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Citrus Fruit Intake Is Associated with Lower Serum Bilirubin Concentration among Women with the *UGT1A1*28* **Polymorphism1,2,3**

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Abstract

UDP-glucuronosyltransferase (UGT) 1A1 glucuronidates bilirubin, estrogens, and xenobiotic compounds. The *UGT1A1*28* polymorphism results in lower promoter activity due to 7 thymineadenine (TA) repeats, rather than the more common 6 TA repeats. Previously, we showed that serum bilirubin, a marker of UGT1A1 activity, was lower among individuals homozygous for the *UGT1A1*28* polymorphism (7/7) when randomized to a high fruit and vegetable (F&V) diet, whereas no effect was seen in individuals with the wild-type (6/6) and heterozygous (6/7) genotypes. Our objective here was to determine if we could detect genotype-diet interactions on bilirubin concentrations in an observational study. Healthy non-smoking men (*n*=146) and women (*n*=147), recruited from the Seattle area, provided blood samples for genotyping and bilirubin measurements. We used multiple linear regression to assess the relationships between *UGT1A1* genotype, bilirubin concentrations and consumption of specific F&V [cruciferous vegetables, citrus fruits, and soy foods (*n*=268)] based on FFQ, and F&V from 6 botanical families [*Cruciferae*, *Rosaceae*, *Rutaceae*, *Umbelliferae*, *Solanaceae* and *Leguminosae* (*n*=261)] based on 3 d food records. We observed a significant interaction of *UGT1A1* genotype and citrus consumption among women. Women with the 7/7 genotype who consumed 0.5 or more daily servings of citrus fruit or foods from the *Rutaceae* botanical family had ~30% lower serum bilirubin than those with the same genotype who consumed less, while 6/6 and 6/7 genotypes did not differ by consumption (*P* for interaction = 0.006 and 0.03 respectively). These results suggest that citrus consumption may increase UGT1A1 activity among women with the 7/7 genotype.

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Introduction

 UDP -glucuronosyltransferases 6 (UGT) conjugate endogenous and exogenous compounds with 5'-diphosphoglucuronic acid to form glucuronidated compounds that are more watersoluble and easily excreted. Two families of human UGT—UGT1A and UGT2B glucuronidate a wide range of substrates. UGT1A enzymes conjugate estrogens, bilirubin, and xenobiotic compounds, and UGT2B enzymes glucuronidate bile acids, androgens, and drugs; however, there is overlapping substrate specificity among members of both families (1–5). Here we focus on UGT1A1 which metabolizes dietary carcinogens, such as 2-amino-1 methyl-6-phenylimidazo[4,5-b]pyridine (PhIP), 17β-estradiol, and is the primary UGT that glucuronidates bilirubin (5–7).

Genetic polymorphisms that alter enzyme expression and/or activity and affect carcinogen clearance have been identified in *UGT* (8–10). The *UGT1A1*28* polymorphism, characterized by 7 thymine-adenine (TA) repeats rather than the more common 6 TA repeats, results in lower promoter activity and is the genetic basis for mild unconjugated hyperbilirubinemia associated with reduced hepatic bilirubin glucuronidation (Gilbert syndrome) (11,12). Individuals homozygous for the *UGT1A1*28* variant alleles (7/7) have been shown to have higher circulating concentrations of serum bilirubin (13,14), as well as reduced amounts of conjugated metabolites of the carcinogens benzo(a)pyrene [benzo(a)pyrene-7,8-dihydrodiol(−); (13)] and PhIP [N-hydroxy-PhIP (1)]. These data suggest that individuals with the *UGT1A1*28* alleles may be at increased risk of cancer due to higher or more prolonged carcinogen exposure. Results from population-based case-control studies have shown that pre-menopausal women with the *UGT1A1**28 polymorphism have a higher risk of breast cancer than women with the homozygous wild-type alleles (6/6) (15,16), perhaps due to higher circulating estradiol concentrations associated with reduced UGT1A1 activity (17).

Many phytochemicals induce UGT (18–20). We showed previously, in observational (21) and controlled feeding studies (22), that serum bilirubin concentrations were lower among individuals with the 7/7 genotype when exposed to fruits and vegetables (F&V); whereas, no effect was seen in individuals with the wild-type $(6/6)$ and heterozygous $(6/7)$ genotypes. In the present cross-sectional study, we also use serum bilirubin as an endogenous marker of UGT1A1 activity. Citrus fruit, cruciferous vegetables and soy (and/or phytochemicals contained in these foods) induce UGT activity (23–29). Our objective was to determine whether habitual consumption [as assessed by FFQ] of these foods and recent consumption [as measured by 3 d food records (3DFR)] of foods from several botanical families that contain phytochemicals that induce UGT (19) (*Cruciferae*, *Rosaceae*, *Solanaceae*, *Leguminosae*, *Rutaceae* and *Umbelliferae*), are associated with serum bilirubin concentrations. We also tested whether associations between diet and bilirubin differ by *UGT1A1* genotype.

