doi:10.1177/1947603514554992.

# Management of Osteoarthritis with Avocado/Soybean Unsaponifiables

Blaine A. Christiansen, PhD<sup>1</sup>, Simi Bhatti<sup>2</sup>, Ramin Goudarzi, PharmD<sup>3</sup>, and Shahin Emami, PharmD, PhD, HDR<sup>4</sup>

Blaine A. Christiansen: bchristiansen@ucdavis.edu; Simi Bhatti: simi@formulationtech.com; Ramin Goudarzi: ramin.goudarzi@pharminusa.com

<sup>1</sup>University of California-Davis Health System, Department of Orthopaedic Surgery, Lawrence J. Ellison Musculoskeletal Research Center, 4635 2nd Ave, Suite 2000, Sacramento, CA 95817, Phone: 916-734-3974, Fax: 916-734-5750

<sup>2</sup>Formulation Technology Inc. 571 Armstrong Way Oakdale, CA 95361 Ph: 209- 847-0331 Fax: 209-847-1975

<sup>3</sup>Pharmin USA, LLC, 2375 Lindbergh Avenue San Jose, CA 95128 Phone: 408-326-2378, Fax: 408-998-9777

<sup>4</sup>Corresponding author: shahin.emami1@gmail.com

#### **Abstract**

Osteoarthritis (OA) is a painful and life-altering disease that severely limits the daily activity of millions of Americans, and is one of the most common causes of disability in the world. With obesity on the rise and the world's population living longer, the prevalence of OA is expected to increase dramatically in the coming decades, generating burdensome socioeconomic costs. This review summarizes current pharmaceutical, non-pharmaceutical, and prospective new treatments for OA, with primary focus on the dietary supplement Avocado/Soybean Unsaponifiables (ASU). ASU modulates OA pathogenesis by inhibiting a number of molecules and pathways implicated in OA. Anticatabolic properties prevent cartilage degradation by inhibiting the release and activity of matrix metalloproteinases (MMP-2,3,13) and increasing tissue inhibitors of these catabolic enzymes (TIMP-1). ASU also inhibits fibrinolysis by stimulating the expression of plasminogen activator inhibitor (PAI-1). Anabolic properties promote cartilage repair by stimulating collagen and aggrecan synthesis via inhibition of inflammatory cytokines such as IL1, IL6, IL8, TNF, ERK, and PGE2. Chondroprotective effects are mediated by correcting growth factor abnormalities, increasing TGFβ while decreasing vascular endothelial growth factor (VEGF) in synovial fluid. ASU also inhibits cholesterol absorption and endogenous cholesterol biosynthesis, which mediate reactive oxygen species pathology in chondrocytes. At the clinical level, ASU reduces pain and stiffness while improving joint function, resulting in decreased dependence on analgesics.

#### Keywords

Osteoarthritis; cartilage; dietary supplements; avocado soybean unsaponifiables (ASU); Arthrocen

#### Introduction

Osteoarthritis (OA) is a chronic synovial joint disease, characterized by two main features: 1) progressive damage of articular cartilage, bone remodeling and new bone formation (osteophytes and subchondral bone sclerosis), and 2) synovial inflammation and fibrosis of ligaments, tendons, menisci, and capsules. All joints may be affected, but the most commonly involved are knees, hands, and hips (Fig. 1). While chronic OA used to be regarded as a "wear and tear disease," researchers now believe that low grade inflammation and growth of blood vessels and nerves from the subchondral bone into articular cartilage, as well as metabolic disorders, play a major role in disease pathology. [1-4] Patients with OA suffer from pain, inflammation, and limited joint function. Pharmacological interventions are mostly palliative, focusing on alleviation of symptoms or slowing disease progression until damaged hip or knee joints are eventually replaced. [5-10] Women are more severely impacted than men by knee osteoarthritis.<sup>[11]</sup> Differences in knee anatomy (narrower femurs, thinner patellae, larger quadriceps angles, and differences in tibial condylar size), previous knee trauma, genetic and hormonal influences may play a role. Other factors like age, and obesity, are also common factors. In general, women present for treatment in more advanced stages of osteoarthritis and have more debilitating pain than men. Women also have less cartilage volume and greater cartilage wear, and overall differences in mechanical alignment.

## **Current Pharmacologic Therapies**

Pain medications currently used to treat the symptoms of OA include acetaminophen, topical capsaicin, topical and oral nonsteroidal anti-inflammatory drugs (NSAIDs i.e. naproxen and ibuprofen), and the synthetic opioids tramadol and codeine. However, each of these therapies has potential drawbacks that may limit their widespread use. Analgesics can be addictive, while acetaminophen can have serious side effects, such as kidney and liver damage. [12] Patients who do not respond to acetaminophen may be prescribed NSAIDs. Treatment with NSAIDs, which inhibit cyclooxigenaeses (COX1 and COX2) thereby blocking prostaglandin synthesis, improves quality of life and decreases pro-inflammatory cytokines including interleukin 6 (IL6), vascular endothelial growth factor (VEGF), and tumor necrosis factor alpha (TNFa) in synovial fluid and mitogen-activated protein kinases (MAPKs) in knee OA.[13] However, NSAIDs can also cause serious side effects, including upper gastrointestinal (GI) toxicity (dyspepsia, ulcers, perforation, obstructions and bleeding) and liver dysfunction. As such, they are typically prescribed for the shortest possible duration at the lowest effective dose. To reduce the risk of these upper GI complications, the US FDA has approved the use of the NSAID HZT501 (Duexis®), a drug containing 800 mg ibuprofen, in combination with 26.6 mg famotidine, a histamine H2receptor antagonist. [14] Alternatively the NSAID celecoxib has less risk of upper GI complications by selectively inhibiting the isoenzyme COX-2, which is specific to inflamed

tissue, versus COX-1, which is constitutive in many tissues including the GI tract. It should be noted that daily treatment with celecoxib is more effective in patients with normal BMI than obese patients.<sup>[15]</sup>

Intra-articular injection of corticosteroids (GC) is recommended to relieve inflammation and pain in OA joints. However, GC injections are short acting, prone to adverse side effects, and have limited disease-modifying effects. For patients with knee OA, viscosupplementation with hyularonin may be used to replaces shock absorbing and lubricant material in the joint fluid, but the effects are similarly short-lived.

## **Current Non-Pharmacologic Therapies**

Currently, guidelines for OA management are available from numerous organizations, including the American Academy of Orthopedic Surgeons (AAOS), the American College of Rheumatology (ACR), the American Geriatrics Society (AGS), the American Pain Society (APS), and the Osteoarthritis Research Society International (OARSI) in the US, and the European League Against Rheumatism (EULAR) and the United Kingdom's National Institute for Health and Clinical Excellence (NICE) in Europe. Collectively, these guidelines reflect the experience of physicians across a variety of medical disciplines. While all generally use the same data sources (i.e., evidence-based research, expert opinion, patient experience, and cost-effectiveness analysis), they differ in focus. For instance, the AAOS and AGS guidelines reflect the perspective of specialists in orthopaedic surgery, geriatrics, and pain management, while the EULAR and OARSI guidelines primarily emphasize the findings of experts in rheumatology. The NICE guidelines are developed jointly by physicians and other healthcare professionals working in conjunction with a range of clinical researchers. In addition, the scope varies, with some guidelines (e.g., AAOS, ACR, EULAR, and OARSI) addressing specific types of OA (i.e., knee, hip, or hand) and others (e.g., AGS, APS, and NICE) addressing OA more generally. As such, recommendations can vary widely; for instance, guidelines for use of NSAIDS. [10, 16-19] Recommended nonpharmacologic interventions range between therapeutic exercises, patient education, transcutaneous electrical nerve stimulation, acupuncture, orthotics and insoles, heat and cryotherapy, patellar tapping, and weight control. In an effort to evaluate these varying guidelines, The Appraisal of Guidelines Research and Evaluation (AGREE II) scored 17 clinical practice guidelines (CPGs) including EULAR, NICE, OARSI, AAOS, and ACR, on six different measures-- D1: scope and purpose, D2: stakeholder involvement, D3: rigor of development, D4: clarity and presentation, D5: applicability, and D6: editorial independence. [20] The general clinical management recommendations tended to be similar among high-quality CPGs, although interventions addressed varied. Non-pharmacological management interventions were superficially addressed in more than half of the selected CPGs.

## **Prospective New Treatments**

New non-invasive, disease-modifying therapies for OA are lacking and needed by millions of patients. A number of prospective new treatments targeting proinflammatory mediators,

cytokines, bone turnover, angiogenic and neurogenic factors are being investigated, with varying success in clinical trials and clinical use.<sup>[21]</sup>

Interleukin 1 (IL1) may prove a effective target, as IL1 induces matrix metalloproteinase (MMP) production, resulting in the degradation of aggrecan and other matrix constituents. IL1 also induces high levels of COX2 and prostaglandin E2 (PGE2), which may explain the pain associated with OA degeneration. [22] The drug diacerein, an inhibitor of IL1, may modify both disease symptoms and disease-structure in OA. Oral diacerein has proven effective in reducing pain, although evidence from clinical trials and scientific literature suggest that the effectiveness in OA is weak. It can be used in conjunction with NSAIDs or viscosupplementation therapies for additive effects due to its alternative mechanism of action. The most common side effects of diacerein are gastrointestinal, such as diarrhea and changes in the color of urine. Meanwhile, the IL1-receptor antagonists, anakinra and orthokin are reported to improve Western Ontario and McMaster Universities Arthritis Index (WOMAC) scores, [23–25]. In addition, the IL1 $\beta$  antibody "gevokizumab" is in phase II clinical trials for safety and biological activity in the treatment of hand OA. [26, 27]

Nerve growth factor (NGF) has also been recognized as an important mediator of chronic pain in OA. Tanezumab, a monoclonal antibody against  $\beta$ -NGF receptor tyrosine kinase (TrkA) inhibits NGF action and reduces pain in patients. [28] Two randomized phase III clinical trials indicate that tanezumab provides superior pain relief while improving physical function and global disease assessment scores in patients with painful hip OA. [27, 29–33] While in most cases tanezumab is well tolerated, the unexpected occurrence of rapid destructive arthropathies suggests there may be safety issues. Alternatively, using the drug adalimumab to inhibit TNF $\alpha$ , which upregulates  $\beta$ -NGF, does not improve global disease assessment scores in OA of the hand. [27, 34]

Several studies have explored therapies aimed at modifying bone turnover for treatment of osteoarthritis. Strontium ranelate (SrRa), an element similar to calcium, is easily taken up by the body and incorporated into bones in place of calcium. SrRa is currently indicated for the prevention of fracture in severe osteoporosis. The SEKOIA (SrRa Efficacy in Knee OsteoarthrItis triAl) trial, a 3-year randomized, double-blind, placebo-controlled trial, evaluated the efficacy, safety and disease-modifying effects of SrRa given at 1-2 g/day in patients with knee OA. MRI data indicate that SrRa significantly reduced cartilage volume loss and bone marrow lesion progression. Symptoms also improved in terms of pain and physical function after 6 and 12 months respectively, [35] while treatment was deemed safe and well tolerated. These data indicate that SrRa could be a promising new symptom and disease-modifying treatment for OA. Additionally, there is a need for further investigations to establish the optimal dosage and to better clarify the mechanism of action of SrRa in OA. [35–37] Several clinical studies have investigated the effects of anti-resorptive therapies such as bisphosphonates on OA symptoms. A study by Carbone et al. found that alendronate use in OA patients decreased bone abnormalities and attenuated knee pain, yet cartilage degeneration was still present in the MRI scans of treated patients.<sup>[38]</sup> Spector et al. determined that risedronate use led to significant improvements in WOMAC scores and preservation of knee joint space compared to placebo in a one-year randomized control trial involving patients with moderate OA. [39] However, a 2-year randomized control trial of

risedronate treatment revealed contradictory results, with no significant improvement of WOMAC score or joint space retention in the knee.<sup>[40]</sup> Similarly, Nishii *et al.* observed no inhibition of osteoarthritis progression in treated hip OA patients after 2 years of ALN treatment.<sup>[41]</sup> Therefore, in spite of the growing body of clinical work investigating the subject, no definitive conclusion can be reached on the practicality of using bisphosphonates to treat patients with OA.

