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## Recognition of influenza H3N2 variant virus by human neutralizing antibodies

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### Abstract

Since 2011, over 300 human cases of infection, especially in exposed children, with the influenza A H3N2 variant (H3N2v) virus that circulates in swine in the US have been reported. The structural and genetic basis for the lack of protection against H3N2v induced by vaccines containing seasonal H3N2 antigens is poorly understood. We isolated 17 human monoclonal antibodies (mAbs) that neutralized H3N2v virus from subjects experimentally immunized with an

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#### Author contributions

SB, TN, JAF, NJT, BSK, AC, JAR, RJW, ABW, and JEC designed the research study and experiments. SB, TN, JAF, NK, NJT, BSK, HGK, VS, RML, GS, AC, JAR, and ABW conducted laboratory experiments and acquired and analyzed data. BSK and RJW conducted animal studies. JCS gave statistical review. SE, LL, and KME conducted the parental vaccine study and acquired PBMCs. SB and JEC wrote the manuscript. All authors read and commented on the manuscript.

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H3N2v candidate vaccine. Six mAbs exhibited very potent neutralizing activity ( $IC_{50} < 200$  ng/ml) against the H3N2v virus but not against current human H3N2 circulating strains. Fine epitope mapping and structural characterization of antigen-antibody complexes revealed that H3N2v specificity was attributable to amino acid polymorphisms in the 150-loop and the 190-helix antigenic sites on the hemagglutinin protein. H3N2v-specific antibodies also neutralized human H3N2 influenza strains naturally circulating between 1995 and 2005. These results reveal a high level of antigenic relatedness between the swine H3N2v virus and previously circulating human strains, consistent with the fact that early human H3 seasonal strains entered the porcine population in the 1990s and reentered the human population, where they had not been circulating, as H3N2v about a decade later. The data also explain the increased susceptibility to H3N2v viruses in young children, who lack prior exposure to human seasonal strains from the 1990s.

## Introduction

Annual outbreaks of influenza A viruses (IAVs) in humans are a major global health problem, causing more than 250,000 deaths every year (1). In addition to yearly epidemics, novel influenza viruses originating from other animals periodically cross the species barrier to humans and cause pandemics with high morbidity and mortality rates. IAVs are enveloped viruses that contain the antigenic hemagglutinin (HA) and neuraminidase (NA) surface glycoproteins. HA encodes the receptor-binding site (RBS) and fusion peptide essential for attachment and entry into the host cell and is the primary target for potent neutralizing antibodies (2). The globular head domain that contains the sialic acid-binding (SA-binding) pocket is the major antigenic portion of the HA and tolerates high sequence variability. As a consequence, influenza viruses undergo constant antigenic drift that allows escape from antibody-mediated immunity. There are currently 18 known subtypes of IAVs that fall into 2 broad groups based on the HA sequences and phylogeny (3). Of these, only H1 and H3 subtypes currently circulate in humans. Preferential binding of particular HA molecules to different types of SA receptors on host cells is the major determinant of host specificity (4). The HA of avian IAVs has high affinity for  $\alpha$  2,3-linked SA, whereas human influenza viruses have high affinity for  $\alpha$  2,6-linked SA (4–7).

The IAV genome is segmented, and the virus is capable of superinfecting cells with a heterologous IAV in a single animal. These features allow for reassortment of the influenza genome in intermediate hosts, such as swine or poultry, enabling emergence of strains that are capable of crossing the species barrier to humans (8). In particular, swine may act as a mixing vehicle for IAVs, because their upper respiratory tract epithelial cells possess both  $\alpha$  2,3- and  $\alpha$  2,6-linked SA receptors, which allow infection with both avian and human IAV (6). Although swine influenza viruses do not generally infect humans, sporadic cases of human infections with swine H1N1 and H3N2 have been documented since 1958 (9). Reassorted swine influenza viruses that are capable of infecting humans can cause severe disease and pose a pandemic threat due to lack of preexisting immunity to the virus. The H1N1 influenza pandemic in 2009–2010 was associated with a virus of swine origins and is an example of a swine virus that was able to transmit easily in the human population and cause disease (10).

Influenza viruses that circulate in pigs are designated “variant” viruses when they cause human infections. Swine-origin IAV H3N2v viruses containing the matrix gene from the 2009 H1N1 pandemic virus were first detected in humans in July 2011. Since then, there have been at least 345 reported cases of human infections with H3N2v viruses, with a high prevalence in children (11–13). A recent study showed that all children <5 years old and >80% up to 14 years old lack protective serum antibody titers against H3N2v (14). Most cases of H3N2v-associated disease have been associated with exposure to swine, with very limited human-human transmission (12). H3N2v is antigenically distinct from the currently circulating H3N2 seasonal strains, and it has been determined that vaccination with 2010–2011 annual trivalent inactivated virus does not induce neutralizing antibodies against the variant H3N2 virus (14). Lack of preexisting immunity to the variant virus, especially in children, may be a major concern if a highly transmissible H3N2v outbreak occurs (14–16).

Here, we describe the characterization of human mAbs to H3N2v HA isolated from individuals vaccinated with an experimental monovalent inactivated H3N2v vaccine candidate. We used these mAbs to define the molecular basis of strain specificity or cross-reactivity for human neutralizing antibodies recognizing the HA of H3 seasonal or emerging H3 variant viruses. The results indicate that polymorphisms in the 150 helix and the 190 loop, located near the RBS on HA, play a major role in escape of H3N2v virus from immunity induced by seasonal H3N2 vaccines. Furthermore, our results reveal that the HA protein of H3N2v strains is antigenically similar to the human H3N2 IAV strains that circulated during the late 1990s, during which several H3N2 spillover events have been suggested to occur from humans into US swine (17–19).

## Results

### Isolation of H3N2v-reactive human mAbs from vaccinated donors

Healthy adult donors received 2 doses of subvirion H3N2v vaccine (15 µg of HA/dose) 21 days apart in an open-label trial, the results of which were reported previously (20). Peripheral blood samples were obtained from volunteers after informed consent on the day of vaccination (day 0) and 21 days after the second dose of vaccine (day 42). Cryopreserved peripheral blood mononuclear cells (PBMCs) were immortalized by EBV transformation, and we collected supernatants from the resulting lymphoblastoid cell lines. Supernatants were screened by ELISA for binding to recombinant HA protein from the H3N2v strain A/Minnesota/11/2010 (designated here as the MNv strain) or one of two representative H3N2 seasonal strains, A/Victoria/361/2011 (designated here as the Victoria strain) or A/Wisconsin/67/2005. The frequency of H3N2 seasonal or variant-reactive B cell lines was reported previously in the description of the vaccine trial results, and those studies indicated a significant B cell response to the H3N2v HA among the vaccinated individuals (20). The majority of the B cells that secreted H3 HA-reactive antibodies on day 42 recognized the variant HA specifically, with limited cross-reactivity between variant virus-reactive antibodies secreted by B cells and those secreted by seasonal virus-reactive B cells (Supplemental Figure 1; supplemental material available online with this article; doi: 10.1172/jci.insight.86673DS1). Transformed B cell lines with supernatants that showed reactivity against the MNv HA in ELISA were selected for fusion to generate human

hybridoma cell lines secreting mAbs. We used PBMCs collected on study day 42 from a total of 12 subjects to isolate 36 cloned hybridomas secreting mAbs. The IgG subclass and light chain type of the 17 neutralizing antibodies are presented along with the donor number for the sample from which they were isolated (Figure 1).