Subjects and Methods

Subjects

293 healthy, non-smoking men (*n*=146) and women (*n*=147), aged 20–40 y, were recruited from the Seattle area via advertisements in university newspapers, flyers displayed in campus buildings, and targeted mailings of individuals identified from the Washington State Department of Licensing. Participants completed an eligibility questionnaire and were excluded based on the following criteria: 1) medical history of gastrointestinal, hepatic, or renal disorders; 2) current or planned pregnancy or lactation; 3) major dietary and/or weight change

⁶Abbreviations used: AhR, aryl hydrocarbon receptor; ER, estrogen receptor; FHCRC, Fred Hutchinson Cancer Research Center; F&V, fruit and vegetable; PhIP, 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine; TA, thymine-adenine; UGT, UDPglucuronosyltransferase; UM-NCC, University of Minnesota Nutrition Coordinating Center; 3DFR, 3 d food record

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 $(>4.5 \text{ kg})$ in the past year; 4) antibiotic use within the past 3 mo; 5) BMI >30 or <18 ; 6) current use of over-the-counter, recreational, and prescription drugs (including oral contraceptives); 7) regular exposure (including occupational) to passive smoke or organic solvents; 8) alcohol intake >2 drinks/d (720 mL beer, 240 mL wine or 90 mL hard liquor); 9) no interest in participating in the subsequent controlled feeding trial; 10) exercise regimens that require or result in significant short-term dietary changes. Participants were asked to discontinue use of all multivitamins and dietary supplements 1 wk prior to their participation in the study. The study was approved by the Institutional Review Board at the Fred Hutchinson Cancer Research Center (FHCRC) and informed written consent was obtained from all participants.

Data and sample collection

Participants completed a FFQ reporting on their dietary intake within the past 3 mo, food records on 3 consecutive d and a health and demographics questionnaire. Body weight and height were measured and a 12 h fasting blood sample was drawn from all participants to provide peripheral leukocyte DNA for *UGT* genotyping and serum for bilirubin measurements.

FFQ

Dietary data were collected from 274 (94%) participants using a FFQ developed by the FHCRC Nutrition Assessment Shared Resource. Completed FFQ were analyzed for daily servings of specific foods and daily consumption of nutrients using the database from the University of Minnesota Nutrition Coordinating Center (UM-NCC), Minneapolis, MN (19) and F&V (30). Servings of all F&V were adjusted for serving size and usual frequency of consumption. Daily servings (113 g) of cruciferous vegetables were calculated by summation of adjusted daily servings (as described above) of broccoli, cabbage, cauliflower, Brussels sprouts and coleslaw. Daily servings of citrus fruits were calculated by summation of adjusted daily servings of orange (1 orange), grapefruit (1/2 grapefruit), orange juice (177 mL), and grapefruit juice (177 mL). Daily servings of soy products were calculated by summation of daily servings (adjusted for serving size) of tofu (86 g), miso soup (237 mL), and soy milk (237 mL).

Food records

3DFR were collected from 289 (99%) participants and analyzed as previously described (31). Participants were trained by a registered dietitian on how to keep food records. The 3DFR were analyzed by trained nutritionists using the UM-NCC Nutrition Data System for Research that incorporates a comprehensive food product list and nutrient database (32) to estimate daily intake of total F&V (together and separately), as well as those defined by botanical family, based on standard serving sizes (228 g raw, 113 g cooked or canned, 118 mL juice, etc.). We focused on daily servings of plants from the *Cruciferae*, *Rosaceae*, *Solanaceae*, *Leguminosae*, *Rutaceae* and *Umbelliferae* families because they include foods which contain phytochemicals previously shown to induce UGT (20). Cruciferous vegetables (i.e., *Cruciferae*), such as broccoli and cabbage, are rich in sulfur-containing glucosinolates. Foods in the *Umbelliferae* (e.g., carrots, parsley), *Solanaceae* (e.g., tomato, eggplant) and *Rutaceae* (e.g., citrus fruits) families contain carotenoids and many flavonoids. Fruit, such as apples and berries, in the *Rosaceae* family are also abundant in flavonoids, and legumes (i.e., *Leguminosae*), such as lentils and beans, are rich in lignans.

Determination of UGT1A1 promoter genotypes and serum bilirubin measurements

Genotyping for the *UGT1A1*28* polymorphism was performed as previously described (22). Serum total and direct (conjugated) bilirubin concentrations were quantified using a Cobas MIRA Plus centrifugal analyzer (Roche Diagnostic Systems). Indirect (unconjugated) bilirubin was calculated as the difference between total and direct bilirubin.