Anti-depressants have shown promising preliminary results for treatment of pain associated with OA by increasing serotonin levels in the brain. Serotonin-norepinephrine reuptake inhibitors duloxetine (Cymbalta) and milnacipran significantly improve pain in OA, [27, 42]. An open-label trial also suggested analgesic effectiveness of methotrexate, and anti-inflammatory drug that acts by inhibiting the metabolism of folic acid, demonstrating that up to 20 mg/week for 6 months achieved OARSI responder criteria in knee OA and warranted a randomized controlled trial. [43]

Other treatments are aimed at improving disease pathology by building cartilage. The small molecule kartogenin was identified in an image-based high-throughput screen to promote chondrocyte differentiation. It shows chondroprotective effects in vitro and is efficacious in two animal models of OA. Kartogenin induces chondrogenesis by disrupting the interaction between filamin A and the transcription factor core-binding factor b subunit (CBF\$), thereby altering CBFβ-RUNX1 and possibly RUNX2 transcriptional programs.<sup>[44]</sup> Autologous injection of platelet-rich plasma (PRP) has been used to stimulate cartilage repair and healing in OA patients, [27, 45, 46] but the presence of other growth factors in PRP may be problematic. Furthermore, Bone morphogenic protein 7 (BMP7), FGF-8, and botulium toxin A (BoNT-A), are used in the treatment of knee OA. [47] BoNT-A has an analgesic effect by temporarily suppressing acetylcholine secretion at pre-synaptic nueuromuscular junctions, and appears to be effective and safe for the management of advanced knee OA. However, these results cannot be generalized to patients with mild knee joint pain or nonspecific soft tissue pain in the knee joint region. Further research is necessary to investigate possible complications such as aggravation of infection, effect on muscle strength, and neuropathic joint degeneration.

Current nonsurgical and reconstructive surgical therapies are unsuccessful in reversing OA. Recently a phase I trial was reported in which chondrocytes were modified via intra-articular DNA injection to produce TGF1 in patients with advanced knee OA, [48]. Intra-articular injection of adipose-derived stem cell (ADSC) therapy in the new European program is also under investigation. [49] ADSC induced the release of trophic factors that exerted anti-inflammatory effects on both synoviocytes and chondrocytes, with no MMP1, MMP3 or MMP13 production, suggesting safe and effective use of ADSCs for clinical applications. However, both treatments need proof-of-concept studies in larger patient populations. Alternatively, intra-articular injection of human mesenchymal stem cells can lead to articular cartilage protection through the SDF-1/CXCR4 axis. [50–54]

## **Dietary Supplements**

Natural products can be safer than prescription medications with less undesirable side effects. Dietary supplements including avocado soybean unsaponifiables (ASU), chondroitin sulfate, hyaluronan and glucosamine sulfate have been reported to modify EULAR symptoms for the treatment of OA.<sup>[55, 56]</sup> They are used to treat mild to moderate pain, and alleviate symptoms to reduce the consumption of NSAIDs.

Several trials for chondroitin sulfate, glucosamine sulfate, and hyaluronan  $(C_{14}H_{21}NO_{11})^n$  are in process. [56, 57] Chondroitin sulfate, glucosamine sulfate and hyaluronan are building blocks for proteoglycan synthesis, and major constituents of the ECM in cartilage and synovial fluid. [58] They are produced by chondrocytes and syonivocytes or obtained through diet. [59–65] Hyaluronan and hyaluronic acid (Hyalgan® hylan-GF20 / Synvisc®) can be injected into the knee joint of patients with oateoarthritis who cannot tolerate NSAIDs, or are awaiting joint surgery. [66] A recent report indicates that viscosupplementation with Hylan-GF20 slows type II collagen degradation and joint inflammation in patients with OA. [67] However Hylan-GF20 was not present in granulomas, an indicator of inflammation, raising the question of clinical significance in pain reduction, [68]. Also, viscosupplementation with hyaluronic acid itself does not significantly improve disease outcome, and little is known about long-term effects.

The efficacy of glucosamine and/or chondroitin in treating knee OA pain was evaluated in the Glucosamine/Chondroitin Arthritis Intervention Trial (GAIT), funded by the National Center for Complementary and Alternative Medicine (NCCAM) and the National Institute of Arthritis and Musculoskeletal and Skin Diseases (NIAMS). Patients were treated daily for 24 weeks with glucosamine alone (1500 mg), chondroitin sulfate alone (1200 mg), glucosamine and chondroitin sulfate combined (same doses), a placebo, or celecoxib (200 mg), which served as a positive control. While there were no statistically significant differences between any of the experimental treatments and the placebo overall, patients with moderate-to-severe pain given both glucosomine and chondroitin sulfate did show improvement (79% experienced pain reduction versus 54% for placebo). Because of the small size of this subgroup these findings should be considered preliminary and need to be confirmed in further studies. [61, 69–71]

Glucosamine does not appear to slow arthritis progression over the long term, and has many potential complications. The most common adverse effects are epigastric pain or tenderness, heartburn, diarrhea, and nausea. Glucosamine also may cause allergic reactions in patients with seafood allergies, as a product of lobster, crab, and shrimp shells. Glucosamine may interact with various pharmaceuticals, such as warfarin (Coumadin) and diabetes medications, dangerously modifying their efficacy.

Similarly, chondroitin sulfate appears not to provide meaningful benefit for patients with OA, and their combination has not proven effective for either pain management or functional improvement. OARSI and NICE no longer recommend the use of glucosamine or chondroitin sulfate alone, or in combination, if no effects are observed after six months or radiographic changes are marginal. [55, 61, 62, 72]

A network meta-analysis of 10 trials in 3803 patients by Juni and collaborators in 2012 found no clinically significant improvements in OA pain alleviation or JSN parameters with glucosamine, chondroitin, or combined treatment compared to placebo. Despite these results, many patients believe otherwise, potentially due to the natural course of disease, regression to the mean or the placebo effect. The authors conclude that such patients should be permitted to use these supplements if they cover the cost themselves, since neither of these preparations was found to be dangerous.<sup>[73]</sup>

SierraSil is a dietary supplement marketed for joint pain relief that is derived from the mineral-rich clay found in the high Sierra Mountains in the US. Clinical trial testing short-term efficacy of SierraSil at doses of 2 and 3g per day failed to show sustained benefits over placebo, and iron toxicity has been reported.<sup>[74]</sup>

Some OA patients experience pain relief from topical creams containing capsaicin, the active component of chili peppers. However, use of these creams may introduce side effects such as burning, stinging, and redness of the skin and eyes.<sup>[10]</sup>

## **Avocado and Soybean Unsaponifiables**

Avocado/soybean unsaponifiables (ASUs) are natural vegetable extracts made from avocado and soybean oils, consisting of the leftover fraction (approximately 1%) that cannot be made into soap after saponification chemical reaction. ASU is composed of one third avocado and two thirds soybean unsaponifiables (A1S2U). The major components of ASU are phytosterols  $\beta$ -sitosterol, campesterol, and stigmasterol, which are rapidly incorporated into cells. ASU is a complex mixture of many compounds including fat-soluble vitamins, sterols, triterpene alcohols and possibly furan fatty acids. The identity of the active component(s) remains unknown. The sterols content of ASU preparations are the primary contributors to biological activity in articular chondrocytes<sup>[75]</sup>. Preclinical *in vitro* and *in vivo* studies have demonstrated that ASUs have beneficial effects on OA.<sup>[76–90]</sup>

ASU possesses chondroprotective, anabolic and anticatabolic properties. It inhibits the breakdown of cartilage and promotes cartilage repair by inhibiting a number of molecules and pathways implicated in OA (Fig. 2). ASU stimulates the synthesis of collagen and aggrecan by inhibiting inflammatory cytokines such as IL1, IL6, IL8, TNF, and PGE2 through modulation of NF-kappaB.<sup>[91–94]</sup> The combination of ASU and epigallocatechin gallate (EGCG, a major component of green tea catechins), affect an array of inflammatory molecules including expression of COX-2 and production of PGE2 in chondrocytes.<sup>[95]</sup> COX-2 regulates the production of PGE2; both are mediators involved in the process of cartilage breakdown. ASU also inhibits the release and activity of collagenase (MMP2) and stromelysin 1 (MMP3) in cultured chondrocytes,<sup>[77, 96]</sup> increases tissue inhibitors of metalloproteinases (TIMP-1),<sup>[79, 97]</sup> and inhibits IL-1 induced ERK but not p38 or JNK in chondrocytes *in vitro*.<sup>[86]</sup>

*In vitro* studies show that ASU inhibits fibrinolysis by stimulating the expression of plasminogen activator inhibitor (PAI-1). [98] PAI-1 inhibits tissue plasminogen activator and urokinase (uPA), thereby blocking plasminogen activation and inhibiting fibrinolysis (the physiological breakdown of blood clots). This fibrinolytic and tissue destructive proteinase

cascade may play a role in OA joint inflammation via altered expression of uPA receptors.<sup>[99]</sup>

ASUs alters growth factor levels implicated in OA pathogenesis, increasing TGF- $\beta1$  and TGF- $\beta2$  in the canine knee joint fluid, [100] to repair cartilage and decreasing VEGF, which is markedly elevated in synovial fluid of patients. [13, 97] In a study of implant osseointegration in rat tibiae, ASU administration improved markers of bone growth, including bone morphogenic protein 2 (BMP-2) and transforming growth factor beta 1(TGF- $\beta1$ ), though histomorphometric analysis of implant osseointegration was only slightly improved. [101] ASU also inhibits cholesterol absorption and endogenous cholesterol biosynthesis. [102] 60% of patients with OA exhibit high levels of oxidized low-density lipoproteins (oLDL) in serum, which mediates reactive oxygen species (ROS) activity in chondrocytes and OA pathology. [103, 104] Treating patients with a daily dose of 300 mg ASU for 3 months decreased oLDL levels. [105]