### Binding and neutralization profile of H3N2v mAbs

The neutralization potential of the clones in this antibody panel against MNv virus was determined by microneutralization assay using the MNv virus. Seventeen of thirty-six mAbs exhibited neutralizing activity against the variant virus when tested in concentrations as high as 10  $\mu\text{g/ml}$  (Figure 1). The  $\text{IC}_{50}$  values are shown as a heat map, with increased color intensity corresponding to an increase in neutralizing potency. Six mAbs (found at the top of Figure 1 and designated as H3v-126, H3v-71, H3v-47, H3v-104, H3v-98, and H3v-81) showed very potent neutralization against the virus, with  $\text{IC}_{50}$  values of less than 100 ng/ml. We also determined the ability of 4 mAbs (H3v-98, H3v-104, H3v-71, and H3v-45) to neutralize 4 H3N2 strains representing each antigenic cluster circulating in swine, A/Swine/Texas/4199-2/98 (cluster I), A/Swine/Colorado/23619/99 (cluster II), A/Swine/Oklahoma/18089/99 (cluster III), and A/Ohio/13/2012 (cluster IV) (Supplemental Table 1). H3v-98 and H3v-104 showed potent inhibiting activity against 3 of 4 strains. H3v-71 and H3v-45 exhibited activity against 2 strains at low concentrations.

The 17 H3N2v-neutralizing mAbs were tested for binding to MNv and 3 seasonal strains A/Perth/16/2009, Victoria, and A/Texas/50/2012. All of the antibodies had half-maximal effective concentrations ( $\text{EC}_{50}$ ) for binding below 100 ng/ml to MNv HA (Figure 1). Increasing intensity of the orange cell fill color in the  $\text{EC}_{50}$  column corresponds to increasing binding for the indicated HA. The antibodies displayed a differential binding pattern — 9 mAbs bound specifically to the MNv HA, while the other 8 bound to both the variant HA and a combination of seasonal HAs (Figure 1). Interestingly, all but one (H3v-47) of the potently neutralizing mAbs exhibited a variant-specific binding phenotype without detectable cross-reactivity for the HA of H3 seasonal strains (Figure 1) or other HA subtypes (Supplemental Table 2).

We determined the nucleotide sequence of the antibody heavy chain variable gene regions (Table 1). The 17 neutralizing antibodies had unique HCDR3 sequences, indicating that the mAbs represented independent clones. We also performed next-generation sequence analysis of antibody gene repertoires for a subset of donors (donors 10, 27, 37, and 41). Interestingly, we were able to identify on the day of immunization (day 0) a member of the clonal lineage for 2 of the 6 mAbs obtained from these donors, suggesting that the clone was induced by prior infection (Supplemental Table 3). The frequency of those clones was greatly expanded on day 7, the expected day of peak plasmablast circulation in peripheral blood (Supplemental Table 3). A clonal lineage of one of the clones, H3v-104, is shown in Supplemental Figure 2, with the amino acid sequence alignment of 296 clonal variants shown in Supplemental Table 4.

### Potently neutralizing mAbs block the RBS

Due to the HA specificity demonstrated by the mAbs, we anticipated that the majority of the potently neutralizing mAbs mediated neutralization by binding to the less conserved head domain of HA. We performed hemagglutination inhibition (HAI) assays in order to identify mAbs that interfered with the receptor-binding function of HA. A total of 9 of the 17 neutralizing mAbs exhibited HAI activity against the MNv virus, suggesting that these mAbs function by blocking virus binding to the SA receptor (Figure 1). The most potent mAbs that displayed variant-specific binding phenotype also exhibited HAI activity, suggesting that the variant-specific polymorphisms around the RBS in the head domain region on MNv HA play a major role in determining the unique antigenic profile of the variant virus.

### In vivo efficacy of H3v-126, H3v-104, and H3v-71

We tested representative mAbs as prophylaxis in a mouse challenge study. Groups of 6- to 8-week-old female DBA/2J mice at 5 animals per group were injected with 100  $\mu$ g of individual mAbs by the intraperitoneal route on the day prior to virus challenge. Controls ( $n = 10$ ) were injected with PBS. The modestly increased body weight for all 3 groups of mAb-treated animals when compared with the PBS-treated animals was not statistically different (Figure 2).

### Competition-binding studies

In order to determine if the neutralizing mAbs bound to common or diverse epitopes on HA, we performed competition-binding assays using biolayer interferometry with all of the neutralizing antibodies. Fifteen of the seventeen neutralizing mAbs were classified into competition-binding groups based on their ability to block other mAbs from binding to the HA; we were unable to detect good binding signal for H3v-81 or H3v-7 with biolayer interferometry. The 15 neutralizing mAbs tested fell into 3 major competition-binding groups, with some overlap between the groups designated groups 1 and 2 and a distinct group 3 (Figure 3). Notably, 4 of the most potent neutralizing clones (H3v-98, H3v-104, H3v-126, and H3v-71) that also exhibited HAI activity and variant specificity fell into the same competition-binding group, group 1.

### Mutagenesis experiments and electron microscopy revealed regions on the H3N2v HA important for its immune escape from antibodies induced by seasonal H3N2 viruses

The 6 most potently neutralizing mAbs that displayed a variant-specific phenotype (i.e., mAbs that bound specifically to the MNv HA and not to the seasonal HAs) were chosen for fine epitope mapping. We sought to determine the residues important for immune escape of the H3N2v virus from antibodies induced by seasonal vaccines. We considered that the HA of MNv has 52 polymorphisms as compared with the Victoria seasonal strain. We initially performed a mutagenesis screen by introducing variant-specific polymorphisms into a cDNA encoding the Victoria HA protein to identify mutations that would enhance binding to the variant-specific mAbs. 17 variant-specific polymorphisms were introduced into Victoria HA as single or double mutations. Three double-mutant HA molecules, Victoria I202V/T203I,

A163E/L164Q, and R142G/N144V, enhanced binding to mAbs H3v-98 and H3v-104 in comparison to wild-type Victoria HA (Supplemental Figure 3).

Based on the results of the initial mutagenesis screen, we targeted the region around the RBS and introduced residues from seasonal strains into the H3N2v HA to identify residues that disrupted binding to wild-type MNv. The EC<sub>50</sub> values for MNv HA mutants that disrupt binding to each antibody are shown with representative binding curves (Figure 4). Mutation of Y155T/N156K/L157S disrupted binding to both mAb H3v-71 and mAb H3v-81, whereas mutant MNv N158G/Y159S/K160T did not bind to mAb H3v-71 and MNv K189N/T192I/N193R did not bind to H3v-81. A single-mutation Y137S disrupted binding to H3v-141. In addition, a triple alanine mutant, MNv L194A/Y195A/V196A, did not bind to mAbs H3v-98, H3v-104, H3v-71, H3v-81, and H3v-104.