Statistical analysis

Four individuals (1%) did not provide any dietary information and were excluded from analyses. Six (2%) additional participants were excluded because they had *UGT1A1* genotypes other than 6/6, 6/7, or 7/7. Of the remaining 283 study participants, 261 (92%) and 268 (95%) completed 3DFR on 3 consecutive d within 1 wk prior to serum collection and FFQ, respectively. Serum bilirubin concentrations were log-transformed prior to analysis to normalize distributions. Daily servings of total F&V and total botanicals were divided into three groups (<4, 4–5, and >5) and used as grouped linear variables in statistical models. Daily servings of all other dietary variables were split dichotomously $\left(\frac{1}{2}$ or $\frac{1}{2}+\right)$ for analysis. Multiple linear regression analysis was used to determine whether demographic characteristics [age $($30 \text{ or } 30+$ y), sex, race (Caucasian, Asian or other), and duration of physical activity$ (≤6 or >6 h/wk)] and *UGT1A1* genotype (used as a grouped linear variable to test for significant linear trend) were associated with serum bilirubin concentrations. Differences in daily intake of each food group by *UGT1A1* genotype (used as a categorical variable—6/6, 6/7 or 7/7) were assessed using linear regression.

Multiple linear regression analysis was also used to assess the association of $F\&V$ consumption (independent variables; total F&V or botanicals and each F&V variable tested individually) with each serum bilirubin measure. These analyses were adjusted *a priori* for *UGT1A1* genotype, sex, and total F&V or energy (for total F&V and total botanicals) intake. The following covariates were assessed as potential confounders or predictors in all regression models: age (continuous), body wt (continuous), duration of physical activity (≤ 6 or > 6 h/wk), race/ethnicity (Caucasian, Asian or other), total energy intake (continuous), and season (winter, spring, summer, or fall; for 3DFR variables only). Predictors were included in models if the *P*-value for the test that the regression coefficient(s) for the variable term(s) was equal to zero was significant (*P*<0.05). Confounders were included if their addition to the regression model changed the main effect(s) estimate(s) by 10% or greater. To determine whether dietary effects on bilirubin concentrations differed by genotype, interactions of each dietary variable (coded as grouped linear) with *UGT1A1* genotype (coded as grouped linear) was assessed by testing the null hypothesis that the regression coefficient(s) for the interaction term(s) was equal to zero. Statistical tests were two-sided and *P*<0.05 was considered statistically significant. All statistical analyses were performed using Stata 9.0 (StataCorp).

Results

Sex, race, and *UGT1A1* genotype were associated with total, direct, and indirect serum bilirubin (Table 1). Men had statistically significantly higher serum bilirubin concentrations than women $(P \le 0.001)$. These measures were also statistically significantly higher among Asians than Caucasians $(P < 0.01)$; whereas, total and indirect bilirubin concentrations were significantly lower among races other than Caucasian $(P < 0.05)$. There was a statistically significant linear trend of increasing total, direct, and indirect bilirubin (*P* < 0.001) with higher numbers of *UGT1A1*28* alleles. Age was not associated with total and indirect bilirubin; however, individuals \geq 30 y had slightly lower concentrations of direct (conjugated) bilirubin than those who were <30 y ($P = 0.02$), after adjusting for all other covariates. Duration of physical activity was not associated with any bilirubin measures. There were no statistically significant differences in mean daily intake of each F&V group or botanical family by *UGT1A1* genotype (Supplemental Table 1).

Total F&V consumption was not statistically significantly associated with any measure of serum bilirubin (Supplemental Table 2). However, among the individual F&V groups, there was a statistically significant interaction between *UGT1A1* genotype and citrus fruit ($P = 0.006$; **Table 4**) and *Rutaceae* (*P* = 0.03; **Table 5**) consumption associated with total and indirect (data not shown) bilirubin among women but not men: women with the 7/7 genotype who

consumed 0.5 or more daily servings of citrus fruit or *Rutaceae* had lower (~30%) total and indirect bilirubin concentrations than those who consumed less. In contrast, bilirubin measures did not differ by consumption of these food groups among men or women with the 6/6 and 6/7 genotypes. Multiple regression models that included citrus consumption, *UGT1A1* genotype, race, age, total F&V consumption, and the citrus -*UGT1A1* genotype interaction terms accounted for \sim 30% (\mathbb{R}^2 values) of the variation in total and indirect bilirubin, respectively.

Discussion

We observed a statistically significant interaction between *UGT1A1* genotype and consumption of citrus, which was associated with lower serum bilirubin among women with the 7/7 genotype. These results are consistent with previous studies that showed bilirubin concentrations were lower among individuals with the 7/7 genotype who were exposed to cruciferous vegetables (21) or a combination of soy, citrus fruit, and cruciferous vegetables (22).