At the clinical level, ASU reduces pain and stiffness while improving function in joints, resulting in decreased dependence on analgesics. ASU efficacy and safety during and post treatment have been assessed in various randomized, double blind, multicenter trials in patients with symptomatic knee or hip OA. Two studies conducted over a 3 month period report that standard treatment with 300 mg/day of ASU improved indices of pain, stiffness, and physical function, as measured by WOMAC, and decreased analgesic drug demanded in patients with OA.[106–109] A third trial conducted over 6 months reports similarly improved function compared to placebo, measured by the Lequense Functional Index, with persistent effects after termination of treatment.<sup>[108]</sup> In a 6-month trial on patients with femorotibial gonarthrosis, ASU was as effective as 400 mg of chondroitin sulfate three times per day, as measured by WOMAC. [110] Most recently a three-year randomized trial on patients with hip OA, performed under the ACR criteria (minimum of 1-4 mm hip JSW on the pelvic radiographs), reports excellent safety, but no significant reduction in the mean rate of JSN after one year. However, analyzing the results under different parameters reveals a significant 20% reduction in the rate of progression in patients with severe hip OA (p =0.04), indicating a potential structure modifying effect of ASU,[111] as confirmed in the ERADIAS study. In a clinical trial of patients with hip OA, the effects of ASU treatment over 3 years were evaluated by radiography to identify joint pathology and disease progression on the structural level. Although JSN was not statistically significant between ASU and placebo treatment, secondary analysis of disease progression, measured by JSN (0.5 mm) or total hip replacements, indicated 20% improvement with ASU (42.2% versus 51.4% of placebo group, p=0.054). Computerized image analysis also showed significant histological differences not detectable by traditional scoring methods. [112] In sheep, ASU treatment following cartilage insult improved articular integrity, as measured by toluidine blue staining, after 6 months compared to untreated animals. These improvements were the result of decreased catabolism and increased anabolism of cartilage by ASU.<sup>[78]</sup> Indeed, ASU reduces inflammation mediated cartilage degradation by reducing IL-1, PGE2 and MMP-3 production, while also inducing proteoglycan, non-collagenous protein (NCP) and collagen synthesis within 72 hours of administration to bovine cells in culture.<sup>[75]</sup> A recent study in patients with nonspecific dorsalgia demonstrated analgesic effect of Piascledine with positive outcome after one month. [113] However, a randomized, double blind, placebo-

controlled clinical trial carried out in 14 obese adult volunteers over 3 months reports no significant effect on these parameters, as measured by hyperglycemic-hyperinsulinemic clamp technique.<sup>[114]</sup>

Four double-blind placebo-controlled randomized human clinical trials (RCTs) evaluate ASU's impact on knee and hip OA.[109] Two of these indicated that ASU treatment decreased NSAID intake over 3 months. [106, 107] Another found that ASU improves Lequense Functional Index (LFI) compared to placebo over the course of 6 months, and also that improvements took 2 months to take effect, and subside after treatment ended.<sup>[108]</sup> Alternatively, a long-term study indicated no significant difference in JSN, or other parameters of disease, after 2 years of ASU treatment, [80] indicating that the beneficial impact of ASU on OA may be limited to short-term effects. However, this study also focused on identifying structure-modifying effects, versus symptom-modifying effects; while these two different measures of OA severity often correlate, ASU may impact each uniquely. Evidence for symptom modifying effects of ASU is much stronger, and thus an alternative explanation for these contradictory findings is that while ASU does not improve structural damage of OA, as measured in this study, it does improve symptoms such as pain and mobility, as measured in previous studies. [82] In a study of chronic nonspecific back pain, treatment with ASU (piascledine) combined with the NSAID artrosiline (320 mg/day) showed significant analgesic effect over NSAID treatment alone. The positive effect of ASU was demonstrated after one month of treatment. The authors suggest that further RCTs are needed to confirm results. To this end, the ERADIAS trial determined whether ASU Expansience<sup>TM</sup> treatment slowed the radiological progression of hip OA.<sup>[113]</sup> As for safety, none of the four RCTs reported significant differences in adverse effects between ASU and placebo.

Factors like BMI, severity of disease and activity level may influence the effect of ASU, as these conditions exacerbate inflammatory conditions and mechanical stresses that contribute to OA. Adipose tissue plays an important role by producing metabolic factors with catabolic and proinflammatory properties, including cytokines, chemokines and adipokines (IL-6, and TNF-α, IL-8, IFN gamma), which orchestrate pathophysiological processes in OA. Soluble mediators produced by adipocytes may also modulate chondrocyte metabolism and contribute to cartilage degradation. ASU may counteract these inflammatory processes by inhibiting the translocation of the transcription factor NF-κB from the cytoplasm to the nucleus, which controls transcription of many proinflammatory factors. [86, 115] As such, ASU acts as an anabolic agent in vitro, reducing the production of pro-inflammaotry mediators, including IL-1, IL-6, IL-8, macrophage inflammatory protein-1, NO, MMP-13, TNF-α, and COX2/PGE2<sup>[88, 94, 115, 116]</sup> from various cell types. In mice, ASU decreases proinflammatory interferon-gamma (IFNy) and IL4 production, in the context of parasitic diseases.[117, 118] While more studies need to be conducted to show the effects of ASU in patients with varying BMI, the anti-inflammatory effects of ASU are likely to protect cartilage from obesity-associated inflammatory degradation and improve OA symptoms. Indeed, ASU significantly decreased the rate of OA progression to 40% compared with 50% in the placebo group in one study. [111] However, this study showed that ASU did not influence the rate of OA progression in the obese subset of patients with mean symptom duration 4 and BMI of 27 kg/m<sup>2</sup>.

However, excessive inflammation associated with obesity may also impede efficacy, as it does with celecoxib (NSAID) treatment, which is not as effective in obese patients (BMI in excess of 30 kg/m²). The influence of obesity, and how it influences ASU efficacy may also depend on the parameters used to measure and define disease. In a study examining the relationship between BMI and OA in patients scheduled to undergo hip replacement, increasing BMI was associated with increasing levels of pain and functional disability, but not radiographic joint damage. Thus, obesity might influence some aspects of disease and treatment, but not others. This should be taken into account when designing and assessing studies intended to examine the impact of obesity on treatment efficacy. [119]

ASU has anti-inflammatory effects in mice when administered in conjunction with the anti-parasitic drug Praziquantel, reducing inflammatory cytokines interferon-gamma (IFN $\gamma$ ) and IL4, as well as granuloma size, while increasing cidal activity.<sup>[117, 118]</sup> ASU also protects gingival elastic fibers from degradation by human leukocyte elastase,<sup>[120]</sup> hypodermatitis,<sup>[121]</sup> and ischemic damage.<sup>[122]</sup>

A recent electronic databases analysis demonstrated the benefits and harms of oral medicinal plant products in treating osteoarthritis. The authors used standard methods for trial selection and data extraction, and assessed the quality of the body of evidence using the GRADE approach for major outcomes such as pain, function, radiographic joint changes, quality of life, withdrawals due to adverse events, total adverse events, and serious adverse events. The ASU product Piasclidine® formed a small and clinically questionable improvement in symptoms, compared to placebo after three to 12 months treatment. Radiographic joint changes, as change in joint space width (JSW) did not differ between ASU 300 mg treatment and placebo. Moderate-quality evidence from a single study confirmed possible benefits of ASU 600 mg over placebo. There is no evidence that Piasclidine significantly improves joint structure, and limited evidence that it prevents joint space narrowing. Authors suggest further investigations are required to determine optimum daily doses producing clinical benefits without adverse events. [123]

ASU is considered as drug in most countries, and therefore is prescribed by physicians. However, in United States it is classified as dietary supplement, and can be purchased as over-the-counter supplements, Avoca ASU (ASU-NMX1000, Nutramax Laboratories Inc., Edgewood, MD, USA), and Maximize ASU 300 / SierraSil (Maximum International Inc. Pompano Beach, FL). Avoca ASU, a combination of ASU and glucosamine sulfate, has been shown to suppress TNF $\alpha$ , IL1 $\beta$ , COX2, iNOS, PGE2, NF $\kappa$ B activation and nitrite production in articular chondrocytes and monocytes/macrophages, reducing pain and inflammation in OA patients. [115, 116, 124] However, conflicting reports indicate the complete absence of specific ASU molecules in Avoca ASU when compared with Piascledine. [75, 125–127]

Questions remain about the efficacy and safety of ASUs for treatment of OA (Table 4). Macaigne and colleagues published a case report in 2004 describing a female with lymphatic colitis associated with Piascledine treatment.<sup>[128]</sup> Further prospective multicenter studies are warranted to investigate whether other microscopic colitis cases<sup>[129]</sup> are observed in patients treated with Piascledine. Avoca ASU that contains glucosamine can induce allergic reaction

in people with shellfish allergy. Even in very small quantities, these people may experience mild symptoms, such as hives or nasal congestion, or more-severe, even life-threatening symptoms.

An alternative ASU formulation is Arthrocen (Pharmin, USA, LLC. San Jose, CA). Arthrocen is an extract from avocado and soybean oils that does not contain any ingredients of animal origin, artificial flavor, sweetener, or color; each capsule contains 100 mg unsaponifiable persea gratissima unsaponifiable (avocado) and 200 mg unsaponifiable glycine max (soybean) extracts, Silica, magnesium stearate (E470b-manufactured from vegetable oil), and gelatin fines.