We also performed electron microscopy (EM) of MNv HA in complex with Fab portions of the 3 most potent variant-specific antibodies, H3v-104, H3v-126, and H3v-71 (Figure 5, A–C), along with the Fab of a stem-binding antibody CR9114 (used as reference in each complex). The EM reconstructions revealed that all 3 antibodies bound to the RBS on HA, with overlapping footprints (Figure 5D). This finding was consistent with the competition-binding data and demonstrated that H3v-126, H3v-71, and H3v-104 all fall under the same competition-binding group (group 1). The H3v-104 mAb displayed a comparatively broader footprint on HA, extending below the RBS, supporting our previous observation that the Victoria R142G/N144V mutant enhanced binding to mAb H3v-104 but not to H3v-126 or H3v-71. H3v-71 bound similarly to H3v-126 but appeared to interact more with the upper rim of the RBS (Figure 5D). This finding could explain why mutations in the 190 helix and 150 loop affected binding of H3v-71 but not of H3v-126. Collectively, the results from mutagenesis and EM structural studies indicated that variant-specific HA residues residing in 2 major structural features surrounding the RBS, the 150 loop and the 190 helix, principally account for antigenic distinction of the variant virus from current seasonal strains.

### **The HA of H3N2v virus is antigenically related to that of older human seasonal H3N2 IAIVs**

The majority of the severe cases of influenza infection caused by H3N2v viruses to date in the US have occurred in children, suggesting partial immunity in adults. We sought to identify the nature of preexisting immunity to the variant virus in the adult population that might provide partial protection against severe disease. Soluble HA proteins belonging to 12 different seasonal H3N2 strains that circulated in humans from 1968 to 2013 were cloned, expressed, and assayed for binding to 6 potentially neutralizing mAbs with variant-specific binding and neutralizing phenotypes. Interestingly, 3 mAbs, H3v-98, H3v-104, and H3v-71, showed strong binding to H3N2 viruses that circulated between 1997 and 2004 but not to the HA of H3 subtype strains that circulated before or after that period (Supplemental Figure 4). To determine if these mAbs had the potential to neutralize these older seasonal strains, we performed microneutralization assays to test activity against 12 viruses that circulated between 1968 and 2013. Indeed, H3v-98 and H3v-104 neutralized all of the strains tested between 1995 and 2005, whereas H3v-71 neutralized all of the viruses between 1997 and 2002 (Figure 6). We used a representative panel of live-virus H3 seasonal strains for the

neutralization assays that matched as closely as possible the strains that we had used to make recombinant HA molecules for the seasonal strain HA-binding assays. Although we did not detect binding for mAbs H3v-98 or H3v-104 to the HA from A/New York/55/2004 or A/Hiroshima/52/2005, we did observe potent neutralization against the A/Wisconsin/67/2005 strain that was available for neutralization testing. In summary, prior infection with seasonal H3N2 strains that circulated during 2004 and 2005 might induce antibodies that cross-react with H3N2v viruses but likely does not guarantee the presence of protective antibodies against H3N2v because of antigenic heterogeneity.

## Discussion

We report here the isolation of human mAbs against H3N2v virus and the use of them to determine the molecular basis for the antigenic distinction between H3 seasonal strains and the H3N2v viruses. Seventeen antibodies neutralized MNv H3N2v at concentrations of less than 10  $\mu\text{g/ml}$ . Four of the seventeen neutralizing mAbs exhibited ultrapotent neutralizing activity ( $\text{IC}_{50} < 10 \text{ ng/ml}$ ). Three of these four ultrapotent neutralizing H3N2v mAbs also displayed potent inhibiting activity against swine H3N2 strains belonging to different antigenic clusters. Collectively, these results suggest that some of these H3N2v mAbs could be used in humans as therapeutics against many H3N2 strains circulating in swine in the case of zoonotic transmission event.

The H3 HA-binding breadth of IgG secreted by memory B cells from individuals vaccinated with monovalent inactivated H3N2v revealed that there is limited cross-reactivity between antibodies secreted by variant-reactive B cells and H3 seasonal virus-reactive B cells. About half of the H3v-reactive clones that were isolated displayed specific binding to the MNv HA, with no detectable cross-reactivity for HAs from 3 recent seasonal H3N2 strains. The potentially neutralizing variant-specific antibodies clustered into the same competition-binding group and used the same virus neutralization mechanism by blocking the RBS on HA. This finding suggested that common antibodies induced by seasonal trivalent inactivated influenza virus (TIV) vaccination that recognize immunodominant epitopes adjacent to the RBS on HA fail to contribute to cross-protection against variant virus. A recent study showed that the molecular basis for antigenic drift in human H3N2 seasonal strains from 1968 to 2006 was attributed to 7 single amino acid substitutions at positions 145, 155, 156, 158, 159, 189, and 193 in HA (21). Six of these residues are located in the antigenic site B (upper rim of RBS). Fine-epitope mapping of 5 potentially neutralizing H3N2v-specific mAbs revealed a similar pattern in context to H3N2v antigenicity. The residues important for variant specificity of these mAbs were located primarily in the 150-loop and 190-helix antigenic elements near the RBS.

Historically, swine IAVs have caused sporadic human infections, but these outbreaks had been mostly self-limiting. The 2009 H1N1 pandemic was an example of a swine IAV variant, containing the same subtype of HA and NA as that of the circulating human strain, which caused a human pandemic. A recent rise in swine-origin H3N2v human infections has raised concerns of another swine-origin influenza virus pandemic (11, 13). Surveillance of IAVs in live-animal markets in Minnesota that included human, swine, and environmental samples provided evidence of interspecies transmission of IAVs from swine to human,

signifying a potential risk to persons attending live markets (22). Moreover, H3N2 was identified as the predominant subtype circulating in the swine population; a majority of these swine H3N2 isolates had the same genomic constellation as the H3N2v viruses isolated previously from humans (22). Vaccination with seasonal TIV does not provide protection against H3N2v viruses, indicating these swine-origin viruses have the potential to spread within unprotected human populations (14).

The HA from the swine-origin 2009 pandemic H1N1 strain is antigenically similar to human H1N1 viruses that circulated between 1918 and 1943 (23). It is hypothesized that IAV classical swine H1N1 was introduced into the domestic swine population sometime during the 1918 human pandemic and remained relatively static in swine for greater than 80 years until the end of the 20th century (24, 25). In light of the 2009 pandemic, swine have been thought to act as a potential frozen reservoir of human IAVs, which can result in pandemics through reintroduction of the human-origin virus into a then susceptible human population at a remote time decades later (23). The reactivity of the human mAbs that we isolated from H3N2v vaccines revealed that the same phenomenon may be occurring now with H3 viruses, as human H3 seasonal strains from the past may be harbored in an antigenically static manner in farm swine in the US.