Phytochemicals found in citrus fruits increase UGT activity (28,33–35). Small changes in UGT activity may result from direct phytochemical binding to the enzyme's active site; however, the primary mechanism is hypothesized to be through induction of *UGT* gene expression (19). Many phytochemicals can regulate multiple transcription factors. Quercetin, a flavonoid found in citrus fruit, activates the nuclear factor-erythroid 2-related factor 2 transcription factor, that binds to the antioxidant response element (36,37), and induces *UGT1A1* expression (38, 39). Therefore, consumption of citrus fruit associated with greater bilirubin glucuronidation may be a result of induced *UGT1A1* expression/activity.

Previously, in a study in which participants were recruited on the basis of high and low F&V consumption, we observed that *Cruciferae* intake was associated with lower bilirubin concentrations among individuals with the 7/7 genotype (21); however, we did not observe this relationship in the current study. Although the range of *Cruciferae* consumption among participants with the 7/7 genotype in the current study is slightly wider than that among those in the previous study [(0–1.0) versus (0.2–0.7), respectively], mean intake of *Cruciferae* was ~60% lower. This level of consumption in our current study was possibly too low to reliably detect differences in total serum bilirubin concentrations among participants with the 7/7 genotype. In addition, among individuals with the 7/7 genotype, mean total bilirubin concentration in the current study was ~30% lower than in the previous study, which may have also affected our ability to detect differences.

As previously reported, women had lower serum bilirubin than men, independent of *UGT1A1* genotype (21,22,40,41), presumably due to estrogen- and/or progesterone-mediated *UGT1A1* transcription (42–44). Results from several studies suggest that estrogen receptor (ER)-mediated increases in transcriptional activity of the arylhydrocarbon receptor (AhR) affect *UGT1A1* (45–47). Phytochemicals in citrus foods activate the AhR (48–51) to increase *UGT1A1* transcription (39). Thus, lower bilirubin concentrations among women may result from increased *UGT1A1* expression associated with ER/AhR cross-talk.

The mechanism for the effects of F&V consumption only among women with the 7/7 genotype is unclear. Extra TA repeats associated with the *UGT1A1*28* polymorphism result in low promoter activity due to reduced binding of TATA-binding protein (11,52) and other transcription factors, including the AhR (53). Recently, it was proposed that the AhR can serve as a positive transcriptional co-regulator of the ER (54). Perhaps, increased AhR/ER cross-talk among women who consume F&V can somewhat overcome the transcriptional defect associated with the *UGT1A1*28* polymorphism.

The collection of detailed dietary and demographic and behavioral data from study participants is the primary strength of this study. Changes in UGT expression/activity can be detected within an h of phytochemical exposure (55,56); thus, recent consumption from 3DFR may be most relevant for detecting associations between F&V intake and bilirubin concentrations. However, because study participants tend to overestimate F&V consumption when reporting their usual intake on FFQ (30), the FFQ may further separate the low- and high-consumers and this may have increased our power to detect differences in bilirubin concentrations by citrus fruit consumption compared to those associated with *Rutaceae* consumption. This may have contributed to the stronger association with citrus fruit consumption.

Glucuronidated compounds are rapidly excreted. It is possible that changes in bilirubin concentrations, used here as a surrogate for UGT activity, are diluted compared to those occurring in relevant tissues, such as the liver and intestine, which are in direct contact with phytochemicals released during F&V digestion. Perhaps the most significant limitation of this study is the small sample size among strata of women and men with the rare 7/7 genotype, which probably limited our ability to distinguish differences in bilirubin glucuronidation. Our decision to categorize continuous F&V intake may have also decreased our power to detect associations between bilirubin concentrations and F&V consumption; however, similar associations were observed using continuous citrus fruit intake. Another potential issue is the large number of comparisons performed during the statistical analysis, which increases the likelihood of observing a significant association due to chance. A total of 33 statistical tests were performed. With an α -significance level of 0.05, \sim 2 associations would have been statistically significant by chance alone, and, thus, these results should be interpreted with caution. However, our result for citrus fruit was at the $P = 0.006$ level, and that level of significance among any one of 33 comparisons has a likelihood of occurrence by chance of less than 20%.