In general, FDA does not held dietary supplements to the stringent standards of pharmaceutical manufacture. If ASU is to be widely used for the treatment of OA, serious consideration should be given to their current regulatory status in order to ensure potency, purity, and as well as the excipients. Many studies have demonstrated substantial variation between the content listed on the labels of these products and the actual content. The sterols content of ASU have been demonstrated to have biological activities in culture and in animal models. This approach allowed us to compare the contents of 3 commercial supplements. We found multiple peaks were present in the Piascledine-300 (Expanscience) mass spectrometry analysis (Agilent 7890 GC System, 7693 Auto Sampler, Agilent VL MSD with triple Axis Detector, Brea, California), compared to the Arthrocen 300 mg (PharminUSA) or ASU300-Avocado Soy Unsaponifiable with Sierra 600 mg (Maximize, Maximum Int.) preparations (Figure 2). Similar results were found for Piascledine-300<sup>[75]</sup> with mass spectra sterol content of C20H30O2, C20H28O2, sitosterol, stigmasterol, campesterol, squalene, beta-tocopherol, desmethyl tocopherol, oleic acid docosane, alphaamyrin, and cholesterol. In 2 letters to the editor, Msika<sup>[126]</sup> and Henroitin<sup>[125]</sup> claimed that the exact ingredients and preparation of ASU-Expanscience was an intellectual proprietary, protected by patent. Msika further emphasized that the tocopherols, sterols and patented specific molecules from avocado contribute to the originality of the product, different from natural avocado unsaponifiables. In contrast, they analyzed Dasuquin with MSM, Dasuquin, and Avoca ASU (Nutramax), and compared them with ASU Expanscience. The analyses revealed content of products were significantly different from those indicated on the Nutramax labels—with no citrostadienol, and brassicasterol present in ASU Expanscience. Contrary to that they found contents included high level of rapeseed oil and unsaponifiables products with very low tocopherol, and without respected ratio of 1:2 for avocado to soybean unsaponifiables. The original ASU Piascledine®300 pills contains 100 mg of avocado unsaponifiables and 200 mg of soybean unsaponifiables. The difference in sterol content is based on the A/S ratio and avocado-specific modified unsaponifiables obtained by a patented process. Henrotin et al., have shown ASU effects are best when the ASU ratio are 2:1<sup>[92]</sup>. In support, Henroitin recommended additional studies to ensure efficacy of Nutramax products<sup>[125]</sup>. These issues and reported adverse effects of ASU (Table 5<sup>[128, 130]</sup>) raises concerns about the content and purity of ASU supplements on the market, with implications for patient safety.

### Conclusion

Osteoarthritis inflicts pain and physical limitation on millions of people. Improving joint function and patient activity is a central public health concern to improve quality and length of life. The aim is not only to treat pain, but also to prevent the onset of disease. There is no cure for OA and even symptomatic treatment options are scarce, dominated by pain management and surgical intervention. ASUs may prove to be an effective treatment option for symptomatic OA, as they have been shown to possess chondroprotective, anabolic, and anticatabolic properties, as well as anti-inflammatory properties. At the clinical level, ASUs reduce pain and stiffness while improving joint function. Importantly, ASUs are a natural, slow-acting agent that do not merely address acute pain, but actively prevent progression of OA symptoms. Further studies are required to determine the specific mechanisms and target molecules of ASU function on OA at the cellular and metabolic level.

## Acknowledgement

We are grateful to Susan Eastman, health librarian at Stanford Hospital Health Library. We are grateful to many authors for their generous contributions while writing this review. We also apologize those authors of many relevant papers whose work are not cited due to space constraints. We also acknowledge PL for providing us photos and X-ray images of his hands and knees.

Dr. Christiansen is funded by the National Institute of Arthritis and Musculoskeletal and Skin Diseases, part of the National Institutes of Health, under Award Number AR062603, and by the Department of Defense – Congressionally Directed Medical Research Programs, under Award Number PR110178.

#### **Competing Interest Statement**

Simi Bhatti and Ramin Goudarzi are employees of Formulation Technology Inc. and Pharmin USA, LLC, respectively. However, neither Formulation Technology nor Pharmin USA contributed funds or resources to this study or the co-authors.

#### References

- 1. Li Y, Wei X, Zhou J, Wei L. The age-related changes in cartilage and osteoarthritis. Biomed Res Int. 2013; 2013:916530. [PubMed: 23971049]
- 2. Bijlsma JW, Berenbaum F, Lafeber FP. Osteoarthritis: an update with relevance for clinical practice. Lancet. 2011; 377(9783):2115–2126. [PubMed: 21684382]
- 3. Loeser RF. Aging processes and the development of osteoarthritis. Curr Opin Rheumatol. 2013; 25(1):108–113. [PubMed: 23080227]
- 4. Houard X, Goldring MB, Berenbaum F. Homeostatic mechanisms in articular cartilage and role of inflammation in osteoarthritis. Curr Rheumatol Rep. 2013; 15(11):375. [PubMed: 24072604]
- 5. Lequesne M, Brandt K, Bellamy N, Moskowitz R, Menkes CJ, Pelletier JP, Altman R. Guidelines for testing slow acting drugs in osteoarthritis. J Rheumatol Suppl. 1994; 41:65–71. discussion 72–3. [PubMed: 7799389]
- 6. Jordan KM, Arden NK, Doherty M, Bannwarth B, Bijlsma JW, Dieppe P, Gunther K, Hauselmann H, Herrero-Beaumont G, Kaklamanis P, Lohmander S, Leeb B, Lequesne M, Mazieres B, Martin-Mola E, Pavelka K, Pendleton A, Punzi L, Serni U, Swoboda B, Verbruggen G, Zimmerman-Gorska I, Dougados M. EULAR Recommendations 2003: an evidence based approach to the management of knee osteoarthritis: Report of a Task Force of the Standing Committee for International Clinical Studies Including Therapeutic Trials (ESCISIT). Ann Rheum Dis. 2003; 62(12):1145–1155. [PubMed: 14644851]
- 7. Zhang W, Doherty M, Arden N, Bannwarth B, Bijlsma J, Gunther KP, Hauselmann HJ, Herrero-Beaumont G, Jordan K, Kaklamanis P, Leeb B, Lequesne M, Lohmander S, Mazieres B, Martin-Mola E, Pavelka K, Pendleton A, Punzi L, Swoboda B, Varatojo R, Verbruggen G, Zimmermann-

- Gorska I, Dougados M. EULAR evidence based recommendations for the management of hip osteoarthritis: report of a task force of the EULAR Standing Committee for International Clinical Studies Including Therapeutics (ESCISIT). Ann Rheum Dis. 2005; 64(5):669–681. [PubMed: 15471891]
- 8. Zhang W, Moskowitz RW, Nuki G, Abramson S, Altman RD, Arden N, Bierma-Zeinstra S, Brandt KD, Croft P, Doherty M, Dougados M, Hochberg M, Hunter DJ, Kwoh K, Lohmander LS, Tugwell P. OARSI recommendations for the management of hip and knee osteoarthritis, Part II: OARSI evidence-based, expert consensus guidelines. Osteoarthritis Cartilage. 2008; 16(2):137–162. [PubMed: 18279766]
- Richette P. Pharmacological therapies for osteoarthritis. Therapie. 2011; 66(5):383–390. [PubMed: 22031681]
- 10. Hochberg MC, Altman RD, April KT, Benkhalti M, Guyatt G, McGowan J, Towheed T, Welch V, Wells G, Tugwell P. American College of Rheumatology 2012 recommendations for the use of nonpharmacologic and pharmacologic therapies in osteoarthritis of the hand, hip, and knee. Arthritis Care Res (Hoboken). 2012; 64(4):465–474. [PubMed: 22563589]
- 11. Hame SL, Alexander RA. Knee osteoarthritis in women. Curr Rev Musculoskelet Med. 2013; 6(2): 182–187. [PubMed: 23471773]
- Curatolo M, Bogduk N. Pharmacologic pain treatment of musculoskeletal disorders: current perspectives and future prospects. Clin J Pain. 2001; 17(1):25–32. [PubMed: 11289086]
- 13. Gallelli L, Galasso O, Falcone D, Southworth S, Greco M, Ventura V, Romualdi P, Corigliano A, Terracciano R, Savino R, Gulletta E, Gasparini G, De Sarro G. The effects of nonsteroidal anti-inflammatory drugs on clinical outcomes, synovial fluid cytokine concentration and signal transduction pathways in knee osteoarthritis. A randomized open label trial. Osteoarthritis Cartilage. 2013; 21(9):1400–1408. [PubMed: 23973155]
- 14. Bello AE. DUEXIS((R)) (ibuprofen 800 mg, famotidine 26.6 mg): a new approach to gastroprotection for patients with chronic pain and inflammation who require treatment with a nonsteroidal anti-inflammatory drug. Ther Adv Musculoskelet Dis. 2012; 4(5):327–339. [PubMed: 23024710]
- 15. Sands GH, Brown PB, Essex MN. The Efficacy of Continuous Versus Intermittent Celecoxib Treatment in Osteoarthritis Patients with Body Mass Index >/=30 and <30 kg/m(2.). Open Rheumatol J. 2013; 7:32–37. [PubMed: 23919092]
- Hauk L. Treatment of Knee Osteoarthritis: A Clinical Practice Guideline from the AAOS. Am Fam Physician. 2014; 89(11):918–920. [PubMed: 25077402]
- 17. Wise J. NICE keeps paracetamol in UK guidelines on osteoarthritis. BMJ. 2014; 348:g1545. [PubMed: 24530969]
- 18. McAlindon TE, Bannuru RR, Sullivan MC, Arden NK, Berenbaum F, Bierma-Zeinstra SM, Hawker GA, Henrotin Y, Hunter DJ, Kawaguchi H, Kwoh K, Lohmander S, Rannou F, Roos EM, Underwood M. OARSI guidelines for the non-surgical management of knee osteoarthritis. Osteoarthritis Cartilage. 2014; 22(3):363–388. [PubMed: 24462672]
- 19. Balmaceda CM. Evolving guidelines in the use of topical nonsteroidal anti-inflammatory drugs in the treatment of osteoarthritis. BMC Musculoskelet Disord. 2014; 15:27. [PubMed: 24444047]
- 20. Brosseau L, Rahman P, Toupin-April K, Poitras S, King J, De Angelis G, Loew L, Casimiro L, Paterson G, McEwan J. A systematic critical appraisal for non-pharmacological management of osteoarthritis using the appraisal of guidelines research and evaluation II instrument. PLoS One. 2014; 9(1):e82986. [PubMed: 24427268]
- 21. Mobasheri A. The future of osteoarthritis therapeutics: emerging biological therapy. Curr Rheumatol Rep. 2013; 15(12):385. [PubMed: 24170255]
- 22. Abraham CL, Maas SA, Weiss JA, Ellis BJ, Peters CL, Anderson AE. A new discrete element analysis method for predicting hip joint contact stresses. J Biomech. 2013; 46(6):1121–1127. [PubMed: 23453394]
- 23. Dougados M, Nguyen M, Berdah L, Mazieres B, Vignon E, Lequesne M. Evaluation of the structure-modifying effects of diacerein in hip osteoarthritis: ECHODIAH, a three-year, placebocontrolled trial. Evaluation of the Chondromodulating Effect of Diacerein in OA of the Hip. Arthritis Rheum. 2001; 44(11):2539–2547. [PubMed: 11710710]

24. Henrotin Y, Sanchez C, Balligand M. Pharmaceutical and nutraceutical management of canine osteoarthritis: present and future perspectives. Vet J. 2005; 170(1):113–123. [PubMed: 15993795]