During late 1990s, there were several spillovers of H3N2 virus from humans to pigs resulting in the introduction of H3N2 viruses into the US swine population (17–19). The H3N2v viruses that were isolated from humans in 2011 were phylogenetically closer to seasonal strains that circulated during late 1990s than to current H3 human strains (19, 26). We hypothesized that the swine-origin H3N2v viruses might be antigenically related to the seasonal H3N2 viruses from late 1990s, and thus the H3N2v-specific antibodies that showed no reactivity against the current seasonal strains might recognize the older seasonal viruses. Indeed, we found that 3 potentially neutralizing variant-specific mAbs (H3v-98, H3v-104, and H3v-71) had the ability to neutralize seasonal strains that circulated between 1997 and 2002. Additionally, H3v-98 and H3v-104 also neutralized strains from 1995 and 2005. These results indicate that the H3N2v viruses that caused human infections from 2011 to 2013 are antigenically related to the human ancestral H3N2 strains that circulated from 1995 to 2005. Taken together, the data suggest that children who were born after 2005 do not have any preexisting cross-reactive immunity against H3N2v virus because they were not exposed to the earlier H3 seasonal strains. Therefore, these children should be the priority target population for vaccination to prevent an H3N2v outbreak.

The phenomenon of antigenic similarity between swine IAVs and ancestral human strains might be explained by several mechanisms. First, swine may act as a relatively static reservoir of human IAVs. Second, differences in the location of the immunodominant antibody epitopes on HA for swine and humans might allow preservation of antigenic epitopes targeted by humans. We compared the HA1 sequences from H3N2v Minnesota virus and the seasonal Victoria 2011 virus with that of the Sydney 1997 strain (a possible ancestor to the H3N2v Minnesota virus). The HA1 subunit of H3N2v Minnesota and H3N2 Victoria strains has 37 or 32 polymorphisms, respectively, when compared with the HA1 sequence of H3N2 Sydney 1997, suggesting that these strains have drifted at a relatively similar rate in swine compared with the drift in humans. Thirteen of these positions that are



situated primarily in the upper or lower rim of the RBS (antigenic sites A and B) showed variation in both Minnesota and Victoria HA. In contrast, there were comparatively more polymorphisms in the H3N2v Minnesota HA sequence in antigenic sites C and E, whereas polymorphisms in the 220-loop (antigenic site D) were seen predominantly in the Victoria 2011 strain. These host-based differences in the observed immunodominance pattern provide a possible explanation of why H3N2 viruses in the swine population retain antigenic features of their human ancestral strains. This recurring phenomenon, in which swine populations act as a static/divergent antigenic reservoir of previously circulating human IAVs should thus be taken into serious consideration for pandemic preparedness.

## Methods

### Influenza viruses

The seed stock of H3N2v strain MNv was obtained from Terrence Tumpey (US CDC). The working stocks used for microneutralization assay and HA inhibition assays were made from the supernatant of virus-infected MDCK cell culture monolayers in plain Dulbecco's Modified Eagle Medium (Gibco DMEM, Invitrogen, 11965) with 2 µg/ml of TPCK-trypsin. The seasonal H3N2 strains A/Fujian/411/2002 (FR-1146), A/Perth/16/2009 (FR-370), A/Wisconsin/67/2005 (FR-397), and A/Texas/50/2012 (FR-1210) were provided by the Influenza Reagent Resource (<http://www.influenzareagentresource.org/>) of the US CDC. Two H3N2 seasonal strains, Victoria (NR-44022) and A/Sydney/5/1997 (NR-12278), were obtained from BEI Resources.

### Recombinant soluble HA proteins

Sequences encoding the HA genes of interest were optimized for expression, and cDNAs were synthesized (Genscript) as soluble trimeric constructs by replacing the transmembrane and cytoplasmic domain sequences with cDNAs encoding the GCN4 trimerization domain and a His-tag at the C-terminus. Synthesized genes were subcloned into the pcDNA3.1(+) mammalian expression vector (Invitrogen). HA protein was expressed by transient transfection of 293F cells with polyethylenimine transfection reagent and was grown in expression medium (Freestyle 293 Expression Medium; Invitrogen, 12338). The supernatants were harvested after 7 days, filter-sterilized with a 0.4-µm filter, and purified with HisTrap TALON FF crude columns (GE Healthcare Life Sciences).

### PBMC isolation and hybridoma generation

A cohort of 25 donors was vaccinated twice, 21 days apart, with 15 µg of HA/0.5-ml dose of reassortant MNv NYMC X-203, as part of NIH-sponsored clinical research trials of this experimental vaccine (DMID protocol 12-0011). The details of the clinical trial were described previously (20). PBMCs from these donors were isolated from day 0 (day of first vaccination) and protocol day 42 (3 weeks after the second dose of vaccine) by density gradient separation on Ficoll and cryopreserved. The human lymphocytes from day 42 were thawed and immortalized by transformation with EBV substrain B95.8 in the presence of CpG10103, cyclosporin A, and a Chk2 inhibitor (27). The cells were plated in a 384-well plate, and 8 days later the supernatants from these transformed B cells were used to screen for the presence of antibodies that bound to soluble H3N2 MNv HA using capture ELISA.

The positive wells containing B cells secreting anti-H3N2v antibodies were expanded onto irradiated human PBMC feeder layers for 4 days and then fused with HMMA2.5 myeloma cells using a Cytopulse PA4000 electrofusion device. After fusion, human hybridomas were selected in medium with HAT solution containing ouabain, and several rounds of limiting dilution passages were performed in 384-well culture plates to isolate cell lines of the hybridomas with the highest level of secretion of IgG (27).

### **mAb production and purification**

The hybridoma cell lines with the highest level for IgG expression for each clone were selected as single cells using flow cytometric sorting to obtain clones secreting mAbs. Once hybridoma clones were obtained following sorting and growth in a 384-well plate, we expanded them first into wells of a 48-well plate and then further into a 75-cm<sup>2</sup> flask to 70% confluency in hybridoma growth medium (ClonaCell-HY medium E from STEMCELL Technologies, 03805). The cells then were washed and expanded equally to four 225-cm<sup>2</sup> flasks for antibody expression in serum-free medium (GIBCO Hybridoma-SFM, Invitrogen, 12045084). The supernatant was harvested after 3 weeks, filtered with a 0.4- $\mu$ m filter, and the monoclonal IgGs were purified by affinity chromatography using protein G columns (GE Life Sciences, Protein G HP Columns).

### **EC<sub>50</sub> binding analysis**

The EC<sub>50</sub> concentration for each antibody was determined as described previously (28). Briefly, we performed ELISA using plates coated with the HA of interest at 2  $\mu$ g/ml overnight at 4°C and then blocked with 5% nonfat dry milk, 2% goat serum, and 0.1% Tween-20 in PBS for 1 hour. Three-fold dilutions of the mAb starting from 10  $\mu$ g/ml were added to the wells and incubated for 1 hour, followed by incubation for 1 hour at 1:4,000 dilution of anti-human IgG alkaline phosphatase conjugate (Meridian Life Science, W99008A). The plates were washed 3 times between each step with PBS containing 0.1% Tween-20. Phosphatase substrate solution (1 mg/ml p-nitrophenol phosphate in 1 M Tris aminomethane) was added to the plates and incubated for 1 hour, and the optical density values were measured at 405-nm wavelength on a BioTek plate reader. Each dilution was done in triplicate, and the EC<sub>50</sub> values were calculated in Prism software (GraphPad) using nonlinear regression analysis.