UGT1A1 glucuronidates several compounds, such as estradiol and PhIP, that have been associated with cancer; thus, changes in UGT1A1 expression/activity could affect cancer risk. Several studies have suggested that individuals with the *UGT1A1*28* polymorphism are at increased risk of certain cancers (15,16,57,58) and toxicity from certain drugs and have a reduced ability to clear potential carcinogens (1,8,13,59,60). The results of this study suggest that women with the 7/7 genotype who consume F&V, particularly citrus fruit, may have higher UGT1A1 activity than women with the 7/7 genotype who do not consume F&V. Our results suggest that certain components in F&V may increase UGT1A1 activity among individuals with this variant genotype and thus potentially improve clearance of certain carcinogens and influence cancer susceptibility.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Literature Cited

- 1. Girard H, Thibaudeau J, Court MH, Fortier LC, Villeneuve L, Caron P, Hao Q, von Moltke LL, Greenblatt DJ, Guillemette C. UGT1A1 polymorphisms are important determinants of dietary carcinogen detoxification in the liver. Hepatology 2005;42:448–57. [PubMed: 15986396]
- 2. Guillemette C, Belanger A, Lepine J. Metabolic inactivation of estrogens in breast tissue by UDPglucuronosyltransferase enzymes: an overview. Breast Cancer Res 2004;6:246–54. [PubMed: 15535854]
- 3. Malfatti MA, Felton JS. Human UDP-glucuronosyltransferase 1A1 is the primary enzyme responsible for the N-glucuronidation of N-hydroxy-PhIP in vitro. Chem Res Toxicol 2004;17:1137–44. [PubMed: 15310245]

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- 4. Senafi SB, Clarke DJ, Burchell B. Investigation of the substrate specificity of a cloned expressed human bilirubin UDP-glucuronosyltransferase: UDP-sugar specificity and involvement in steroid and xenobiotic glucuronidation. Biochem J 1994;303(Pt 1):233–40. [PubMed: 7945246]
- 5. Tukey RH, Strassburg CP. Human UDP-glucuronosyltransferases: metabolism, expression, and disease. Annu Rev Pharmacol Toxicol 2000;40:581–616. [PubMed: 10836148]
- 6. Guillemette C. Pharmacogenomics of human UDP-glucuronosyltransferase enzymes. Pharmacogenomics J 2003;3:136–58. [PubMed: 12815363]
- 7. Miners JO, McKinnon RA, Mackenzie PI. Genetic polymorphisms of UDP-glucuronosyltransferases and their functional significance. Toxicology 2002;181–182:453–6. [PubMed: 12505351]
- 8. Burchell B. Genetic variation of human UDP-glucuronosyltransferase: implications in disease and drug glucuronidation. Am J Pharmacogenomics 2003;3:37–52. [PubMed: 12562215]
- 9. Lampe JW. Diet, genetic polymorphisms, detoxification, and health risks. Altern Ther Health Med 2007;13:S108–11. [PubMed: 17405687]
- 10. Nagar S, Remmel RP. Uridine diphosphoglucuronosyltransferase pharmacogenetics and cancer. Oncogene 2006;25:1659–72. [PubMed: 16550166]
- 11. Bosma PJ, Chowdhury JR, Bakker C, Gantla S, de Boer A, Oostra BA, Lindhout D, Tytgat GN, Jansen PL, et al. The genetic basis of the reduced expression of bilirubin UDPglucuronosyltransferase 1 in Gilbert's syndrome. N Engl J Med 1995;333:1171–5. [PubMed: 7565971]
- 12. Burchell B, Hume R. Molecular genetic basis of Gilbert's syndrome. J Gastroenterol Hepatol 1999;14:960–6. [PubMed: 10530490]
- 13. Fang JL, Lazarus P. Correlation between the UDP-glucuronosyltransferase (UGT1A1) TATAA box polymorphism and carcinogen detoxification phenotype: significantly decreased glucuronidating activity against benzo(a)pyrene-7,8-dihydrodiol(−) in liver microsomes from subjects with the UGT1A1*28 variant. Cancer Epidemiol Biomarkers Prev 2004;13:102–9. [PubMed: 14744740]
- 14. Lampe JW. Health effects of vegetables and fruit: assessing mechanisms of action in human experimental studies. Am J Clin Nutr 1999;70:475S–90S. [PubMed: 10479220]
- 15. Adegoke OJ, Shu XO, Gao YT, Cai Q, Breyer J, Smith J, Zheng W. Genetic polymorphisms in uridine diphospho-glucuronosyltransferase 1A1 (UGT1A1) and risk of breast cancer. Breast Cancer Res Treat 2004;85:239–45. [PubMed: 15111762]
- 16. Guillemette C, Millikan RC, Newman B, Housman DE. Genetic polymorphisms in uridine diphosphoglucuronosyltransferase 1A1 and association with breast cancer among African Americans. Cancer Res 2000;60:950–6. [PubMed: 10706110]
- 17. Sparks R, Ulrich CM, Bigler J, Tworoger SS, Yasui Y, Rajan KB, Porter P, Stanczyk FZ, Ballard-Barbash R, et al. UDP-glucuronosyltransferase and sulfotransferase polymorphisms, sex hormone concentrations, and tumor receptor status in breast cancer patients. Breast Cancer Res 2004;6:R488– 98. [PubMed: 15318931]
- 18. Moon YJWX, Morris ME. Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. Toxicology in Vitro 2006;20:187–210. [PubMed: 16289744]
- 19. Saracino MR, Lampe JW. Phytochemical regulation of UDP-glucuronosyltransferases: implications for cancer prevention. Nutr Cancer 2007;59:121–41. [PubMed: 18001207]
- 20. Surh YJ. Cancer chemoprevention with dietary phytochemicals. Nat Rev Cancer 2003;3:768–80. [PubMed: 14570043]
- 21. Peterson S, Bigler J, Horner NK, Potter JD, Lampe JW. Cruciferae interact with the UGT1A1*28 polymorphism to determine serum bilirubin levels in humans. J Nutr 2005;135:1051–5. [PubMed: 15867280]
- 22. Chang JL, Bigler J, Schwarz Y, Li SS, Li L, King IB, Potter JD, Lampe JW. UGT1A1 polymorphism is associated with serum bilirubin concentrations in a randomized, controlled, fruit and vegetable feeding trial. J Nutr 2007;137:890–7. [PubMed: 17374650]
- 23. Appelt LC, Reicks MM. Soy induces phase II enzymes but does not inhibit dimethylbenz[a] anthracene-induced carcinogenesis in female rats. J Nutr 1999;129:1820–6. [PubMed: 10498753]
- 24. Dingley KH, Ubick EA, Chiarappa-Zucca ML, Nowell S, Abel S, Ebeler SE, Mitchell AE, Burns SA, Steinberg FM, Clifford AJ. Effect of dietary constituents with chemopreventive potential on

