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# Coronary Artery Disease Associated Transcription Factor TCF21 Regulates Smooth Muscle Precursor Cells that Contribute to the Fibrous Cap

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# Abstract

TCF21 is a basic helix-loop-helix transcription factor that has recently been implicated as contributing to susceptibility to coronary heart disease based on genome wide association studies. In order to identify transcriptionally regulated target genes in a major disease relevant cell type, we performed siRNA knockdown of TCF21 in in vitro cultured human coronary artery smooth muscle cells and compared the transcriptome of siTCF21 versus siCONTROL treated cells. The raw (FASTQ) as well as processed (BED) data from 3 technical replicates per treatment has been deposited with Gene Expression Omnibus (GSE44461)

# **Specifications**

Organism/cell line/tissue	Human Coronary Artery Smooth Muscle Cells(HCASMC)
Sex	Male/female
Sequencer or array type	Illumina HiSeq2000
Data format	Raw and analyzed

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Experimental factors	siRNA treatment: siTCF21; siCONTROL
Experimental features	HCASMCs were purchased from Lonza and cultured according to the manufacturer's recommendations. Cells were serum-starved and transfected with siRNA pools from OriGene. Total RNA was collected and RiboZero libraries generated and sequenced on an Illumina HiSeq2000 instrument. The resulting reads were mapped to hg19 with TopHat(1)/Bowtie2(2). Differentially expressed genes were identified using either DESeq (3) and edgeR(4).
Consent	manufacturer's informed donor consent
Sample source location	n/a

## Direct link to deposited data

http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE44461

#### **Experimental Design, Materials and Methods**

#### Cell culture

Human primary Coronary Artery Smooth Muscle Cells (HCASMC, Lonza #CC-2583 Lot 200212 and Cell Applications # 350-05a Lot 1508) were cultured in Smooth Muscle Growth Medium-2 including hEGF, insulin, hFGF-B and FBS, but without antibiotics (Lonza, #CC-3182). For RNA-Seq studies donor-pooled HCASMC were transfected with 300 nM TCF21 Trilencer-27 Human siRNA (OriGene #SR304753C) or Trilencer-27 Universal Scrambled Negative Control siRNA (OriGene #SR30004) at 80% confluence using the Amaxa Basic Nucleofector Kit for Primary Mammalian Smooth Muscle Cells (Lonza #VPI-1004) at a density of  $1 \times 106$  cells per 100 µL sample using Nucleofector Program U-025. Cells were changed to medium with supplements at 18 hours post-transfection and cultured for an additional 48 hours.

#### **RNA** sequencing

Total RNA from either siTCF21 or siCTRL treated samples was depleted for ribosomal RNA with the Ribo-Zero magnetic kit from Epicentre (Illumina #MRZH116), libraries generated with the Epicentre ScriptSeq v2 RNA-Seq library preparation kit (Illumina #SSV21106) and thereafter sequenced as 100bp paired-end reads on an Illumina HiSeq 2000 instrument. The resulting data has been deposited at GEO under accession number GSE44461. Reads resulting from RNA-Sequencing of siCTRL and siTCF21 treated HCASMC were mapped using software tools TopHat+Bowtie2. Differential expression level between samples was analyzed using the software tools DESeq and edgeR at an FDR 0.05, with intersection of the 466 and 430 respective identified genes providing a group of 380 common genes.

### References

- Trapnell C, Pachter L, Salzberg SL. Tophat: Discovering splice junctions with rna-seq. Bioinformatics. 2009; 25:1105–1111. [PubMed: 19289445]
- Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. Genome Biol. 2009; 10:R25. [PubMed: 19261174]
- 3. Anders S, Huber W. Differential expression analysis for sequence count data. Genome Biol. 2010; 11:R106. [PubMed: 20979621]

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4. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics. 2010; 26:139–140. [PubMed: 19910308]

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