### **HAI and neutralization assays to determine IC<sub>50</sub> values**

Neutralization potential of all of the mAbs was determined by microneutralization assay and HAI assay. For microneutralization, 50  $\mu$ l of 2-fold serial dilutions of each antibody starting at 20  $\mu$ g/ml was incubated with 50  $\mu$ l of 100 TCID<sub>50</sub> of the virus in viral growth medium (VGM) for 1 hour at room temperature. VGM consists of plain DMEM with 2  $\mu$ g/ml of TPCK-trypsin and 50  $\mu$ g/ml gentamicin. The MDCK cell monolayer cultures were washed 2 times with 100  $\mu$ l PBS containing 0.1% Tween-20, and the virus-antibody mixture then was added to cells and incubated for 32 hours at 37°C. The cells were washed again and fixed with 100  $\mu$ l of 80% methanol/20% PBS. The presence of influenza nucleoprotein in the fixed cells was determined by ELISA using a 1:8,000 dilution of mouse anti-NP antibody (BEI Resources, NR 4282) as the primary antibody and a 1:4,000 dilution of goat anti-mouse alkaline phosphate conjugate as the secondary antibody (ThermoFisher Scientific,

31320). Each dilution was tested in duplicate and the half-maximal inhibitory concentration ( $IC_{50}$ ) was determined by nonlinear regression analysis of  $\log_{10}$  (inhibitor) vs. response function, using Prism software (GraphPad). An  $IC_{50}$  value of 2  $\mu\text{g/ml}$  was used as the threshold to determine the presence of functional neutralization. For performing the HAI assay, we used turkey red blood cells (Rockland) that were diluted to 0.5% in Alsever's solution (Sigma, A3551). 25  $\mu\text{l}$  of 4 hemagglutination units of virus were incubated with 25  $\mu\text{l}$  of 2-fold dilutions of the mAb, starting at 10  $\mu\text{g/ml}$  in PBS for 1 hour at 37°C. 50  $\mu\text{l}$  of the virus-antibody mixture was incubated with turkey red blood cells for 1 hour at room temperature. The HAI titer was defined as the highest dilution of antibody that inhibited hemagglutination of red blood cells. Each dilution was performed in duplicate.

### **In vivo efficacy of H3N2v mAbs**

On the day prior to infection, 6- to 8-week-old female DBA/2J mice (Jackson Laboratories, 000671) ( $n = 5$ ) were injected by i.p. route with 1, 10, or 100  $\mu\text{g}$  of antibody H3v-71, H3v-104, or H3v-126 in 50  $\mu\text{l}$  PBS. Controls were treated with PBS ( $n = 10$ ). On day 0, mice were inoculated intranasally with  $10^7$  PFU of A/Minnesota/10/11 X-203 virus under isoflurane anesthesia and then weighed daily. All treated mice survived for the duration of the study. One animal treated with PBS alone died.

### **Competition-binding groups**

Biolayer interferometry using an Octet Red instrument (ForteBio) was used to confirm mAb-HA binding and to perform competition-binding assays. The HA was loaded onto ForteBio Ni-NTA tips at a concentration of 25  $\mu\text{g/ml}$ , and binding to 2 successively applied mAbs at 100  $\mu\text{g/ml}$  was tested. All of the dilutions were made in 1X kinetic buffer (ForteBio, 18-5032). The actual binding signal for each mAb was obtained after 300 seconds of a single association step of the mAb on to HA. If binding of the first antibody blocked the binding of the second antibody by reducing its actual binding signal by more than 70%, it was defined as a competitor. If binding of the first antibody did not block the binding of the second antibody by reducing its actual binding signal by less than 30%, it was defined as a noncompetitor. A signal reduction between 30% and 70% was defined as partial blocking.

### **Site-directed mutagenesis of genes encoding HA proteins**

Primers for site-directed mutagenesis were designed using the Agilent QuikChange Primer Design program (Agilent Technologies). The QuikChange Lightning Multi-site Mutagenesis kit (Agilent, 210515-5) was used to introduce multiple mutations into cDNAs encoding the HA genes of H3N2 MNv or H3N2 Victoria, according to the manufacturer's instructions. These mutant HAs were tested for antibody binding in ELISAs, as above, to determine  $EC_{50}$  values for binding and to identify amino acids that comprise the epitope.

### **EM**

SEC-purified MNv complexed with Fab H3v-104 and Fab CR9114 was diluted to 14.7  $\mu\text{g/ml}$ , applied to freshly glow-discharged 400-mesh carbon-coated copper grids, and negatively stained with 2% uranyl formate. Similar complexes containing MNv and Fab CR9114 were then prepared separately with both Fab H3v-126 and Fab H3v-71 at 7.2  $\mu\text{g/ml}$

and 5.8  $\mu\text{g/ml}$ , respectively. The complexes containing Fab H3v-104 or Fab H3v-126 were imaged at  $\times 92,000$  magnification on an FEI Talos at 200 keV, resulting in a pixel size of 1.57  $\text{\AA}/\text{pixel}$  (calibrated using catalase crystal diffraction), with a dose of 23.96  $\text{e}/\text{\AA}^2$ . The complex containing Fab H3v-71 was imaged at  $\times 52,000$  magnification on an FEI Tecnai T12 at 120 kV TEM, resulting in a pixel size of 2.05  $\text{\AA}/\text{pixel}$ , with a dose of 25.41  $\text{e}/\text{\AA}^2$ . All data were collected using Legikon Multi-Scale Imaging (MSI-raster 3.1) software (29). The Talos is equipped with an FEI Ceta 4 k  $\times$  4 k CMOS and the T12 TEM is equipped with a Teitz F416 4 k  $\times$  4 k CMOS.

### EM data processing

For the complex containing Fab H3v-104 DoGpicker was used to automatically select particles from 450 raw micrographs that were then binned by 2, resulting in a 3.14  $\text{\AA}/\text{pixel}$  size, and placed into 128  $\times$  128 pixel boxes (30). Particles were aligned with Iterative MRA-MSA and ISAC, resulting in a final stack of 3,618 raw particles (31, 32). Class averages from ISAC were used to create a common lines initial model in EMAN2 (33). Model refinement was conducted in EMAN, resulting in a 21.3  $\text{\AA}$  resolution reconstruction based on a 0.5 FSC cutoff value (34). The same processing pipeline was used to prepare a reconstruction of the complex containing Fab H3v-126 from 306 raw micrographs and a final stack of 3,202 particles. Because of variable occupancy of the Fab several rounds of MRA-MSA were conducted to isolate particles containing a single-bound Fab, H3v-126. A model of HA bound with Fab CR9114 only was used as an initial model for refinement, resulting in a resolution of 19.8  $\text{\AA}$ . The complex containing Fab H3v-71 also had variable Fab occupancy. Five rounds of MRA-MSA and one round of ISAC were conducted to isolate particles with only 1 Fab H3v-71 bound, resulting in a final stack of 3,931 particles in 192 pixel boxes. A model of HA bound with Fab CR9114 only was used as an initial model for refinement, resulting in a resolution of 27.2  $\text{\AA}$ .