adduct formation of a low dose of the heterocyclic amines PhIP and IQ and phase II hepatic enzymes. Nutr Cancer 2003;46:212–21. [PubMed: 14690798]

- 25. Elegbede JA, Maltzman TH, Elson CE, Gould MN. Effects of anticarcinogenic monoterpenes on phase II hepatic metabolizing enzymes. Carcinogenesis 1993;14:1221–3. [PubMed: 8508509]
- 26. Hecht SS, Carmella SG, Murphy SE. Effects of watercress consumption on urinary metabolites of nicotine in smokers. Cancer Epidemiol Biomarkers Prev 1999;8:907–13. [PubMed: 10548320]
- 27. Pantuck EJ, Pantuck CB, Anderson KE, Wattenberg LW, Conney AH, Kappas A. Effect of brussels sprouts and cabbage on drug conjugation. Clin Pharmacol Ther 1984;35:161–9. [PubMed: 6692645]
- 28. Pfeiffer E, Treiling CR, Hoehle SI, Metzler M. Isoflavones modulate the glucuronidation of estradiol in human liver microsomes. Carcinogenesis 2005;26:2172–8. [PubMed: 16051636]
- 29. van der Logt EM, Roelofs HM, Nagengast FM, Peters WH. Induction of rat hepatic and intestinal UDP-glucuronosyltransferases by naturally occurring dietary anticarcinogens. Carcinogenesis 2003;24:1651–6. [PubMed: 12869420]
- 30. Kristal AR, Vizenor NC, Patterson RE, Neuhouser ML, Shattuck AL, McLerran D. Precision and bias of food frequency-based measures of fruit and vegetable intakes. Cancer Epidemiol Biomarkers Prev 2000;9:939–44. [PubMed: 11008912]
- 31. Horner NK, Kristal AR, Prunty J, Skor HE, Potter JD, Lampe JW. Dietary determinants of plasma enterolactone. Cancer Epidemiol Biomarkers Prev 2002;11:121–6. [PubMed: 11815409]
- 32. Schakel SF, Sievert YA, Buzzard IM. Sources of data for developing and maintaining a nutrient database. J Am Diet Assoc 1988;88:1268–71. [PubMed: 3171020]
- 33. Siess MH, Le Bon AM, Suschetet M. Dietary modification of drug-metabolizing enzyme activities: dose-response effect of flavonoids. J Toxicol Environ Health 1992;35:141–52. [PubMed: 1602520]
- 34. Siess MH, Mas JP, Canivenc-Lavier MC, Suschetet M. Time course of induction of rat hepatic drugmetabolizing enzyme activities following dietary administration of flavonoids. J Toxicol Environ Health 1996;49:481–96. [PubMed: 8968409]
- 35. Sun XY, Plouzek CA, Henry JP, Wang TT, Phang JM. Increased UDP-glucuronosyltransferase activity and decreased prostate specific antigen production by biochanin A in prostate cancer cells. Cancer Res 1998;58:2379–84. [PubMed: 9622078]
- 36. Kluth D, Banning A, Paur I, Blomhoff R, Brigelius-Flohe R. Modulation of pregnane X receptorand electrophile responsive element-mediated gene expression by dietary polyphenolic compounds. Free Radic Biol Med 2007;42:315–25. [PubMed: 17210444]
- 37. Tanigawa S, Fujii M, Hou DX. Action of Nrf2 and Keap1 in ARE-mediated NQO1 expression by quercetin. Free Radic Biol Med 2007;42:1690–703. [PubMed: 17462537]
- 38. Petri N, Tannergren C, Holst B, Mellon FA, Bao Y, Plumb GW, Bacon J, O'Leary KA, Kroon PA, et al. Absorption/metabolism of sulforaphane and quercetin, and regulation of phase II enzymes, in human jejunum in vivo. Drug Metab Dispos 2003;31:805–13. [PubMed: 12756216]
- 39. Sugatani J, Yamakawa K, Tonda E, Nishitani S, Yoshinari K, Degawa M, Abe I, Noguchi H, Miwa M. The induction of human UDP-glucuronosyltransferase 1A1 mediated through a distal enhancer module by flavonoids and xenobiotics. Biochem Pharmacol 2004;67:989–1000. [PubMed: 15104253]
- 40. Lampe JW, Bigler J, Horner NK, Potter JD. UDP-glucuronosyltransferase (UGT1A1*28 and UGT1A6*2) polymorphisms in Caucasians and Asians: relationships to serum bilirubin concentrations. Pharmacogenetics 1999;9:341–9. [PubMed: 10471066]
- 41. Zucker SD, Horn PS, Sherman KE. Serum bilirubin levels in the U.S. population: gender effect and inverse correlation with colorectal cancer. Hepatology 2004;40:827–35. [PubMed: 15382174]
- 42. Anderson GD. Sex differences in drug metabolism: cytochrome P-450 and uridine diphosphate glucuronosyltransferase. J Gend Specif Med 2002;5:25–33. [PubMed: 11859684]
- 43. Jeong H, Choi S, Song JW, Chen H, Fischer JH. Regulation of UDP-glucuronosyltransferase (UGT) 1A1 by progesterone and its impact on labetalol elimination. Xenobiotica 2008;38:62–75. [PubMed: 18098064]