- 25. Yang KG, Raijmakers NJ, van Arkel ER, Caron JJ, Rijk PC, Willems WJ, Zijl JA, Verbout AJ, Dhert WJ, Saris DB. Autologous interleukin-1 receptor antagonist improves function and symptoms in osteoarthritis when compared to placebo in a prospective randomized controlled trial. Osteoarthritis Cartilage. 2008; 16(4):498–505. [PubMed: 17825587]
- Baltzer AW, Ostapczuk MS, Stosch D, Seidel F, Granrath M. A new treatment for hip osteoarthritis: clinical evidence for the efficacy of autologous conditioned serum. Orthop Rev (Pavia). 2013; 5(2):59–64. [PubMed: 23888203]
- 27. Smelter E, Hochberg MC. New treatments for osteoarthritis. Curr Opin Rheumatol. 2013; 25(3): 310–316. [PubMed: 23425965]
- 28. Lane NE, Schnitzer TJ, Birbara CA, Mokhtarani M, Shelton DL, Smith MD, Brown MT. Tanezumab for the treatment of pain from osteoarthritis of the knee. N Engl J Med. 2010; 363(16): 1521–1531. [PubMed: 20942668]
- 29. Chevalier X, Eymard F, Richette P. Biologic agents in osteoarthritis: hopes and disappointments. Nat Rev Rheumatol. 2013; 9(7):400–410. [PubMed: 23545735]
- 30. Brown MT, Murphy FT, Radin DM, Davignon I, Smith MD, West CR. Tanezumab reduces osteoarthritic hip pain: results of a randomized, double-blind, placebo-controlled phase III trial. Arthritis Rheum. 2013; 65(7):1795–1803. [PubMed: 23553790]
- 31. Spierings EL, Fidelholtz J, Wolfram G, Smith MD, Brown MT, West CR. A phase III placebo- and oxycodone-controlled study of tanezumab in adults with osteoarthritis pain of the hip or knee. Pain. 154(9):1603–1612. [PubMed: 23707270]
- 32. Seidel MF, Wise BL, Lane NE. Nerve growth factor: an update on the science and therapy. Osteoarthritis Cartilage. 2013; 21(9):1223–1228. [PubMed: 23973134]
- 33. Sanga P, Katz N, Polverejan E, Wang S, Kelly KM, Haeussler J, Thipphawong J. Efficacy, safety, and tolerability of fulranumab, an anti-nerve growth factor antibody, in the treatment of patients with moderate to severe osteoarthritis pain. Pain. 2013; 154(10):1910–1919. [PubMed: 23748114]
- 34. Teich N. Topical application of TNF-blockers. Dtsch Med Wochenschr. 2013; 138(8):381–385. [PubMed: 23404327]
- 35. Pelletier JP, Roubille C, Raynauld JP, Abram F, Dorais M, Delorme P, Martel-Pelletier J. Disease-modifying effect of strontium ranelate in a subset of patients from the Phase III knee osteoarthritis study SEKOIA using quantitative MRI: reduction in bone marrow lesions protects against cartilage loss. Ann Rheum Dis. 2013
- 36. Bruyere O, Reginster JY, Bellamy N, Chapurlat R, Richette P, Cooper C. Clinically meaningful effect of strontium ranelate on symptoms in knee osteoarthritis: a responder analysis. Rheumatology (Oxford). 2014; 53(8):1457–1464. [PubMed: 24667161]
- 37. Tenti S, Cheleschi S, Guidelli GM, Galeazzi M, Fioravanti A. What about strontium ranelate in osteoarthritis? Doubts and securities. Mod Rheumatol. 2014
- 38. Carbone LD, Nevitt MC, Wildy K, Barrow KD, Harris F, Felson D, Peterfy C, Visser M, Harris TB, Wang BW, Kritchevsky SB. The relationship of antiresorptive drug use to structural findings and symptoms of knee osteoarthritis. Arthritis and rheumatism. 2004; 50(11):3516–3525. [PubMed: 15529367]
- 39. Spector TD, Conaghan PG, Buckland-Wright JC, Garnero P, Cline GA, Beary JF, Valent DJ, Meyer JM. Effect of risedronate on joint structure and symptoms of knee osteoarthritis: results of the BRISK randomized, controlled trial [ISRCTN01928173]. Arthritis research & therapy. 2005; 7(3):R625–R633. [PubMed: 15899049]
- 40. Buckland-Wright JC, Messent EA, Bingham CO 3rd, Ward RJ, Tonkin C. A 2 yr longitudinal radiographic study examining the effect of a bisphosphonate (risedronate) upon subchondral bone loss in osteoarthritic knee patients. Rheumatology. 2007; 46(2):257–264. [PubMed: 16837470]
- 41. Nishii T, Tamura S, Shiomi T, Yoshikawa H, Sugano N. Alendronate treatment for hip osteoarthritis: prospective randomized 2-year trial. Clin Rheumatol. 2013; 32(12):1759–1766. [PubMed: 23881439]
- 42. Chappell AS, Desaiah D, Liu-Seifert H, Zhang S, Skljarevski V, Belenkov Y, Brown JP. A double-blind, randomized, placebo-controlled study of the efficacy and safety of duloxetine for the

- treatment of chronic pain due to osteoarthritis of the knee. Pain Pract. 2011; 11(1):33–41. [PubMed: 20602715]
- 43. Wenham CY, Grainger AJ, Hensor EM, Caperon AR, Ash ZR, Conaghan PG. Methotrexate for pain relief in knee osteoarthritis: an open-label study. Rheumatology (Oxford). 2013; 52(5):888–892. [PubMed: 23300331]
- 44. Johnson K, Zhu S, Tremblay MS, Payette JN, Wang J, Bouchez LC, Meeusen S, Althage A, Cho CY, Wu X, Schultz PG. A stem cell-based approach to cartilage repair. Science. 2012; 336(6082): 717–721. [PubMed: 22491093]
- 45. Zhu Y, Yuan M, Meng HY, Wang AY, Guo QY, Wang Y, Peng J. Basic science and clinical application of platelet-rich plasma for cartilage defects and osteoarthritis: a review. Osteoarthritis Cartilage. 2013; 21(11):1627–1637. [PubMed: 23933379]
- 46. Andia I, Maffulli N. Platelet-rich plasma for managing pain and inflammation in osteoarthritis. Nat Rev Rheumatol. 2013; 9(12):721–730. [PubMed: 24080861]
- 47. Chou CL, Lee SH, Lu SY, Tsai KL, Ho CY, Lai HC. Therapeutic effects of intra-articular botulinum neurotoxin in advanced knee osteoarthritis. J Chin Med Assoc. 2010; 73(11):573–580. [PubMed: 21093825]
- 48. Madry H, Cucchiarini M. Advances and challenges in gene-based approaches for osteoarthritis. J Gene Med. 2013; 15(10):343–355. [PubMed: 24006099]
- Jorgensen C. ADIPOA: cell therapy with stromal adipocytes cells. Rev Med Interne. 2011;
   32(Suppl 2):S203. [PubMed: 22018931]
- 50. Shen W, Chen J, Zhu T, Chen L, Zhang W, Fang Z, Heng BC, Yin Z, Chen X, Ji J, Chen W, Ouyang HW. Intra-articular injection of human meniscus stem/progenitor cells promotes meniscus regeneration and ameliorates osteoarthritis through stromal cell-derived factor-1/CXCR4-mediated homing. Stem Cells Transl Med. 2014; 3(3):387–394. [PubMed: 24448516]
- 51. Emadedin M, Aghdami N, Taghiyar L, Fazeli R, Moghadasali R, Jahangir S, Farjad R, Baghaban Eslaminejad M. Intra-articular injection of autologous mesenchymal stem cells in six patients with knee osteoarthritis. Arch Iran Med. 2012; 15(7):422–428. [PubMed: 22724879]
- 52. Sato M, Uchida K, Nakajima H, Miyazaki T, Guerrero AR, Watanabe S, Roberts S, Baba H. Direct transplantation of mesenchymal stem cells into the knee joints of Hartley strain guinea pigs with spontaneous osteoarthritis. Arthritis Res Ther. 2012; 14(1):R31. [PubMed: 22314040]
- 53. Koh YG, Choi YJ. Infrapatellar fat pad-derived mesenchymal stem cell therapy for knee osteoarthritis. Knee. 2012; 19(6):902–907. [PubMed: 22583627]
- 54. Shen W, Chen J, Zhu T, Yin Z, Chen X, Chen L, Fang Z, Heng BC, Ji J, Chen W, Ouyang HW. Osteoarthritis prevention through meniscal regeneration induced by intra-articular injection of meniscus stem cells. Stem Cells Dev. 2013; 22(14):2071–2082. [PubMed: 23461527]
- 55. Richette P. Management of osteoarthritis: oral therapies. Rev Prat. 2012; 62(5):654–660. [PubMed: 22730795]
- 56. Reginster JY, Gillot V, Bruyere O, Henrotin Y. Evidence of nutriceutical effectiveness in the treatment of osteoarthritis. Curr Rheumatol Rep. 2000; 2(6):472–477. [PubMed: 11123100]
- 57. Hochberg M, Chevalier X, Henrotin Y, Hunter DJ, Uebelhart D. Symptom and structure modification in osteoarthritis with pharmaceutical-grade chondroitin sulfate: what's the evidence? Curr Med Res Opin. 2013; 29(3):259–267. [PubMed: 23186102]
- 58. Hui AY, McCarty WJ, Masuda K, Firestein GS, Sah RL. A systems biology approach to synovial joint lubrication in health, injury, and disease. Wiley Interdiscip Rev Syst Biol Med. 2012; 4(1): 15–37. [PubMed: 21826801]
- Reginster JY, Deroisy R, Rovati LC, Lee RL, Lejeune E, Bruyere O, Giacovelli G, Henrotin Y, Dacre JE, Gossett C. Long-term effects of glucosamine sulphate on osteoarthritis progression: a randomised, placebo-controlled clinical trial. Lancet. 2001; 357(9252):251–256. [PubMed: 11214126]
- 60. Pavelka K, Gatterova J, Olejarova M, Machacek S, Giacovelli G, Rovati LC. Glucosamine sulfate use and delay of progression of knee osteoarthritis: a 3-year, randomized, placebo-controlled, double-blind study. Arch Intern Med. 2002; 162(18):2113–2123. [PubMed: 12374520]
- 61. Clegg DO, Reda DJ, Harris CL, Klein MA, O'Dell JR, Hooper MM, Bradley JD, Bingham CO 3rd, Weisman MH, Jackson CG, Lane NE, Cush JJ, Moreland LW, Schumacher HR Jr, Oddis CV,