### Antibody heavy and light chain variable gene sequence analysis

Antibody heavy and light chain genes for each of the neutralizing mAbs were cloned from the hybridoma lines after single-cell flow cytometric sorting to biologically clone the cell lines. RNA was extracted from these hybridoma clones using the RNeasy mini kit (Qiagen, 74106), followed by RT-PCR amplification of antibody gene cDNAs. PCR products encoding antibody heavy or light chain genes were cloned individually into the pGEM-T vector, and Sanger nucleotide sequence analysis was used to determine the antibody cDNA sequences. Analysis of variable gene sequences was performed using the international ImMunoGeneTics (IMGT) information system (<http://imgt.org/>). We also performed next-generation sequence analysis of antibody gene repertoires from 4 selected donors and defined in detail clonal lineage for 1 clone. First, antibody heavy chain variable gene sequences were obtained using RT-PCR and Illumina MiSeq 2  $\times$  300 paired-end amplicon sequence analysis from PBMCs collected on day 0, 7, or 42 after immunization. For donor 41 and the H3v-104 clone, we identified those sequences sharing use of the  $V_{\text{H}}1-69$  and  $J_{\text{H}}4$  gene segments that encode H3v-104; the complete repertoire sequence database of donor 41 is available at the NCBI Sequence Read Archive at accession SRP075907. Then, CD-HIT was used to identify the HCDR3 sequences of 296 sequences that clustered within 85% identity with the HCDR3 sequence of H3v-104. We built a Phylip lineage using the

alakazam and shazam packages of the Change-O software suite and visualized the network in Cytoscape.

### Statistics

The IC<sub>50</sub> values were calculated after log transformation of antibody concentrations using a 3-parameter nonlinear fit analysis of antibody log<sub>10</sub> concentration vs. response with R<sup>2</sup> values > 0.85. The EC<sub>50</sub> values were calculated after log transformation of antibody concentrations using sigmoidal dose-response nonlinear fit analysis with R<sup>2</sup> values of > 0.85. Change in mouse weights over days 1–8 were compared by antibody using repeated-measures analysis of covariance, controlling for weight at day 0. All statistics were analyzed using Prism software version 5 (GraphPad). A *P* value less than 0.05 was considered significant.

### Study approval

PBMCs were collected at the Emory University Vaccine Treatment and Evaluation Unit after informed consent from otherwise healthy subjects with prior history of experimental H3N2v subunit vaccination, as described in the Methods. The protocol and consent form were approved prior to study by the Emory University Institutional Review Board Committee, Atlanta, Georgia, USA. The animal protocol covering the H3N2v influenza virus challenge infections of passively immunized mice was reviewed and approved by the Institutional Animal Care and Use Committee at St. Jude Children's Research Hospital.

### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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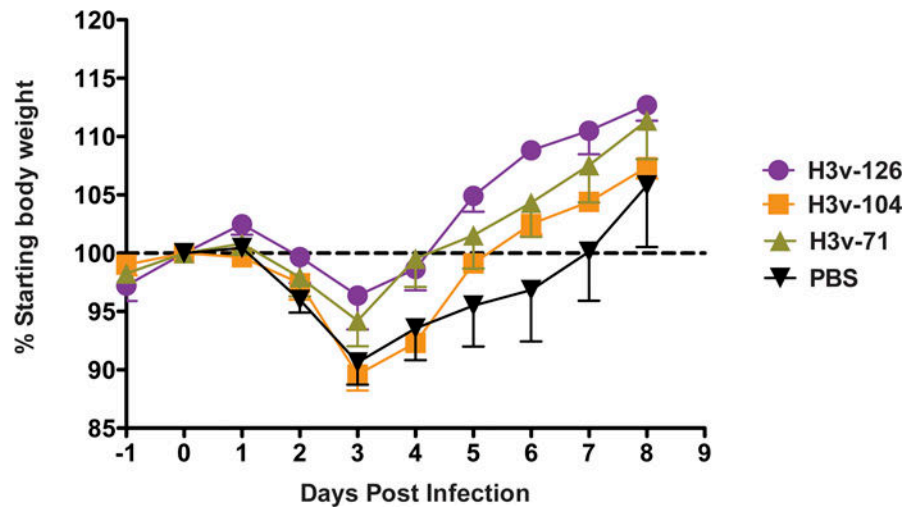
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Monoclonal antibody	Subject number	IgG subclass	Light chain	EC <sub>50</sub> (ng/mL) for indicated strain				H3N2v IC <sub>50</sub> (ng/mL)	H3N2v HAI (µg/mL)
				H3N2v Minnesota 2010	H3N2 Perth 2009	H3N2 Victoria 2011	H3N2 Texas 2012		
H3v-126	27	1	λ	3	>	>	>	3	0.16
H3v-71	37	1	λ	10	>	>	>	7	0.31
H3v-47	10	1	κ	4	18	26	38	8	>5.0
H3v-104	41	1	κ	14	>	>	>	9	0.16
H3v-98	37	1	κ	19	>	>	>	14	0.32
H3v-81	37	1	λ	29	>	>	>	59	1.25
H3v-141	54	1	κ	25	>	>	>	137	1.25
H3v-45	9	3	λ	14	>	>	>	257	1.25
H3v-95	37	3	λ	9	159	1	239	402	2.5
H3v-62	37	1	κ	4	>	1,625	>	438	>5.0
H3v-84	37	1	κ	2	88	5	135	482	>5.0
H3v-11	1	1	κ	32	>	144	>	564	5.0
H3v-21	1	1	κ	11	>	352	>	697	>5.0
H3v-86	37	1	κ	27	357	13	368	956	>5.0
H3v-7	1	1	κ	34	>	>	>	1,226	>5.0
H3v-9	2	1	κ	38	>	>	>	1,301	>5.0
H3v-79	37	1	λ	86	353	>	219	1,602	>5.0

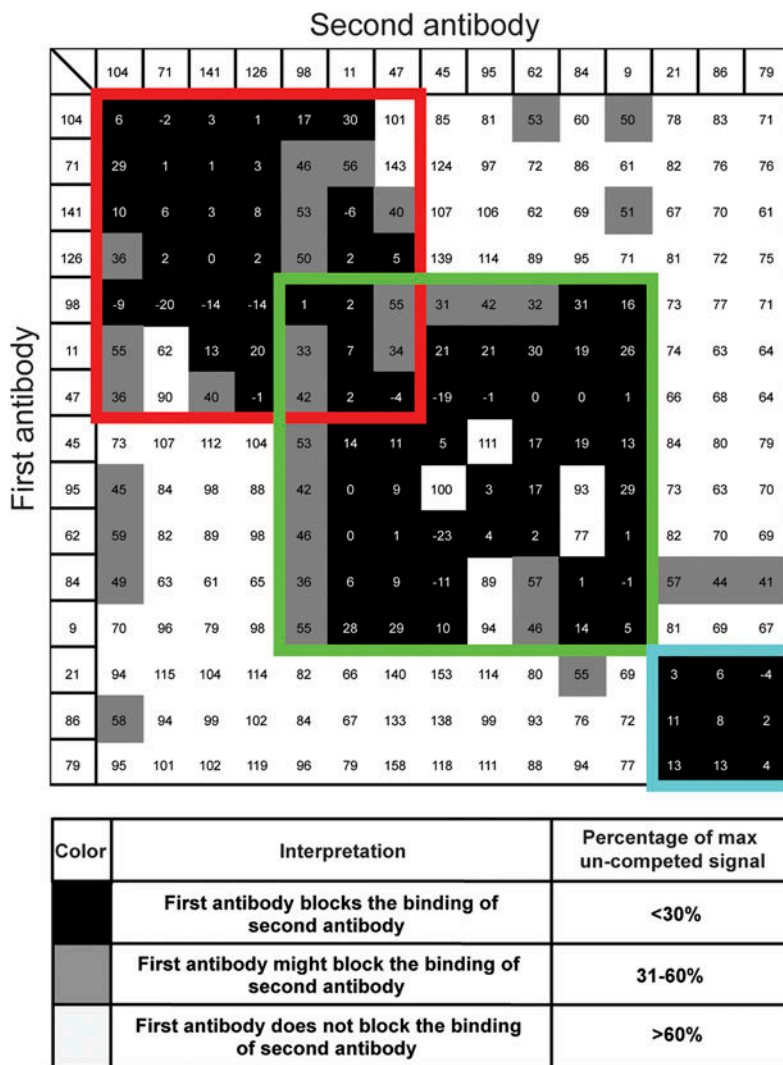
**Figure 1. Characterization of 17 neutralizing monoclonal antibodies (mAbs)**