- 44. Stahlberg N, Merino R, Hernandez LH, Fernandez-Perez L, Sandelin A, Engstrom P, Tollet-Egnell P, Lenhard B, Flores-Morales A. Exploring hepatic hormone actions using a compilation of gene expression profiles. BMC Physiol 2005;5:8. [PubMed: 15953391]
- 45. Angus WG, Larsen MC, Jefcoate CR. Expression of CYP1A1 and CYP1B1 depends on cell-specific factors in human breast cancer cell lines: role of estrogen receptor status. Carcinogenesis 1999;20:947–55. [PubMed: 10357772]
- 46. Matthews J, Gustafsson JA. Estrogen receptor and aryl hydrocarbon receptor signaling pathways. Nucl Recept Signal 2006;4:e016. [PubMed: 16862222]
- 47. Thomsen JS, Wang X, Hines RN, Safe S. Restoration of aryl hydrocarbon (Ah) responsiveness in MDA-MB-231 human breast cancer cells by transient expression of the estrogen receptor. Carcinogenesis 1994;15:933–7. [PubMed: 8200098]
- 48. Amakura Y, Tsutsumi T, Nakamura M, Kitagawa H, Fujino J, Sasaki K, Toyoda M, Yoshida T, Maitani T. Activation of the aryl hydrocarbon receptor by some vegetable constituents determined using in vitro reporter gene assay. Biol Pharm Bull 2003;26:532–9. [PubMed: 12673038]
- 49. Ciolino HP, Daschner PJ, Yeh GC. Dietary flavonols quercetin and kaempferol are ligands of the aryl hydrocarbon receptor that affect CYP1A1 transcription differentially. Biochem J 1999;340(Pt 3): 715–22. [PubMed: 10359656]
- 50. Pohl C, Will F, Dietrich H, Schrenk D. Cytochrome P450 1A1 expression and activity in Caco-2 cells: modulation by apple juice extract and certain apple polyphenols. J Agric Food Chem 2006;54:10262–8. [PubMed: 17177569]
- 51. Zhang S, Qin C, Safe SH. Flavonoids as aryl hydrocarbon receptor agonists/antagonists: effects of structure and cell context. Environ Health Perspect 2003;111:1877–82. [PubMed: 14644660]
- 52. Hsieh TY, Shiu TY, Huang SM, Lin HH, Lee TC, Chen PJ, Chu HC, Chang WK, Jeng KS, et al. Molecular pathogenesis of Gilbert's syndrome: decreased TATA-binding protein binding affinity of UGT1A1 gene promoter. Pharmacogenet Genomics 2007;17:229–36. [PubMed: 17496722]
- 53. Sugatani J, Mizushima K, Osabe M, Yamakawa K, Kakizaki S, Takagi H, Mori M, Ikari A, Miwa M. Transcriptional regulation of human UGT1A1 gene expression through distal and proximal promoter motifs: implication of defects in the UGT1A1 gene promoter. Naunyn Schmiedebergs Arch Pharmacol 2008;377:597–605. [PubMed: 18172616]
- 54. Ohtake F, Baba A, Fujii-Kuriyama Y, Kato S. Intrinsic AhR function underlies crosstalk of dioxins with sex hormone signalings. Biochem Biophys Res Commun 2008;370:541–6. [PubMed: 18358233]
- 55. Basu NK, Ciotti M, Hwang MS, Kole L, Mitra PS, Cho JW, Owens IS. Differential and special properties of the major human UGT1-encoded gastrointestinal UDP-glucuronosyltransferases enhance potential to control chemical uptake. J Biol Chem 2004;279:1429–41. [PubMed: 14557274]
- 56. Williams JA, Ring BJ, Cantrell VE, Campanale K, Jones DR, Hall SD, Wrighton SA. Differential modulation of UDP-glucuronosyltransferase 1A1 (UGT1A1)-catalyzed estradiol-3-glucuronidation by the addition of UGT1A1 substrates and other compounds to human liver microsomes. Drug Metab Dispos 2002;30:1266–73. [PubMed: 12386134]
- 57. Girard H, Butler LM, Villeneuve L, Millikan RC, Sinha R, Sandler RS, Guillemette C. UGT1A1 and UGT1A9 functional variants, meat intake, and colon cancer, among Caucasians and African-Americans. Mutat Res 2008;644:56–63. [PubMed: 18675828]
- 58. Shatalova EG, Walther SE, Favorova OO, Rebbeck TR, Blanchard RL. Genetic polymorphisms in human SULT1A1 and UGT1A1 genes associate with breast tumor characteristics: a case-series study. Breast Cancer Res 2005;7:R909–21. [PubMed: 16280036]
- 59. Iyer L, Das S, Janisch L, Wen M, Ramirez J, Karrison T, Fleming GF, Vokes EE, Schilsky RL, Ratain MJ. UGT1A1*28 polymorphism as a determinant of irinotecan disposition and toxicity. Pharmacogenomics J 2002;2:43–7. [PubMed: 11990381]
- 60. Peters U, Sinha R, Bell DA, Rothman N, Grant DJ, Watson MA, Kulldorff M, Brooks LR, Warren SH, DeMarini DM. Urinary mutagenesis and fried red meat intake: influence of cooking temperature, phenotype, and genotype of metabolizing enzymes in a controlled feeding study. Environ Mol Mutagen 2004;43:53–74. [PubMed: 14743346]