- Wolfe F, Molitor JA, Yocum DE, Schnitzer TJ, Furst DE, Sawitzke AD, Shi H, Brandt KD, Moskowitz RW, Williams HJ. Glucosamine, chondroitin sulfate, and the two in combination for painful knee osteoarthritis. N Engl J Med. 2006; 354(8):795–808. [PubMed: 16495392]
- 62. Henrotin Y, Mobasheri A, Marty M. Is there any scientific evidence for the use of glucosamine in the management of human osteoarthritis? Arthritis Res Ther. 2012; 14(1):201. [PubMed: 22293240]
- 63. Altman RD, Abramson S, Bruyere O, Clegg D, Herrero-Beaumont G, Maheu E, Moskowitz R, Pavelka K, Reginster JY. Commentary: osteoarthritis of the knee and glucosamine. Osteoarthritis Cartilage. 2006; 14(10):963–966. [PubMed: 16857394]
- 64. Henrotin Y, Chevalier X, Herrero-Beaumont G, McAlindon T, Mobasheri A, Pavelka K, Schon C, Weinans H, Biesalski H. Physiological effects of oral glucosamine on joint health: current status and consensus on future research priorities. BMC Res Notes. 2013; 6:115. [PubMed: 23531101]
- 65. Henrotin Y, Lambert C. Chondroitin and glucosamine in the management of osteoarthritis: an update. Curr Rheumatol Rep. 2013; 15(10):361. [PubMed: 23955063]
- 66. Rutjes AW, Juni P, da Costa BR, Trelle S, Nuesch E, Reichenbach S. Viscosupplementation for osteoarthritis of the knee: a systematic review and meta-analysis. Ann Intern Med. 2012; 157(3): 180–191. [PubMed: 22868835]
- 67. Henrotin Y, Chevalier X, Deberg M, Balblanc JC, Richette P, Mulleman D, Maillet B, Rannou F, Piroth C, Mathieu P, Conrozier T. Early decrease of serum biomarkers of type II collagen degradation (Coll2-1) and joint inflammation (Coll2-1 NO(2)) by hyaluronic acid intra-articular injections in patients with knee osteoarthritis: a research study part of the Biovisco study. J Orthop Res. 2013; 31(6):901–907. [PubMed: 23423846]
- 68. Waddell DD, Beyer A, Thompson TL, Morawiak J, Elkins C, Rosenberg A, Spitzer A. No Conclusive Evidence that Histologically Found Granulomas and Acute Local Reactions Following Hylan G-F 20 Injections Are Related or Have Clinical Significance. J Knee Surg. 2013
- 69. Sawitzke AD, Shi H, Finco MF, Dunlop DD, Harris CL, Singer NG, Bradley JD, Silver D, Jackson CG, Lane NE, Oddis CV, Wolfe F, Lisse J, Furst DE, Bingham CO, Reda DJ, Moskowitz RW, Williams HJ, Clegg DO. Clinical efficacy and safety of glucosamine, chondroitin sulphate, their combination, celecoxib or placebo taken to treat osteoarthritis of the knee: 2-year results from GAIT. Ann Rheum Dis. 2010; 69(8):1459–1464. [PubMed: 20525840]
- 70. Sawitzke AD, Shi H, Finco MF, Dunlop DD, Bingham CO 3rd, Harris CL, Singer NG, Bradley JD, Silver D, Jackson CG, Lane NE, Oddis CV, Wolfe F, Lisse J, Furst DE, Reda DJ, Moskowitz RW, Williams HJ, Clegg DO. The effect of glucosamine and/or chondroitin sulfate on the progression of knee osteoarthritis: a report from the glucosamine/chondroitin arthritis intervention trial. Arthritis Rheum. 2008; 58(10):3183–3191. [PubMed: 18821708]
- 71. The NIH Glucosamine/Chondroitin Arthritis Intervention Trial (GAIT). J Pain Palliat Care Pharmacother. 2008; 22(1):39–43. [PubMed: 19062354]
- 72. Miller KL, Clegg DO. Glucosamine and chondroitin sulfate. Rheum Dis Clin North Am. 2011; 37(1):103–118. [PubMed: 21220090]
- 73. Wandel S, Juni P, Tendal B, Nuesch E, Villiger PM, Welton NJ, Reichenbach S, Trelle S. Effects of glucosamine, chondroitin, or placebo in patients with osteoarthritis of hip or knee: network meta-analysis. BMJ. 2010; 341:c4675. [PubMed: 20847017]
- 74. Miller MJ, Mehta K, Kunte S, Raut V, Gala J, Dhumale R, Shukla A, Tupalli H, Parikh H, Bobrowski P, Chaudhary J. Early relief of osteoarthritis symptoms with a natural mineral supplement and a herbomineral combination: a randomized controlled trial [ISRCTN38432711]. J Inflamm (Lond). 2005; 2:11. [PubMed: 16242032]
- Lippiello L, Nardo JV, Harlan R, Chiou T. Metabolic effects of avocado/soy unsaponifiables on articular chondrocytes. Evid Based Complement Alternat Med. 2008; 5(2):191–197. [PubMed: 18604259]
- 76. Thiers MH. Unsaponifiable constituents of avocado and soya oils. Treatment of certain forms of arthralgia. J Med Lyon. 1972; 53(222):195–198. [PubMed: 5065287]
- 77. Mauviel A, Loyau G, Pujol JP. Effect of unsaponifiable extracts of avocado and soybean (Piascledine) on the collagenolytic action of cultures of human rheumatoid synoviocytes and rabbit

- articular chondrocytes treated with interleukin-1. Rev Rhum Mal Osteoartic. 1991; 58(4):241–245. [PubMed: 1647544]
- 78. Cake MA, Read RA, Guillou B, Ghosh P. Modification of articular cartilage and subchondral bone pathology in an ovine meniscectomy model of osteoarthritis by avocado and soya unsaponifiables (ASU). Osteoarthritis Cartilage. 2000; 8(6):404–411. [PubMed: 11069724]
- 79. Kut-Lasserre C, Miller CC, Ejeil AL, Gogly B, Dridi M, Piccardi N, Guillou B, Pellat B, Godeau G. Effect of avocado and soybean unsaponifiables on gelatinase A (MMP-2), stromelysin 1 (MMP-3), and tissue inhibitors of matrix metalloproteinase (TIMP- 1 and TIMP-2) secretion by human fibroblasts in culture. J Periodontol. 2001; 72(12):1685–1694. [PubMed: 11811504]
- 80. Lequesne M, Maheu E, Cadet C, Dreiser RL. Structural effect of avocado/soybean unsaponifiables on joint space loss in osteoarthritis of the hip. Arthritis Rheum. 2002; 47(1):50–58. [PubMed: 11932878]
- 81. Kucharz EJ. Application of avocado/soybean unsaponifiable mixtures (piascledine) in treatment of patients with osteoarthritis. Ortop Traumatol Rehabil. 2003; 5(2):248–251. [PubMed: 18034015]
- 82. Ernst E. Avocado-soybean unsaponifiables (ASU) for osteoarthritis a systematic review. Clin Rheumatol. 2003; 22(4–5):285–288. [PubMed: 14576991]
- 83. Angermann P. Avocado/soybean unsaponifiables in the treatment of knee and hip osteoarthritis. Ugeskr Laeger. 2005; 167(33):3023–3025. [PubMed: 16109242]
- 84. Henrotin YE, Deberg MA, Crielaard JM, Piccardi N, Msika P, Sanchez C. Avocado/soybean unsaponifiables prevent the inhibitory effect of osteoarthritic subchondral osteoblasts on aggrecan and type II collagen synthesis by chondrocytes. J Rheumatol. 2006; 33(8):1668–1678. [PubMed: 16832844]
- 85. Kawcak CE, Frisbie DD, McIlwraith CW, Werpy NM, Park RD. Evaluation of avocado and soybean unsaponifiable extracts for treatment of horses with experimentally induced osteoarthritis. Am J Vet Res. 2007; 68(6):598–604. [PubMed: 17542691]
- 86. Gabay O, Gosset M, Levy A, Salvat C, Sanchez C, Pigenet A, Sautet A, Jacques C, Berenbaum F. Stress-induced signaling pathways in hyalin chondrocytes: inhibition by Avocado-Soybean Unsaponifiables (ASU). Osteoarthritis Cartilage. 2008; 16(3):373–384. [PubMed: 17707661]
- 87. Christensen R, Bartels EM, Astrup A, Bliddal H. Symptomatic efficacy of avocado-soybean unsaponifiables (ASU) in osteoarthritis (OA) patients: a meta-analysis of randomized controlled trials. Osteoarthritis Cartilage. 2008; 16(4):399–408. [PubMed: 18042410]
- 88. Boileau C, Martel-Pelletier J, Caron J, Msika P, Guillou GB, Baudouin C, Pelletier JP. Protective effects of total fraction of avocado/soybean unsaponifiables on the structural changes in experimental dog osteoarthritis: inhibition of nitric oxide synthase and matrix metalloproteinase-13. Arthritis Res Ther. 2009; 11(2):R41. [PubMed: 19291317]
- 89. Dinubile NA. A potential role for avocado- and soybean-based nutritional supplements in the management of osteoarthritis: a review. Phys Sportsmed. 2010; 38(2):71–81. [PubMed: 20631466]
- 90. Altinel L, Sahin O, Kose KC, Bas O, Ozen OA, Saritas ZK, Pamuk K. Healing of osteochondral defects in canine knee with avocado/soybean unsaponifiables: a morphometric comparative analysis. Eklem Hastalik Cerrahisi. 2011; 22(1):48–53. [PubMed: 21417987]
- 91. Lamaud E, Robert AM, Wepierre J. Biochemical effects of unsaponifiable lipidic components of avocado and soya bean administered percutaneously on the connective tissue components of hairless rat skin. Int J Cosmet Sci. 1979; 1(4):213–219. [PubMed: 19467069]
- 92. Henrotin YE, Labasse AH, Jaspar JM, De Groote DD, Zheng SX, Guillou GB, Reginster JY. Effects of three avocado/soybean unsaponifiable mixtures on metalloproteinases, cytokines and prostaglandin E2 production by human articular chondrocytes. Clin Rheumatol. 1998; 17(1):31–39. [PubMed: 9586676]
- 93. Mauviel A, Daireaux M, Hartmann DJ, Galera P, Loyau G, Pujol JP. Effects of unsaponifiable extracts of avocado/soy beans (PIAS) on the production of collagen by cultures of synoviocytes, articular chondrocytes and skin fibroblasts. Rev Rhum Mal Osteoartic. 1989; 56(2):207–211. [PubMed: 2727601]
- 94. Henrotin YE, Sanchez C, Deberg MA, Piccardi N, Guillou GB, Msika P, Reginster JY. Avocado/soybean unsaponifiables increase aggrecan synthesis and reduce catabolic and proinflammatory