The antibodies are arranged in the order of neutralization potency (column 9) with the most potent antibodies at the top. Seventeen mAbs isolated by human B cell hybridoma generation exhibited neutralization potential (shown as half-maximal inhibitory concentration [IC<sub>50</sub>]) at <5 µg/ml against the H3N2v virus by microneutralization assay. Nine antibodies exhibited hemagglutinin inhibition (HAI) activity, indicating that they disrupt receptor-binding function of the virus. The mAbs were tested for binding against HA from H3N2v or 3 seasonal strains (shown as half-maximal effective concentration [EC<sub>50</sub>]). The > symbol indicates that binding was not detected at the maximum concentration tested (2 µg/ml). The experiments for determining EC<sub>50</sub> (*n* = 4), IC<sub>50</sub> (*n* = 3), and HAI (*n* = 3) were conducted twice independently.





**Figure 2. Prophylactic efficacy of H3v-104, H3v-126, and H3v-71 in mice**  
Groups of mice ( $n = 5$ ) were treated with 100  $\mu\text{g}$  of individual mAbs 24 hours before challenge with mouse-adapted A/Minnesota/11/2010 X203 virus. Controls ( $n = 10$ ) were injected with PBS. The weight loss of mice was measured daily for 14 days after inoculation (day 0).

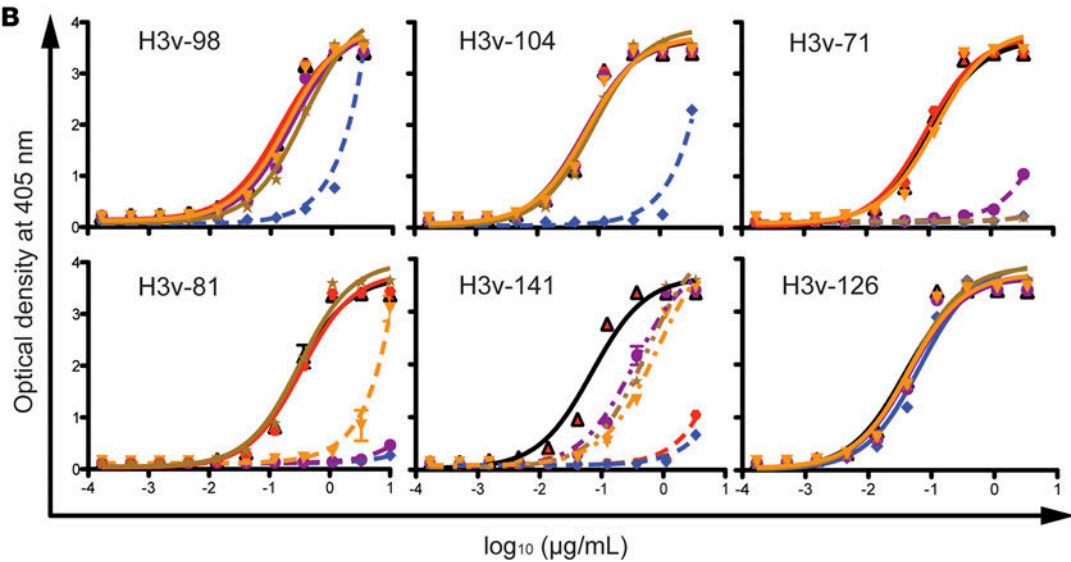


**Figure 3. Competition binding of neutralizing antibodies to H3N2v A/Minnesota/11/2010 hemagglutinin (HA) protein**

Biolayer interferometry was used to perform competition-binding assays. The HA was loaded onto Ni-NTA tips, and binding to 2 successive antibodies was tested. The binding signal for each antibody was obtained from a single association step of the mAb onto HA. If binding of the first antibody blocked the binding of the second antibody by reducing its binding signal by more than 70%, it was defined as a competitor, indicated in black. The values in the table indicate the percentage of the maximum un-competed binding signal. The red box indicates group 1, the binding group comprising the potently neutralizing mAbs. The green box and the blue box represent group 2 (partially overlaps with group 1) and group 3, respectively. The experiment was conducted twice independently.