Table 1

Concentrations of serum total, direct, and indirect bilirubin among study participants grouped by demographic characteristics and *UGT1A1* genotype

¹ Bilirubin data were transformed ln(x+1). Back-transformed means and 95% CI (in parentheses) are presented.

2 Significant associations with all bilirubin measures (*P* ≤ 0.01) after adjusting for all other variables in table using multiple linear regression

3 Significant association with direct bilirubin concentration (*P* = 0.02) after adjusting for all other variables in table using multiple linear regression

4 Significant linear associations with all bilirubin measures (*P* < 0.001) after adjusting for all other variables in table using multiple linear regression with grouped linear *UGT1A1* genotype

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Total serum bilirubin among women and men stratified by *UGT1A1* genotype and daily servings of foods assessed by FFQ: interaction Total serum bilirubin among women and men stratified by *UGT1A1* genotype and daily servings of foods assessed by FFQ: interaction of genotype and diet *1*

Women Men

Women

Men

2

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*2*Adjusted for race, age, and total energy (total fruits & vegetables) or total fruits & vegetables (soy products, citrus fruit, and cruciferous vegetables) using multiple linear regression

 2 Adjusted for race, age, and total energy (total fruits & vegetables) or total fruits & vegetables (soy products, citrus fruit, and cruciferous vegetables) using multiple linear regression

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Total serum bilirubin among women and men stratified by *UGT1A1* genotype and daily servings of foods assessed by 3DFR: interaction Total serum bilirubin among women and men stratified by *UGT1A1* genotype and daily servings of foods assessed by 3DFR: interaction

 NIH-PA Author ManuscriptNIH-PA Author Manuscript **Women Men**

Women

Men

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*2*Adjusted for race, age, and total energy (total botanicals) or total botanicals (models for all other botanical families) using multiple linear regression

 2 Adjusted for race, age, and total energy (total botanicals) or total botanicals (models for all other botanical families) using multiple linear regression