- mediator production by human osteoarthritic chondrocytes. J Rheumatol. 2003; 30(8):1825–1834. [PubMed: 12913942]
- 95. Ownby SL, Fortuno LV, Au AY, Grzanna MW, Rashmir-Raven AM, Frondoza CG. Expression of pro-inflammatory mediators is inhibited by an avocado/soybean unsaponifiables and epigallocatechin gallate combination. J Inflamm (Lond). 2014; 11(1):8. [PubMed: 24678847]
- 96. Khayyal MT, el-Ghazaly MA. The possible "chondroprotective" effect of the unsaponifiable constituents of avocado and soya in vivo. Drugs Exp Clin Res. 1998; 24(1):41–50. [PubMed: 9604147]
- 97. Cinelli M, Guiducci S, Del Rosso A, Pignone A, Del Rosso M, Fibbi G, Serrati S, Gabrielli A, Giacomelli R, Piccardi N, Matucci Cerinic M. Piascledine modulates the production of VEGF and TIMP-1 and reduces the invasiveness of rheumatoid arthritis synoviocytes. Scand J Rheumatol. 2006; 35(5):346–350. [PubMed: 17062432]
- 98. Boumediene K, Felisaz N, Bogdanowicz P, Galera P, Guillou GB, Pujol JP. Avocado/soya unsaponifiables enhance the expression of transforming growth factor beta1 and beta2 in cultured articular chondrocytes. Arthritis Rheum. 1999; 42(1):148–156. [PubMed: 9920025]
- 99. Belcher C, Fawthrop F, Bunning R, Doherty M. Plasminogen activators and their inhibitors in synovial fluids from normal, osteoarthritis, and rheumatoid arthritis knees. Ann Rheum Dis. 1996; 55(4):230–236. [PubMed: 8733439]
- 100. Altinel L, Saritas ZK, Kose KC, Pamuk K, Aksoy Y, Serteser M. Treatment with unsaponifiable extracts of avocado and soybean increases TGF-beta1 and TGF-beta2 levels in canine joint fluid. Tohoku J Exp Med. 2007; 211(2):181–186. [PubMed: 17287602]
- 101. de Oliveira GJ, de Paula LG, Spin-Neto R, Stavropoulos A, Spolidorio LC, Marcantonio E Jr, Marcantonio RA. Effect of avocado/soybean unsaponifiables on osseointegration: a proof-of-principle preclinical in vivo study. Int J Oral Maxillofac Implants. 2014; 29(4):949–957. [PubMed: 25032777]
- 102. Chevallier F, Lutton C, Sulpice JC, D'Hollander F. Influence of the daily ingestion of a total unsaponifiable extract from avocado and soy bean oils on cholesterol metabolism in the rat. Pathol Biol (Paris). 1975; 23(3):225–230. [PubMed: 1096036]
- 103. Zushi S, Akagi M, Kishimoto H, Teramura T, Sawamura T, Hamanishi C. Induction of bovine articular chondrocyte senescence with oxidized low-density lipoprotein through lectin-like oxidized low-density lipoprotein receptor 1. Arthritis Rheum. 2009; 60(10):3007–3016. [PubMed: 19790061]
- 104. Kishimoto H, Akagi M, Zushi S, Teramura T, Onodera Y, Sawamura T, Hamanishi C. Induction of hypertrophic chondrocyte-like phenotypes by oxidized LDL in cultured bovine articular chondrocytes through increase in oxidative stress. Osteoarthritis Cartilage. 2010; 18(10):1284– 1290. [PubMed: 20633688]
- 105. Zborovskii AB, Akhverdian Iu R, Sivordova LE, Simakova ES, Zavodovskii BV. Efficiency of unsaponifiable compounds of soya beans and avocado in health care personnel with osteoarthrosis in Volgograd. Med Tr Prom Ekol. 2013; (2):41–44. [PubMed: 23785818]
- 106. Blotman F, Maheu E, Wulwik A, Caspard H, Lopez A. Efficacy and safety of avocado/soybean unsaponifiables in the treatment of symptomatic osteoarthritis of the knee and hip. A prospective, multicenter, three-month, randomized, double-blind, placebo-controlled trial. Rev Rhum Engl Ed. 1997; 64(12):825–834. [PubMed: 9476272]
- 107. Appelboom T, Schuermans J, Verbruggen G, Henrotin Y, Reginster JY. Symptoms modifying effect of avocado/soybean unsaponifiables (ASU) in knee osteoarthritis. A double blind, prospective, placebo-controlled study. Scand J Rheumatol. 2001; 30(4):242–247. [PubMed: 11578021]
- 108. Maheu E, Mazieres B, Valat JP, Loyau G, Le Loet X, Bourgeois P, Grouin JM, Rozenberg S. Symptomatic efficacy of avocado/soybean unsaponifiables in the treatment of osteoarthritis of the knee and hip: a prospective, randomized, double-blind, placebo-controlled, multicenter clinical trial with a six-month treatment period and a two-month followup demonstrating a persistent effect. Arthritis Rheum. 1998; 41(1):81–91. [PubMed: 9433873]
- 109. Ameye LG, Chee WS. Osteoarthritis and nutrition. From nutraceuticals to functional foods: a systematic review of the scientific evidence. Arthritis Res Ther. 2006; 8(4):R127. [PubMed: 16859534]

110. Pavelka K, Coste P, Geher P, Krejci G. Efficacy and safety of piascledine 300 versus chondroitin sulfate in a 6 months treatment plus 2 months observation in patients with osteoarthritis of the knee. Clin Rheumatol. 2010; 29(6):659–670. [PubMed: 20179981]

- 111. Maheu E, Cadet C, Marty M, Moyse D, Kerloch I, Coste P, Dougados M, Mazieres B, Spector TD, Halhol H, Grouin JM, Lequesne M. Randomised, controlled trial of avocado-soybean unsaponifiable (Piascledine) effect on structure modification in hip osteoarthritis: the ERADIAS study. Ann Rheum Dis. 2014; 73(2):376–384. [PubMed: 23345601]
- 112. Shimizu C, Coutts RD, Healey RM, Kubo T, Hirasawa Y, Amiel D. Method of histomorphometric assessment of glycosaminoglycans in articular cartilage. J Orthop Res. 1997; 15(5):670–674. [PubMed: 9420595]
- 113. Merkulova DM, Onsin AA, Merkulov YA. Piascledine in the treatment of chronic dorsalgia. Zh Nevrol Psikhiatr Im S S Korsakova. 2013; 113:18–22. [PubMed: 24107875]
- 114. Martinez-Abundis E, Gonzalez-Ortiz M, Mercado-Sesma AR, Reynoso-von-Drateln C, Moreno-Andrade A. Effect of avocado soybean unsaponifiables on insulin secretion and insulin sensitivity in patients with obesity. Obes Facts. 2013; 6(5):443–448. [PubMed: 24135894]
- 115. Heinecke LF, Grzanna MW, Au AY, Mochal CA, Rashmir-Raven A, Frondoza CG. Inhibition of cyclooxygenase-2 expression and prostaglandin E2 production in chondrocytes by avocado soybean unsaponifiables and epigallocatechin gallate. Osteoarthritis Cartilage. 2010; 18(2):220– 227. [PubMed: 19748608]
- 116. Au RY, Al-Talib TK, Au AY, Phan PV, Frondoza CG. Avocado soybean unsaponifiables (ASU) suppress TNF-alpha, IL-1beta, COX-2, iNOS gene expression, and prostaglandin E2 and nitric oxide production in articular chondrocytes and monocyte/macrophages. Osteoarthritis Cartilage. 2007; 15(11):1249–1255. [PubMed: 17845860]
- 117. Soliman MF. Evaluation of avocado/soybean unsaponifiable alone or concurrently with praziquantel in murine schistosomiasis. Acta Trop. 2012; 122(3):261–266. [PubMed: 22342904]
- 118. Ndjonka D, Rapado LN, Silber AM, Liebau E, Wrenger C. Natural products as a source for treating neglected parasitic diseases. Int J Mol Sci. 2013; 14(2):3395–3439. [PubMed: 23389040]
- 119. Lubbeke A, Duc S, Garavaglia G, Finckh A, Hoffmeyer P. BMI and severity of clinical and radiographic signs of hip osteoarthritis. Obesity (Silver Spring). 2009; 17(7):1414–1419. [PubMed: 19197252]
- 120. Kut C, Assoumou A, Dridi M, Bonnefoix M, Gogly B, Pellat B, Guillou GB, Godeau G. Morphometric analysis of human gingival elastic fibres degradation by human leukocyte elastase protective effect of avocado and soybean unsaponifiables (ASU). Pathol Biol (Paris). 1998; 46(7):571–576. [PubMed: 9842576]
- 121. Chaze J. Treatment of hypodermatitis of the leg with unsaponifiable extracts of avocado and soya. Phlebologie. 1972; 25(3):315–318. [PubMed: 4662060]
- 122. Yaman M, Eser O, Cosar M, Bas O, Sahin O, Mollaoglu H, Fidan H, Songur A. Oral administration of avocado soybean unsaponifiables (ASU) reduces ischemic damage in the rat hippocampus. Arch Med Res. 2007; 38(5):489–494. [PubMed: 17560453]
- 123. Cameron M, Chrubasik S. Oral herbal therapies for treating osteoarthritis. Cochrane Database Syst Rev. 2014; 5:CD002947. [PubMed: 24848732]
- 124. Frondoza CG, Heinecke LF, Grzanna MW, Au AY, Ownby SL. Modulation of cytokine-induced prostaglandin E(2) production in cultures of articular chondrocytes obtained from carpal joints of camels (Camelus dromedarius). Am J Vet Res. 2011; 72(1):51–58. [PubMed: 21194335]
- 125. Henrotin Y. Avocado/soybean unsaponifiable (ASU) to treat osteoarthritis: a clarification. Osteoarthritis Cartilage. 2008; 16(9):1118–1119. author reply 1120. [PubMed: 18304841]
- 126. Msika P, Baudouin C, Saunois A, Bauer T. Avocado/soybean unsaponifiables, ASU EXPANSCIENCE, are strictly different from the nutraceutical products claiming ASU appellation. Osteoarthritis Cartilage. 2008; 16(10):1275–126. [PubMed: 18420430]
- 127. Frondoza CG. Response to letter to editor entitled: "Avocado/soybean unsaponifiables, ASU Expanscience, are strictly different from the nutraceutical products claiming ASU appellation" (4365). Osteoarthritis and Cartilage. 2008; 16:1590–1591. [PubMed: 18539488]
- 128. Macaigne G, Ozon N, Dikov D, Auriault ML, Deplus R. Piascledine-associated lymphocytic colitis. Gastroenterol Clin Biol. 2004; 28(4):412–413. [PubMed: 15146164]

129. Macaigne G, Lahmek P, Locher C, Lesgourgues B, Costes L, Nicolas MP, Courillon-Mallet A, Ghilain JM, Bellaiche G, de Montigny-Lehnardt S, Barjonet G, Vitte RL, Faroux R, Lambare B, Fleury A, Pariente A, Nahon S. Microscopic Colitis or Functional Bowel Disease With Diarrhea: A French Prospective Multicenter Study. Am J Gastroenterol. 2014

130. Olivier P, Montastruc JL. Post-marketing safety profile of avocado-soybean unsaponifiables. Presse Med. 2010; 39(10):e211–e216. [PubMed: 20937576]

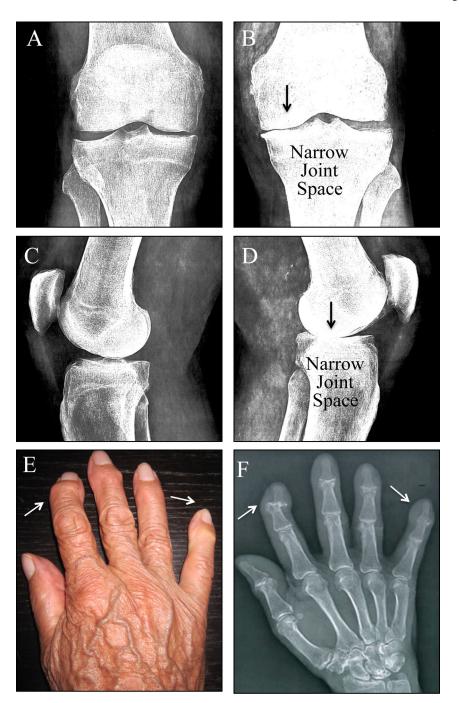


Figure 1. Anteroposterior (A–B) and cross-table lateral (C–D) of knee osteoarthritis.