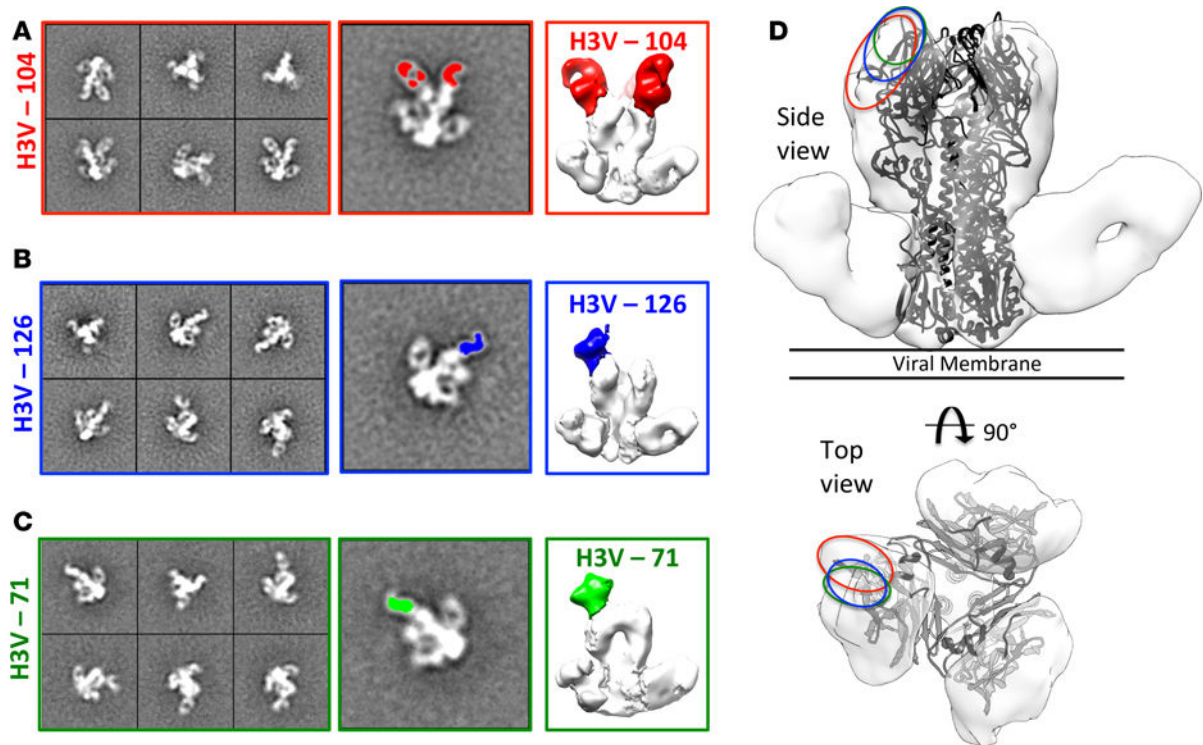
**A**

Graph Legend	H3N2v MN mutants	EC <sub>50</sub> (μg/mL)					
		H3v-98	H3v-104	H3v-71	H3v-81	H3v-141	H3v-126
▲	MN wild-type	0.13	0.05	0.09	0.29	0.07	0.04
●	MN Y137S	0.13	0.06	0.09	0.34	>	0.04
●	MN Y155T/N156K/L157S	0.19	0.06	>	>	0.30	0.04
★	MN N158G/Y159S/K160T	0.30	0.08	>	0.32	0.45	0.04
★	MN K189N/T192I/N193R	0.18	0.06	0.12	>	0.61	0.04
◆	MN L194A/Y195A/V196A	2.89	9.00	>	>	>	0.06

**B**

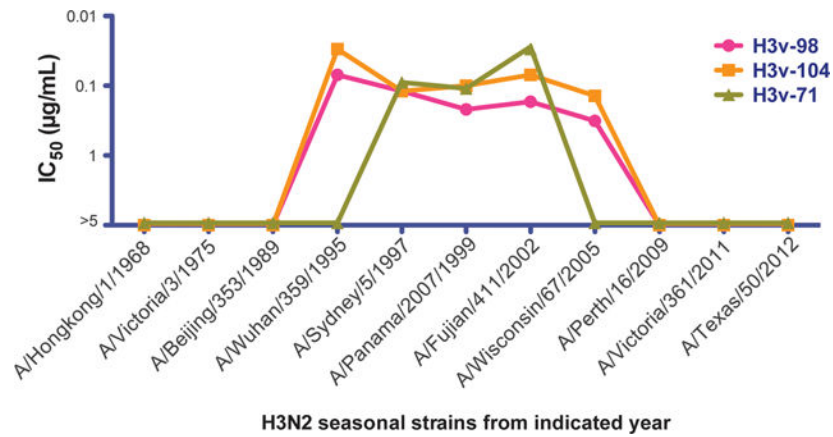
**Figure 4. Binding of H3 variant-specific antibodies to mutated Minnesota hemagglutinin (HA) proteins**

Mutagenesis of MNv HA was performed to determine antigenic residues important for recognition by variant-specific mAbs. Mutants of H3N2v A/Minnesota/11/2010 (MN) HA were generated by site-directed mutagenesis, and the half-maximal effective concentration (EC<sub>50</sub>) values were determined by performing ELISA with serial dilutions of each antibody against the mutant HAs. The table (A) shows EC<sub>50</sub> values and the graph (B) shows binding curves. The mutants that disrupted binding completely or decreased the EC<sub>50</sub> by greater than 5-fold are represented as dashed or dotted lines, respectively, and are indicated by red EC<sub>50</sub> values in the table. The > symbol indicates that binding was not detected at the maximum concentration tested (10 μg/ml). The experiments for determining the EC<sub>50</sub> ( $n = 4$ ) values were performed twice independently.



**Figure 5. Negative stain EM images of hemagglutinin-Fab complexes**

In each case the stem-binding antibody CR9114 was added to the complex in order to improve 3D reconstructions. **(A)** Reference-free 2D class averages of complex containing Fab 104 (left), single class average with Fab colored in red (middle), and 3D reconstruction (right). **(B)** Reference-free 2D class averages of complex containing Fab 126 (left), single class average with Fab colored in blue (middle), and 3D reconstruction (right). **(C)** Reference-free 2D class averages of complex containing Fab 71 (left), single class average with Fab colored in green (middle), and 3D reconstruction (right). **(D)** Side and top views of HA-Fab 126-CR9114 with Fab 126 removed and crystal structure of H3V (4FNK) fitted. Binding sites of the 3 antibodies described in A–C is highlighted using colors corresponding to Fabs.



**Figure 6. Neutralization of H3N2 seasonal strains**

The monoclonal antibodies were tested for neutralization activity by microneutralization assay against 11 seasonal strains isolated between 1968 and 2012. The half-maximal inhibitory concentration (IC<sub>50</sub>) values for H3v-98 (pink), H3v-104 (orange), or H3v-71 (green) are indicated on the y axis, and the H3N2 strains used for neutralization are indicated on the x axis. The IC<sub>50</sub> values are represented as baseline if neutralization was not detected at any concentration less than 5 µg/ml. The microneutralization assay for determining the IC<sub>50</sub> ( $n = 3$ ) values was performed twice independently.

**Table 1**

Antibody variable genes encoding 17 H3v-reactive human mAbs

mAb	Heavy chain		HCDR3 length	HCDR3 amino acid sequence
	V <sub>H</sub>	D J <sub>H</sub>		
H3v-126	2-70D*04	6-6*01 4*02	16	ARTDGGSISSAAYFES
H3v-71	1-69*01	3-10*02 5*02	16	AREGLGSVIIGPWFDP
H3v-47	1-69*05	2-2*01 3*02	21	ARGASKVEPAAPAYSDAFDM
H3v-104	1-69*13	5-12*01 4*02	16	ARDYYRGEFSGYDFES
H3v-98	3-23*01	1-1*01 4*02	13	AKSSFTRKGSPEFY
H3v-81	3-49*04	1-7*01 6*02	19	SREAAANWNPYHYSNMGMDV
H3v-141	3-15*01	3-10*01 4*02	15	TTDNSFYYSYGGYFDH
H3v-45	3-9*01	1-26*01 5*02	17	AKDGASGGTYEAGFDP
H3v-95	3-21*01	5-18*01 3*02	15	ARDLSVYSYGGAFDI
H3v-62	3-30*04	2-15*01 5*02	14	ARRFCTGGSCYLDP
H3v-84	3-48*02	3-16*01 4*02	14	ARDGAVVFGVPPFT
H3v-11	1-18*01	2-21*01 4*02	17	ARRSRRAWGLSKQGPLDY
H3v-21	1-2*02	5-18*01 4*02	14	ARGYNLGLVLLFDY
H3v-86	6-1*01	3-22*01 4*02	15	ARGIQHWMMVVVAFDH
H3v-7	3-33*04	2-15*01 4*02	15	AKDRDGGVARAPLDY
H3v-9	4-3401	3-3*01 4*02	20	ARGRPSDESWSGYLDNNGFDF
H3v-79	1-69*02	6-19*01 6*02	22	AVRAFSTAVAGKGPWHYYGMDV