Symptomatic knee OA typically presents with narrowing of the joint space and bone spurs (arrows). During the development of OA, articular cartilage breaks down over time and become thin. As a result the bone surfaces rub against each other, further damaging the cartilage and bone, and causing pain. Joints with late stage OA are often painful, warm to the touch, possibly red, swollen, and have notable loss of function. When cartilage wears away, bones may no longer be protected from rubbing and grinding against each other,

causing pain and subchondral cysts. **Osteoarthritis of the hand (E–F).** Osteoarthritis of the interphalangeal joints of the hand. Presence of bony swelling and deformity due to osteophytes at the distal interphalangeal joints (Join Swelling and Heberden's nodes).

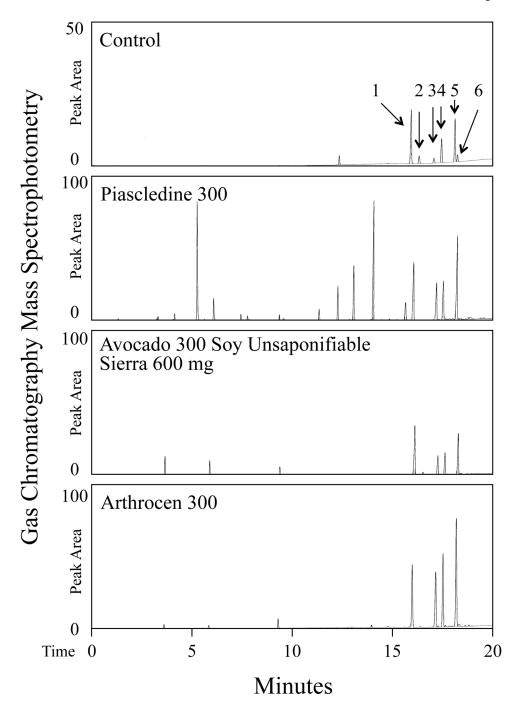


Figure 2. Gas Chromatography-Mass Spectrometry analysis of major sterol components of Piascledine 300, Avocado 300 Soy Unsaponifiable with Sierra 600 mg, and Arthrocen 300. Control sample exhibited characteristic peaks corresponding to 1- Dihydrocholesterol (5 $\alpha$ -Cholestan-3 $\beta$ -ol; internal control Sigma Aldrich), 2- Brass (Brassicasterol), 3- Camp (Campesterol), 4- Stigmn (Stigmastanol), 5-  $\beta$ -Sito ( $\beta$ -sitosterol), 6- Stigma (Stigmasterol).

Christiansen et al. Page 24

Table 1

Inhibitory effects of Avocado Soybean Unsaponifiable on inflammatory and catabolic mediators of osteoarthritis

Molecular Mediator	Target Tissue/Organ	Organism	Assay	References
Interleukin-1 beta (IL1β)	Synoviocytes Chondrocytes	Mice Rabbit Human	In Vitro	77, 81, 116
Interleukin-(IL4)	Chondrocyte		In Vitro	
Interleukin-6 (IL-6)	Chondrocyte	Human	In Vitro	81, 92
Interleukin-8 (IL-8)	Chondrocyte	Human	In Vitro	92
Macrophage inhibitory protein-1beta (MIP-1β)	Chondrocyte	Human	In Vitro	87, 116
MMP-2 (also known as Collagenase, or Gelatinase-2)	Fibroblasts, Chondrocyte	Human	In Vitro	79, 81, 92
MMP-3 (also known as Stromelysin)	Fibroblasts Chondrocyte  + Subchondral bone osteoblasts (SBO)	Mice Human	In Vitro	79, 81, 84, 86, 92, 101
MMP-13 Collagenase-3	Chondrocytes Chondrocytes + SBO	Mice Human	In Vivo	84, 86
Tissue inhibitors of MMP (TIMP-1)	Fibroblasts + SBO	Human	In Vitro	79, 84
COX2	Chondrocytes Monocyte/Macrophage-like cells Chondrocytes + SBO	Equine Human	In Vitro	84, 116, 115
Prostagladine-2 (PGE2)	Hyalin Chondrocytes Monocyte/Macrophage-like cells	Mice Equine Human	In Vitro	86, 92, 116
NF-kB	Hyalin Chondrocytes Nuclear translocation of p65	Mice Equine Human	In Vitro	86, 115
ERK1/2	Hyalin Chondrocytes	Mice Human	In Vitro	86
$\text{TNF-}\alpha$	Chondrocytes	Human	In Vitro	84
iNOS	Chondrocytes Monocyte/Macrophage-like cells	Human	In Vitro	84, 116
NO	Chondrocytes Monocyte/Macrophage-like cells	Human	In Vitro	116
oLDL	Osteoblasts	Human	Serum	104
Fibronectin	Chondrocytes	Human	In Vitro	81
Alkaline phosphatase	Osteoblasts	Human	In Vitro	

Table 2

Stimulatory effects of Avocado Soybean Unsaponifiable on anti-inflammatory, anabolic mediators that protect against osteoarthritis

Molecular Mediator	Target Tissue/Organ	Organism	Assay	References
Collagen synthesis	Articular synoviocytes Chondrocytes, Skin fibroblasts	Rabbit Bovine Human	In Vitro	81, 84, 93
Collagen II mRNA	Chondrocytes + Subchondral Bone Osteoblasts (SBO)	Human	In Vitro	84
Agrecan Proteoglycan	Chondrocytes SBO	Equine Human	In Vitro	84, 94
TGF-β1	Knee joint fluid Osseointegration in tibiae	Rat Canine	In Vivo In Vitro	84, 98, 100, 101
TGF-β2	Knee joint fluid	Canine	In Vitro In Vivo	98, 100
TGF-β3	Chondrocytes + SBO	Human	In Vitro	84
BMP-2	Osseointegration in tibiae	Rat	In Vivo	101
Osteocalcin	Chondrocytes + SBO	Human	In Vitro	84
Chondroprotector	Bone Implant	Rat	In Vivo	96
Delayed destruction of the joints	Radiological evaluation	Human	In Vivo	81
Plasminogen activator inhibitor 1 (PAI-1)	Chondrocytes Osteoblasts	Bovine	In Vitro	98

 Table 3

 Content analysis of supplements containing Avocado Soybean Unsaponifiable

Company/ Manufacturer	Brand Name	Dosage Form	Other Ingredients	Excipients on the label
Nuramax Laboratories, Inc.	Avoca ASU	Avocado/Soyben Unsaponifiables, non-shellfish glucosamine, NMX1000 <sup>®</sup>	OptiMSM® Methylsulfonylmeth Green tea extract	
Helseudsalg Faaborg Denmark	AvoSol	Avocado 100 mg / Soy 200 mg Unsaponifiables	Vitamin C 30 mg	Glucose syrup, ox gelatin, soy protein isolate, extract rich in tocopherol, silicon dioxide, magnesium salts from fatty acids.
Dr. Theos Official USA	Avosoy	Avocado-Soybean Unsaponifiables	Vitamin C 60 mg Vitamin E 30 U Manganese 2mg	Cellulose, dicalcium phosphate, sodium crosscarmelose, silicon dioxide, gum acacia, vegetable stearic acid, film coating, magnesium stearate
Dr. Theos Official USA	Avosoy Complete	Avocado-Soybean Unsaponifiables 300mg Glucosamine 1500mg Porcine chondroitin 800mg	Vitamin C 60 mg Vitamin E 30 U Manganese 2mg	
Swanson Health Products. Fargo, ND, USA	AvoVida	100 mg Unsaponifiables Persea gratissima Glycine max	30% β-sitosterol, campesterol, stigmasterol	Soy protein isolate, mixed tocopherols, silica Microcrystalline cellulose (plant fiber), gelatin, magnesium stearate.
Pharmin, USA, Llc. Formulation Technology, USA	Arthrocen 300 mg	Avocado 100 mg / Soy 200 mg Unsaponifiables	Persea gratissima Glycine max	Silica, magnesium stearate (E470b manufactured from vegetable oil), and gelatin fines.
Nuramax Laboratories, Inc.	Cosamin ASU	Avocado/Soyben Unsaponifiables	Glucosamine Sulafate Chondoitine Sulfate	
Maximum International USA	Maximize	ASU300-Avocado Soy Unsaponifiables SierraSil : 600 mg	Iron: 1.1 mg (from SierraSil) Iron toxicity recommendation on the container.	Microcrystalline cellulose, maltodextrin, croscarmellose sodium, silicon dioxide, stearic acid, hydroxypropylm ethylcellulose, hydroxypropylcellulos, magnesium stearate, polyethylene glycol.
Expanscience Lab., Courbevoie, France Pharmascience, Montreal Canada. Pharma Inv. Chile SA, Santiago Solvay Ins.Biol. Chenioterapicosells Microsules Y Bernabo Siegfried, Rhein	Piascledine 300	Avocado/Soybean Unsaponifiable	Not described	Butylated hydroxytoluene (BHT) 0.05 mg/Capsule Colloidal anhydrous silica

Christiansen et al. Page 27

Table 4
Adverse effects of Avocado Soybean Unsaponifiable

Organ	Side Effects	Frequencies	Drug withdrawal	References
Skin	Eczema Hives Photosensitivity Hypersensiticity syndrom	32.5%		130
Liver	Liver injury Bilirubin ALKP GGTP	16.2%	Return to Normal	130
Gastrointestinal	Regurgitation Heartburn Nausea Epigastric pain Dyspepsia Diarrhea Constipation Microscopic colitis	12%	Return to Normal	106, 129, 130
Coagulation	Platelets	6.8%		130

ALKP: alkaline phosphatases GGTP: Gamma-glutamyl transpeptidase